

SUPPORTING INFORMATION

Copper-Catalyzed Three-Component Aminofluorination of Alkenes and 1,3-Dienes: Direct Entry to Diverse β -Fluoro Alkylamines

Guangshou Feng,[§] Colton K. Ku,[§] Jiaqi Zhao, and Qiu Wang*

Department of Chemistry, Duke University, Durham NC 27708
Email: qiu.wang@duke.edu

Table of Contents

1. General Methods	S1
2. Optimization Screens of Alkene Aminofluorination Reaction Conditions	S1
3. Preparation of Starting Materials	S4
4. Scope of alkenes and compound characterization data	S9
5. Scope of amines and compound characterization data.....	S19
6. Amino fluorination of 1,3-dienes and compound characterization data	S26
7. Synthetic applications.....	S32
8. Mechanism studies	S39
9. References	S41
10. Copies of the NMR spectra	S42
11. X-ray crystallography information.....	S283

1. General Methods

General Procedures

Round-bottom flasks and stir bars were dried either with a propane torch or in an oven at 140 °C overnight and cooled/stored in a desiccator filled with Drierite. Optimization and substrate screens were performed in 10-mL FEP Tubes without prior drying, and a Teflon-coated micro stir bar. All other reactions were performed in round-bottom flasks with rubber septa and Teflon-coated stir bars. Plastic syringes were used for the transfer of pure solvents, while glass pipets were used for transfer of crude reaction mixtures. Analytical thin-layer chromatography (TLC) was performed using aluminum plates coated with a 0.25 mm layer of 230–400 mesh silica gel with fluorescent indicator (254 nm). TLC plates were visualized by exposure to ultraviolet light and treatment with mainly vanillin or phosphomolybdic acid (PMA) stain. Organic solutions were concentrated under reduced pressure using a rotary evaporator and flash chromatography performed using 60 Å silica gel and HPLC-grade solvents.

Materials

Commercial reagents and anhydrous solvents were used as received. Specific anhydrous solvents (CH₂Cl₂, Toluene, Dioxane, and THF) were obtained from the Innovative Technologies solvent purification system. Commercially available substrates were purchased in >95% purity and used without further purification.

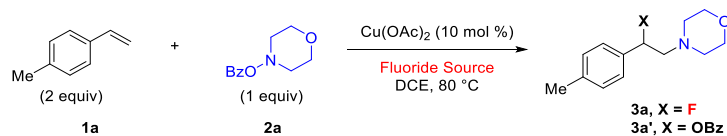
Instrumentation

Nuclear magnetic resonance spectra were recorded on either a Varian 400 MHz spectrometer or Bruker 500 MHz spectrometer at room temperature. Chemical shifts for ¹H NMR are reported in parts per million (ppm, δ) and referenced to residual protium in CDCl₃ (δ 7.26). NMR values are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, m = multiplet, br = broad), coupling constant (Hz), and integration. Infrared spectroscopic data are reported in wavenumbers (cm⁻¹) with selected peaks shown. High-resolution mass spectra were obtained using a liquid chromatography-electrospray ionization and time-of-flight mass spectrometer. The X-ray single crystal data and refinement was obtained from UNC Chapel Hill department of chemistry X-ray Core Laboratory.

2. Optimization Screens for Alkene Aminofluorination Reaction Conditions

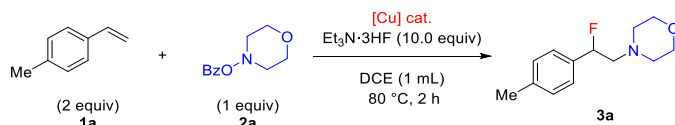
General procedure 1(GP1) for optimization screening

To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylamine and copper catalyst. Solvent (1.0 mL), 4-methylstyrene, and fluorinating reagent were sequentially added via syringe. The mixture was then stirred and heated at the indicated temperature until the consumption of *O*-benzoylhydroxylamine, verified by TLC analysis. The resulting reaction mixture was cooled to room temperature and quenched by the addition of Et₃N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was either purified by silica column chromatography or added to CDCl₃ (0.5 mL) with CH₂Br₂ (0.1 mmol) as a quantitative internal standard for NMR analysis.

Table S1. Screening of fluoride sources.^a

Entry	Fluoride source	Fluoride (equiv)	3a (%) ^b	3a' (%) ^b
1	$\text{Py}(\text{HF})_x$	2.0	0	0
2	AgF	2.0	0	0
3	CsF	2.0	0	11
4	TBAF	2.0	0	0
5	$\text{Et}_3\text{N}\cdot 3\text{HF}$	2.0	40	6
6	$\text{Et}_3\text{N}\cdot 3\text{HF}$	10.0	50 (50)^c	trace

^aReaction conditions: **1a** (2.0 equiv), **2a** (0.2 mmol, 1.0 equiv), fluoride source (2 or 10 equiv), $\text{Cu}(\text{OAc})_2$ (10 mol %), DCE (1.0 mL), 80 °C, 2 h. ^bYields determined by ¹H NMR spectroscopy with CH_2Br_2 as a quantitative internal standard. ^cIsolated yield.

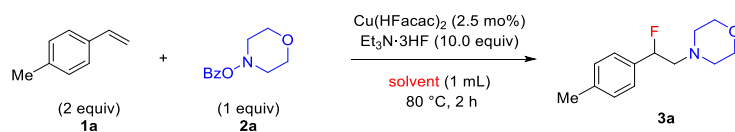
Table S2. Screening of copper catalysts.^a

Entry	$[\text{Cu}]$ cat.	3a (%) ^b
1	$[\text{Cu}(\text{OTf})_2]\text{PhMe}$	50
2	$\text{Cu}(\text{OAc})$	50
3	CuCl_2	40
4	CuCl	44
5	$\text{Cu}(\text{eh})_2$	60
6	$\text{Cu}(\text{MeCN})_4\text{PF}_6$	52
7	IPrCuCl	62 (68%)^c
8	CuF_2	54
9	CuBr	52
10	CuI	12
11	CuCN	30
12	CuTC	52
13	$\text{Cu}(\text{PhCO}_2)_2$	52
14	$\text{Cu}(\text{NO}_2)_2\cdot 3\text{H}_2\text{O}$	40
15	CuSCN	52
16	CuSO_4	54
17	$\text{CuSO}_4\cdot 5\text{H}_2\text{O}$	50
18	CuO	68 (67) ^c

19	CuO ₂	58
20	Cu(OTf) ₂	54
21	CuBr ₂	42
22	Cu(MeCN)BF ₄	54
23	Cu(OAc) ₂	48 (47) ^c
24	Cu(OAc) ₂	50 (50) ^c
25	Cu(TFA) ₂	59
26	Cu(OPiv) ₂	65
27	Cu(acac)₂	76 (76)^c
28	Cu(HFacac)₂	81 (83)^c
29	Cu(HFacac)₂ (2.5 mol %)	82 (83)^c

^a Reaction conditions: **1a** (2.0 equiv), **2a** (0.2 mmol, 1.0 equiv), fluoride source (10 equiv), copper cat. (2.5–10 mol %), DCE (1.0 mL), 80 °C, 2 h. ^bYields determined by ¹H NMR spectroscopy with CH₂Br₂ as a quantitative internal standard. ^cIsolated yields.

Table S3. Screening of solvents.^a

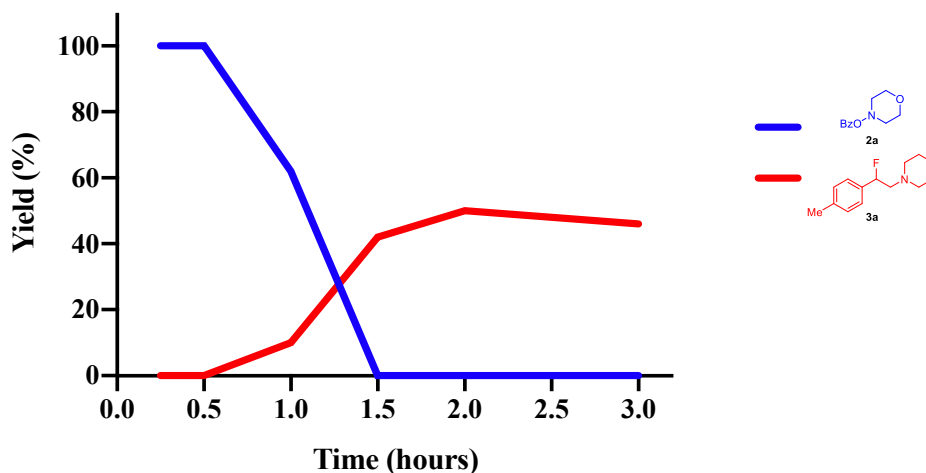


Entry	Solvent	3a (%) ^b
1	DME	30
2	1,4-Dioxane	46
3	THF	24
4	Toluene	20
5	DCE	83
6	MeCN	40

^aReaction conditions: **1a** (2.0 equiv), **2a** (0.2 mmol, 1.0 equiv), Et₃N·3HF (10 equiv), solvent (1.0 mL), 80 °C, 2 h.

^bYields determined by ¹H NMR spectroscopy with CH₂Br₂ as a quantitative internal standard.

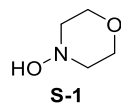
Reaction progress and completion time testing



To determine reaction progress and completion time of the above transformation, seven reactions with 4-methylstyrene **1a** (0.2 mmol scale) and *O*-benzoyl hydroxymorpholine **2a** were subject to **GPI**, and quenched at 0.25, 0.5, 1.0, 1.5, 2.0, 2.5, and 3.0 h, respectively. Crude NMR yields using CH₂Br₂ as a quantitative internal standard were used to generate the above graph, which indicates complete consumption of the hydroxylamine occurs within 1.5 h and the formation of the product finishes within 2.0 h.

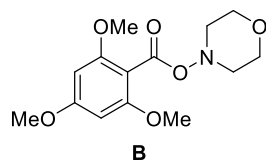
3. Preparation of Starting Materials

3.1. Preparation of hydroxylamine derivatives and compound characterizations.



Morpholin-4-ol (S-1). To a 250-mL round-bottom flask was added NaOMe (1.59 g, 28 mmol, 1.4 equiv) and MeOH (70 mL). In a separate 100-mL round-bottom flask was added *O*-benzoyl-*N*-hydroxymorpholine **2a** (4.14 g, 20 mmol, 1.0 equiv) and Et₂O (60 mL). The dissolved hydroxylamine ester was then added via syringe to the solution of NaOMe. The mixture was allowed to stir for 1 h, and then directly dried *in vacuo*. The crude mixture was purified by flash column chromatography (30% ethyl acetate–hexanes to 40% ethyl acetate–hexanes) to afford morpholin-4-ol (**S-1**) as colorless oil (1.33 g, 69%). Spectroscopic data match the commercially available *N*-hydroxylamine (CAS 5765-63-9).

¹H NMR (400 MHz, CDCl₃) δ 6.56 (br, 1H), 3.91 (d, *J* = 12.0 Hz, 2H), 3.76–3.47 (m, 2H), 3.14 (d, *J* = 11.0 Hz, 2H), 2.72 (td, *J* = 11.0, 3.2 Hz, 2H).



Morpholino 2,4,6-trimethoxybenzoate (B). To a 100-mL round-bottom flask was added 2,4,6-trimethoxybenzoic acid (0.849 g, 4.0 mmol, 1.0 equiv), CH₂Cl₂ (10 mL), oxalyl chloride (406 μL,

4.8 mmol, 1.2 equiv), and dimethylformamide (0.2 mL). Upon the discontinuation of gas evolution, the mixture was directly concentrated under reduced pressure. The mixture was next diluted with CH₂Cl₂ (10 mL), and morpholin-4-ol (433 mg, 4.2 mmol, 1.05 equiv) was added over 5 min, followed by the addition of triethylamine (585 μL, 4.2 mmol, 1.05 equiv). Upon consumption of acid starting material (monitored by TLC), the reaction was quenched through the addition of saturated aqueous NH₄Cl (10 mL) and H₂O (20 mL). The organic layers were separated, and washed with saturated aqueous NH₄Cl (10 mL × 3), saturated aqueous NaHCO₃ (10 mL × 3), H₂O (10 mL × 3), and brine (10 mL × 2). The organic layer was dried over Na₂SO₄, and filtered. The filtrate was concentrated *in vacuo*. Purification by flash column chromatography (20% ethyl acetate–hexanes to 60% ethyl acetate–hexanes) gave **B** as a white solid (493 mg, 42%). Spectroscopic data matched the reported literature.¹

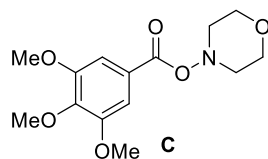
R_f = 0.33 (10% MeOH, 40% ethyl acetate, 50% hexanes).

¹H NMR (500 MHz, CDCl₃) δ 6.08 (s, 2H), 3.98–3.75 (m, 13H), 3.44 (d, *J* = 9.3 Hz, 2H), 3.04–2.90 (m, 2H).

¹³C NMR (125 MHz, CDCl₃) δ 164.5, 162.9, 158.9, 104.3, 90.6, 65.8, 56.8, 55.9, 55.4.

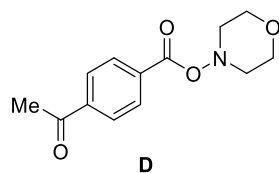
FTIR (thin film): cm⁻¹ 2965, 2845, 1748, 1607, 1590, 1458, 1243, 1228, 1129, 1039.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₄H₂₀NO₆⁺ 298.1285; found: 298.1287.



Morpholino 3,4,5-trimethoxybenzoate (C). To a 100-mL round-bottom flask was added 3,4,5-trimethoxybenzoic acid (1.02 g, 4.8 mmol, 1.2 equiv), CH₂Cl₂ (15 mL), and 1,1'-carbonyldiimidazole (778 mg, 4.8 mmol, 1.2 equiv). The mixture was allowed to stir overnight, until the formation of CO₂ gas halted. Morpholin-4-ol (413 mg, 4.0 mmol, 1.0 equiv) was next added in CH₂Cl₂ (5 mL). The mixture was allowed to stir until the consumption of hydroxylamine (monitored by TLC). The solvent was removed *in vacuo*. The crude mixture was diluted with ethyl acetate (10 mL) and H₂O (10 mL). The organic layer was separated, washed with H₂O (10 mL × 2), saturated aqueous NH₄Cl (10 mL × 3), brine (10 mL × 2), dried over Na₂SO₄, and filtered. The filtrate was concentrated *in vacuo* to afford the crude product. Purification by flash column chromatography (40% ethyl acetate–hexanes to 50% ethyl acetate–hexanes) gave **C** as a white solid (716 mg, 60%). Spectroscopic data matched the reported literature.²

¹H NMR (400 MHz, CDCl₃): δ 7.25 (s, 2H), 4.02–3.78 (m, 13H), 3.52–3.39 (m, 2H), 3.11–2.96 (m, 2H).



Morpholino 4-acetylbenzoate (D). To a 100-mL round-bottom flask was added 4-acetylbenzoic acid (0.657 g, 4.0 mmol, 1.0 equiv), CH₂Cl₂ (10 mL), oxalyl chloride (406 μL, 4.8 mmol, 1.2 equiv), and dimethylformamide (0.2 mL). Upon the discontinuation of gas evolution, the mixture was directly concentrated under reduced pressure. The mixture was next diluted with CH₂Cl₂ (10 mL), and morpholin-4-ol (433 mg, 4.2 mmol, 1.05 equiv) was added over 5 min, followed by the addition of triethylamine (585 μL, 4.2 mmol, 1.05 equiv). Upon consumption of acid starting material (monitored by TLC), the reaction was quenched by the addition of saturated aqueous NH₄Cl (10 mL) and H₂O (20 mL). The separated organic layer was washed with saturated aqueous NH₄Cl (10 mL × 3), saturated aqueous NaHCO₃ (10 mL × 3), H₂O (10 mL × 3), and brine (10 mL × 2), dried over Na₂SO₄, and filtered. The

filtrate was concentrated *in vacuo*. Purification by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes) gave **D** as a white solid (545 mg, 55%).

R_f = 0.18 (30% ethyl acetate–hexanes).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.09 (d, J = 8.4 Hz, 2H), 8.01 (d, J = 8.4 Hz, 2H), 4.08–3.72 (m, 4H), 3.57–3.34 (m, 2H), 3.14–2.90 (m, 2H), 2.65 (s, 3H).

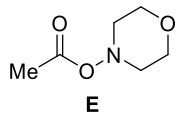
$^{13}\text{C NMR}$ (125 MHz, CDCl_3): δ 197.3, 163.7, 140.4, 132.9, 129.7, 128.2, 65.8, 57.0, 26.8.

FTIR (thin film): cm^{-1} 2967, 2900, 2858, 1737, 1685, 1404, 1240, 1100, 1083, 857, 763.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{16}\text{NO}_4^+$ 250.1074; found: 250.1077.

General procedure 2 (GP2) for the preparation of hydroxylamine starting materials E, F and G.

To a 15-mL round-bottom flask was added morpholin-4-ol (433 mg, 4.2 mmol, 1.05 equiv), dimethylaminopyridine (9.0 mg, 0.075 mmol, 0.019 equiv), freshly distilled CH_2Cl_2 (2 mL), and triethylamine (0.59 mL, 4.2 mmol, 1.05 equiv). To the reaction flask at 0 °C was added the solution of acyl chloride (4.0 mmol, 1.0 equiv) in CH_2Cl_2 (2 mL). The mixture was allowed to stir at room temperature for 30 min, until the consumption of acyl chloride (monitored by TLC). The reaction mixture was then washed with aqueous solution of HCl (1 M, 5 mL x 2). The separated organic layer was dried over Na_2SO_4 , filtered, and the filtrate was concentrated *in vacuo*. Purification by column chromatography (5% MeOH, 30% ethyl acetate, 65% hexanes unless otherwise noted) generated the desired product.



Morpholino acetate (E) Preparation *via* **GP2** gave **E** as a light-yellow oil (375 mg, 65%).

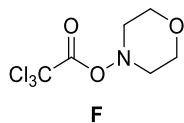
R_f = 0.34 (50% Ethyl acetate–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 3.85 (d, J = 10.9 Hz, 2H), 3.77–3.65 (br. m, 2H), 3.23 (d, J = 9.6 Hz, 2H), 2.87–2.76 (br. m, 2H), 1.99 (s, 3H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 168.8, 65.6, 56.7, 19.5.

FTIR (thin film): cm^{-1} 1756, 1210, 1102, 1003, 891, 857.

HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_6\text{H}_{11}\text{NO}_3\text{Na}^+$ 168.0631; found: 168.0628.



Morpholino 2,2,2-trichloroacetate (F). Preparation *via* **GP2** and purification by column chromatography (20% ethyl acetate–hexanes) gave **F** as a yellow solid (726 mg, 73%).

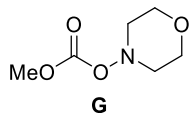
R_f = 0.34 (50% ethyl acetate–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 3.97 (d, J = 11.9 Hz, 2H), 3.82–3.72 (br. m, 2H), 3.43 (d, J = 9.9 Hz, 2H), 3.11–3.01 (br. m, 2H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 160.0, 88.9, 65.6, 56.8.

FTIR (thin film): cm^{-1} 1777, 1460, 1267, 1194, 1098, 962, 865, 812, 674.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_6\text{H}_9^{35}\text{Cl}_3\text{NO}_3^+$ 247.9643; found: 247.9643.



Methyl morpholino carbonate (G). Preparation via **GP2** gave **G** as a light-yellow oil (496 mg, 77%).

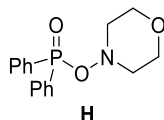
$R_f = 0.48$ (50% ethyl acetate–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 3.91 (d, $J = 11.3$ Hz, 2H), 3.80 (s, 3H), 3.77–3.70 (br. m, 2H), 3.34 (d, $J = 9.7$ Hz, 2H), 3.00–2.85 (br. m, 2H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 154.8, 65.6, 56.8, 54.9.

FTIR (thin film): cm^{-1} 1767, 1440, 1268, 1229, 1100, 1052, 860.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_6\text{H}_{12}\text{NO}_4^+$ 162.0761; found: 162.0761.



Morpholino diphenylphosphinate (H). **H** was synthesized following a previously reported method with modification.³ To a 10-mL round-bottom flask added morpholin-4-ol (412 mg, 4.0 mmol, 1.0 equiv), triethylamine (1.39 mL, 10 mmol, 2.5 equiv) and CH_2Cl_2 (0.5 mL). To the reaction flask at 0 °C, was added diphenylphosphinyl chloride (1.92 mL, 4.8 mmol, 1.2 equiv). The reaction mixture was allowed to stir at room temperature, until the consumption of morpholin-4-ol. Solvents were then removed under reduced pressure. To the residue, was added diethyl ether (10 mL) and decanted for four times. The combined extracts were filtered through a silica/celite pad and concentrated in *vacuo* to give **H** as a white solid (308 mg, 1.01 mmol, 25%).

$R_f = 0.13$ (50% ethyl acetate–hexanes).

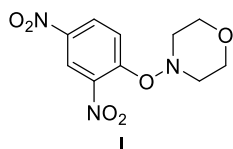
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.84 (d, $J = 8.0$ Hz, 2H), 7.82 (d, $J = 8.0$ Hz, 2H), 7.55–7.50 (m, 2H), 7.47–7.41 (m, 4H), 3.82 (d, $J = 11.8$ Hz, 2H), 3.58–3.50 (br. m, 2H), 3.25 (d, $J = 10.3$ Hz, 2H), 3.04–2.96 (br. m, 2H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 132.2 (d, $J_{\text{C-P}} = 2.7$ Hz), 131.9 (d, $J_{\text{C-P}} = 9.9$ Hz), 130.7 (d, $J_{\text{C-P}} = 135.3$ Hz), 128.31 (d, $J_{\text{C-P}} = 12.9$ Hz), 65.8, 58.7 (d, $J_{\text{C-P}} = 2.7$ Hz).

$^{31}\text{P NMR}$ (CDCl_3 , 202 MHz): δ 33.63–33.36 (m).

FTIR (thin film): cm^{-1} 1438, 1227, 1128, 1020, 866, 787, 726, 693, 546, 530.

