## **Supplementary Information**

# Single Cell Profiling of Primary and Paired Metastatic Lymph Node Tumors in Breast Cancer Patients

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### Supplementary Fig. 1. Overview of major clusters in PT and MLNT (related to Fig. 1)

**a**. Hematoxylin and eosin (HE) staining of PT and MLNT tissues in 3 representative patients. Original magnification, 10x, scale bar = 1 mm or 200x, scale bar = 50  $\mu$ m. **b**. UMAP visualization of the entire data set before (left) and after (right) batch alignment. Cells were colored by batch effect (patients). **c&d**. UMAP plots showing the expression pattern of marker genes (c) or major cell types (d). **e**. The proportion of tissues within major cell types. The color represents different tissues. The *x*-axis is the fraction of tissues, and the *y*-axis is the major cell types. **f**. The proportion of major cell types. The *x*-axis is the fraction of cell types, and the *y*-axis presents different patients. **g**. Percentage of spots representing positive detected tumor cells in LNMT. Source data are provided as a Source Data file.



# Supplementary Fig. 2. Characterization of T cells and NK cells in PT and MLNT (related to Fig. 2)

**a**. A UMAP embedding plot showing representative markers of T cells and NK cells. **b**. Violin plots show a set of known marker genes that distinguish various clusters. **c**. Bar plots showing the percentages of T cells and NK cell clusters between 2 tissues (PT: n = 8 samples, LNM: n = 8 samples). Statistical testing was performed by two-sided Wilcoxon test. Data are presented as mean values +/- SD. **d&e**. Cumulative distribution plot showing the distribution of cytotoxic scores (e) or exhausted scores (f) in each CD8 T subtype. A rightward shift of the curve indicates increased state scores. **f**. The variance (bottom) and z score (top) of the first 20 PCs. The expression matrix of CD8 T cells from PT and MLNT was calculated for the first 20 PCs. To compare the difference between PTs and MLNTs in each PC, z scores from unpaired t test were measured between the tumor and lymph node of each PC. **g**. The top 20 genes that contribute to PC2 variance. Genes were ranked by PC2 loadings. Source data are provided as a Source Data file.



## Supplementary Fig. 3. Characterization of TCR in PT and MLNT (related to Fig. 3)

**a.** Bar plot showing the number of T cells for TCR analysis in each patient. Very rare immune cells were present in Patient 2 and Patient 4; thus, these 2 samples were removed from downstream analysis. **b.** Pie charts showing the fraction of clonal size of all clonotypes. Red represents unique clonotypes, green represents clonal size clonotypes of 2, and blue represents clonal size clonotypes of 3 or above. **c&d**. Developmental transition ability (b) or clonal expansion levels (c) of CD4 T cells and CD8 T cells clusters quantified by STARTRAC-transition indices for each patient (n = 6). In the box plots, the center line corresponds to the median, box corresponds to the interquartile range (IQR), and whiskers  $1.5 \times IQR$ . Source data are provided as a Source Data file.



Supplementary Fig. 4. Characterization of migrated T cells in PT and MLNT (related to Fig. 4)

**a&b.** Top 8 enriched pathways for up-regulated genes for matched CD8 T cells (a) or matched Tconvs (b) in LNMT vs PT. **c**. Volcano plot showing differentially expressed genes between LNMT vs PT for matched Tregs. *P* value <0.05, log 2 (fold change)  $\ge$  0.25. Statistical testing was performed by a two-sided Wilcoxon test. Source data are provided as a Source Data file.



