Supporting Information: Quantum chemical study of a radical relay mechanism for the HydG-catalyzed synthesis of a $Fe(II)(CO)_2(CN)$ cysteine precursor to the H-cluster of [FeFe] hydrogenase

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1 Computational methods

Structural modeling. The initial HydG structure for the simulations was obtained from the published crystal structure, PDB ID $4WCX$.¹ Herein, only Chain A of the protein dimer was employed in the study as it included the dangler iron. When preparing the simulation input structure, five missing residues in the N-terminal region were omitted and seven missing residues (345-351) were manually added in the middle of the chain. Because the SAM cofactor was missing from Chain A, we modeled its structure by substituting the SAM structure from Chain C for the corresponding methionine residue in chain A by superposition. The crystal structure included a free alanine (ALA) and sulfur atom (H2S) bonded to the auxiliary cluster, which we interpreted to be identical to the cysteine ligand to the dangler Fe from experimental evidence.² In order to add the missing tyrosine substrate, the crystal

structure of the tryptophan lyase NosL (pdb ID: $4R34$)³ was used as a template to place the tyrosine substrate into HydG by superposition with the tryptophan substrate in NosL; the interactions between tyrosine and HydG were then optimized using a docking calculation.⁴ The protonation of all residues was decided based on comparing the experimental pH value (7.0) with standard side chain pKa values, and His265 was protonated as HID ($N\delta$ is protonated) as N_f is coordinated to the dangler iron. Given the tyrosine substrate, two protonation states were considered in this study, shown in Scheme S1.

Details of the MM molecular dynamics simulations. Before starting any QM/MM simulations, the structure was relaxed by running classical molecular dynamics (MD) using molecular mechanics (MM) force fields in the AMBER software package. ⁵ The AMBER-FB15 protein force field⁶ and TIP3P-FB water model⁷ were used for the protein and water molecules in the system, the GAFF small molecule force field⁸ was used to model the SAM cofactor, and the Fe-S clusters used a force field model published previously for MD simulations of [FeFe] hydrogenases. ⁹ These force fields are mutually compatible based on past studies that used the Fe-S cluster force field together with AMBER-family force fields, 10,11 and the high accuracy of AMBER-FB15 and TIP3P-FB when used together. ⁶

The MM MD simulations used a simulation time step of 1 fs. Harmonic energy restraints were added to selected interatomic distances in order to ensure the force field does not change the coordination environment around the transition metal centers. The force constant was set to 50 kcal/mol/ \AA^2 and the restrained distances include the distances between the dangler iron and the atoms coordinated to it (two oxygen atoms from two water molecules, the O, N, and S atoms from the cysteine ligand, and the N ϵ in His265). The cutoff values for shortrange electrostatics and van der Waals interactions were set to 12 Å , and the particle-mesh Ewald method was used for long range summation of electrostatic interactions. ¹² Covalent bond lengths involving hydrogen were constrained using the SHAKE algorithm. ¹³ A Langevin thermostat algorithm with a collision frequency of 1.0 ps^{-1} was employed for temperature control. In the simulation procedure, energy minimization was carried out first, followed by gradual heating from 0 K to 300 K using a 200 ps MD simulation at constant volume (NVT), and this was followed by a 200 ps equilibration simulation at constant temperature and pressure (NPT) where a Berendsen barostat was added. Next, a 50 ns MD simulation was carried out under NVT conditions, and the final structure was used as the starting point of hybrid QM/MM simulations.

Hybrid QM/MM simulations at the canonical cluster. The QM/MM simulations were carried out using the Q-Chem and AMBER software packages.^{5,14,15} Due to the complexity of the HydG catalytic reaction, different QM regions were chosen based on which reaction step was studied. Figure S1, left panel shows the selection of QM regions for two reactions occurring at the rSAM Fe-S cluster; in these reaction steps, the broken symmetry approximation was used to model the high spin states and antiferromagnetic coupling of the Fe atoms. Standard electrostatic embedding was applied for the electrostatic interactions between the QM and MM regions, and a pseudo-bond and pseudo-atom approach¹⁶ was used to treat the covalent bonds between the QM and MM regions.