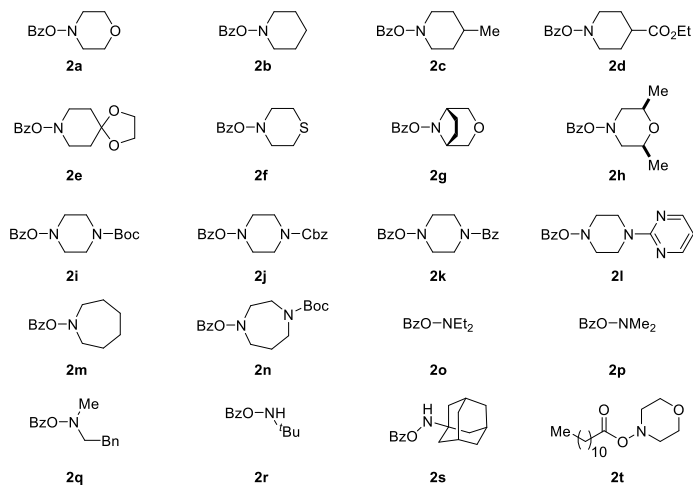
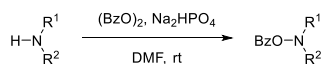
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{19}\text{NO}_3\text{P}^+$ 304.1097; found: 304.1092.



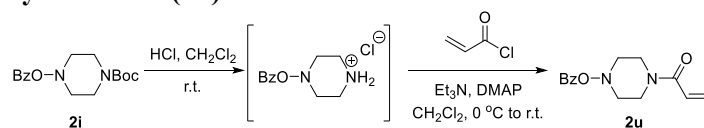
4-(2,4-Dinitrophenoxy)morpholine (I). Prepared following a previously reported method.⁴ Spectroscopic data were in agreement.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.81 (s, 1H), 8.41 (d, $J = 9.3$ Hz, 1H), 7.90 (d, $J = 9.3$ Hz, 1H), 4.06 (d, $J = 11.4$ Hz, 2H), 3.75 (t, $J = 11.4$ Hz, 2H), 3.33 (d, $J = 10.3$ Hz, 2H), 3.16 (t, $J = 10.3$ Hz, 2H).

O-Benzoylhydroxylamines below were synthesized according to the literature procedure.⁵



4-Acryloylpiperazin-1-yl benzoate (**2u**)



To a 25-mL round-bottom flask was added *tert*-butyl 4-(benzoyloxy)piperazine-1-carboxylate **2i** (306 mg, 1.0 mmol, 1.0 equiv), CH_2Cl_2 (4.0 mL), the solution of HCl in dioxane (4M, 2 mL, 8.0 mmol, 8.0 equiv). The reaction mixture was allowed to stir at room temperature for 8 h, and then concentrated under reduced pressure. The crude solid was used for the next step without further purification.

To a solution of the crude residue in CH_2Cl_2 (4 mL) at 0 °C, was added trimethylamine (202 mg, 2.0 mmol, 2.0 equiv), DMAP (2.4 mg, 0.02 mmol, 0.02 equiv), and acryloyl chloride (91 mg, 1.0 mmol, 1.0 equiv). Then mixture was allowed to warm to room temperature and stir overnight. The mixture was diluted with CH_2Cl_2 (20 mL). The organic layer was washed with water (10 mL \times 2) and the brine (10 mL). The organic layer was dried over Na_2SO_4 , filtered and concentrated *in vacuo*. Purification by flash column chromatography (30% ethyl acetate–hexanes to 50% ethyl acetate–hexanes) gave **2u** as a colorless oil (225 mg, 87% over two steps).

R_f = 0.10 (50% ethyl acetate–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.97 (t, J = 19.7 Hz, 2H), 7.58 (t, J = 7.4 Hz, 1H), 7.45 (t, J = 7.4 Hz, 2H), 6.58 (dd, J = 16.8, 10.6 Hz, 1H), 6.33 (dd, J = 16.8, 1.8 Hz, 1H), 5.74 (dd, J = 10.6, 1.8 Hz, 1H), 4.67–4.38 (m, 1H), 4.22–3.82 (m, 1H), 3.70–3.57 (m, 1H), 3.57–3.44 (m, 2H), 3.42–3.25 (m, 1H), 3.09–2.58 (m, 2H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 165.4, 164.5, 133.4, 129.5, 128.9, 128.6, 128.5, 127.0, 56.1, 55.7, 43.9, 40.3.

FTIR (thin film): cm^{-1} 1736, 1647, 1613, 1438, 1246, 1085, 1019, 709.

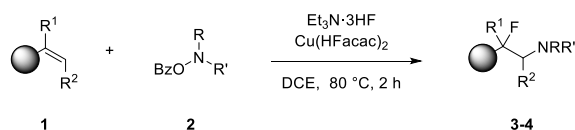
HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3\text{Na}^+$ 283.1053; found: 283.1047.

3.2. Preparation of olefin substrates and compound characterization.

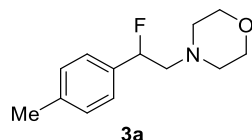
Olefins (**1a–1q** and **1u–1z**) were purchased from commercial sources and used directly without further purification. **1r–1s**,⁶ and **1t**⁷ are known alkenes.

4. Scope of alkenes and compound characterization data

General procedure 3 (GP3) for alkene aminofluorination reactions



To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylamine **2** (0.2 mmol, 1.0 equiv) and Cu(HFacac)₂ (2.4 mg, 2.5 mol%). DCE (1.0 mL), alkene **1** (0.4 mmol, 2.0 equiv), and Et₃N·3HF (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et₃N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography.



4-(2-Fluoro-2-(*p*-tolyl)ethyl)morpholine (3a). Synthesized using GP3 from **1a** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (37.3 mg, 83%).

R_f = 0.14 (30% ethyl acetate–hexanes).

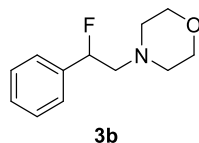
¹H NMR (500 MHz, CDCl₃) δ 7.23 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 5.62 (ddd, J = 48.9, 8.8, 2.4 Hz, 1H), 3.77–3.72 (m, 4H), 2.91 (ddd, J = 17.1, 14.2, 8.8 Hz, 1H), 2.68–2.53 (m, 5H), 2.36 (s, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 138.3, 135.7 (d, J_{C-F} = 20.0 Hz), 129.1, 125.6, 92.6 (d, J_{C-F} = 173.0 Hz), 66.9, 64.9 (d, J_{C-F} = 23.4 Hz), 54.1, 21.1.

¹⁹F NMR (471 MHz, CDCl₃) δ -175.11.

FTIR (thin film): cm⁻¹ 2955, 2854, 2808, 1453, 1138, 1115, 1008, 869, 816, 536.

HRMS (ESI) m/z : [M+H]⁺ Calcd for C₁₃H₁₉FNO⁺ 224.1445; found: 224.1451.



4-(2-Fluoro-2-phenylethyl)morpholine (3b). Synthesized using GP3 from **1b** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (27.2 mg, 65%).

R_f = 0.12 (30% ethyl acetate–hexanes).

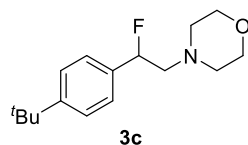
¹H NMR (500 MHz, CDCl₃) δ 7.40–7.22 (m, 5H), 5.60 (ddd, J = 48.9, 8.8, 2.3 Hz, 1H), 3.69 (t, J = 4.7 Hz, 4H), 2.98–2.72 (m, 1H), 2.67–2.58 (m, 1H), 2.58–2.41 (m, 4H).

¹³C NMR (125 MHz, CDCl₃) δ 138.7 (d, J_{C-F} = 20.6 Hz), 128.5, 128.5, 125.6 (d, J_{C-F} = 6.9 Hz), 92.6 (d, J_{C-F} = 173.7 Hz), 66.9, 64.9 (d, J_{C-F} = 23.1 Hz), 54.1.

¹⁹F NMR (471 MHz, CDCl₃) δ -176.90.

FTIR (thin film): cm^{-1} 2956, 2854, 2809, 1495, 1452, 1115, 1008, 914, 727, 698.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{12}\text{H}_{17}\text{FNO}^+$ 210.1289; found: 210.1291.



4-(2-(4-(*tert*-Butyl)phenyl)-2-fluoroethyl)morpholine (3c). Synthesized using **GP3** from **1c** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (39.2 mg, 74%).

R_f = 0.20 (30% ethyl acetate–hexanes).

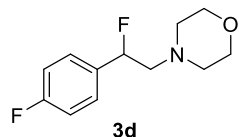
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.33 (d, J = 8.2 Hz, 2H), 7.20 (d, J = 8.2 Hz, 2H), 5.58 (ddd, J = 49.1, 9.0, 2.1 Hz, 1H), 3.69 (t, J = 4.7 Hz, 4H), 2.95–2.76 (m, 1H), 2.67–2.58 (m, 1H), 2.58–2.43 (m, 4H), 1.24 (s, 9H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 151.6, 135.6 (d, $J_{\text{C-F}}$ = 20.1 Hz), 125.4, 125.3 (d, $J_{\text{C-F}}$ = 6.5 Hz), 92.5 (d, $J_{\text{C-F}}$ = 172.8 Hz), 66.9, 64.8 (d, $J_{\text{C-F}}$ = 23.1 Hz), 54.0, 34.6, 31.3.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -175.11.

FTIR (thin film): cm^{-1} 2960, 2858, 2809, 1453, 1269, 1117, 1009, 870, 835, 71.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{25}\text{FNO}^+$ 266.1915; found: 266.1914.



4-(2-Fluoro-2-(4-fluorophenyl)ethyl)morpholine (3d). Synthesized using **GP3** from **1d** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (29.1 mg, 63%).

R_f = 0.10 (30% ethyl acetate–hexanes).

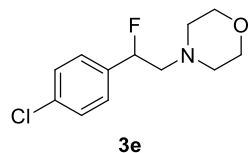
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.32 (dd, J = 8.0, 5.6 Hz, 2H), 7.08–7.03 (m, 2H), 5.62 (ddd, J = 48.5, 8.6, 2.8 Hz, 1H), 3.79–3.66 (m, 4H), 2.89 (ddd, J = 17.3, 14.2, 8.6 Hz, 1H), 2.67–2.54 (m, 5H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 162.7 (dd, $J_{\text{C-F}}$ = 247.0, 2.1 Hz), 134.6 (dd, $J_{\text{C-F}}$ = 20.6, 3.2 Hz), 127.4 (dd, $J_{\text{C-F}}$ = 8.1, 6.9 Hz), 115.4 (d, $J_{\text{C-F}}$ = 21.6 Hz), 92.1 (d, $J_{\text{C-F}}$ = 173.8 Hz), 66.9, 64.8 (d, $J_{\text{C-F}}$ = 23.4 Hz), 54.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -113.32, -175.06.

FTIR (thin film): cm^{-1} 2957, 2855, 2810, 1605, 1511, 1454, 1222, 1115, 1009, 834, 539.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{12}\text{H}_{16}\text{F}_2\text{NO}^+$ 228.1195; found: 228.1201.

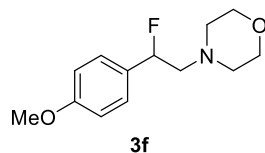


4-(2-(4-Chlorophenyl)-2-fluoroethyl)morpholine (3e). Synthesized using **GP3** from **1e** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (22.9 mg, 47%).

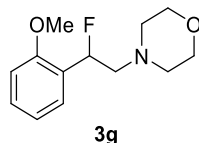
R_f = 0.12 (30% ethyl acetate–hexanes).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.20 (d, J = 8.4 Hz, 2H), 7.13 (d, J = 8.4 Hz, 2H), 5.47 (ddd, J = 48.5, 8.5,

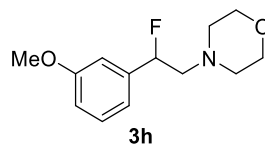
2.6 Hz, 1H), 3.56–3.61 (t, $J = 4.5$ Hz, 4H), 2.72 (ddd, $J = 17.6, 14.2, 8.5$ Hz, 1H), 2.61–2.31 (m, 5H).
 ^{13}C NMR (125 MHz, CDCl_3) δ 137.2 (d, $J_{\text{C-F}} = 20.4$ Hz), 134.3, 128.7, 126.9, 92.0 (d, $J_{\text{C-F}} = 174.4$ Hz), 66.9, 64.7 (d, $J_{\text{C-F}} = 23.3$ Hz), 54.1.
 ^{19}F NMR (471 MHz, CDCl_3) δ -177.29.
FTIR (thin film): cm^{-1} 2956, 2854, 2809, 1493, 1453, 1115, 1091, 1010, 880, 824, 719, 537.
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{12}\text{H}_{16}\text{ClFNO}^+$ 244.0899; found: 244.0906.



4-(2-Fluoro-2-(4-methoxyphenyl)ethyl)morpholine (3f). Synthesized using **GP3** from **1f** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a yellow oil (38.0 mg, 79%).
 $R_f = 0.10$ (30% ethyl acetate–hexanes).
 ^1H NMR (500 MHz, CDCl_3) δ 7.27 (d, $J = 8.4$ Hz, 2H), 6.90 (d, $J = 8.4$ Hz, 2H), 5.60 (ddd, $J = 48.7, 8.8, 2.7$ Hz, 1H), 3.81 (s, 3H), 3.72–3.76 (t, $J = 4.5$ Hz, 4H), 2.92 (ddd, $J = 16.7, 14.1, 8.8$ Hz, 1H), 2.73–2.46 (m, 5H).
 ^{13}C NMR (125 MHz, CDCl_3) δ 159.8, 130.8 (d, $J_{\text{C-F}} = 20.3$ Hz), 127.1 (d, $J_{\text{C-F}} = 6.2$ Hz), 113.9, 92.4 (d, $J = 172.3$ Hz), 66.9, 64.7 (d, $J_{\text{C-F}} = 24.0$ Hz), 55.3, 54.1.
 ^{19}F NMR (471 MHz, CDCl_3) δ -172.14.
FTIR (thin film): cm^{-1} 2956, 2853, 2810, 1613, 1515, 1248, 1116, 1033, 832.
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{19}\text{FNO}_2^+$ 240.1394; found: 240.1400.



4-(2-Fluoro-2-(2-methoxyphenyl)ethyl)morpholine (3g). Synthesized using **GP3** from **1g** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a yellow oil (26.3 mg, 55%).
 $R_f = 0.12$ (30% ethyl acetate–hexanes).
 ^1H NMR (500 MHz, CDCl_3) δ 7.41 (d, $J = 7.5$ Hz, 1H), 7.34–7.27 (m, 1H), 7.02–6.97 (m, 1H), 6.87 (d, $J = 8.0$ Hz, 1H), 6.04 (ddd, $J = 49.1, 8.8, 1.8$ Hz, 1H), 3.83 (s, 3H), 3.76 (t, $J = 4.6$ Hz, 4H), 2.80 (ddd, $J = 19.3, 14.2, 8.8$ Hz, 1H), 2.72–2.55 (m, 5H).
 ^{13}C NMR (125 MHz, CDCl_3) δ 155.3 (d, $J_{\text{C-F}} = 5.6$ Hz), 129.2 (d, $J_{\text{C-F}} = 1.4$ Hz), 127.3, 125.9 (d, $J_{\text{C-F}} = 9.6$ Hz), 120.6, 110.2, 87.9 (d, $J_{\text{C-F}} = 171.9$ Hz), 67.0 (2C), 63.8 (d, $J_{\text{C-F}} = 22.5$ Hz), 55.3, 54.0.
 ^{19}F NMR (471 MHz, CDCl_3) δ -185.00.
FTIR (thin film): cm^{-1} 2955, 2853, 1601, 1586, 1490, 1454, 1288, 1267, 1116, 1034, 869, 783, 698.
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{19}\text{FNO}_2^+$ 240.1394; found: 240.1400.



4-(2-Fluoro-2-(3-methoxyphenyl)ethyl)morpholine (3h). Synthesized using **GP3** from **1h** and *O*-

benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (28.6 mg, 60%).

R_f = 0.10 (30% ethyl acetate–hexanes).

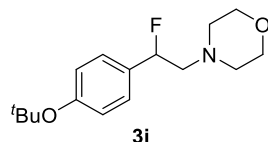
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.31–7.26 (m, 1H), 6.92–6.84 (m, 3H), 5.63 (ddd, J = 49.0, 8.8, 2.5 Hz, 1H), 3.82 (s, 3H), 3.77–3.73 (m, 4H), 2.90 (ddd, J = 17.6, 14.3, 8.8 Hz, 1H), 2.72–2.53 (m, 5H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 159.7, 140.3 (d, $J_{\text{C-F}}$ = 20.0 Hz), 129.6, 117.7, 113.9 (d, $J_{\text{C-F}}$ = 1.4 Hz), 111.1 (d, $J_{\text{C-F}}$ = 7.5 Hz), 92.5 (d, $J_{\text{C-F}}$ = 174.4 Hz), 66.9, 64.9 (d, $J_{\text{C-F}}$ = 22.9 Hz), 55.2, 54.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -177.08.

FTIR (thin film): cm^{-1} 2956, 2853, 2807, 1603, 1493, 1241, 1116, 1031, 754.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{19}\text{FNO}_2^+$ 240.1394; found: 240.1401.



4-(2-(4-(tert-Butoxy)phenyl)-2-fluoroethyl)morpholine (3i). Synthesized using **GP3** from **1i** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a yellow oil (40.0 mg, 71%).

R_f = 0.13 (30% ethyl acetate–hexanes).

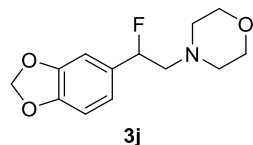
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.24 (d, J = 8.4 Hz, 2H), 6.99 (d, J = 8.4 Hz, 2H), 5.61 (ddd, J = 48.8, 8.9, 2.5 Hz, 1H), 3.77–3.70 (m, 4H), 2.92 (ddd, J = 17.0, 14.2, 8.9 Hz, 1H), 2.70–2.53 (m, 5H), 1.35 (s, 9H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 155.7 (d, $J_{\text{C-F}}$ = 2.1 Hz), 133.4 (d, $J_{\text{C-F}}$ = 20.2 Hz), 126.4 (d, $J_{\text{C-F}}$ = 6.4 Hz, 2C), 124.0, 92.5 (d, $J_{\text{C-F}}$ = 172.6 Hz), 78.7, 67.0, 64.8 (d, $J_{\text{C-F}}$ = 23.5 Hz), 54.1, 28.8.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -173.54.

FTIR (thin film): cm^{-1} 2974, 2854, 2808, 1719, 1508, 1365, 1236, 1160, 1116, 895, 853, 549.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{25}\text{FNO}_2^+$ 282.1864; found: 282.1869.



4-(2-(Benzo[*d*][1,3]dioxol-5-yl)-2-fluoroethyl)morpholine (3j). Synthesized using **GP3** from **1j** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a pale-yellow oil (35.8 mg, 71%).

R_f = 0.14 (30% ethyl acetate–hexanes).

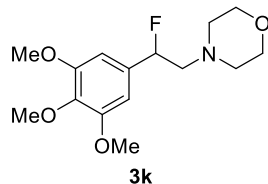
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 6.78 (s, 1H), 6.73 (d, J = 0.7 Hz, 2H), 5.90 (s, 2H), 5.48 (ddd, J = 48.5, 8.7, 2.6 Hz, 1H), 3.72–3.56 (m, 4H), 2.89–2.72 (m, 1H), 2.61–2.58 (m, 1H), 2.58–2.42 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 147.9, 132.6 (d, $J_{\text{C-F}}$ = 20.4 Hz), 119.5 (d, $J_{\text{C-F}}$ = 7.3 Hz), 108.2, 106.3 (d, $J_{\text{C-F}}$ = 6.5 Hz), 101.2, 92.5 (d, $J_{\text{C-F}}$ = 173.7 Hz), 66.9, 64.8 (d, $J_{\text{C-F}}$ = 24.0 Hz), 54.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -171.98.

FTIR (thin film): cm^{-1} 2894, 2855, 2812, 1717, 1490, 1444, 1247, 1115, 1036, 931, 870, 716.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{17}\text{FNO}_3^+$ 254.1187; found: 254.1189.



4-(2-Fluoro-2-(3,4,5-trimethoxyphenyl)ethyl)morpholine (3k). Synthesized using **GP3** from **1k** and *O*-benzoylhydroxymorpholine **2a**. IPrCuCl (5 mol %) was used instead of the Cu(HFacac)₂. Isolated by flash column chromatography (25% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a pale-yellow oil (27.0 mg, 45%).

R_f = 0.20 (50% ethyl acetate–hexanes).

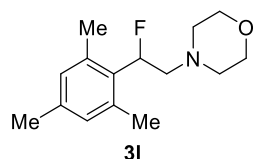
¹H NMR (500 MHz, CDCl₃) δ 6.56 (s, 2H), 5.59 (ddd, J = 48.7, 8.6, 2.2 Hz, 1H), 3.87 (s, 6H), 3.84 (s, 3H), 3.78–3.72 (m, 4H), 2.98–2.81 (m, 1H), 2.73–2.65 (m, 1H), 2.65–2.50 (m, 4H).

¹³C NMR (125 MHz, CDCl₃) δ 153.4, 138.0, 134.4 (d, J_{C-F} = 20.4 Hz), 102.6 (d, J_{C-F} = 7.2 Hz), 92.9 (d, J_{C-F} = 174.5 Hz), 66.9, 64.9 (d, J_{C-F} = 23.2 Hz), 60.9, 56.2, 54.1.

¹⁹F NMR (471 MHz, CDCl₃) δ -174.42.

FTIR (thin film): cm⁻¹ 2939, 2841, 1591, 1454, 1236, 1117, 1007, 870, 705.

HRMS (ESI) m/z : [M+H]⁺ Calcd for C₁₅H₂₃FNO₄⁺ 300.1606; found: 300.1600.



4-(2-Fluoro-2-mesitylethyl)morpholine (3l). Synthesized using **GP3** from **1l** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale oil (25.6 mg, 51%).

R_f = 0.14 (30% ethyl acetate–hexanes).

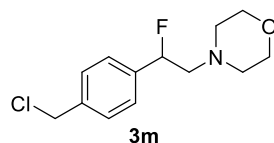
¹H NMR (500 MHz, CDCl₃) δ 6.83 (s, 2H), 6.03 (ddd, J = 48.4, 9.3, 2.7 Hz, 1H), 3.78–3.74 (m, 4H), 3.15 (td, J = 14.4, 9.3 Hz, 1H), 2.67–2.57 (m, 4H), 2.50 (ddd, J = 33.1, 14.4, 2.7 Hz, 1H), 2.38–2.34 (m, 6H), 2.26 (s, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 137.7, 135.7 (d, J_{C-F} = 3.6 Hz), 131.4 (d, J_{C-F} = 17.8 Hz), 129.9, 90.7 (d, J_{C-F} = 172.9 Hz), 66.9, 62.2 (d, J_{C-F} = 22.7 Hz), 54.1, 20.8, 20.4, 20.4.