### Supplementary Fig. 5. Myeloid cells in PT and MLNT (related to Fig. 5).

**a.** A UMAP plot showing the expression pattern of selected marker genes for myeloid clusters. **b.** A UMAP plot showing DC clusters color-coded by tissues. Different cell clusters are circled. **c.** A violin plot showing the *ISG15* expression level of DC clusters. **d.** A heatmap showing the difference of enriched pathway analyzed by GSVA between PT and MLNT in the DC-C3-*LAMP3* cluster. **e.** A UMAP plot showing the projection of GSE114727 dataset<sup>1</sup> according to the "ingest" function. Cells are colored according to tissue type (left) or myeloid markers, *LYZ* (right). **f.** A boxplot of the M1 (left panel) and M2 (right panel) signature of macrophage clusters. (Macro-C1-*APOC1* n = 627 cells, Macro-C2-*SLC40A1* n = 468 cells, Macro-C3-*VCAN* n = 540 cells, Macro-C4-*CXCL11* n = 126 cells, Macro-C5-*SPP1* n = 542 cells, Macro-C6-*CCL3* = 475 cells). In the box plots, the center line corresponds to the median, the box corresponds to the IQR, and whiskers  $1.5 \times IQR$ . **g.** Scatter plots show that M1 correlated with the M2 signature in PTs and LNMTs. The correlation coefficient and *P*-value were calculated with two-sided Pearson rank correlation. Source data are provided as a Source Data file.



Supplementary Fig. 6. Characterization of PLA2G2A+ CAFs (related to Fig. 6)

**a**. A UMAP plot showing the marker genes for 3 types of stromal cells. Yellow indicates higher expression and black indicates lower expression. **b**. A dot plot showing marker genes across TEC and CAF clusters. Dot sizes indicate the fraction of expressed cells and are colored according to the normalized expression level. **c**. A UMAP plot showing the *PLA2G2A* in the TEC and CAF data sets. Yellow indicates higher expression and black indicates lower expression. **d**. Scatter plots showing *PLA2G2A* correlated with immune cell markers in the TCGA-BRCA dataset (n = 640). Lines are fit for each select pair. The correlation coefficient and *P*-value were calculated with two-sided Pearson rank correlation. Source data are provided as a Source Data file.



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#### Supplementary Fig. 7. Characterization of epithelial cells (related to Fig. 7)

**a**. A histogram showing the distribution of malignant scores in epithelial cells and reference cells. Orange indicates the malignant score distribution of malignant epithelial cells, blue indicates nonmalignant epithelial cells, and grey indicates reference cells. b. Cells were classified into malignant epithelial cells, nonmalignant cells, and non-epithelial cells. A boxplot the showing number of features (left) and epithelial scores (right) of the 3 types. (Non-epithelial: n=100422 cells, Non-malignant epithelium: n = 12069 cells, Malignant epithelium: n = 6354 cells). In the box plots, the center line corresponds to the median, the box corresponds to the IQR, and whiskers  $1.5 \times IQR$ . c&d. Upset plot showing the overlapping of the upregulated genes in PT (c) and the upregulated genes in LNMT (d). Differential analysis was calculated for each patient. P value < 0.05, log2 (fold  $change \ge 0.5$ . e. Volcano plot showing the differentially expressed genes between PT and LNMT in malignant cells from patient 5. P value < 0.05, log2 (fold change)  $\ge$  0.5. Statistical testing was performed by a two-sided Wilcoxon test. f&g. Volcano plot showing the differentially expressed genes between PT and LNMT in malignant cells for CNV cluster 1(f) or CNV cluster 2 (g) from patient 8. P value < 0.05, log2 (fold change)  $\ge 0.5$ . Statistical testing was performed by a two-sided Wilcoxon test. g. GSEA analysis showed that genes rank in PT vs LNMT of cluster 1 (left panel) and cluster 2 (right panel) in patient 8 are enriched in the antigen presentation pathway. The left represents a high gene rank in PT. Statistical testing was performed by permutation test. The Pvalues were corrected with Benjamini-Hochberg adjustment. h. Circos plot showing 6 CNV clusters of malignant epithelial cells according to CNVs similarity for patient 5. Cells were colored by tissue. i. The bar plot shows the frequency of 5 CNV clusters in patient 5. j. Violin plots showing the MHC I antigen presentation signature (Up) and MHC II antigen presentation signature (Down) of 5 CNV clusters from patient 5. Source data are provided as a Source Data file.

#### Supplementary References:

1 Azizi, E. *et al.* Single-Cell Map of Diverse Immune Phenotypes in the Breast Tumor Microenvironment. *Cell* **174**, 1293-1308 e1236, doi:10.1016/j.cell.2018.05.060 (2018).