The QM region was treated using density functional theory (DFT) using the unrestricted B3LYP density functional approximation, which we deemed appropriate for the canonical cluster as the reaction steps here involved mostly organic species. A hybrid basis set was used comprising the LANL2DZ basis set and pseudopotential for Fe atoms and the $6-31G^*$ basis set for all other atoms. The choice of the relatively small basis was necessary in order to enable the QM/MM umbrella sampling described later, which involved running >10,000 serial individual calculations. To validate the accuracy of using this basis, we carried out potential energy scans using the larger def-TZVP basis; a comparison of energy profiles shows that the choice of basis set affects the barrier height by 0-2 kcal/mol (Figure S2).

The QM/MM free energy profiles were generated using an umbrella sampling approach where 15 ps of MD simulations were carried out at multiple windows along the reaction coordinate. The cutoff values for the non-bonded interactions were set to 12 \AA , and the time step was set to 1 fs. In order to compute QM/MM free energy profiles, an umbrella sampling approach was adopted. Multiple independent simulations were carried out corresponding to values of the chosen reaction coordinate; in this study most of the spacing between simulations was set to 0.2 Å. Individual umbrella sampling runs were modified in order to maximize the thermodynamic overlap between windows while keeping computational cost affordable; 20 windows were used for tyrosine radical generation, 45 windows for the 2-D umbrella sampling of tyrosine decomposition, 20 windows for DHG radical formation, and 12 windows for DHG decomposition.

The initial structures of each umbrella sampling window were determined using a series of constrained energy minimizations. A harmonic potential was added to each simulation to ensure the simulation trajectory remains close to the reaction coordinate. The umbrella sampling QM/MM MD simulations were carried out with an added harmonic potential to ensure the simulation trajectory remains close to the reaction coordinate. The force constants of the harmonic potentials were chosen according to the slope of the energy profile from the constrained minimizations and ranged from 5 to 80 kcal/mol/ \AA ². QM/MM MD simulations were carried out for 15 ps for each window, and then the weighted histogram analysis method (WHAM) procedure^{17,18} was used for data in the last 10 ps to determine the free energy profile from the biased trajectories. This procedure produces relative free energies with a statistical error on the order of 1 kcal/mol.

Cluster model calculations at the auxiliary cluster. The reactions at the auxiliary cluster required a greater number of atoms, electrons, and basis functions to be treated simultaneously at the QM level. Because this increased the computational cost significantly, we could not carry out QM/MM umbrella sampling calculations for these reaction steps, and instead used a cluster model that included the atoms shown in blue in Figure S1, right panel. In this model, the dangler Fe is coordinated to the tridentate cysteine ligand, 5 methylimidazole (as a model for the His265 side chain), and two water molecules. The cysteine S atom bridges the dangler Fe and the Fe4S⁴ auxiliary cluster, and coordination of cysteine residues to the other three Fe atoms are modeled as MeS (methanethiol) groups.

To initiate the catalytic cycle, the initial equivalent of COOH• and CN[−] ligands are placed in close proximity to the dangler iron. Figure S3 indicates that the dangler Fe is adjacent to a large cavity in the TIM barrel normally occupied by water molecules, indicating there is sufficient space for the movement and reorientation of ligands.

The QM calculations at the auxiliary cluster were carried out using the TeraChem package,^{19,20} which includes graphics processing unit (GPU)-accelerated implementations of density functional theory (DFT) and implicit solvent models. During geometry optimizations, the B3LYP functional and mixed LANL2DZ ECP/6-31G* basis set was adopted, same as the QM/MM calculations. A few optimizations focusing on spin crossover employed the $B3LYP^*$ functional²¹ instead, which reduced the percentage of Hartree-Fock exchange from 20% to 15% and improved accuracy for spin crossover enthalpies of iron-containing complexes.²² After optimizing the geometries, single point energies were computed along the minimum energy pathway to further improve accuracy; these employed a hybrid functional that combines 5% HF / 95% B88 exchange and P86 correlation, here called BP86x5,²³ and a larger triple- ζ basis set called ma-def2-TZVP(-f)_LTZ+.²⁴ This basis combines def2-TZVP²⁵ with $l \geq 3$ basis functions removed for non-Fe atoms, augmented by a minimal set of diffuse functions,²⁶ and the LANL2TZ+ ECP/basis set for Fe atoms.²⁷ Empirical dispersion corrections of the D3(BJ) form were used, adopting the model parameters developed for BP86.²⁸ A switching Gaussian polarizable continuum model 29–31 was used with standard Bondi radii and a dielectric constant of 78.4 (equivalent to water), because we observed the auxiliary cluster in our MM MD simulations to be solvated by 15-20 water molecules.