¹⁹F NMR (471 MHz, CDCl₃) δ -181.57.

FTIR (thin film): cm⁻¹ 2959, 2854, 2809, 1678, 1612, 1452, 1142, 1116, 1050, 1007, 871, 852, 731, 572.

HRMS (ESI) m/z : [M+H]⁺ Calcd for C₁₅H₂₃FNO⁺ 252.1758; found: 252.1765.



4-(2-(4-(Chloromethyl)phenyl)-2-fluoroethyl)morpholine (3m). Synthesized using **GP3** from **1m** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a yellow oil (26.8 mg, 52%).

R_f = 0.16 (30% ethyl acetate–hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 5.67 (ddd, J = 48.7, 8.7, 2.6 Hz, 1H), 4.58 (s, 2H), 3.76–3.74 (m, 4H), 2.89 (ddd, J = 17.4, 14.2, 8.7 Hz, 1H), 2.76–2.54 (m, 5H).

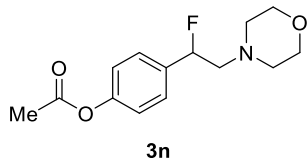
¹³C NMR (125 MHz, CDCl₃) δ 138.9 (d, J_{C-F} = 20.1 Hz), 137.7, 128.7, 125.9 (d, J_{C-F} = 7.0 Hz), 92.2 (d,

$J_{C-F} = 174.3$ Hz), 66.8, 64.8 (d, $J_{C-F} = 23.0$ Hz), 54.0, 45.7.

^{19}F NMR (471 MHz, CDCl_3) δ -177.64.

FTIR (thin film): cm^{-1} 2984, 2843, 2801, 1493, 1350, 1123, 1056, 968, 867, 813, 727.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{18}\text{ClFNO}^+$ 258.1056; found: 258.1061.



4-(1-Fluoro-2-morpholinoethyl)phenyl acetate (3n). Synthesized using **GP3** from **1n** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (30% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a pale oil (27.0 mg, 50%).

$R_f = 0.11$ (30% ethyl acetate–hexanes).

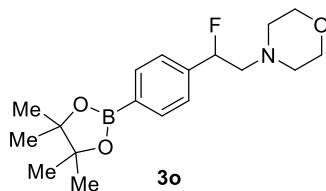
^1H NMR (500 MHz, CDCl_3) δ 7.36 (d, $J = 8.4$ Hz, 2H), 7.10 (d, $J = 8.4$ Hz, 2H), 5.65 (ddd, $J = 48.6, 8.7, 2.3$ Hz, 1H), 3.76–3.74 (m, 4H), 2.89 (ddd, $J = 17.4, 14.2, 8.7$ Hz, 1H), 2.72–2.53 (m, 5H), 2.30 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 169.4, 150.6, 136.3 (d, $J_{C-F} = 20.4$ Hz), 126.7, 121.7, 92.2 (d, $J_{C-F} = 174.2$ Hz), 66.9, 64.8 (d, $J_{C-F} = 23.1$ Hz), 54.1, 21.1.

^{19}F NMR (471 MHz, CDCl_3) δ -176.30.

FTIR (thin film): cm^{-1} 2929, 2854, 1755, 1509, 1369, 1191, 1116, 1009, 911, 882, 869.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{19}\text{FNO}_3^+$ 268.1344; found: 268.1350.



4-(2-Fluoro-2-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)ethyl)morpholine (3o). Synthesized using **GP3** from **1o** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (40.2 mg, 60%).

$R_f = 0.20$ (33% ethyl acetate–hexanes).

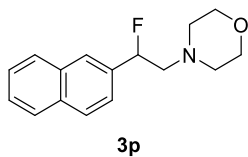
^1H NMR (500 MHz, CDCl_3) δ 7.82 (d, $J = 7.8$ Hz, 2H), 7.34 (d, $J = 7.8$ Hz, 2H), 5.67 (ddd, $J = 48.9, 8.6, 2.2$ Hz, 1H), 3.82–3.61 (m, 4H), 3.00–2.83 (m, 1H), 2.72–2.64 (m, 1H), 2.64–2.45 (m, 4H), 1.34 (s, 12H).

^{13}C NMR (125 MHz, CDCl_3) δ 141.7 (d, $J_{C-F} = 20.1$ Hz), 134.9, 128.3 (d, $J_{C-F} = 11.6$ Hz), 124.7 (d, $J_{C-F} = 7.1$ Hz), 92.5 (d, $J_{C-F} = 174.5$ Hz), 83.9, 66.9, 64.8 (d, $J_{C-F} = 22.9$ Hz), 54.0, 24.8.

^{19}F NMR (471 MHz, CDCl_3) δ -178.83.

FTIR (thin film): cm^{-1} 2976, 2854, 2808, 1614, 1357, 1142, 1009, 857, 658.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{18}\text{H}_{28}^{[11]\text{B}}\text{FNO}_3^+$ 336.2141; found: 336.2140.



4-(2-Fluoro-2-(naphthalen-2-yl)ethyl)morpholine (3p). Synthesized using **GP3** from **1p** and *O*-benzoylhydroxymorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to

30% ethyl acetate–hexanes), as a pale-yellow oil (32.2 mg, 62%).

R_f = 0.30 (50% ethyl acetate–hexanes).

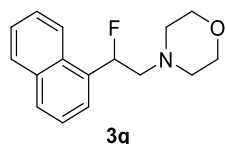
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.86 (dd, J = 9.3, 5.1 Hz, 3H), 7.82 (s, 1H), 7.56–7.47 (m, 2H), 7.45 (d, J = 8.6 Hz, 1H), 5.83 (ddd, J = 48.8, 8.6, 2.4 Hz, 1H), 3.84–3.66 (m, 4H), 3.14–2.88 (m, 1H), 2.81–2.67 (m, 1H), 2.67–2.55 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 136.1 (d, $J_{\text{C-F}}$ = 19.6 Hz), 133.3, 133.1, 128.4, 128.1, 127.8, 126.5, 126.4, 124.8 (d, $J_{\text{C-F}}$ = 8.1 Hz), 123.2 (d, $J_{\text{C-F}}$ = 5.7 Hz), 92.8 (d, $J_{\text{C-F}}$ = 173.9 Hz), 66.9, 64.9 (d, $J_{\text{C-F}}$ = 23.1 Hz), 54.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -176.57.

FTIR (thin film): cm^{-1} 2937, 2865, 2816, 1717, 1274, 1110, 1011, 872, 824, 741.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{19}\text{FNO}^+$ 260.1445; found: 260.1439.



4-(2-Fluoro-2-(naphthalen-1-yl)ethyl)morpholine (3q). Synthesized using **GP3** from **1q** and *O*-benzoylhydroxylmorpholine **2a**. IPrCuCl (5 mol %) was used instead of the $\text{Cu}(\text{HFacac})_2$. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 40% ethyl acetate–hexanes), as a pale oil (34.4 mg, 66%).

R_f = 0.30 (50% ethyl acetate–hexanes).

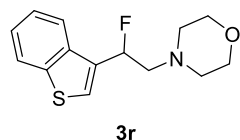
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.94 (d, J = 8.2 Hz, 1H), 7.89 (dd, J = 8.0, 1.6 Hz, 1H), 7.84 (d, J = 8.2 Hz, 1H), 7.62 (d, J = 8.0 Hz, 1H), 7.59–7.47 (m, 3H), 6.42 (ddd, J = 48.4, 8.7, 2.1 Hz, 1H), 3.82–3.78 (t, J = 4.5 Hz, 4H), 3.02 (ddd, J = 19.0, 14.6, 8.7 Hz, 1H), 2.86 (ddd, J = 36.0, 14.6, 2.1 Hz, 1H), 2.78–2.58 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 134.4 (d, $J_{\text{C-F}}$ = 18.7 Hz), 133.6, 129.6 (d, $J_{\text{C-F}}$ = 4.1 Hz), 129.0, 128.8 (d, $J_{\text{C-F}}$ = 1.3 Hz), 126.5, 125.8, 125.3, 123.2 (d, $J_{\text{C-F}}$ = 11.2 Hz), 122.6, 91.0 (d, $J_{\text{C-F}}$ = 174.2 Hz), 67.0, 64.6 (d, J = 22.7 Hz), 54.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -180.07.

FTIR (thin film): cm^{-1} 3050, 2955, 2853, 2809, 1675, 1452, 1137, 1009, 798, 775.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{19}\text{FNO}^+$ 260.1445; found: 260.1449.



4-(2-(Benzo[*b*]thiophen-3-yl)-2-fluoroethyl)morpholine (3r). Synthesized using **GP3** from **1r** and *O*-benzoylhydroxylmorpholine **2a**. IPrCuCl (5 mol %) was used instead of the $\text{Cu}(\text{HFacac})_2$. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (23.9 mg, 45%).

R_f = 0.25 (50% ethyl acetate–hexanes).

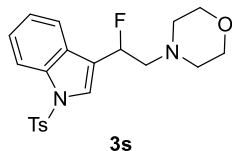
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.88 (d, J = 7.4 Hz, 1H), 7.83 (d, J = 7.4 Hz, 1H), 7.50 (s, 1H), 7.45–7.33 (m, 2H), 6.04 (ddd, J = 48.1, 8.2, 1.7 Hz, 1H), 3.86–3.70 (m, 4H), 3.20–3.00 (m, 1H), 2.98–2.80 (m, 1H), 2.72–2.56 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 140.6, 136.7, 133.6 (d, $J_{\text{C-F}}$ = 21.2 Hz), 124.7, 124.4, 123.9 (d, $J_{\text{C-F}}$ = 8.5 Hz), 123.0, 121.9, 89.0 (d, $J_{\text{C-F}}$ = 172.3 Hz), 66.9, 63.2 (d, $J_{\text{C-F}}$ = 22.9 Hz), 54.2.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -175.77.

FTIR (thin film): cm^{-1} 2923, 2854, 1597, 1447, 1371, 1175, 1118, 977, 748, 576.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{14}H_{17}FNOS^+$ 266.1009; found: 266.1010.



4-(2-Fluoro-2-(1-tosyl-1H-indol-3-yl)ethyl)morpholine (3s). Synthesized using **GP3** from **1s** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 40% ethyl acetate–hexanes), as a pale-yellow oil (26.2 mg, 33%).

R_f = 0.20 (50% ethyl acetate–hexanes).

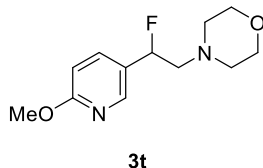
1H NMR (500 MHz, $CDCl_3$) δ 7.98 (d, J = 8.3 Hz, 1H), 7.78 (d, J = 8.3 Hz, 2H), 7.63 (d, J = 2.4 Hz, 1H), 7.60 (d, J = 7.6 Hz, 1H), 7.34 (t, J = 7.6 Hz, 1H), 7.27–7.21 (m, 3H), 5.91 (dd, J = 48.4, 6.6 Hz, 1H), 5.91 (dd, J = 48.4, 6.6 Hz, 1H), 3.75 (t, J = 4.5 Hz, 5H), 3.09 (ddd, J = 17.9, 14.2, 8.2 Hz, 1H), 2.86 (dd, J = 30.2, 14.2 Hz, 1H), 2.74–2.49 (m, 4H), 2.35 (s, 3H).

^{13}C NMR (125 MHz, $CDCl_3$) δ 145.3, 135.2, 135.1, 130.0, 128.4, 128.3, 126.9, 125.2, 123.7 (d, J_{C-F} = 9.2 Hz), 123.5, 120.1, 113.8, 87.2 (d, J_{C-F} = 169.6 Hz), 66.9, 62.8 (d, J_{C-F} = 22.9 Hz), 54.1, 21.6.

^{19}F NMR (471 MHz, $CDCl_3$) δ -175.43.

FTIR (thin film): cm^{-1} 2924, 2853, 2811, 1673, 1138, 1428, 1115, 1008, 866, 760, 733.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{21}H_{24}FN_2O_3S^+$ 403.1486; found: 403.1489.



4-(2-Fluoro-2-(6-methoxypyridin-3-yl)ethyl)morpholine (3t). Synthesized using **GP3** from **1t** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (31.7 mg, 66%).

R_f = 0.14 (30% ethyl acetate–hexanes).

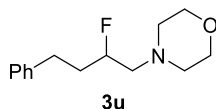
1H NMR (500 MHz, $CDCl_3$) δ 8.14 (s, 1H), 7.59 (dd, J = 8.6, 2.1 Hz, 1H), 6.76 (d, J = 8.6 Hz, 1H), 5.61 (ddd, J = 48.0, 8.3, 3.0 Hz, 1H), 3.94 (s, 3H), 3.73 (t, J = 4.7 Hz, 4H), 2.93 (ddd, J = 17.1, 14.1, 8.3 Hz, 1H), 2.74–2.46 (m, 1H), 2.74–2.46 (m, 4H).

^{13}C NMR (125 MHz, $CDCl_3$) δ 164.5 (d, J_{C-F} = 1.4 Hz), 144.9 (d, J_{C-F} = 7.3 Hz), 136.4 (d, J_{C-F} = 5.0 Hz), 127.0 (d, J_{C-F} = 20.8 Hz), 111.0, 90.6 (d, J_{C-F} = 172.7 Hz), 66.9, 64.1 (d, J_{C-F} = 24.0 Hz), 54.1, 53.6.

^{19}F NMR (471 MHz, $CDCl_3$) δ -174.28.

FTIR (thin film): cm^{-1} 2948, 2853, 1609, 1495, 1392, 1286, 1116, 1022, 869, 832.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{12}H_{18}FN_2O_2^+$ 241.1347; found: 241.1342.



4-(2-Fluoro-4-phenylbutyl)morpholine (3u). Synthesized using **GP3** from **1u** and *O*-benzoylhydroxylmorpholine **2a**. $IPrCuCl$ (5 mol %) was used instead of the $Cu(HFacac)_2$. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a pale oil (5.5 mg, 12%).

R_f = 0.10 (50% ethyl acetate–hexanes).

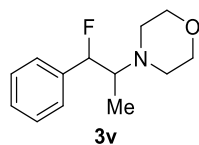
¹H NMR (500 MHz, CDCl₃) δ 7.32–7.27 (m, 2H), 7.23–7.18 (m, 3H), 4.81–4.52 (m, 1H), 3.75–3.58 (m, 4H), 2.88–2.78 (m, 1H), 2.75–2.68 (m, 1H), 2.68–2.58 (m, 1H), 2.55–2.41 (m, 5H), 2.07–1.95 (m, 1H), 1.95–1.78 (m, 1H).

¹³C NMR (125 MHz, CDCl₃) δ 141.2, 128.5, 128.8, 126.1, 91.5 (d, J_{C-F} = 170.0 Hz), 66.9, 62.8 (d, J_{C-F} = 20.5 Hz), 54.3, 35.2 (d, J_{C-F} = 20.8 Hz), 31.2 (d, J_{C-F} = 4.6 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -182.13.

FTIR (thin film): cm⁻¹ 2952, 2853, 1717, 1453, 1273, 1116, 1035, 867, 699.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₄H₂₁FNO⁺ 238.1602; found: 238.1600.



4-(1-Fluoro-1-phenylpropan-2-yl)morpholine (3v). Synthesized using **GP3** from (*E*)-**1v** and *O*-benzoylhydroxylmorpholine **2a**. ¹H-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 2.5:1. Isolated as a mixture of two diastereomers by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (32.2 mg, 60%).

Synthesized using **GP3** from (*Z*)-**1v** and *O*-benzoylhydroxylmorpholine **2a**. ¹H-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 1.8:1. Isolated as a mixture of two diastereomers by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale-yellow oil (25.0 mg, 56%).

R_f = 0.20 (33% ethyl acetate–hexanes).

Major isomer (3v-I): ¹H NMR (500 MHz, CDCl₃) δ 7.45–7.29 (m, 5H), 5.36 (dd, J = 47.9, 8.3 Hz, 1H), 3.72 (t, J = 4.0 Hz, 4H), 3.13–2.92 (m, 1H), 2.80–2.58 (m, 4H), 0.84 (d, J = 6.9 Hz, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 138.5 (d, J = 20.1 Hz), 128.5 (d, J = 2.0 Hz), 128.3, 126.7 (d, J = 6.4 Hz), 95.8 (d, J = 176.5 Hz), 67.6, 63.5 (d, J = 21.8 Hz), 49.6, 11.2 (d, J = 6.2 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -175.16.

FTIR (thin film): cm⁻¹ 2955, 2851, 1720, 1451, 1267, 1114, 999, 751, 699.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₃H₁₉FNO⁺ 224.1445; found: 224.1447.

The relative stereochemistry of major isomer **3v-I** has been determined by X-ray analysis. More information of the X-ray analysis is provided on S283.

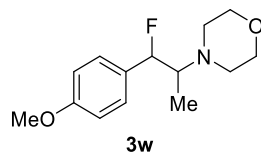
Minor isomer (3v-II): ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.34 (m, 2H), 7.30 (3, J = 6.6 Hz, 3H), 5.72 (dd, J = 48.1, 2.2 Hz, 1H), 3.71 (t, J = 4.5 Hz, 4H), 2.83 (dq, J = 13.4, 6.8, 2.2 Hz, 1H), 2.73–2.57 (m, 4H), 1.04 (dd, J = 6.8, 1.1 Hz, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 139.5 (d, J = 20.7 Hz), 128.2, 127.7, 125.1 (d, J = 8.5 Hz), 94.7 (d, J = 178.7 Hz), 67.5, 64.9 (d, J = 22.0 Hz), 49.9, 7.6 (d, J = 6.7 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -193.68.

FTIR (thin film): cm⁻¹ 2924, 2853, 1669, 1450, 1254, 1199, 1117, 955, 984, 720, 699.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₃H₁₉FNO⁺ 224.1445; found: 224.1448.



4-(1-Fluoro-1-(4-methoxyphenyl)propan-2-yl)morpholine (3w). Synthesized using **GP3** from **1w** and *O*-benzoylhydroxylmorpholine **2a**. ¹H-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 1.8:1. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 15% ethyl acetate–hexanes), as a pale-yellow oil (26.8 mg, 53%).

Minor isomer (3w-I): *R_f* = 0.40 (33% ethyl acetate–hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.26 (d, *J* = 8.5 Hz, 2H), 6.90 (d, *J* = 8.5 Hz, 2H), 5.30 (dd, *J* = 47.3, 8.1 Hz, 1H), 3.82 (s, 3H), 3.73 (t, *J* = 4.4 Hz, 3H), 3.09–2.98 (m, 1H), 2.78–2.64 (m, 4H), 0.82 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 159.8 (d, *J_{C-F}* = 1.8 Hz), 130.6 (d, *J_{C-F}* = 20.7 Hz), 128.1 (d, *J_{C-F}* = 5.7 Hz), 113.8, 95.4 (d, *J_{C-F}* = 175.9 Hz), 67.5, 63.5 (d, *J_{C-F}* = 22.1 Hz), 55.3, 49.6, 11.3 (d, *J_{C-F}* = 5.8 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -170.96.

FTIR (thin film): cm⁻¹ 2933, 2851, 1613, 1514, 1249, 1115, 1033, 996, 831.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₄H₂₁FNO₂⁺ 254.1551; found: 254.1545.

Major isomer (3w-II): *R_f* = 0.35 (33% ethyl acetate–hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.22 (d, *J* = 8.5 Hz, 2H), 6.89 (d, *J* = 8.5 Hz, 2H), 5.64 (d, *J* = 47.3 Hz, 1H), 3.81 (s, 3H), 3.70 (t, *J* = 4.5 Hz, 4H), 2.88–2.74 (m, 1H), 2.74–2.54 (m, 4H), 1.06 (d, *J* = 6.6 Hz, 3H).

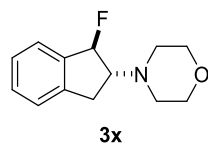
¹³C NMR (125 MHz, CDCl₃) δ 159.2, 126.6 (d, *J_{C-F}* = 8.2 Hz), 113.6, 94.8 (d, *J_{C-F}* = 177.1 Hz), 67.4, 64.8 (d, *J_{C-F}* = 22.9 Hz), 55.3, 49.9, 7.8.

¹⁹F NMR (471 MHz, CDCl₃) δ -189.81.

FTIR (thin film): cm⁻¹ 2924, 2852, 1613, 1514, 1453, 1247, 1115, 1033, 981, 828.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₄H₂₁FNO₂⁺ 254.1551; found: 254.1548.

The assignments of relative stereochemistry of two diastereomers **3w-I** and **3w-II** are based on the ¹H, ¹³C, and ¹⁹F-NMR analysis in comparison to **3v-I** and **3v-II**.



4-(1-Fluoro-2,3-dihydro-1H-inden-2-yl)morpholine (3x). Synthesized using **GP3** from **1x** and *O*-benzoylhydroxylmorpholine **2a**. ¹H-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 10:1. Isolated as a mixture of two diastereomers by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale-yellow oil (26.5 mg, 60%).

R_f = 0.20 (33% ethyl acetate–hexanes).

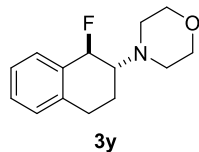
¹H NMR (500 MHz, CDCl₃) δ 7.41 (d, *J* = 7.1 Hz, 1H), 7.34–7.25 (m, 2H), 7.22 (d, *J* = 7.2 Hz, 1H), 6.00 (dd, *J* = 57.7, 5.5 Hz, 1H), 3.79 (t, *J* = 4.7 Hz, 4H), 3.35–3.09 (m, 2H), 2.90–2.76 (m, 1H), 2.72 (s, 2H), 2.65–2.56 (m, 2H).

¹³C NMR (125 MHz, CDCl₃) δ 139.5 (d, *J_{C-F}* = 5.3 Hz), 139.0 (d, *J_{C-F}* = 20.5 Hz), 129.4 (d, *J_{C-F}* = 2.3 Hz), 127.3 (d, *J_{C-F}* = 1.8 Hz), 124.8, 124.6, 99.0 (d, *J_{C-F}* = 179.9 Hz), 73.3 (d, *J_{C-F}* = 17.4 Hz), 66.9, 52.4, 34.2 (d, *J_{C-F}* = 5.6 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -170.49 (major), -180.50 (minor).

FTIR (thin film): cm⁻¹ 2949, 2850, 1483, 1356, 1146, 1115, 1001, 896, 751, 664.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₃H₁₇FNO⁺ 222.1289; found: 222.1295.



4-(1-Fluoro-1,2,3,4-tetrahydronaphthalen-2-yl)morpholine (3y). Synthesized using **GP3** from **1y** and morpholino dodecanoate **2t**. Only one diastereomer (dr >20:1) was observed by ¹H-NMR analysis of the crude reaction mixture. Isolated by flash column chromatography (5% ethyl acetate–hexanes to 15% ethyl acetate–hexanes), as a pale-yellow oil (24.5 mg, 52%).

R_f = 0.30 (20% ethyl acetate–hexanes).

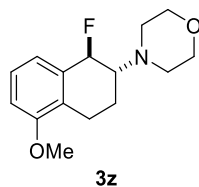
¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, *J* = 4.0 Hz, 1H), 7.25 (dd, *J* = 5.7, 3.5 Hz, 2H), 7.12 (d, *J* = 4.0 Hz, 1H), 5.71 (dd, *J* = 52.7, 7.9 Hz, 1H), 3.76 (t, *J* = 4.3 Hz, 4H), 3.04–2.92 (m, 1H), 2.90–2.84 (m, 2H), 2.83–2.71 (m, 4H), 2.12 (ddt, *J* = 12.2, 8.0, 4.2 Hz, 1H), 1.76 (ddd, *J* = 24.5, 12.2, 8.0 Hz, 1H).

¹³C NMR (125 MHz, CDCl₃) δ 137.0 (d, *J*_{C-F} = 3.6 Hz), 134.4 (d, *J*_{C-F} = 17.7 Hz), 128.4, 128.4 (d, *J*_{C-F} = 5.7 Hz), 128.2, 126.5, 89.8 (d, *J*_{C-F} = 173.4 Hz), 67.3, 65.1 (d, *J*_{C-F} = 17.4 Hz), 50.1, 28.5, 23.4 (d, *J*_{C-F} = 6.7 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -167.52.

FTIR (thin film): cm⁻¹ 2923, 2852, 1734, 1454, 1251, 1117, 1012, 746.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₄H₁₉FNO⁺ 236.1445; found: 236.1442.



4-(1-Fluoro-5-methoxy-1,2,3,4-tetrahydronaphthalen-2-yl)morpholine (3z). Synthesized using **GP3** from **1z** and *O*-benzoylhydroxymorpholine **2a**. Only one diastereomer (dr >20:1) was observed by ¹H-NMR analysis of the crude reaction mixture. Isolated by flash column chromatography (5% ethyl acetate–hexanes to 12% ethyl acetate–hexanes), as a pale-yellow oil (28.6 mg, 54%).

R_f = 0.20 (20% ethyl acetate–hexanes).

¹H NMR (500 MHz, CDCl₃) δ 7.23 (t, *J* = 8.0 Hz, 1H), 7.10 (d, *J* = 7.8 Hz, 1H), 6.78 (d, *J* = 8.1 Hz, 1H), 5.68 (dd, *J* = 52.5, 8.0 Hz, 1H), 3.82 (s, 3H), 3.75 (t, *J* = 4.6 Hz, 4H), 2.97–2.83 (m, 2H), 2.81–2.68 (m, 4H), 2.64–2.51 (m, 1H), 2.18–2.05 (m, 1H), 1.77–1.63 (m, 1H).

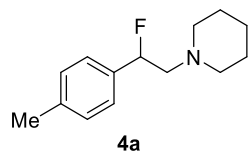
¹³C NMR (125 MHz, CDCl₃) δ 156.5, 136.0 (d, *J*_{C-F} = 18.3 Hz), 127.0, 126.0 (d, *J*_{C-F} = 3.7 Hz), 120.0 (d, *J*_{C-F} = 8.0 Hz), 109.3, 89.7 (d, *J*_{C-F} = 173.8 Hz), 67.5, 64.3 (d, *J*_{C-F} = 17.1 Hz), 55.4, 50.1, 22.6 (d, *J*_{C-F} = 1.4 Hz), 22.5 (d, *J*_{C-F} = 2.3 Hz).

¹⁹F NMR (471 MHz, CDCl₃) δ -168.61.

FTIR (thin film): cm⁻¹ 2951, 2855, 1587, 1473, 1263, 1111, 1069, 941, 790, 746.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₅H₂₁FNO₂⁺ 266.1551; found: 266.1550.

5. Scope of amines and compound characterization data



1-(2-Fluoro-2-(*p*-tolyl)ethyl)piperidine (4a). Synthesized using **GP3** from **1a** and *O*-benzoylhydroxylpiperidine **2b**. Isolated by flash column chromatography (5% ethyl acetate–hexanes to 10% ethyl acetate–hexanes), as a pale oil (21.0 mg, 47%).

R_f = 0.30 (20% ethyl acetate–hexanes).

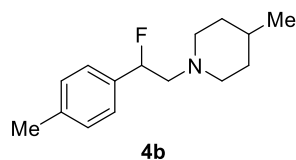
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.23 (d, J = 8.0 Hz, 2H), 7.17 (d, J = 8.0 Hz, 2H), 5.64 (ddd, J = 49.0, 8.9, 2.0 Hz, 1H), 3.01–2.79 (m, 1H), 2.70–2.56 (m, 1H), 2.56–2.45 (m, 4H), 2.35 (s, 3H), 1.70–1.52 (m, 4H), 1.51–1.37 (m, 2H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 138.1 (d, $J_{\text{C-F}}$ = 1.6 Hz), 136.3 (d, $J_{\text{C-F}}$ = 20.2 Hz), 129.1, 125.6 (d, $J_{\text{C-F}}$ = 6.6 Hz), 92.6 (d, $J_{\text{C-F}}$ = 172.8 Hz), 65.5 (d, $J_{\text{C-F}}$ = 23.4 Hz), 55.0, 26.0, 24.2, 21.2.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -174.77.

FTIR (thin film): cm^{-1} 2932, 2853, 1516, 1452, 1300, 1121, 1017, 880, 814, 759.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{21}\text{FN}^+$ 222.1653; found: 222.1657.



1-(2-Fluoro-2-(*p*-tolyl)ethyl)-4-methylpiperidine (4b). Synthesized using **GP3** from **1a** and *O*-benzoyl-4-methyl-hydroxylpiperidine **2c**. Isolated by flash column chromatography (5% ethyl acetate–hexanes to 10% ethyl acetate–hexanes), as a pale oil (15.3 mg, 32%).

R_f = 0.20 (10% ethyl acetate–hexanes).

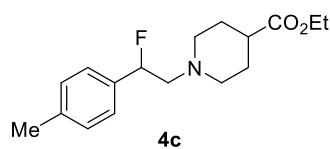
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.23 (d, J = 8.0 Hz, 2H), 7.17 (d, J = 8.0 Hz, 2H), 5.63 (ddd, J = 49.0, 8.9, 2.2 Hz, 1H), 3.08–3.01 (m, 1H), 3.00–2.95 (m, 1H), 2.90 (ddd, J = 17.3, 14.3, 9.0 Hz, 1H), 2.59 (ddd, J = 35.3, 14.3, 2.4 Hz, 1H), 2.35 (s, 3H), 2.19–2.06 (m, 2H), 1.72–1.58 (m, 2H), 1.45–1.23 (m, 3H), 0.93 (d, J = 6.1 Hz, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 138.1 (d, $J_{\text{C-F}}$ = 1.4 Hz), 136.3 (d, $J_{\text{C-F}}$ = 20.0 Hz), 129.1, 125.6 (d, $J_{\text{C-F}}$ = 6.6 Hz), 92.7 (d, $J_{\text{C-F}}$ = 172.8 Hz), 65.1 (d, $J_{\text{C-F}}$ = 23.3 Hz), 54.6, 54.3, 34.3, 30.6, 21.9, 21.2.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -174.83.

FTIR (thin film): cm^{-1} 2947, 2922, 2869, 1721, 1454, 1269, 1125, 1041, 978, 811.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{23}\text{FN}^+$ 236.1809; found: 236.1807.



Ethyl 1-(2-fluoro-2-(*p*-tolyl)ethyl)piperidine-4-carboxylate (4c). Synthesized using **GP3** from **1a** and *O*-benzoyl-4-ethoxycarbonylhydroxylpiperidine **2d**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (35.2 mg, 60%).

R_f = 0.27 (50% ethyl acetate–hexanes).

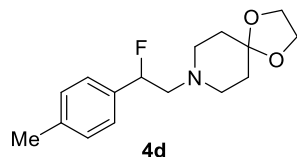
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.22 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 5.61 (ddd, J = 48.8, 8.7, 2.5 Hz, 1H), 4.14 (q, J = 7.1 Hz, 2H), 3.03 (d, J = 11.4 Hz, 1H), 2.95 (d, J = 11.4 Hz, 1H), 2.90 (ddd, J = 17.2, 14.3, 8.7 Hz, 1H), 2.61 (ddd, J = 34.0, 14.3, 2.5 Hz, 1H), 2.35 (s, 3H), 2.29–2.17 (m, 3H), 1.95–1.88 (m, 2H), 1.87–1.75 (m, 2H), 1.25 (t, J = 7.1 Hz, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 175.1, 138.2 (d, $J_{\text{C-F}}$ = 1.9 Hz), 136.0 (d, $J_{\text{C-F}}$ = 20.1 Hz), 129.1, 125.5 (d, $J_{\text{C-F}}$ = 6.7 Hz), 92.7 (d, $J_{\text{C-F}}$ = 172.9 Hz), 64.7 (d, $J_{\text{C-F}}$ = 23.6 Hz), 60.3, 53.4, 53.4, 40.9, 28.3, 21.2, 14.2.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -174.99.

FTIR (thin film): cm^{-1} 2944, 2803, 1729, 1515, 1448, 1284, 1181, 1047, 816, 527.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{17}H_{25}FNO_2^+$ 294.1864; found: 294.1869.



8-(2-Fluoro-2-(*p*-tolyl)ethyl)-1,4-dioxo-8-azaspiro[4.5]decane (4d). Synthesized using **GP3** from **1a** and 1,4-dioxo-8-azaspiro[4.5]decan-8-yl benzoate **2e**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 33% ethyl acetate–hexanes), as a colorless oil (27.7 mg, 50%).

R_f = 0.20 (33% ethyl acetate–hexanes).

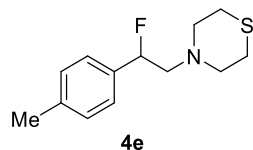
1H NMR ($CDCl_3$, 500 MHz): δ 7.23 (d, J = 8.0 Hz, 2H), 7.18 (d, J = 8.0 Hz, 2H), 5.61 (ddd, J = 49.6, 8.9, 2.3 Hz, 1H), 3.96 (s, 4H), 2.95 (ddd, J = 17.2, 14.3, 8.9 Hz, 1H), 2.70 (br s, 4H), 2.66–2.57 (m, 1H), 2.35 (s, 3H), 1.84–1.75 (m, 4H).

^{13}C NMR ($CDCl_3$, 125 MHz): δ 138.2 (d, J_{C-F} = 1.6 Hz), 136.0 (d, J_{C-F} = 19.9 Hz), 129.1, 125.5 (d, J_{C-F} = 6.5 Hz), 107.0, 92.7 (d, J_{C-F} = 172.9 Hz), 64.2, 64.2 (d, J_{C-F} = 23.6 Hz), 51.8, 34.8, 21.2.

^{19}F NMR ($CDCl_3$, 471 MHz): δ -175.35

FTIR (thin film): cm^{-1} 1143, 1094, 1039, 964, 946, 915, 818.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{16}H_{23}FNO_2^+$ 280.1707; found: 280.1714.



4-(2-Fluoro-2-(*p*-tolyl)ethyl)thiomorpholine (4e). Synthesized using **GP3** from **1a** and *O*-benzoylhydroxylthiomorpholine **2f**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 15% ethyl acetate–hexanes), as a pale oil (33.1 mg, 69%).

R_f = 0.10 (10% ethyl acetate–hexanes).

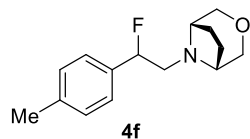
1H NMR (500 MHz, $CDCl_3$) δ 7.22 (d, J = 8.1 Hz, 2H), 7.18 (d, J = 8.1 Hz, 2H), 5.60 (ddd, J = 48.7, 8.5, 2.3 Hz, 1H), 3.02–2.89 (m, 1H), 2.89–2.83 (m, 4H), 2.75–2.69 (m, 4H), 2.69–2.58 (m, 1H), 2.36 (s, 3H).

^{13}C NMR (125 MHz, $CDCl_3$) δ 138.3 (d, J_{C-F} = 1.7 Hz), 135.8 (d, J_{C-F} = 20.1 Hz), 129.2, 125.6 (d, J_{C-F} = 6.5 Hz), 92.5 (d, J_{C-F} = 173.2 Hz), 65.2 (d, J_{C-F} = 23.8 Hz), 55.4, 28.0, 21.2.

^{19}F NMR (471 MHz, $CDCl_3$) δ -175.19.

FTIR (thin film): cm^{-1} 2920, 2808, 1719, 1516, 1454, 1273, 1131, 1040, 957, 815, 755.

HRMS (ESI) m/z : $[M+H]^+$ Calcd for $C_{13}H_{19}FNS^+$ 240.1217; found: 240.1220.



8-(2-Fluoro-2-phenylethyl)-3-oxa-8-azabicyclo[3.2.1]octane (4f). Synthesized using **GP3** from **1a** and 3-oxa-8-azabicyclo[3.2.1]octan-8-yl benzoate **2g**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale-yellow oil (26.0 mg, 52%).

R_f = 0.10 (30% ethyl acetate–hexanes).

1H NMR (500 MHz, $CDCl_3$) δ 7.24 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 5.57 (ddd, J = 48.4, 8.1, 3.1 Hz, 1H), 3.76 (dd, J = 14.4, 10.4 Hz, 2H), 3.49 (ddd, J = 12.6, 10.4, 1.9 Hz, 2H), 3.28–3.24 (m, 1H), 3.00–2.93 (m, 1H), 2.81 (ddd, J = 17.9, 14.1, 8.1 Hz, 1H), 2.51 (ddd, J = 31.7, 14.1, 3.1 Hz, 1H), 2.36 (s,

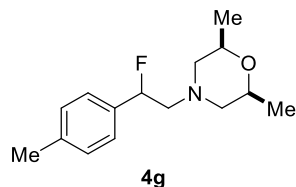
3H), 1.98–1.78 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3) δ 138.2 (d, $J_{\text{C-F}} = 1.9$ Hz), 136.0 (d, $J_{\text{C-F}} = 20.1$ Hz), 129.0, 125.7 (d, $J_{\text{C-F}} = 6.7$ Hz), 94.8 (d, $J_{\text{C-F}} = 172.0$ Hz), 73.1, 73.1, 62.1, 61.8 (d, $J_{\text{C-F}} = 2.2$ Hz), 60.3 (d, $J_{\text{C-F}} = 23.7$ Hz), 25.1, 24.8, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -176.73.

FTIR (thin film): cm^{-1} 2946, 2855, 1516, 1182, 1132, 1008, 981, 883, 816, 641, 543.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{21}\text{FNO}^+$ 250.1602; found: 250.1608.



4-(2-Fluoro-2-(*p*-tolyl)ethyl)-2,6-dimethylmorpholine (4g). Synthesized using **GP3** from **1a** and *O*-benzoyl-2,6-dimethyl-hydroxymorpholine **2h**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 15% ethyl acetate–hexanes), as a pale oil (43.1 mg, 86%).

$R_f = 0.10$ (10% ethyl acetate–hexanes).

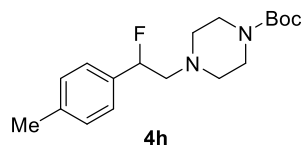
^1H NMR (500 MHz, CDCl_3) δ 7.23 (d, $J = 8.0$ Hz, 2H), 7.18 (d, $J = 8.0$ Hz, 2H), 5.63 (ddd, $J = 49.0, 8.9, 2.1$ Hz, 1H), 3.84–3.53 (m, 2H), 2.92 (d, $J = 9.2$ Hz, 1H), 2.91 – 2.84 (m, 1H), 2.81 (d, $J = 10.6$ Hz, 1H), 2.59 (ddd, $J = 34.9, 14.2, 2.1$ Hz, 1H), 2.36 (s, 3H), 1.92 (dt, $J = 17.8, 10.6$ Hz, 2H), 1.18 (d, $J = 2.7$ Hz, 3H), 1.16 (d, $J = 2.7$ Hz, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 138.3 (d, $J_{\text{C-F}} = 1.7$ Hz), 135.8 (d, $J_{\text{C-F}} = 19.9$ Hz), 129.2, 125.6 (d, $J_{\text{C-F}} = 6.7$ Hz), 92.6 (d, $J_{\text{C-F}} = 172.9$ Hz), 71.7, 71.6, 64.6 (d, $J_{\text{C-F}} = 23.1$ Hz), 60.1, 59.6, 21.2, 19.2, 19.1.

^{19}F NMR (471 MHz, CDCl_3) δ -175.17.

FTIR (thin film): cm^{-1} 2972, 2864, 1516, 1454, 1323, 1142, 1083, 1027, 885, 812, 712.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{23}\text{FNO}^+$ 252.1758; found: 252.1753.



tert-Butyl 4-(2-fluoro-2-(*p*-tolyl)ethyl)piperazine-1-carboxylate (4h). Synthesized using **GP3** from **1a** and *O*-benzoyl-4-*tert*-butoxycarbonyl-hydroxypiperazine **2i**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a pale oil (39.9 mg, 62%).

$R_f = 0.25$ (30% ethyl acetate–hexanes).

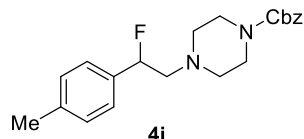
^1H NMR (500 MHz, CDCl_3) δ 7.23 (d, $J = 8.0$ Hz, 2H), 7.16 (d, $J = 8.0$ Hz, 2H), 5.59 (ddd, $J = 48.8, 8.8, 2.3$ Hz, 1H), 3.47–3.44 (m, 4H), 2.87 (ddd, $J = 17.1, 14.3, 8.8$ Hz, 1H), 2.62–2.43 (m, 5H), 2.27 (s, 3H), 1.39 (s, 9H).

^{13}C NMR (125 MHz, CDCl_3) δ 154.7, 138.3, 135.7 (d, $J_{\text{C-F}} = 20.0$ Hz), 129.1, 125.5 (d, $J_{\text{C-F}} = 6.59$ Hz), 92.63 (d, $J_{\text{C-F}} = 173.0$ Hz), 79.6, 64.5 (d, $J_{\text{C-F}} = 23.5$ Hz), 53.4, 28.6, 21.4.

^{19}F NMR (471 MHz, CDCl_3) δ -175.12

FTIR (thin film): cm^{-1} 2975, 2928, 2860, 2811, 1693, 1455, 1419, 1365, 1243, 1170, 1125, 1004, 816.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{17}\text{H}_{25}\text{FNO}_2^+$ 294.1864; found: 294.1869.



2-(4-(2-Fluoro-2-(*p*-tolyl)ethyl)piperazin-1-yl)-1-phenylethan-1-one (4i). Synthesized using **GP3** from **1a** and *O*-benzoyl-4-benzoyloxycarbonyl-hydroxylpiperazine **2j**. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a yellow oil (54.1 mg, 76%).

R_f = 0.38 (50% ethyl acetate–hexanes).

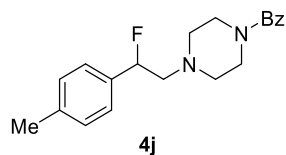
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.39–7.30 (m, 5H), 7.23 (d, J = 8.0 Hz, 2H), 7.18 (d, J = 8.0 Hz, 2H), 5.61 (ddd, J = 48.8, 8.8, 2.3 Hz, 1H), 5.14 (s, 2H), 3.75–3.45 (m, 4H), 2.93 (ddd, J = 17.0, 14.3, 8.8 Hz, 1H), 2.71–2.62 (m, 1H), 2.61–2.57 (m, 4H), 2.36 (s, 3H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 155.2, 138.4, 136.7, 135.6 (d, $J_{\text{C-F}}$ = 20.1 Hz), 129.2, 128.4, 128.0, 127.9, 125.5 (d, $J_{\text{C-F}}$ = 6.7 Hz), 92.6 (d, $J_{\text{C-F}}$ = 173.3 Hz), 67.1, 64.4 (d, $J_{\text{C-F}}$ = 23.6 Hz), 53.3, 43.8, 21.1.

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -175.15.

FTIR (thin film): cm^{-1} 1696, 1455, 1428, 1235, 1123, 1003, 762, 697.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{21}\text{H}_{26}\text{FN}_2\text{O}_2^+$ 357.1973; found: 357.1977.



4-(2-Fluoro-2-(*p*-tolyl)ethyl)piperazin-1-yl(phenyl)methanone (4j). Synthesized using **GP3** from **1a** and *O*-benzoyl-4-benzoylhydroxylpiperazine **2k**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a yellow oil. (45.1 mg, 69%).

R_f = 0.40 (50% ethyl acetate–hexanes).

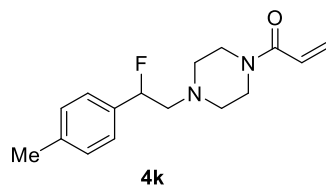
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.40 (s, 5H), 7.22 (d, J = 8.0 Hz, 2H), 7.18 (d, J = 8.0 Hz, 2H), 5.62 (ddd, J = 48.8, 8.7, 2.3 Hz, 1H), 3.82 (br, 2H), 3.46 (br, 2H), 2.94 (ddd, J = 17.0, 14.3, 8.7 Hz, 1H), 2.79–2.46 (m, 5H), 2.35 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.3, 138.4 (d, $J_{\text{C-F}}$ = 1.8 Hz), 135.7, 135.5 (d, $J_{\text{C-F}}$ = 20.0 Hz), 129.6, 129.2, 128.4, 127.0, 125.5 (d, $J_{\text{C-F}}$ = 6.6 Hz), 92.6 (d, $J_{\text{C-F}}$ = 173.2 Hz), 64.2 (d, $J_{\text{C-F}}$ = 23.6 Hz), 53.8, 53.3, 47.6, 42.1, 21.1.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -175.16.