Equilibrium geometries and transition states were optimized using the geomeTRIC software package, 32 which uses a translation-rotation internal coordinate system to efficiently optimize the geometries of multi-molecular systems. After optimization of the transition states, an approximate minimum energy path was obtained by minimizing the energy starting from the TS structure along the imaginary mode with step sizes restricted to $< 0.01 \text{\AA}$. The energy corrections for the TS were obtained the IRCMax approach³³ by tracing over

this path with single-point calculations at the $BP86x5/ma-def2-TZVP-f_LTZ+$ level of theory and choosing the highest energy on the path. The Gibbs free energies of these reaction steps were estimated using vibrational analysis carried out at the reaction endpoints and the transition state and applying the rigid rotor/harmonic oscillator approximation. Although these free energy corrections model the translational entropy using an ideal gas, which is highly approximate and for which numerous corrections have been proposed, $34-36$ we did not find the role of translational entropy to be significant in any reaction steps studied here.

In order to compute standard redox potentials corresponding to electron transfer (ET) and proton-coupled electron transfer (PCET) steps, the free energy change of the reaction was computed in solution. The free energy of the proton at pH 0.0 in aqueous solution was taken to be -11.803 eV, following previous studies,^{37,38} and adjusted to -11.390 eV by adding 59 meV per pH unit according to the Nernst equation. The redox potential was then computed as:

$$
E^{\circ} = \frac{\Delta G(\text{reduced - oxidized})}{nF} - 4.43 \text{ V}
$$
 (1)

where *n* is the number of electrons transferred $(1 \text{ in this study})$, *F* is the Faraday constant $(1 \text{ eV } V^{-1})$, and 4.43 V is the absolute potential of the standard hydrogen electrode.³⁹ In reactions where a chemical reducing agent (dithionite) was used, the overpotential was computed as $\eta = -(E^{\circ} - 0.66V)$ and the resulting value is converted back to a free energy difference in kcal/mol, allowing for the inclusion of electrochemical steps on a reaction free energy diagram. ⁴⁰

EPR properties, in particular the g-tensor and hyperfine tensor (A) eigenvalues, were computed for the structures $5a$, 10 and 11 using the ORCA software package.⁴¹ These calculations used the BP86x5 functional (same as the energies) with the zeroth-order regular relativistic approximation (ZORA) Hamiltonian, ⁴² and a mixed basis set consisting of EPR-III⁴³ for first-row elements and "ZORA-def2-TZVP" for Fe and S, a recontracted version of def2-TZVP⁴⁴ for ZORA calculations. The RIJCOSX method consisting of density fitting for Coulomb and chain-of-spheres approximation for exchange integrals was employed to speed

up the calculations.⁴⁵ The g-tensor was computed using a coupled-perturbed SCF approach⁴⁶ and the hyperfine values were computed from spin-orbit couplings. ⁴⁷ The 3D structure of 5a was used as-is, whereas the 10 and 11 were modified by removing the spectator CH_3SH and HCN ligands (respectively) and re-optimizing the structure at the BP86x5/def2-SV(P) level of theory.

Figure S1: QM regions used for various elementary steps at the rSAM and auxiliary clusters. Left: Atoms shown in brown or red comprise the QM/MM region for the SAM decomposition step, and atoms shown in blue or red comprise the QM/MM region for other reaction steps in the canonical cluster. Right: Atoms in blue comprise the auxiliary cluster model.

Figure S2: The basis set dependence of the three reactions studied using QM/MM, corresponding to Figures 2-4 in the main text, is investigated by driving the reaction coordinate. The larger def-TZVP basis set (blue curve) predicts barrier heights that are 2 kcal/mol lower for both the $C\alpha$ -C β cleavage (top left) and DHG decomposition reactions (bottom left) compared to the 6-31G* basis used in QM/MM umbrella sampling studies. By contrast, the two basis sets give nearly identical energy profiles in DHG radical formation (top right). Schemes of the reactions are shown at bottom right.

Figure S3: The position of the auxiliary cluster and dangler Fe at the end of the HydG TIM barrel. The green surface indicates the interior space of the barrel indicating sufficient space for ligand substitutions to be carried out. The solvent-accessible surface is drawn with a probe radius of 1.4Å.