FTIR (thin film): cm^{-1} 1648, 1414, 1406, 1215, 1114, 754, 598.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{20}\text{H}_{24}\text{FN}_2\text{O}^+$ 327.1867; found: 327.1872.



1-(4-(2-Fluoro-2-(*p*-tolyl)ethyl)piperazin-1-yl)prop-2-en-1-one (4k). Synthesized using **GP3** from **1a** and 4-acryloylpiperazin-1-yl benzoate **2u**. Isolated by flash column chromatography (30% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a pale oil (27.0 mg, 49%).

R_f = 0.10 (50% ethyl acetate–hexanes).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.22 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 6.56 (dd, J = 16.8, 10.6

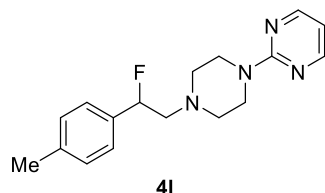
Hz, 1H), 6.28 (dd, $J = 16.8, 1.5$ Hz, 1H), 5.69 (dd, $J = 10.6, 1.5$ Hz, 1H), 5.59 (ddd, $J = 49.0, 8.9, 2.1$ Hz, 1H), 3.88–3.50 (m, 4H), 3.04–2.85 (m, 1H), 2.75–2.66 (m, 1H), 2.66–2.50 (m, 4H), 2.35 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 165.3, 138.5 (d, $J_{\text{C-F}} = 1.2$ Hz), 135.5 (d, $J_{\text{C-F}} = 20.2$ Hz), 129.2, 127.9, 127.4, 125.6 (d, $J_{\text{C-F}} = 6.5$ Hz), 64.2 (d, $J_{\text{C-F}} = 23.7$ Hz), 53.8, 53.2, 45.8, 41.9, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -175.16.

FTIR (thin film): cm^{-1} 2922, 2857, 1642, 1612, 1439, 1233, 1147, 1001, 818, 790, 717.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{22}\text{FN}_2\text{O}^+$ 277.1711; found: 277.1711.



2-(4-(2-Fluoro-2-(*p*-tolyl)ethyl)piperazin-1-yl)pyrimidine (4l). Synthesized using **GP3** from **1a** and 4-(pyrimidin-2-yl)piperazin-1-yl benzoate **2l**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 33% ethyl acetate–hexanes), as a pale oil (28.2 mg, 47%).

$R_f = 0.10$ (30% ethyl acetate–hexanes).

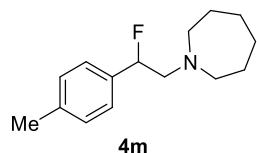
^1H NMR (500 MHz, CDCl_3) δ 8.31 (d, $J = 4.7$ Hz, 2H), 7.25 (d, $J = 7.8$ Hz, 2H), 7.19 (d, $J = 7.8$ Hz, 2H), 6.48 (t, $J = 4.7$ Hz, 1H), 5.67 (ddd, $J = 48.6, 8.3, 1.2$ Hz, 1H), 3.87 (t, $J = 4.9$ Hz, 4H), 3.03–2.88 (m, 1H), 2.74–2.58 (m, 1H), 2.74–2.58 (m, 4H), 2.36 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 161.7, 161.5, 157.7, 138.4 (d, $J_{\text{C-F}} = 1.5$ Hz), 135.8 (d, $J_{\text{C-F}} = 19.8$ Hz), 129.2, 125.6 (d, $J_{\text{C-F}} = 6.5$ Hz), 109.9, 92.7 (d, $J_{\text{C-F}} = 173.0$ Hz), 64.7 (d, $J_{\text{C-F}} = 23.4$ Hz), 53.5, 43.7, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -175.11.

FTIR (thin film): cm^{-1} 2938, 2852, 1583, 1545, 1497, 1358, 1257, 982, 796.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{17}\text{H}_{22}\text{FN}_4^+$ 301.1823; found: 301.1816.



1-(2-Fluoro-2-(*p*-tolyl)ethyl)azepane (4m). Synthesized using **GP3** from **1a** and *O*-benzoylhydroxylazepane **2m**. Isolated by flash column chromatography (hexanes–dichloromethane–ethyl acetate = 15:1:1, v/v/v), as a pale oil (15.1 mg, 32%).

$R_f = 0.10$ (10% ethyl acetate–hexanes).

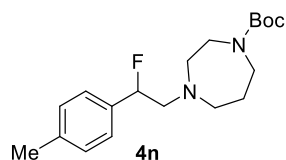
^1H NMR (500 MHz, CDCl_3) δ 7.23 (d, $J = 8.1$ Hz, 2H), 7.17 (d, $J = 8.1$ Hz, 2H), 5.57 (ddd, $J = 48.8, 8.4, 2.6$ Hz, 1H), 3.11–3.01 (m, 1H), 2.88–2.74 (m, 1H), 2.88–2.74 (m, 4H), 2.35 (s, 3H), 1.71–1.54 (m, 8H).

^{13}C NMR (125 MHz, CDCl_3) δ 138.0 (d, $J_{\text{C-F}} = 1.5$ Hz), 136.4 (d, $J_{\text{C-F}} = 20.0$ Hz), 129.1, 125.6 (d, $J_{\text{C-F}} = 6.9$ Hz), 93.2 (d, $J_{\text{C-F}} = 172.8$ Hz), 63.7 (d, $J_{\text{C-F}} = 23.6$ Hz), 55.8, 28.0, 27.1, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -176.95.

FTIR (thin film): cm^{-1} 2923, 2853, 1721, 1451, 1356, 1269, 1135, 1014, 814.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{23}\text{FN}^+$ 236.1809; found: 236.1808.



tert-Butyl 4-(2-fluoro-2-(*p*-tolyl)ethyl)-1,4-diazepane-1-carboxylatez (4n). Synthesized using **GP3** from **1a** and *tert*-butyl 4-(benzyloxy)-1,4-diazepane-1-carboxylate **2n**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a colorless oil (26.9 mg, 40%).

R_f = 0.27 (30% ethyl acetate–hexanes).

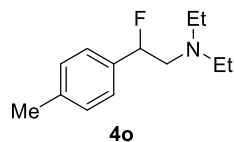
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.21 (d, J = 7.8 Hz, 2H), 7.17 (d, J = 7.8 Hz, 2H), 5.54 (ddd, J = 48.6, 8.3, 2.2 Hz, 1H), 3.52–3.39 (m, 4H), 3.04 (ddd, J = 16.6, 14.5, 8.3 Hz, 1H), 2.86–2.72 (m, 5H), 2.35 (s, 3H), 1.86–1.76 (m, 2H), 1.45 (s, 9H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 155.6, 155.4, 138.2 (d, $J_{\text{C-F}}$ = 7.0 Hz), 135.9 (d, $J_{\text{C-F}}$ = 19.9 Hz), 129.1, 125.5 (d, $J_{\text{C-F}}$ = 6.6 Hz), 93.2 (d, $J_{\text{C-F}}$ = 173.0 Hz), 93.2 (d, $J_{\text{C-F}}$ = 173.4 Hz), 79.2, 76.7, 63.2, 63.0, 56.3, 56.1, 55.2, 55.0, 46.9, 46.4, 46.0, 45.1, 28.5, 27.9, 27.7, 21.1. (Mixture of *cis*- and *trans*-amide rotamers).

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -176.75, -176.95. (The product gives two sets of $^{19}\text{F NMR}$ signals, owing to the presence of *N*-Boc rotamers around the amide.).

FTIR (thin film): cm^{-1} 2930, 1688, 1460, 1411, 1157, 903, 726.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{19}\text{H}_{30}\text{FN}_2\text{O}_2^+$ 337.2286; found: 337.2289.



***N,N*-diethyl-2-fluoro-2-(*p*-tolyl)ethan-1-amine (4o).** Synthesized using **GP3** from **1a** and *O*-benzoylhydroxydiethylamine **2o**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale oil (9.0 mg, 21%).

R_f = 0.20 (30% ethyl acetate–hexanes).

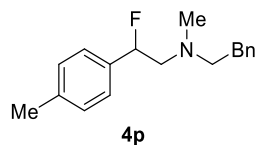
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.23 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 5.55 (ddd, J = 48.6, 8.7, 2.5 Hz, 1H), 2.98 (ddd, J = 16.8, 14.7, 8.7 Hz, 1H), 2.77–2.69 (m, 1H), 2.69–2.62 (m, 4H), 2.35 (s, 3H), 1.06 (t, J = 7.1 Hz, 6H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 138.1 (d, $J_{\text{C-F}}$ = 1.5 Hz), 136.3 (d, $J_{\text{C-F}}$ = 20.5 Hz), 129.1, 125.6 (d, $J_{\text{C-F}}$ = 6.5 Hz), 93.4 (d, $J_{\text{C-F}}$ = 172.9 Hz), 59.4 (d, $J_{\text{C-F}}$ = 23.9 Hz), 47.8, 21.2, 11.8.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -177.06.

FTIR (thin film): cm^{-1} 2970, 2927, 1720, 1450, 1270, 1111, 1069, 816, 711.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{21}\text{FN}^+$ 210.1653; found: 210.1650.



2-Fluoro-*N*-methyl-*N*-phenethyl-2-(*p*-tolyl)ethan-1-amine (4p). Synthesized using **GP3** from **1a** and *O*-benzoyl-*N*-benzyl-*N*-methylhydroxylamine **2q**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale oil (29.8 mg, 55%).

R_f = 0.10 (20% ethyl acetate–hexanes).

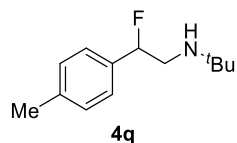
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.31–7.27 (m, 2H), 7.23 (d, J = 7.9 Hz, 2H), 7.18 (d, J = 7.9 Hz, 2H), 7.25–7.17 (m, 3H), 5.59 (ddd, J = 48.7, 8.6, 2.8 Hz, 1H), 3.01 (ddd, J = 16.6, 14.3, 8.6 Hz, 1H), 2.86–2.64 (m, 4H), 2.78–2.64 (m, 1H), 2.48 (s, 3H), 2.36 (s, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 140.3, 138.2 (d, $J_{\text{C-F}}$ = 1.7 Hz), 136.0 (d, $J_{\text{C-F}}$ = 19.8 Hz), 129.2, 128.7, 128.4, 126.0, 125.6 (d, $J_{\text{C-F}}$ = 6.5 Hz), 92.9 (d, $J_{\text{C-F}}$ = 172.9 Hz), 63.5 (d, $J_{\text{C-F}}$ = 23.9 Hz), 59.9, 42.9, 33.8, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -176.41.

FTIR (thin film): cm^{-1} 2924, 2851, 1453, 1122, 1030, 813, 746, 698.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{18}\text{H}_{23}\text{FN}^+$ 272.1809; found: 272.1802.



***N*-(2-Fluoro-2-(*p*-tolyl)ethyl)-2-methylpropan-2-amine (4q).** Synthesized using **GP3** from **1a** and *O*-benzoyl-*N*-(*tert*-butyl)hydroxylamine **2r**. Isolated by flash column chromatography (6% ethyl acetate–hexanes to 12% ethyl acetate–hexanes), as a pale oil (11.2 mg, 26%).

R_f = 0.10 (12% ethyl acetate–hexanes).

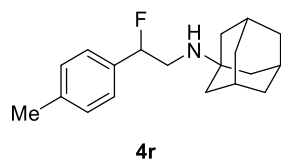
^1H NMR (500 MHz, CDCl_3) δ 7.25 (d, J = 7.9 Hz, 2H), 7.19 (d, J = 7.9 Hz, 2H), 5.51 (ddd, J = 48.9, 9.0, 3.0 Hz, 1H), 3.08 (ddd, J = 15.3, 12.5, 9.1 Hz, 1H), 2.80 (ddd, J = 33.3, 12.5, 3.2 Hz, 1H), 2.36 (s, 3H), 1.12 (s, 9H).

^{13}C NMR (125 MHz, CDCl_3) δ 138.4 (d, $J_{\text{C-F}}$ = 1.7 Hz), 135.7 (d, $J_{\text{C-F}}$ = 19.6 Hz), 129.2, 125.7 (d, $J_{\text{C-F}}$ = 6.5 Hz), 95.0 (d, $J_{\text{C-F}}$ = 170.1 Hz), 50.3, 49.2 (d, $J_{\text{C-F}}$ = 24.1 Hz), 28.9, 21.2.

^{19}F NMR (471 MHz, CDCl_3) δ -180.14.

FTIR (thin film): cm^{-1} 2962, 2924, 1721, 1514, 1361, 1231, 1111, 1014, 815.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{21}\text{FN}^+$ 210.1653; found: 210.1659.



***N*-(2-Fluoro-2-(*p*-tolyl)ethyl)adamantan-1-amine (4r).** Synthesized using **GP3** from **1a** and *N*-(adamantan-1-yl)-*O*-benzoylhydroxylamine **2s**. Isolated by flash column chromatography (5% ethyl acetate–hexanes to 12% ethyl acetate–hexanes), as a pale oil (20.4 mg, 36%).

R_f = 0.15 (12% ethyl acetate–hexanes).

^1H NMR (500 MHz, CDCl_3) δ 7.25 (d, J = 8.0 Hz, 2H), 7.18 (d, J = 8.0 Hz, 2H), 5.49 (ddd, J = 49.0, 9.0, 3.2 Hz, 1H), 3.12 (ddd, J = 15.3, 12.5, 9.1 Hz, 1H), 2.82 (ddd, J = 33.4, 12.5, 3.2 Hz, 1H), 2.36 (s, 3H), 2.07 (s, 3H), 1.66 (d, J = 12.1 Hz, 6H), 1.63 – 1.56 (m, 6H).

^{13}C NMR (125 MHz, CDCl_3) δ 138.4 (d, $J_{\text{C-F}}$ = 1.8 Hz), 135.8 (d, $J_{\text{C-F}}$ = 19.6 Hz), 129.2, 125.7 (d, $J_{\text{C-F}}$ = 6.5 Hz), 95.2 (d, $J_{\text{C-F}}$ = 170.0 Hz), 50.3, 47.1 (d, $J_{\text{C-F}}$ = 24.3 Hz), 42.8, 36.7, 29.6, 21.2.

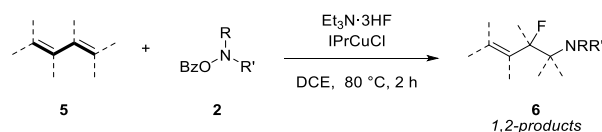
^{19}F NMR (471 MHz, CDCl_3) δ -180.02.

FTIR (thin film): cm^{-1} 2901, 2846, 1515, 1450, 1356, 1310, 1151, 1111, 1033, 814.

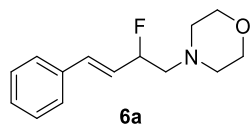
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{19}\text{H}_{27}\text{FN}^+$ 288.2122; found: 288.2123.

6. Aminofluorination of 1,3-dienes and compound characterization data

General procedure 4 (GP4) for aminofluorination of 1,3-dienes



To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylamine **2** (0.2 mmol, 1.0 equiv) and IPrCuCl (4.9 mg, 5 mol %). DCE (1.0 mL), 1,3-diene **5** (0.4 mmol, 2.0 equiv) and Et₃N•3HF (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC analysis, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et₃N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography.



(*E*)-4-(2-Fluoro-4-phenylbut-3-en-1-yl)morpholine (6a). Synthesized using **GP4** from **5a** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale oil (21.2 mg, 45%).

R_f = 0.20 (30% ethyl acetate–hexanes).

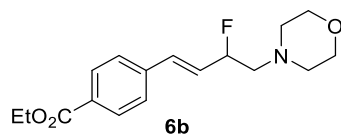
¹H NMR (CDCl₃, 500 MHz): δ 7.40 (d, *J* = 7.5 Hz, 2H), 7.34 (t, *J* = 7.5 Hz, 2H), 7.28 (d, *J* = 7.2 Hz, 1H), 6.70 (dd, *J* = 16.0, 3.3 Hz, 1H), 6.24 (ddd, *J* = 16.0, 13.3, 6.5 Hz, 1H), 5.40–5.11 (m, 1H), 3.75 (t, 4H), 2.79 (ddd, *J* = 18.3, 14.0, 8.0 Hz, 1H), 2.72–2.45 (m, 1H), 2.63–2.45 (m, 4H).

¹³C NMR (125 MHz, CDCl₃) δ 135.9 (d, *J*_{C-F} = 1.4 Hz), 133.1 (d, *J*_{C-F} = 11.5 Hz), 128.7, 128.3, 126.7, 125.5 (d, *J*_{C-F} = 18.4 Hz), 91.7 (d, *J*_{C-F} = 170.0 Hz), 66.9, 63.1 (d, *J* = 22.6 Hz), 54.2.

¹⁹F NMR (CDCl₃, 471 MHz): δ -174.38.

FTIR (thin film): cm⁻¹ 1635, 1451, 1279, 1117, 1010, 745, 695.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₄H₁₉FNO⁺ 236.1445; found: 236.1446.



Ethyl (*E*)-4-(3-fluoro-4-morpholinobut-1-en-1-yl)benzoate (6b). Synthesized using **GP4** from **5b** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 30% ethyl acetate–hexanes), as a yellowish oil (29.0 mg, 47%).

R_f = 0.25 (50% ethyl acetate–hexanes).

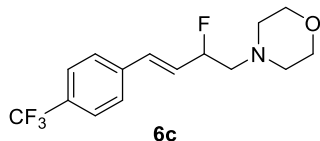
¹H NMR (CDCl₃, 500 MHz): δ 8.00 (d, *J* = 8.1 Hz, 2H), 7.44 (d, *J* = 8.1 Hz, 2H), 6.73 (d, *J* = 16.0 Hz, 1H), 6.35 (ddd, *J* = 16.0, 13.4, 6.4 Hz, 1H), 5.41–5.19 (m, 1H), 4.37 (q, *J* = 7.1 Hz, 2H), 3.74 (t, *J* = 4.4 Hz, 4H), 2.79 (ddd, *J* = 18.7, 14.1, 7.9 Hz, 1H), 2.73–2.63 (m, 1H), 2.58–2.49 (m, 4H), 1.39 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (125 MHz, CDCl₃) δ 166.2, 140.2, 131.7 (d, *J*_{C-F} = 11.8 Hz), 130.0, 129.9, 128.0 (d, *J*_{C-F} = 18.4 Hz), 126.5, 91.3 (d, *J*_{C-F} = 171.5 Hz), 66.9, 62.9 (d, *J*_{C-F} = 22.2 Hz), 61.0, 54.1, 14.3.

¹⁹F NMR (CDCl₃, 471 MHz): δ -176.56.

FTIR (thin film): cm⁻¹ 2958, 2854, 1711, 1607, 1453, 1272, 1106, 1018, 868, 762.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₇H₂₃FNO₃⁺ 308.1657; found: 308.1660.



(E)-4-(2-fluoro-4-(4-(trifluoromethyl)phenyl)but-3-en-1-yl)morpholine (6c). Synthesized using **GP4** from **5c** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 25% ethyl acetate–hexanes), as a yellowish oil (22.6 mg, 38%).

R_f = 0.30 (50% ethyl acetate–hexanes).

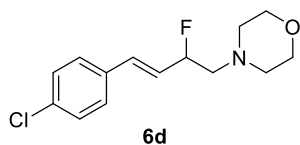
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.58 (d, J = 8.1 Hz, 2H), 7.48 (d, J = 8.1 Hz, 2H), 6.73 (dd, J = 16.0, 1.9 Hz, 1H), 6.34 (ddd, J = 18.8, 15.4, 5.9 Hz, 1H), 5.45–5.10 (m, 1H), 3.75 (t, J = 4.5 Hz, 4H), 2.79 (ddd, J = 18.8, 14.0, 7.8 Hz, 1H), 2.71–2.61 (m, 1H), 2.64–2.56 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 139.5, 131.2 (d, $J_{\text{C-F}}$ = 11.9 Hz), 130.0 (q, $J_{\text{C-F}}$ = 32.7 Hz), 128.3 (d, $J_{\text{C-F}}$ = 18.4 Hz), 126.8, 125.6 (q, $J_{\text{C-F}}$ = 3.7 Hz), 124.1 (q, $J_{\text{C-F}}$ = 271.9 Hz), 91.2 (d, $J_{\text{C-F}}$ = 171.7 Hz), 66.9, 62.9 (d, $J_{\text{C-F}}$ = 22.2 Hz), 54.1.

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -62.61, -176.98.

FTIR (thin film): cm^{-1} 2856, 2814, 1616, 1454, 1323, 1164, 1115, 1066, 1015, 868.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{18}\text{F}_4\text{NO}^+$ 304.1319; found: 304.1321.



(E)-4-(4-(4-chlorophenyl)-2-fluorobut-3-en-1-yl)morpholine (6d). Synthesized using **GP4** from **5d** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 25% ethyl acetate–hexanes), as a yellowish oil (26.3 mg, 49%).

R_f = 0.21 (30% ethyl acetate–hexanes).

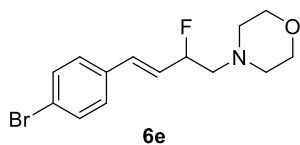
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.32 (d, J = 8.4 Hz, 2H), 7.30 (d, J = 8.4 Hz, 2H), 6.65 (dd, J = 16.0, 2.6 Hz, 1H), 6.22 (ddd, J = 16.0, 13.7, 6.3 Hz, 1H), 5.37–5.03 (m, 1H), 3.74 (t, J = 4.6 Hz, 4H), 2.78 (ddd, J = 18.5, 14.0, 7.9 Hz, 1H), 2.69–2.59 (m, 1H), 2.63–2.53 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 134.5, 134.0, 131.7 (d, $J_{\text{C-F}}$ = 11.8 Hz), 128.9, 127.9, 126.2 (d, $J_{\text{C-F}}$ = 18.4 Hz), 91.5 (d, $J_{\text{C-F}}$ = 170.7 Hz), 66.9, 63.0 (d, $J_{\text{C-F}}$ = 22.2 Hz), 54.2.

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -175.22.

FTIR (thin film): cm^{-1} 2855, 2811, 1491, 1453, 1115, 1089, 1011, 968, 869, 719.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{18}\text{ClFNO}^+$ 270.1056; found: 270.1055.



(E)-4-(4-(4-bromophenyl)-2-fluorobut-3-en-1-yl)morpholine (6e). Synthesized using **GP4** from **5e** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 25% ethyl acetate–hexanes), as a yellowish solid (30.4 mg, 48%).

R_f = 0.22 (30% ethyl acetate–hexanes).

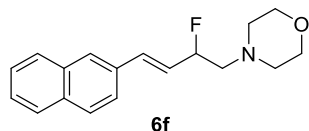
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.45 (d, J = 8.1 Hz, 2H), 7.25 (d, J = 8.1 Hz, 2H), 6.63 (dd, J = 16.0, 2.9 Hz, 1H), 6.23 (ddd, J = 16.0, 13.9, 6.3 Hz, 1H), 5.36–5.09 (m, 1H), 3.74 (t, J = 4.5 Hz, 4H), 2.77 (ddd, J = 18.5, 14.0, 7.9 Hz, 1H), 2.67–2.58 (m, 1H), 2.64–2.39 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3) δ 134.9, 131.8, 131.7 (d, $J_{\text{C-F}} = 12.0$ Hz), 128.2, 126.3 (d, $J_{\text{C-F}} = 18.4$ Hz), 122.1, 91.5 (d, $J_{\text{C-F}} = 170.8$ Hz), 66.9, 63.0 (d, $J_{\text{C-F}} = 22.5$ Hz), 54.2.

^{19}F NMR (CDCl_3 , 471 MHz): δ -175.42.

FTIR (thin film): cm^{-1} 2957, 2854, 1487, 1453, 1115, 1071, 1008, 967, 868, 712.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{18}\text{BrFNO}^+$ 314.0550; found: 314.0550.



(E)-4-(2-Fluoro-4-(naphthalen-2-yl)but-3-en-1-yl)morpholine (6f). Synthesized using **GP4** from **5f** and *O*-benzoylhydroxylmorpholine **2a**. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a yellowish oil (18.4 mg, 33%).

$R_f = 0.20$ (30% ethyl acetate–hexanes).

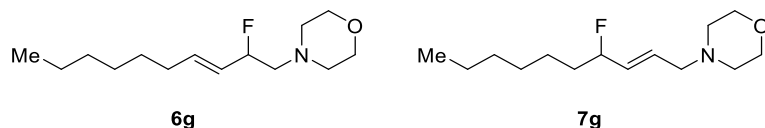
^1H NMR (CDCl_3 , 500 MHz): δ 7.85–7.78 (m, 3H), 7.76 (s, 1H), 7.59 (d, $J = 8.5$ Hz, 1H), 7.51–7.43 (m, 2H), 6.88 (dd, $J = 15.9, 3.0$ Hz, 1H), 6.36 (ddd, $J = 15.9, 13.5, 6.4$ Hz, 1H), 5.56–5.26 (m, 1H), 3.80 (t, $J = 4.4$ Hz, 4H), 2.87 (ddd, $J = 18.1, 14.1, 8.0$ Hz, 1H), 2.8–2.69 (m, 1H), 2.76–2.60 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3) δ 133.5, 133.4 (d, $J_{\text{C-F}} = 11.6$ Hz), 133.3, 133.3, 128.4, 128.1, 127.7, 127.2, 126.4, 126.3, 125.4 (d, $J_{\text{C-F}} = 18.9$ Hz), 123.4, 91.5 (d, $J_{\text{C-F}} = 170.2$ Hz), 66.6, 62.9 (d, $J_{\text{C-F}} = 22.4$ Hz), 54.0.

^{19}F NMR (CDCl_3 , 471 MHz): δ -174.02.

FTIR (thin film): cm^{-1} 2924, 2853, 1716, 1596, 1452, 1271, 1115, 1009, 966, 867, 746.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{18}\text{H}_{21}\text{FNO}^+$ 286.1602; found: 286.1600.



(E)-4-(2-fluorodec-3-en-1-yl)morpholine (6g) and **(E)-4-(4-fluorodec-2-en-1-yl)morpholine (7g).** Synthesized using **GP4** from **5g** and *O*-benzoylhydroxylmorpholine **2a**. ^1H -NMR analysis of the crude reaction mixture indicated the formation of 1,2-product **6g** and 1,4-product **7g** in a ratio of 7:1.

1,2-Product **6g** was isolated by flash column chromatography (5% ethyl acetate–hexanes to 10% ethyl acetate–hexanes) as a yellowish oil (17.1 mg, 35%).

$R_f = 0.20$ (13% ethyl acetate–hexanes).

^1H NMR (CDCl_3 , 500 MHz): δ 5.89–5.75 (m, 1H), 5.61–5.45 (m, 1H), 5.05 (dt, $J = 14.8, 6.5$ Hz, 1H), 3.82–3.65 (t, $J = 4.6, 4\text{H}$), 2.69 (ddd, $J = 17.7, 14.0, 8.2$ Hz, 1H), 2.61–2.51 (m, 4H), 2.57–2.41 (m, 1H), 2.11–2.00 (m, 2H), 1.44–1.33 (m, 2H), 1.3–1.18 (m, 6H), 0.88 (t, $J = 6.9$ Hz, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 136.3 (d, $J_{\text{C-F}} = 11.1$ Hz), 126.4 (d, $J_{\text{C-F}} = 18.7$ Hz), 91.9 (d, $J_{\text{C-F}} = 167.3$ Hz), 66.9, 63.2 (d, $J_{\text{C-F}} = 22.8$ Hz), 54.1, 32.2 (d, $J_{\text{C-F}} = 1.2$ Hz), 31.7, 28.8 (d, $J_{\text{C-F}} = 2.5$ Hz), 22.6, 14.1.

^{19}F NMR (CDCl_3 , 471 MHz): δ -171.31.

FTIR (thin film): cm^{-1} 2925, 2854, 1672, 1454, 1294, 1119, 1009, 968, 869.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{27}\text{FNO}^+$ 244.2071; found: 244.2073.

1,4-Product **7g** was isolated by flash column chromatography (10% ethyl acetate–hexanes to 25% ethyl acetate–hexanes) as a yellowish oil (3.4 mg, 7%).

$R_f = 0.15$ (20% ethyl acetate–hexanes).

^1H NMR (CDCl_3 , 500 MHz): δ 5.82–5.75 (m, 1H), 5.74–5.65 (m, 1H), 4.86 (ddd, $J = 48.8, 12.8, 6.3$ Hz,

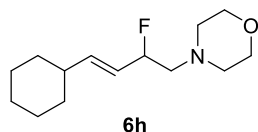
1H), 3.72 (t, $J = 4.6$ Hz, 4H), 3.01 (dd, $J = 5.8, 3.6$ Hz, 2H), 2.54–2.30 (m, 4H), 1.77–1.67 (m, 1H), 1.64–1.57 (m, 1H), 1.37–1.24 (m, 8H), 0.88 (t, $J = 6.8$ Hz, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 132.6 (d, $J_{\text{C-F}} = 20.4$ Hz), 129.6 (d, $J_{\text{C-F}} = 10.5$ Hz), 93.2 (d, $J_{\text{C-F}} = 165.2$ Hz), 66.9, 60.5, 53.5, 35.4 (d, $J_{\text{C-F}} = 22.5$ Hz), 31.7, 29.0, 24.7 (d, $J_{\text{C-F}} = 4.5$ Hz), 22.5, 14.1.

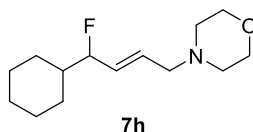
^{19}F NMR (CDCl_3 , 471 MHz): δ -172.83.

FTIR (thin film): cm^{-1} 2927, 2855, 1677, 1454, 1118, 1004, 974, 867.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{27}\text{FNO}^+$ 244.2071; found: 244.2073.



6h



7h

(E)-4-(4-Cyclohexyl-2-fluorobut-3-en-1-yl)morpholine (6h) and (E)-4-(4-Cyclohexyl-4-fluorobut-2-en-1-yl)morpholine (7h). Synthesized using **GP4** from **5h** and *O*-benzoylhydroxymorpholine **2a**. ^1H -NMR analysis of the crude reaction mixture indicated the formation of 1,2-product **6h** and 1,4-product **7h** in a ratio of 8:1.

1,2-Product **6h** was isolated by flash column chromatography (5% ethyl acetate–hexanes to 10% ethyl acetate–hexanes) as a yellowish oil (21.4 mg, 44%).

$R_f = 0.20$ (10% ethyl acetate–hexanes).

^1H NMR (CDCl_3 , 500 MHz): δ 5.76 (ddd, $J = 15.5, 6.0, 4.5$ Hz, 1H), 5.47 (dddd, $J = 15.6, 11.0, 7.1, 1.3$ Hz, 1H), 5.15–4.80 (m, 1H), 3.73 (t, $J = 4.5$ Hz, 4H), 2.69 (ddd, $J = 17.6, 14.0, 8.3$ Hz, 1H), 2.55 (t, $J = 4.5$ Hz, 4H), 2.48 (ddd, $J = 31.9, 14.0, 2.7$ Hz, 1H), 2.10–1.87 (m, 1H), 1.80–1.59 (m, 5H), 1.34–1.21 (m, 2H), 1.20–1.12 (m, 1H), 1.12–1.01 (m, 2H).

^{13}C NMR (125 MHz, CDCl_3) δ 141.6 (d, $J = 10.8$ Hz), 123.9 (d, $J_{\text{C-F}} = 19.0$ Hz), 92.1 (d, $J_{\text{C-F}} = 167.4$ Hz), 66.9, 63.3 (d, $J_{\text{C-F}} = 22.8$ Hz), 54.1, 40.3, 32.5 (d, $J_{\text{C-F}} = 1.8$ Hz), 32.4 (d, $J_{\text{C-F}} = 1.8$ Hz), 26.1, 25.9.

^{19}F NMR (CDCl_3 , 471 MHz): δ -171.47.

FTIR (thin film): cm^{-1} 2923, 2851, 2810, 1620, 1450, 1293, 1140, 1118, 1009, 969, 869.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{25}\text{FNO}^+$ 242.1915; found: 242.1915.

1,4-Product **7h** was isolated by flash column chromatography (10% ethyl acetate–hexanes to 25% ethyl acetate–hexanes) as a yellowish oil (2.4 mg, 5%).

$R_f = 0.10$ (20% ethyl acetate–hexanes).

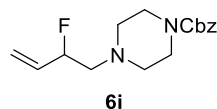
^1H NMR (CDCl_3 , 500 MHz): δ 5.85–5.57 (m, 2H), 4.58 (dt, $J = 48.5, 6.6$ Hz, 1H), 3.72 (t, $J = 4.6$ Hz, 4H), 3.11–2.85 (m, 2H), 2.44 (s, 4H), 1.86 (d, $J = 12.8$ Hz, 1H), 1.81–1.72 (m, 2H), 1.67 (d, $J = 12.2$ Hz, 1H), 1.60–1.49 (m, 1H), 1.30–1.09 (m, 4H), 1.07–0.94 (m, 2H).

^{13}C NMR (125 MHz, CDCl_3) δ 131.1 (d, $J = 20.3$ Hz), 130.7 (d, $J = 12.2$ Hz), 97.2 (d, $J = 167.7$ Hz), 66.9, 60.6, 53.5, 42.3 (d, $J = 21.1$ Hz), 28.2 (d, $J = 4.5$ Hz), 27.9 (d, $J = 5.1$ Hz), 26.3, 25.8, 25.7.

^{19}F NMR (CDCl_3 , 471 MHz): δ -178.48.

FTIR (thin film): cm^{-1} 2923, 2851, 1718, 1450, 1269, 1117, 1069, 969, 869.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{25}\text{FNO}^+$ 242.1915; found: 242.1914.



6i

Benzyl 4-(2-fluorobut-3-en-1-yl)piperazine-1-carboxylate (6i). Synthesized using **GP4** from **5i** and *O*-benzoyl-4-benzoyloxycarbonyl-hydroxypiperazine **2i**. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 25% ethyl acetate–hexanes), as a colorless oil (30.0 mg, 51%).

R_f = 0.50 (50% ethyl acetate–hexanes).

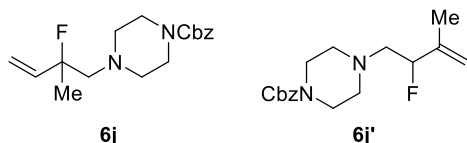
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.44–7.29 (m, 5H), 5.99–5.77 (m, 1H), 5.44–5.32 (m, 1H), 5.27 (d, J = 10.7 Hz, 1H), 5.13 (s, 2H), 5.16–5.00 (m, 1H), 3.66–3.47 (m, 4H), 2.70 (ddd, J = 18.7, 14.2, 8.0 Hz, 1H), 2.61–2.50 (m, 1H), 2.58–2.41 (m, 4H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 155.2, 136.7, 134.6 (d, $J_{\text{C-F}}$ = 19.3 Hz), 128.5, 128.0, 127.9, 117.6 (d, $J_{\text{C-F}}$ = 11.8 Hz), 91.6 (d, $J_{\text{C-F}}$ = 170.9 Hz), 67.1, 62.4 (d, $J_{\text{C-F}}$ = 21.9 Hz), 53.3, 43.8.

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -178.96.

FTIR (thin film): cm^{-1} 2924, 2814, 1696, 1426, 1234, 1121, 1001, 933, 696.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{22}\text{FN}_2\text{O}_2^+$ 293.1660; found: 293.1662.



Benzyl 4-(2-fluoro-2-methylbut-3-en-1-yl)piperazine-1-carboxylate (6j) and benzyl 4-(2-fluoro-3-methylbut-3-en-1-yl)piperazine-1-carboxylate (6j'). Synthesized using **GP4** from **5j** and *O*-benzoyl-4-benzyloxycarbonyl-hydroxypiperazine **2j**. $^1\text{H-NMR}$ analysis of the crude reaction mixture indicated the formation of two 1,2-products **6j** and **6j'** in a ratio of 10:1.

1,2-Product **6j** was isolated by flash column chromatography (2.5% 1,4-dioxane–hexanes), as a colorless oil (27.5 mg, 45%).

R_f = 0.10 (10% 1,4-dioxane–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.45–7.21 (m, 5H), 5.94 (td, J = 17.6, 11.0 Hz, 1H), 5.30 (d, J = 17.4 Hz, 1H), 5.15 (d, J = 11.1 Hz, 1H), 5.13 (s, 2H), 3.49 (t, J = 5.0 Hz, 4H), 2.59–2.33 (m, 2H), 2.59–2.33 (m, 4H), 1.44 (d, J = 21.6 Hz, 3H).

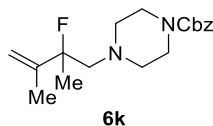
$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 155.3, 139.7 (d, $J_{\text{C-F}}$ = 22.2 Hz), 136.8, 128.5, 128.0, 127.9, 114.0 (d, $J_{\text{C-F}}$ = 11.1 Hz), 96.8 (d, $J_{\text{C-F}}$ = 171.4 Hz), 67.1, 65.7 (d, $J_{\text{C-F}}$ = 22.3 Hz), 54.1, 54.1, 43.9, 23.4 (d, $J_{\text{C-F}}$ = 24.8 Hz).

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -147.99.

FTIR (thin film): cm^{-1} 2926, 2858, 2810, 1698, 1427, 1362, 1242, 1123, 1007, 920, 752.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{17}\text{H}_{24}\text{FN}_2\text{O}_2^+$ 307.1816; found: 307.1816.

Minor product **6j'** was unable to be obtained as a pure product.



Benzyl 4-(2-fluoro-2,3-dimethylbut-3-en-1-yl)piperazine-1-carboxylate (6k). Synthesized using **GP4** from **5k** and *O*-benzoyl-4-benzyloxycarbonyl-hydroxypiperazine **2j**. Isolated by flash column chromatography (2.5% 1,4-dioxane–hexanes to 3.3% 1,4-dioxane–hexanes), as a colorless oil (27.0 mg, 42%).

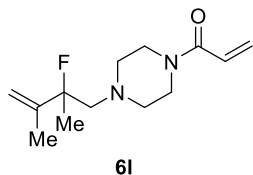
R_f = 0.15 (10% 1,4-dioxane–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.46–7.26 (m, 5H), 5.13 (s, 2H), 5.01 (s, 1H), 4.89 (s, 1H), 3.51 (s, 4H), 2.82–2.34 (m, 2H), 2.82–2.34 (m, 4H), 1.77 (s, 3H), 1.46 (d, J = 22.2 Hz, 3H).

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 155.2, 146.3 (d, $J_{\text{C-F}}$ = 19.9 Hz), 136.8, 128.5, 128.0, 127.9, 110.7 (d, $J_{\text{C-F}}$ = 11.4 Hz), 99.0 (d, $J_{\text{C-F}}$ = 174.4 Hz), 67.1, 64.4 (d, $J_{\text{C-F}}$ = 22.1 Hz), 54.0 (d, $J_{\text{C-F}}$ = 2.0 Hz), 43.7, 23.2 (d, $J_{\text{C-F}}$ = 25.4 Hz), 19.3 (d, $J_{\text{C-F}}$ = 5.5 Hz).

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -146.45.

FTIR (thin film): cm^{-1} 2938, 2861, 2810, 1698, 1427, 1237, 1119, 1094, 1008, 905, 734.
HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{18}\text{H}_{26}\text{FN}_2\text{O}_2^+$ 321.1973; found: 321.1973.



1-(4-(2-Fluoro-2,3-dimethylbut-3-en-1-yl)piperazin-1-yl)prop-2-en-1-one (6l). Synthesized using **GP4** from **5k** and 4-acryloylpiperazin-1-yl benzoate **2u**. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 20% ethyl acetate–hexanes), as a pale oil (21.2 mg, 44%).

R_f = 0.35 (25% ethyl acetate–hexanes).

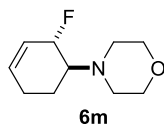
^1H NMR (CDCl_3 , 500 MHz): δ 6.54 (dd, J = 16.8, 10.6 Hz, 1H), 6.27 (dd, J = 16.8, 1.9 Hz, 1H), 5.67 (dd, J = 10.6, 1.9 Hz, 1H), 5.00 (s, 1H), 4.88 (dd, J = 2.3, 1.5 Hz, 1H), 3.59 (d, J = 71.1 Hz, 4H), 2.76–2.37 (m, 2H), 2.76–2.37 (m, 4H), 1.77 (s, 3H), 1.45 (d, J = 22.2 Hz, 3H).

^{13}C NMR (125 MHz, CDCl_3) δ 165.3, 146.4 (d, $J_{\text{C-F}}$ = 20.1 Hz), 127.7, 127.5, 110.5 (d, $J_{\text{C-F}}$ = 11.4 Hz), 99.2 (d, $J_{\text{C-F}}$ = 174.1 Hz), 64.4 (d, $J_{\text{C-F}}$ = 22.0 Hz), 54.3 (d, $J_{\text{C-F}}$ = 41.0 Hz), 45.9, 42.1, 23.3 (d, $J_{\text{C-F}}$ = 25.6 Hz), 19.3 (d, $J_{\text{C-F}}$ = 5.4 Hz).

^{19}F NMR (CDCl_3 , 471 MHz): δ -146.57.

FTIR (thin film): cm^{-1} 2982, 2924, 2809, 1648, 1613, 1439, 1238, 1129, 1007, 905, 791.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{13}\text{H}_{22}\text{FN}_2\text{O}^+$ 241.1711; found: 241.1707.



trans-4-(2-Fluorocyclohex-3-en-1-yl)morpholine (6m). Synthesized using **GP4** from **5m** and *O*-benzoylhydroxymorpholine **2a**. Only *trans*-isomer was observed (dr >20:1) by ^1H -NMR analysis of the crude reaction mixture. Isolated by flash column chromatography (12% ethyl acetate–hexanes to 25% ethyl acetate–hexanes), as a colorless oil (16.5 mg, 45%).

R_f = 0.20 (50% ethyl acetate–hexanes).

^1H NMR (CDCl_3 , 500 MHz): δ 5.87 (dd, J = 9.7, 2.5 Hz, 1H), 5.76–5.64 (m, 1H), 5.28–5.11 (m, 1H), 3.73 (t, J = 4.5 Hz, 4H), 2.81–2.57 (m, 1H), 2.81–2.57 (m, 4H), 2.23–2.09 (m, 2H), 1.92 (dd, J = 8.9, 4.5 Hz, 1H), 1.59–1.47 (m, 1H).

^{13}C NMR (125 MHz, CDCl_3) δ 131.5 (d, $J_{\text{C-F}}$ = 9.5 Hz), 126.3 (d, $J_{\text{C-F}}$ = 21.6 Hz), 88.7 (d, $J_{\text{C-F}}$ = 166.2 Hz), 67.5, 64.4 (d, $J_{\text{C-F}}$ = 16.1 Hz), 49.9, 25.4 (d, $J_{\text{C-F}}$ = 2.9 Hz), 23.1 (d, $J_{\text{C-F}}$ = 7.4 Hz).

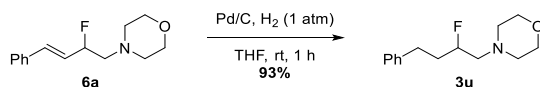
^{19}F NMR (CDCl_3 , 471 MHz): δ -172.49.

FTIR (thin film): cm^{-1} 2924, 2852, 1717, 1451, 1268, 1116, 1015, 901, 854, 732.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{10}\text{H}_{17}\text{FNO}^+$ 186.1289; found: 186.1287.

7. Synthetic applications

7.1. Transformations of β -fluoro homoallylic amines to alkyl-substituted β -fluoroalkylamines



4-(2-Fluoro-4-phenylbutyl)morpholine (3u). To a stirring solution of (*E*)-4-(2-fluoro-4-phenylbut-3-en-1-yl)morpholine **6a** (23.5 mg, 0.1 mmol, 1 equiv) in THF (2.0 mL) was added Pd/C (10% on activated carbon, 4 mg) at room temperature. After the inner atmosphere of the flask was changed to hydrogen by a hydrogen balloon (1atm), the resulting mixture was allowed to stir at room temperature for 1 h. Upon the completion consumption of **6a** (monitored by LC-MS analysis), the reaction was stopped immediately and filtered. The solvent was removed under reduced pressure. The resulting residue was purified by silica gel chromatography (40% ethyl acetate–hexanes), giving **3u** as yellow oil (22.1 mg, 93%).