Figure S4: Energy profile of 5-Ado radical generation, generated by driving the reaction coordinate, defined as RC1 = d(C. . .S) – d(Fe. . .S). Here the activation energy is ≈ 26.3 kcal/mol, which is comparable to the experimental data (23 kcal/mol).

Figure S5: Possible alternative pathways in tyrosine radical formation and the DHG radical formation, (the main mechanism is Figure 2 and 4 in the main text respectively). As for the tyrosine radical formation, the experimental data supports the mechanism that the hydrogen abstraction occurred in the amine hydrogen. Here our calculations also support this mechanism since the other possible H resource, which is the C_{α} , cannot transfer its hydrogen with an energy barrier that as low as the amino group. Regarding the DHG radical formation, the abstraction of the H from amino group gives a more stable product than that of the H connected to the sp3 hybridization carbon.

Scheme 1: Possible protonation states of the tyrosine substrate.

Figure S6: Comparison of alternative tyrosine decomposition mechanisms (the main mechanism is Figure 3 in the main text). Two possible protonation states of tyrosine, denoted TYH and TYY, are shown on the top and bottom respectively. To facilitate a direct comparison, we assume the radical NH2 \bullet + transfers a proton to the Glu side chain prior to to C—C cleavage, which could also be observed in our QM/MM simulations. The energy profiles of the two C—C cleavage mechanisms for each protonation state are shown on the left; blue curves represent the $C\alpha - C\beta$ cleavage and red curves represent the $C\alpha - C(=0)$ cleavage. The frontier orbitals of these two tyrosine models are shown on the right. In the TYH model, the LUMO of the β electron is shared between the NH[•] and the C α —C β σ bond, indicating that the H-atom abstraction may have the effect of weakening the $C\alpha$ — $C\beta$ bond order. This correlates with the lowered barrier of $C\alpha$ — $C\beta$ cleavage in TYH and is not observed in TYY, where the barrier to C_{α} — C_{β} cleavage is significantly higher.

Table S1: The key properties of the various calculated states in the catalytic cycle at the auxiliary cluster. The state labels correspond to Fig. 5 and Fig. 6 in the main text. The charge and spin multiplicity given is for the whole calculated system, including any added COOH• and CN[−] ligands. The state labeled 10 is actually two calculations consisting of a synthon fragment and Fe₄S₄—CN fragment labeled 10.S and 10.C respectively.

State	Charge	Spin	Fe ₄ S ₄	Dangler Fe State	$N(COOH^{\bullet})$	$N(CN^{-})$
Label		Mult.	State			
1	-3	5	$^{+}$	$Fe(II)(Cys)(5-MIm)(H_2O)_2$	$\mathbf{1}$	$\mathbf{1}$
$\overline{2}$	-3	1	$^{+}$	$Fe(II)(Cys)(5-MIm)(CN)(H2O)$	1	$\mathbf{1}$
3	-3	1	$2+$	$Fe(II)(Cys)(5-MIm)(CN)(COOH)$	$\mathbf 1$	$\mathbf{1}$
$\overline{4}$	-2	1	$2+$	$Fe(II)(C_{V}S_{H}^{+})(5-MIm)(CN)(COOH)$	$\mathbf{1}$	$\mathbf{1}$
5	-2	1	$2+$	$Fe(II)(\overline{\text{Cys}})(5\text{-}M\text{Im})(CN)(CO)$	$\mathbf 1$	$\mathbf{1}$
4a	-3	$\overline{2}$	$^{+}$	$Fe(II)(CysH+)(5-MIm)(CN)(COOH)$	$\mathbf 1$	$\mathbf{1}$
5a	-3	$\overline{2}$	$+$	$Fe(II)(Cys)(5-MIm)(CN)(CO)$	1	$\mathbf{1}$
6	-3	1	$+$	$Fe(III)(Cys)(5-MIm)(CN)(COCOOH)$	$\overline{2}$	$\mathbf{1}$
7	-2	$\mathbf{1}$	$2+$	$Fe(II)(CysH+)(5-MIm)(CN)(COCOOH)$	$\overline{2}$	$\mathbf{1}$
8	-2	1	$2+$	$Fe(II)(Cys)(5-MIm)(CN)(CO)$	$\overline{2}$	$\mathbf{1}$
7a	-3	$\overline{2}$	$^{+}$	$Fe(II)(CysH+)(5-MIm)(CN)(COCOOH)$	$\overline{2}$	$\mathbf{1}$
8a	-3	$\overline{2}$	$^{+}$	$Fe(II)(Cys)(5-MIm)(CN)(CO)$	$\overline{2}$	$\mathbf{1}$
9	-3	$\overline{2}$	$^{+}$	$Fe(II)(Cys)\overline{(CN)(CO)_2}$	$\overline{2}$	$\mathbf{1}$
10	-4	$\overline{2}$	$^{+}$	Fe(II)(Cys)(CN)(CO) ₂	$\overline{2}$	$\overline{2}$
10.S	-1	$\mathbf 1$	N/A	Fe(II)(Cys)(CN)(CO) ₂	$\overline{2}$	$\mathbf{1}$
10.C	-3	$\overline{2}$	$^{+}$	N/A	θ	$\mathbf{1}$
11	-3	$\overline{2}$	$^{+}$	N/A	θ	$\mathbf{1}$