R_f = 0.10 (33% ethyl acetate–hexanes).

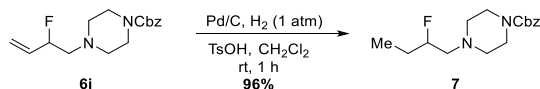
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.32–7.27 (m, 2H), 7.23–7.18 (m, 3H), 4.81–4.52 (m, 1H), 3.75–3.58 (m, 4H), 2.88–2.78 (m, 1H), 2.75–2.68 (m, 1H), 2.68–2.58 (m, 1H), 2.55–2.41 (m, 5H), 2.07–1.95 (m, 1H), 1.95–1.78 (m, 1H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 141.2, 128.5, 128.8, 126.1, 91.5 (d, $J_{\text{C-F}}$ = 170.0 Hz), 66.9, 62.8 (d, $J_{\text{C-F}}$ = 20.5 Hz), 54.3, 35.2 (d, $J_{\text{C-F}}$ = 20.8 Hz), 31.2 (d, $J_{\text{C-F}}$ = 4.6 Hz).

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -182.13.

FTIR (thin film): cm^{-1} 2952, 2853, 1717, 1453, 1273, 1116, 1035, 867, 699.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{14}\text{H}_{21}\text{FNO}^+$ 238.1602; found: 238.1600.



Benzyl 4-(2-fluorobutyl)piperazine-1-carboxylate (7). To a stirring solution of benzyl 4-(2-fluorobut-3-en-1-yl)piperazine-1-carboxylate **6i** (29.2 mg, 0.1 mmol, 1 equiv) in CH_2Cl_2 (2.0 mL) was added Pd/C (10% on activated carbon, 4 mg) and $\text{TsOH}\cdot\text{H}_2\text{O}$ (19 mg, 0.1 mmol, 1.0 equiv) at room temperature. After the inner atmosphere of the flask was changed to hydrogen by a hydrogen balloon (1atm), the resulting mixture was allowed to stir at room temperature for 1 h. Upon the completion consumption of **6i** (monitored by LC-MS analysis), the reaction was stopped immediately and filtered through a plug of activated, neutral Al_2O_3 (Brockman grade I, 58–60Å). The solvent was removed under reduced pressure. The resulting residue was purified by silica gel chromatography (30% ethyl acetate–hexanes), giving **7** as yellow oil (28.2 mg, 96%).

R_f = 0.15 (20% ethyl acetate–hexanes).

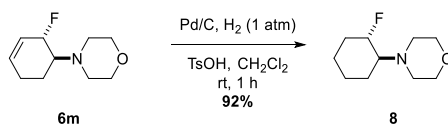
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.50–7.26 (m, 5H), 5.13 (s, 2H), 4.59 (dtdd, J = 50.0, 7.5, 5.0, 2.6 Hz, 1H), 3.53 (t, J = 4.5 Hz, 4H), 2.62 (ddd, J = 19.1, 14.0, 7.6 Hz, 1H), 2.55–2.33 (m, 1H), 2.55–2.33 (m, 4H), 1.77–1.47 (m, 2H), 0.98 (t, J = 7.5 Hz, 3H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 155.2, 136.8, 128.5, 128.0, 127.9, 93.6 (d, $J_{\text{C-F}}$ = 170.0 Hz), 67.1, 62.1 (d, $J_{\text{C-F}}$ = 20.8 Hz), 53.5, 43.8, 26.6 (d, $J_{\text{C-F}}$ = 21.3 Hz), 9.3 (d, $J_{\text{C-F}}$ = 6.1 Hz).

$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -181.26.

FTIR (thin film): cm^{-1} 2935, 2812, 1697, 1426, 1233, 1119, 997, 854, 763, 697.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{24}\text{FN}_2\text{O}_2^+$ 295.1916; found: 295.1816.



4-(2-Fluorocyclohexyl)morpholine (8). To a stirring solution of 4-(2-fluorocyclohex-3-en-1-

yl)morpholine **6m** (18.5 mg, 0.1 mmol, 1 equiv) in CH₂Cl₂ (2.0 mL) was added Pd/C (10% on activated carbon, 4 mg) and TsOH·H₂O (19 mg, 0.1 mmol, 1.0 equiv) at room temperature. After the inner atmosphere of the flask was changed to hydrogen by a hydrogen balloon (1atm), the resulting mixture was allowed to stir at room temperature for 1 h. Upon the completion consumption of **6m** (monitored by LC-MS analysis), the reaction was stopped immediately and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The solvent was removed under reduced pressure. The resulting residue was purified by silica gel chromatography (50% ethyl acetate–hexanes), giving **8** as yellow oil (17.2 mg, 92%).

R_f = 0.20 (50% ethyl acetate–hexanes).

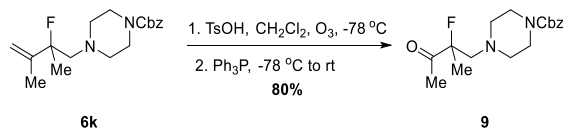
¹H NMR (CDCl₃, 500 MHz): δ 4.52 (dtd, *J* = 50.3, 10.1, 4.8 Hz, 1H), 3.71 (t, *J* = 4.4 Hz, 4H), 2.69 (t, *J* = 4.4 Hz, 4H), 2.50 (dddd, 13.6 Hz, 11.3 Hz, 4.8 Hz, 2.2 Hz, 1H), 2.20–2.00 (m, 1H), 1.95–1.81 (m, 1H), 1.78–1.65 (m, 2H), 1.53–1.38 (m, 1H), 1.33–1.12 (m, 1H), 1.33–1.12 (m, 2H).

¹³C NMR (CDCl₃, 125 MHz): δ 92.0 (d, *J_{C-F}* = 177.3 Hz), 67.7, 67.2 (d, *J_{C-F}* = 15.3 Hz), 49.8, 32.3 (d, *J_{C-F}* = 18.0 Hz), 26.1 (d, *J_{C-F}* = 8.3 Hz), 24.7 (d, *J_{C-F}* = 1.8 Hz), 23.8 (d, *J_{C-F}* = 11.2 Hz).

¹⁹F NMR (CDCl₃, 471 MHz): δ -174.21.

FTIR (thin film): cm⁻¹ 2933, 2854, 1451, 1266, 1116, 1012, 940, 868, 848, 665.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₀H₁₉FNO⁺ 188.1445; found: 188.1444.



Benzyl 4-(2-fluoro-2-methyl-3-oxobutyl)piperazine-1-carboxylate (9). A solution of benzyl 4-(2-fluoro-2,3-dimethylbut-3-en-1-yl)piperazine-1-carboxylate **6k** (32.0 mg, 0.10 mmol, 1.0 equiv) and TsOH·H₂O (28.5 mg, 0.15 mmol, 1.5 equiv) in CH₂Cl₂ was cooled to -78 °C. Ozone was then bubbled through until the solution turned light blue after 10 min. Then triphenylphosphine (39.3, 0.15 mmol, 1.5 equiv) was added. The reaction mixture was allowed to slowly warm up to room temperature and to stir for another 8 h. To the reaction mixture was added water (4 mL) and extracted with dichloromethane (8 mL × 3). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated in *vacuo*. The crude reaction mixture was purified by silica column chromatography (50% ethyl acetate–hexanes), giving **9** as yellow oil (25.8 mg, 80%).

R_f = 0.20 (25% ethyl acetate–hexanes).

¹H NMR (CDCl₃, 500 MHz): δ 7.40–7.28 (m, 5H), 5.11 (s, 2H), 2.79 (dd, *J* = 33.5, 14.4 Hz, 1H), 2.62 (dd, *J* = 15.6, 14.4 Hz, 1H), 2.69–2.54 (m, 2H), 2.49–2.33 (m, 2H), 2.28 (d, *J* = 5.3 Hz, 3H), 1.35 (d, *J* = 21.5 Hz, 3H).

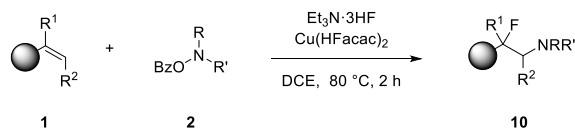
¹³C NMR (CDCl₃, 125 MHz): δ 210.9 (d, *J_{C-F}* = 30.2 Hz), 155.2, 136.7, 128.5, 128.0, 127.9, 102.7 (d, *J_{C-F}* = 184.9 Hz), 67.1, 63.9 (d, *J_{C-F}* = 19.3 Hz), 54.2 (d, *J_{C-F}* = 2.4 Hz), 44.0, 26.7, 20.8 (d, *J_{C-F}* = 23.9 Hz).

¹⁹F NMR (CDCl₃, 471 MHz): δ -156.11.

FTIR (thin film): cm⁻¹ 2860, 2810, 1697, 1425, 1358, 1236, 1098, 1008, 849, 734, 696.

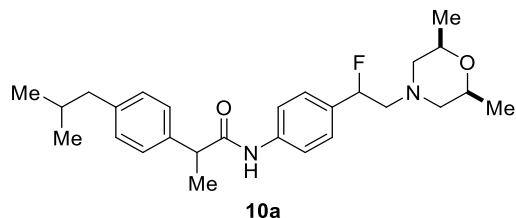
HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₁₇H₂₄FN₂O₃⁺ 323.1766; found: 323.1764.

7.2. Synthesis of the bioactive β-fluoro alkylamine compounds



General procedure 3 (GP3) for alkene aminofluorination reactions.

To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylamine **2** (0.2 mmol, 1.0 equiv) and Cu(HFacac)₂ (2.4 mg, 2.5 mol%). DCE (1.0 mL), alkene **1** (0.4 mmol, 2.0 equiv), and Et₃N•3HF (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et₃N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography.



***N*-(4-(2-(2,6-Dimethylmorpholino)-1-fluoroethyl)phenyl)-2-(4-isobutylphenyl)propanamide (10a).** Synthesized using **GP3** from **1aa** and *O*-benzoyl-2,6-dimethyl-hydroxylmorpholine **2h**. ¹⁹F-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 1:1.

Isolated by flash column chromatography (20% ethyl acetate–hexanes to 33% ethyl acetate–hexanes), as a yellowish oil (55.5 mg, 63%).

R_f = 0.10 (33% ethyl acetate–hexanes).

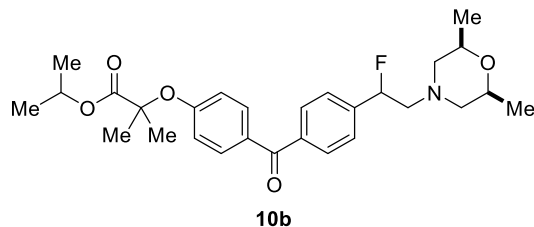
¹H NMR (CDCl₃, 500 MHz): δ 7.43 (d, *J* = 8.3 Hz, 2H), 7.23 (d, *J* = 8.3 Hz, 2H), 7.23 (d, *J* = 8.0 Hz, 2H), 7.15 (d, *J* = 8.0 Hz, 2H), 5.59 (ddd, *J* = 48.6, 8.5, 1.7 Hz, 1H), 3.78–3.65 (m, 2H), 3.78–3.65 (m, 1H), 2.90 (d, *J* = 11.3 Hz, 1H), 2.87–2.81 (m, 1H), 2.79 (d, *J* = 10.4 Hz, 1H), 2.57 (dd, *J* = 33.8, 14.5 Hz, 1H), 2.47 (d, *J* = 7.2 Hz, 2H), 1.97–1.79 (m, 2H), 1.97–1.79 (m, 1H), 1.58 (d, *J* = 7.1 Hz, 3H), 1.16 (d, *J* = 4.1 Hz, 3H), 1.15 (d, *J* = 4.1 Hz, 3H), 0.90 (d, *J* = 6.6 Hz, 6H).

¹³C NMR (CDCl₃, 125 MHz): δ 172.7, 141.2, 138.1, 137.9, 134.4 (d, *J*_{C-F} = 20.3 Hz), 129.9, 127.4, 126.3 (d, *J*_{C-F} = 6.6 Hz), 119.6, 92.2 (d, *J*_{C-F} = 173.6 Hz), 71.6, 64.4 (d, *J*_{C-F} = 23.5 Hz), 59.9, 59.6, 47.8, 45.0, 30.2, 22.4, 19.2, 19.1, 18.5.

¹⁹F NMR (CDCl₃, 471 MHz): δ -175.18, -175.20.

FTIR (thin film): cm⁻¹ 2969, 2632, 2868, 1663, 1601, 1516, 1454, 1411, 1248, 1083, 885, 840, 735.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₂₇H₃₈FN₂O₂⁺ 441.2912; found: 441.2912.



Isopropyl 2-(4-(4-(2-(2,6-dimethylmorpholino)-1-fluoroethyl)benzoyl)phenoxy)-2-methylpropanoate (10b). Synthesized using **GP3** from **1ab** and *O*-benzoyl-2,6-dimethyl-hydroxylmorpholine **2h**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 40% ethyl acetate–hexanes), as a yellowish oil (37.0 mg, 38%).

R_f = 0.20 (33% ethyl acetate–hexanes).

¹H NMR (CDCl₃, 500 MHz): δ 7.76 (d, *J* = 4.1 Hz, 2H), 7.74 (d, *J* = 4.1 Hz, 2H), 7.43 (d, *J* = 8.1 Hz, 2H),

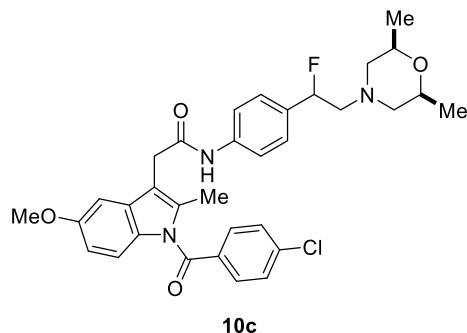
6.86 (d, $J = 8.1$ Hz, 2H), 5.73 (ddd, $J = 48.9, 8.5, 2.3$ Hz, 1H), 5.19–4.95 (m, 1H), 3.79–3.58 (m, 2H), 2.89 (ddd, $J = 22.5, 14.4, 8.5$ Hz, 2H), 2.81 (d, $J = 10.9$ Hz, 1H), 2.66 (ddd, $J = 33.6, 14.4, 2.3$ Hz, 1H), 1.95 (dd, $J = 23.4, 10.8$ Hz, 2H), 1.66 (s, 6H), 1.20 (d, $J = 6.3$ Hz, 6H), 1.18 (d, $J = 3.1$ Hz, 3H), 1.17 (d, $J = 3.1$ Hz, 3H).

^{13}C NMR (CDCl_3 , 125 MHz): δ 195.0, 173.1, 159.7, 142.7 (d, $J_{\text{C-F}} = 20.0$ Hz), 138.2, 132.0, 130.4, 129.9, 125.2 (d, $J_{\text{C-F}} = 7.3$ Hz), 117.2, 92.2 (d, $J_{\text{C-F}} = 175.4$ Hz), 79.4, 71.7, 69.4, 64.4 (d, $J_{\text{C-F}} = 22.5$ Hz), 60.0, 59.7, 25.4, 21.5, 19.2, 19.1.

^{19}F NMR (CDCl_3 , 471 MHz): δ -179.49.

FTIR (thin film): cm^{-1} 2978, 2933, 2807, 1729, 1653, 1598, 1249, 1144, 1102, 929, 854, 767.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{28}\text{H}_{37}\text{FNO}_5^+$ 486.2650; found: 486.2645.



2-(1-(4-Chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)-N-(4-(2-(2,6-dimethylmorpholino)-1-fluoroethyl)phenyl)acetamide (10c). Synthesized using GP3 from **1ac** and *O*-benzoyl-2,6-dimethyl-hydroxylmorpholine **2h** from the reaction on 0.1 mmol scale. Isolated by flash column chromatography (33% ethyl acetate–hexanes to 70% ethyl acetate–hexanes), as a yellowish oil (38.3 mg, 65%).

$R_f = 0.20$ (67% ethyl acetate–hexanes).

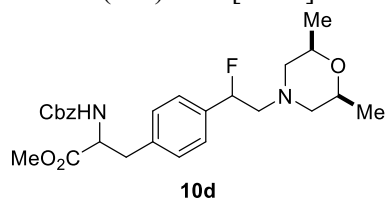
^1H NMR (CDCl_3 , 500 MHz): δ 7.67 (d, $J = 8.2$ Hz, 2H), 7.48 (d, $J = 8.2$ Hz, 2H), 7.43–7.33 (m, 2H), 7.38 (br, 1H), 7.30–7.16 (m, 2H), 6.94 (s, 1H), 6.87 (d, $J = 7.3$ Hz, 1H), 6.71 (d, $J = 7.3$ Hz, 1H), 5.59 (dd, $J = 48.6, 7.5$ Hz, 1H), 3.80 (s, 3H), 3.80 (s, 2H), 3.76–3.64 (m, 2H), 2.88 (d, $J = 10.7$ Hz, 1H), 2.85–2.80 (m, 1H), 2.77 (d, $J = 11.1$ Hz, 1H), 2.64–2.49 (m, 1H), 2.44 (s, 3H), 1.97–1.83 (m, 2H), 1.16 (d, $J = 3.3$ Hz, 3H), 1.15 (d, $J = 3.3$ Hz, 3H).

^{13}C NMR (CDCl_3 , 125 MHz): δ 168.4, 168.3, 156.4, 139.7, 137.6, 136.7, 135.1, 134.9, 133.5, 131.2, 131.0, 130.1, 129.3, 126.3 (d, $J_{\text{C-F}} = 6.6$ Hz), 120.1, 115.2, 112.5, 112.2, 100.8, 92.2 (d, $J_{\text{C-F}} = 173.7$ Hz), 71.6, 64.4 (d, $J_{\text{C-F}} = 23.4$ Hz), 60.0, 59.6, 55.8, 33.4, 19.2, 19.1, 13.4.

^{19}F NMR (CDCl_3 , 471 MHz): δ -175.59.

FTIR (thin film): cm^{-1} 3294, 2929, 2855, 1679, 1603, 1477, 1357, 1322, 1144, 1087, 836, 754.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{33}\text{H}_{36}\text{ClFN}_3\text{O}_4^+$ 592.2373; found: 592.2372.



Methyl 2-(((benzyloxy)carbonyl)amino)-3-(4-(2-(2,6-dimethylmorpholino)-1-fluoroethyl)phenyl)propanoate (10d). Synthesized using GP3 from **1ad** and *O*-benzoyl-2,6-dimethyl-hydroxylmorpholine **2h**. ^{19}F -NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 1:1. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 33% ethyl acetate–hexanes), as a yellowish oil (56.8 mg, 60%).

$R_f = 0.25$ (50% ethyl acetate–hexanes).

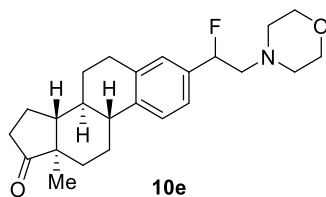
¹H NMR (CDCl₃, 500 MHz): δ 7.39–7.29 (m, 5H), 7.24 (d, *J* = 7.8 Hz, 2H), 7.10 (d, *J* = 7.8 Hz, 2H), 5.62 (ddd, *J* = 48.8, 8.7, 1.9 Hz, 1H), 5.22 (d, *J* = 6.8 Hz, 1H), 5.13–5.01 (m, 2H), 4.66 (dd, *J* = 13.5, 5.8 Hz, 1H), 3.83–3.61 (m, 2H), 3.72 (s, 3H), 3.21–3.02 (m, 2H), 2.90 (d, *J* = 11.0 Hz, 1H), 2.88–2.82 (m, 1H), 2.79 (d, *J* = 11.0 Hz, 1H), 2.57 (dd, *J* = 34.8, 14.3 Hz, 1H), 1.97–1.85 (m, 2H), 1.17 (d, *J* = 2.5 Hz, 3H), 1.16 (d, *J* = 2.5 Hz, 3H).

¹³C NMR (CDCl₃, 125 MHz): δ 171.8, 155.6, 137.7 (d, *J*_{C-F} = 20.1 Hz), 136.2, 136.1, 129.5, 128.6, 128.3, 128.1, 125.8 (d, *J*_{C-F} = 6.7 Hz), 92.4 (d, *J*_{C-F} = 173.7 Hz), 71.7, 71.6, 67.0, 64.5 (d, *J*_{C-F} = 23.0 Hz), 60.1, 59.6, 54.7, 52.4, 37.9, 37.8, 19.2, 19.1.

¹⁹F NMR (CDCl₃, 471 MHz): δ -176.68, -176.68.

FTIR (thin film): cm⁻¹ 3330, 2970, 2870, 1718, 1514, 1454, 1212, 1112, 886, 736, 698.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₂₆H₃₄FN₂O₅⁺ 473.2446; found: 473.2445.



3-(1-Fluoro-2-morpholinoethyl)-13-methyl-6,7,8,9,11,12,13,14,15,16-decahydro-17H-cyclopenta[a]phenanthren-17-one (10e). Synthesized using **GP3** from **1ae** and *O*-benzoylhydroxylmorpholine **2a**. Run the reaction at (0.1 mmol) scale. ¹⁹F-NMR analysis of the crude reaction mixture indicated a diastereomeric ratio (dr) of 1:1. Isolated by flash column chromatography (10% ethyl acetate–hexanes to 40% ethyl acetate–hexanes), as a yellowish oil (33.6 mg, 87%).

R_f = 0.25 (50% ethyl acetate–hexanes).

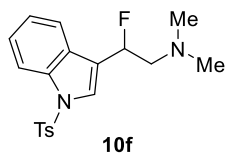
¹H NMR (CDCl₃, 500 MHz): δ 7.30 (d, *J* = 8.1 Hz, 1H), 7.12 (d, *J* = 8.1 Hz, 1H), 7.08 (s, 1H), 5.61 (ddd, *J* = 49.1, 8.9, 1.8 Hz, 1H), 3.75 (t, *J* = 4.6 Hz, 4H), 2.99–2.86 (m, 2H), 2.99–2.86 (m, 1H), 2.69–2.56 (m, 1H), 2.69–2.56 (m, 4H), 2.51 (dd, *J* = 18.1, 8.9 Hz, 1H), 2.41 (dd, *J* = 18.1, 10.1 Hz, 1H), 2.36–2.25 (m, 1H), 2.20–2.09 (m, 1H), 2.10–2.00 (m, 2H), 2.00–1.93 (m, 1H), 1.68–1.57 (m, 2H), 1.57–1.40 (m, 4H), 0.91 (s, 3H).