Figure S7: Several possible pathways of the reduction of COOH radical to CO in the canonical rSAM $Fe₄S₄$ cluster pocket. All of the activation energies are in excess of 30 kcal/mol, which indicates that COOH does not decompose in the cluster pocket. Instead we propose that COOH• diffuses to the auxiliary cluster where reduction to CO occurs at the dangler iron.

Figure S8: Structures of the $Fe(cys)(5-MIm)$ complex used in the multireference DMRG studies.

Table S2: The energy differences between different iron cluster structures calculated by density matrix renormalization group (DMRG).

		$\text{Fe}(H_2O)_2$ $\text{Fe(CO)}(H_2O)$	Fe(CN)(H ₂ O)	Fe(CO)(CN)
Active Space	(20, 18)	(20, 18)	(20, 18)	(22, 20)
$E(HS)$ - $E(LS)$	-68.2	$28.3\,$	25.1	$50.2\,$

Multireference DMRG calculation methods. All four structures were optimized under B3LYP//6-31G*/LANL2DZ(Fe) in Q-Chem, and the DMRG single point energies were calculated using the TZV basis set in PySCF. The active spaces in the calculations were selected from localized molecular orbitals by including the five orbitals with 3d character on the Fe atom and the orbitals with p character on the coordinating ligands $(2p \text{ for } O, C, N)$ and 3p for S) that had an overlap integral of > 0.3 with any of the d orbitals; the orbital selections were then confirmed by visual inspection of isosurface plots. The total number of orbitals and electrons in the DMRG calculations is provided in the table above as (active electrons, active orbitals). The bond dimension of the DMRG calculations is set to 1000, and the number of sweeps is set to the default value of 4. Energies are in units of kcal/mol.

Figure S9: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction 1 -LS + CN⁻ \rightarrow 2 + H₂O where a CN⁻ ligand displaces an aquo ligand. A spectator COOH[•] species is present in the system.

Figure S10: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $2\text{-LS} + \text{COOH}^{\bullet} \rightarrow 3 + \text{H}_2\text{O}$ where COOH^{\bullet} displaces the second aquo ligand. Following this, 3 is protonated to form 4.

Figure S11: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $4 \rightarrow 5 + H₂O$ where the COOH ligand accepts a proton from the cysteine oxygen, then is decomposed to CO and H_2O .

Figure S12: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $5a + COOH^{\bullet} \rightarrow 6$ where the added COOH^{\bullet} forms a C—C bond with the first CO ligand to form a COCOOH ligand to the dangler Fe. Following this, 6 is protonated to form 7.

Figure S13: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $7 \rightarrow 8 + CO + H_2O$ where the COCOOH ligand accepts a proton from the cysteine oxygen, then is decomposed to a free $CO + CO$ ligand + H_2O .

Figure S14: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $8 + CO \rightarrow 9 + 5$ -MIm where the second CO displaces the 5-methylimidazole ligand.

Figure S15: 3D renderings and plot of electronic energies (B3LYP/6-31G* LANL2DZ/PCM) along the minimum energy path of the reaction $10 + CH_3SH \rightarrow 11 + HCN$ where a CH_3SH model of the cysteine side chain displaces the CN ligand to the auxiliary cluster, releasing HCN and turning over the catalytic cycle.

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