¹³C NMR (CDCl₃, 125 MHz): δ 220.7, 140.3, 140.2, 136.8, 136.3 (d, *J*_{C-F} = 19.7 Hz), 136.1 (d, *J*_{C-F} = 19.7 Hz), 126.3 (d, *J*_{C-F} = 6.5 Hz), 126.2 (d, *J*_{C-F} = 6.5 Hz), 125.6, 125.5, 123.1, 123.0, 92.6 (d, *J*_{C-F} = 173.2 Hz), 92.5 (d, *J*_{C-F} = 173.2 Hz), 67.0, 65.0 (d, *J*_{C-F} = 23.5 Hz), 64.9 (d, *J*_{C-F} = 23.5 Hz), 54.1, 50.5, 48.0, 44.4, 38.2, 38.1, 35.9, 31.6, 29.5, 29.4, 26.3, 25.7, 21.6, 13.9.

¹⁹F NMR (CDCl₃, 471 MHz): δ -175.26, -175.53.

FTIR (thin film): cm⁻¹ 2929, 2855, 1736, 1453, 1257, 1116, 1008, 869, 734.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₂₄H₃₃FNO₂⁺ 386.2490; found: 386.2489.



2-Fluoro-N,N-dimethyl-2-(1-tosyl-1H-indol-3-yl)ethan-1-amine (10f). Synthesized using **GP3** from **1af** and *O*-benzoyl-*N,N*-dimethylhydroxylamine **2p**. Isolated by flash column chromatography (20% ethyl acetate–hexanes to 50% ethyl acetate–hexanes), as a yellowish oil (20.9 mg, 26%).

R_f = 0.20 (50% ethyl acetate–hexanes).

¹H NMR (CDCl₃, 500 MHz): δ 7.97 (d, *J* = 8.3 Hz, 1H), 7.77 (d, *J* = 7.9 Hz, 2H), 7.62 (d, *J* = 2.3 Hz, 1H), 7.59 (d, *J* = 8.3 Hz, 1H), 7.34 (t, *J* = 7.7 Hz, 1H), 7.26 (t, *J* = 7.7 Hz, 2H), 7.23 (d, *J* = 7.9 Hz, 2H), 5.85 (ddd, *J* = 48.4, 8.5, 2.2 Hz, 1H), 3.05 (ddd, *J* = 17.0, 14.0, 8.6 Hz, 1H), 2.76 (ddd, *J* = 32.1, 14.0, 2.5 Hz,

1H), 2.40 (s, 6H), 2.35 (s, 3H).

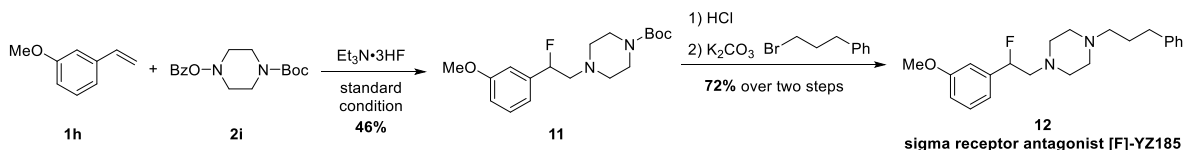
¹³C NMR (CDCl₃, 125 MHz): δ 145.2, 135.1 (d, *J*_{C-F} = 9.0 Hz), 130.0, 128.4 (d, *J*_{C-F} = 2.7 Hz), 126.9, 125.1, 123.7 (d, *J*_{C-F} = 9.3 Hz), 123.4, 120.2, 120.1, 120.0, 113.8, 87.3 (d, *J*_{C-F} = 169.8 Hz), 63.5 (d, *J*_{C-F} = 22.8 Hz), 46.0, 21.6.

¹⁹F NMR (CDCl₃, 471 MHz): δ -176.88.

FTIR (thin film): cm⁻¹ 2923, 2853, 1596, 1446, 1370, 1172, 1019, 974, 746.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₉H₂₂FN₂O₂S⁺ 361.1381; found: 361.1377.

7.3. Synthesis of the sigma receptor antagonist derivative [F]-YZ185



To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoyl-4-*tert*-butoxycarbonylhydroxylpiperazine **2i** (0.2 mmol, 1.0 equiv) and Cu(HFacac)₂ (2.4 mg, 2.5 mol%). DCE (1.0 mL), 1-methoxy-3-vinylbenzene **1h** (0.4 mmol, 2.0 equiv), and Et₃N·3HF (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was then stirred at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine, verified by TLC (20% ethyl acetate–hexanes) analysis. The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et₃N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al₂O₃ (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography (50% ethyl acetate–hexanes), giving **11** as yellowish oil (30.7 mg, 46%).

tert-Butyl 4-(2-fluoro-2-(3-methoxyphenyl)ethyl)piperazine-1-carboxylate (**11**).

R_f = 0.15 (25% ethyl acetate–hexanes).

¹H NMR (CDCl₃, 500 MHz): δ 7.28 (t, *J* = 7.8 Hz, 1H), 6.88 (d, *J* = 7.8 Hz, 2H), 6.78 (s, 1H), 5.62 (ddd, *J* = 48.9, 8.7, 2.2 Hz, 1H), 3.81 (s, 3H), 3.47 (t, *J* = 4.6 Hz, 4H), 2.91 (ddd, *J* = 17.6, 14.3, 8.7 Hz, 1H), 2.65 (ddd, *J* = 34.3, 14.3, 2.5 Hz, 1H), 2.60–2.49 (m, 4H), 1.46 (s, 9H).

¹³C NMR (CDCl₃, 125 MHz): δ 159.7, 154.7, 140.3 (d, *J*_{C-F} = 19.8 Hz), 129.6, 117.7 (d, *J*_{C-F} = 7.1 Hz), 113.9, 111.1 (d, *J*_{C-F} = 7.8 Hz), 92.6 (d, *J*_{C-F} = 174.6 Hz), 79.7, 64.6 (d, *J*_{C-F} = 23.0 Hz), 55.3, 53.4, 28.4.

¹⁹F NMR (CDCl₃, 471 MHz): δ -177.09.

FTIR (thin film): cm⁻¹ 2933, 1692, 1421, 1365, 1245, 1169, 1125, 1004, 868, 779, 698.

HRMS (ESI) m/z: [M+H]⁺ Calcd for C₁₈H₂₈FN₂O₃⁺ 339.2079; found: 339.2076.

To a 25-mL round-bottom flask was added *tert*-butyl 4-(2-fluoro-2-(3-methoxyphenyl)ethyl)piperazine-1-carboxylate **11** (33.8 mg, 0.1 mmol, 1.0 equiv), 1,4-dioxane (2.0 mL), HCl in 1,4-dioxane (4 M, 0.2 mL, 0.8 mmol, 8.0 equiv). The reaction was allowed to stir at room temperature for 8 h, and then concentrated under reduced pressure. The crude solid was used for the next step without further purification.

To a solution of the crude residue in DMSO (2 mL), was added potassium carbonate (27.6 mg, 0.2 mmol, 2.0 equiv), 1-bromo-3-phenylpropane (23.9 mg, 0.12 mmol, 1.2 equiv). The mixture was allowed to stir at room temperature for overnight. The reaction was quenched by the addition of water (2 mL). The mixture was extracted with ethyl acetate (5 mL × 3). The combined organic layers were washed with water (10 mL × 2) and the brine (10 mL). The organic layer was dried over Na₂SO₄, filtered and concentrated *in vacuo*. Purification by flash column chromatography (30% ethyl acetate–hexanes to 50% ethyl acetate–hexanes) gave **12** as colorless oil (25.6 mg, 72% over two steps).

1-(2-Fluoro-2-(3-methoxyphenyl)ethyl)-4-(3-phenylpropyl)piperazine (12).

R_f = 0.15 (50% ethyl acetate–hexanes).

$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.28 (dd, J = 8.5, 7.0 Hz, 3H), 7.19 (d, J = 7.1 Hz, 3H), 6.94–6.80 (m, 3H), 5.62 (ddd, J = 49.1, 8.8, 1.9 Hz, 1H), 3.81 (s, 3H), 2.91 (ddd, J = 17.6, 14.3, 8.9 Hz, 1H), 2.78–2.46 (m, 1H), 2.78–2.46 (m, 8H), 2.78–2.46 (m, 2H), 2.45–2.34 (m, 2H), 1.84 (q, J = 7.7 Hz, 2H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 159.7, 142.1, 140.5 (d, $J_{\text{C-F}}$ = 19.7 Hz), 129.6, 128.4, 128.3, 125.8, 117.8 (d, $J_{\text{C-F}}$ = 6.7 Hz), 113.9, 111.1 (d, $J_{\text{C-F}}$ = 7.5 Hz), 92.6 (d, $J_{\text{C-F}}$ = 174.4 Hz), 64.6 (d, $J_{\text{C-F}}$ = 22.8 Hz), 58.0, 55.3, 53.6, 53.1, 33.7, 28.5.

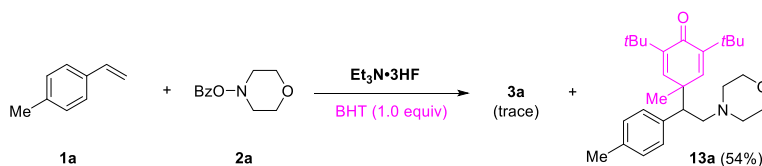
$^{19}\text{F NMR}$ (CDCl_3 , 471 MHz): δ -177.01.

FTIR (thin film): cm^{-1} 2934, 2809, 1699, 1602, 1493, 1454, 1267, 1158, 1030, 784, 748.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{22}\text{H}_{30}\text{FN}_2\text{O}^+$ 357.2337; found: 357.2337.

8. Mechanism studies

8.1. Control experiment in the presence of 2,6-di-*tert*-butyl-4-methylphenol (BHT)



To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylmorpholine **2a** (41.4 mg, 0.2 mmol, 1.0 equiv), 2,6-di-*tert*-butyl-4-methylphenol (BHT, 44.1 mg, 0.2 mmol, 1.0 equiv) and $\text{Cu}(\text{HFacac})_2$ (2.4 mg, 2.5 mol %). DCE (1.0 mL), 4-methylstyrene **1a** (0.4 mmol, 2.0 equiv) and $\text{Et}_3\text{N}\cdot 3\text{HF}$ (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et_3N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al_2O_3 (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography (20% ethyl acetate–hexanes), giving **13a** as yellow oil (46.1 mg, 54%).

2,6-Di-*tert*-butyl-4-methyl-4-(2-morpholino-1-(*p*-tolyl)ethyl)cyclohexa-2,5-dien-1-one (13a).

R_f = 0.31 (33% ethyl acetate–hexanes).

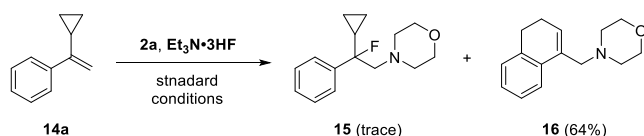
$^1\text{H NMR}$ (CDCl_3 , 500 MHz): δ 7.08 (d, J = 7.9 Hz, 2H), 6.98 (d, J = 7.9 Hz, 2H), 6.56 (d, J = 2.8 Hz, 1H), 6.43 (d, J = 2.8 Hz, 1H), 3.61–3.50 (m, 4H), 2.92 (dd, J = 8.1, 4.5 Hz, 1H), 2.53 (dd, J = 12.9, 8.1 Hz, 1H), 2.48 (dd, J = 12.9, 4.5 Hz, 1H), 2.32 (s, 3H), 2.28–2.24 (m, 4H), 1.26 (s, 9H), 1.15 (s, 9H), 1.07 (s, 3H).

$^{13}\text{C NMR}$ (CDCl_3 , 125 MHz): δ 186.3, 146.5, 146.4, 144.3, 137.3, 136.4, 129.2, 128.5, 66.9, 60.1, 53.9, 52.5, 42.4, 34.9, 34.8, 29.5, 29.4, 25.2, 21.0.

FTIR (thin film): cm^{-1} 1656, 1637, 1456, 1362, 1249, 1118, 910, 868, 734.

HRMS (ESI) m/z : $[\text{M}+\text{H}]^+$ Calcd for $\text{C}_{28}\text{H}_{42}\text{NO}_2^+$ 424.3210; found: 424.3215.

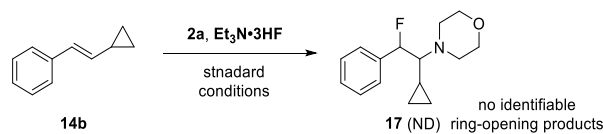
8.2. Radical clock experiments



To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylmorpholine **2a** (41.4 mg, 0.2 mmol, 1.0 equiv) and $\text{Cu}(\text{HFacac})_2$ (2.4 mg, 2.5 mol %). DCE (1.0 mL), (1-cyclopropylvinyl)benzene **14a** (0.4 mmol, 2.0 equiv), and $\text{Et}_3\text{N}\cdot\text{3HF}$ (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et_3N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al_2O_3 (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was purified by silica column chromatography (10% ethyl acetate–hexanes), giving **16** as clear oil (29.4 mg, 64%)

4-((3,4-Dihydronaphthalen-1-yl)methyl)morpholine (**16**).

^1H NMR (CDCl_3 , 500 MHz): δ 7.57 (d, $J = 7.6$ Hz, 1H), 7.22–7.11 (m, 3H), 6.02 (t, $J = 4.2$ Hz, 1H), 3.75–3.66 (m, 4H), 3.30 (s, 2H), 2.76 (t, $J = 8.0$ Hz, 2H), 2.48 (br s, 4H), 2.35–2.26 (m, 2H). Spectroscopic data match a previous report.⁸

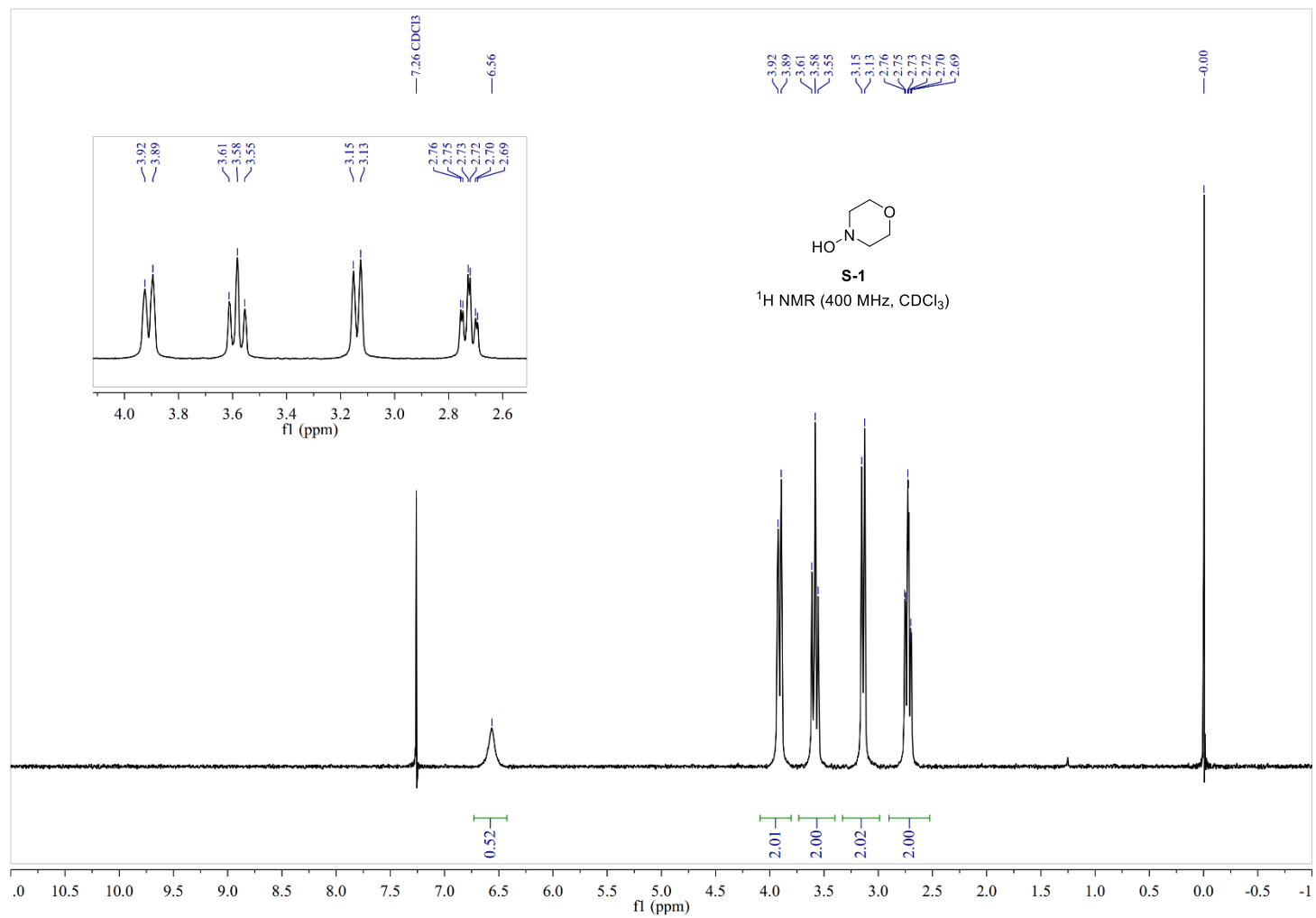


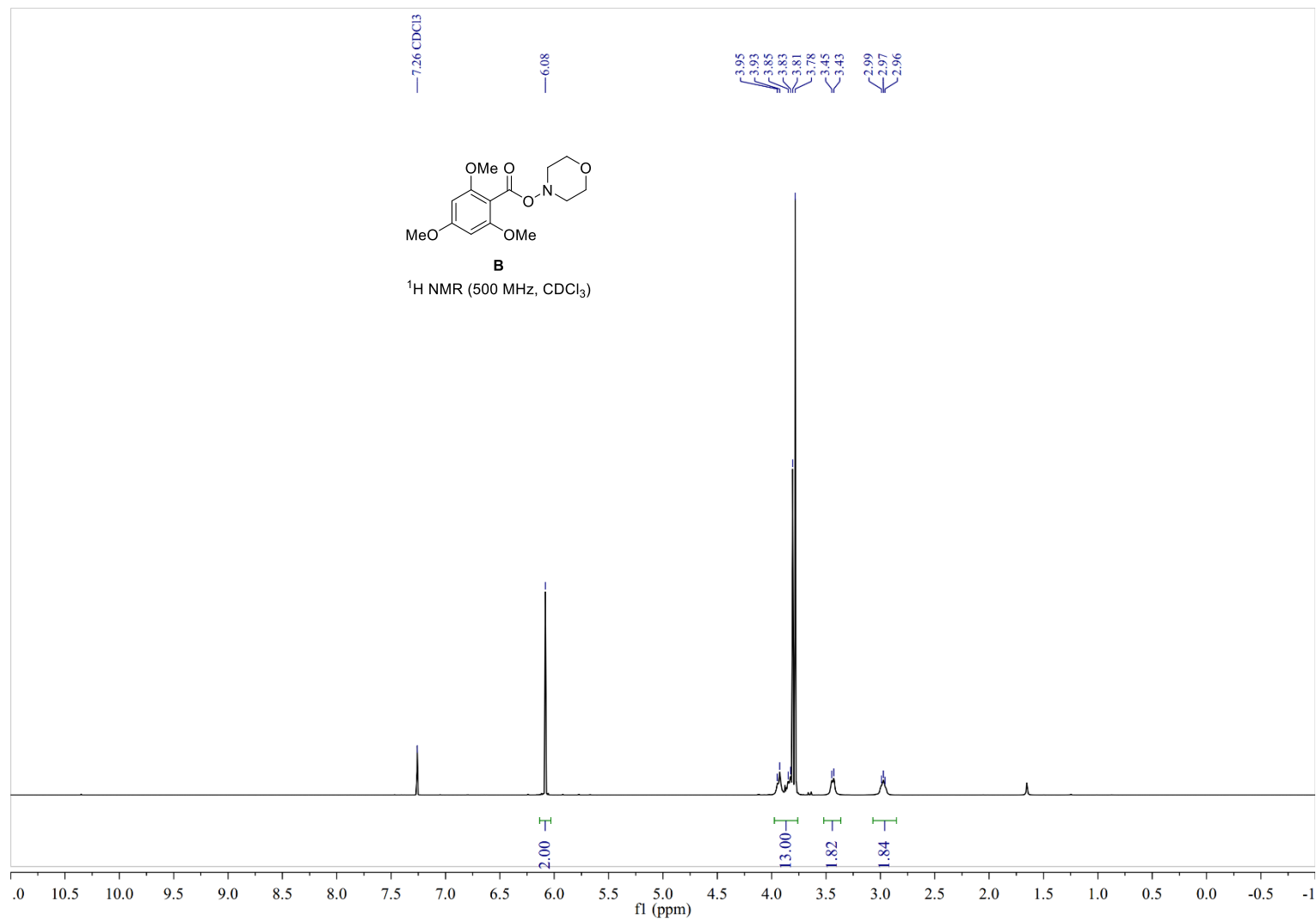
To a 10-mL FEP tube with Teflon-coated micro stir bar was added *O*-benzoylhydroxylmorpholine **2a** (41.4 mg, 0.2 mmol, 1.0 equiv) and $\text{Cu}(\text{HFacac})_2$ (2.4 mg, 2.5 mol %). DCE (1.0 mL), (*E*)-(2-cyclopropylvinyl)benzene **14b** (0.4 mmol, 2.0 equiv), and $\text{Et}_3\text{N}\cdot\text{3HF}$ (322 mg, 2 mmol, 10 equiv) were sequentially added. The mixture was allowed to stir at 80 °C for 2 h until the consumption of *O*-benzoylhydroxylamine (verified by TLC, 20% ethyl acetate–hexanes). The resulting reaction mixture was cooled to room temperature and quenched through the addition of Et_3N (0.5 mL). The solution was then diluted with ethyl acetate to a final volume of 5.0 mL and filtered through a plug of activated, neutral Al_2O_3 (Brockman grade I, 58–60Å). The filtrate was concentrated under reduced pressure, providing the crude reaction mixture. The crude reaction mixture was analyzed by LC-MS, ^1H NMR and ^{19}F NMR spectrometry, none of which shows identifiable ring-opening product **17**.

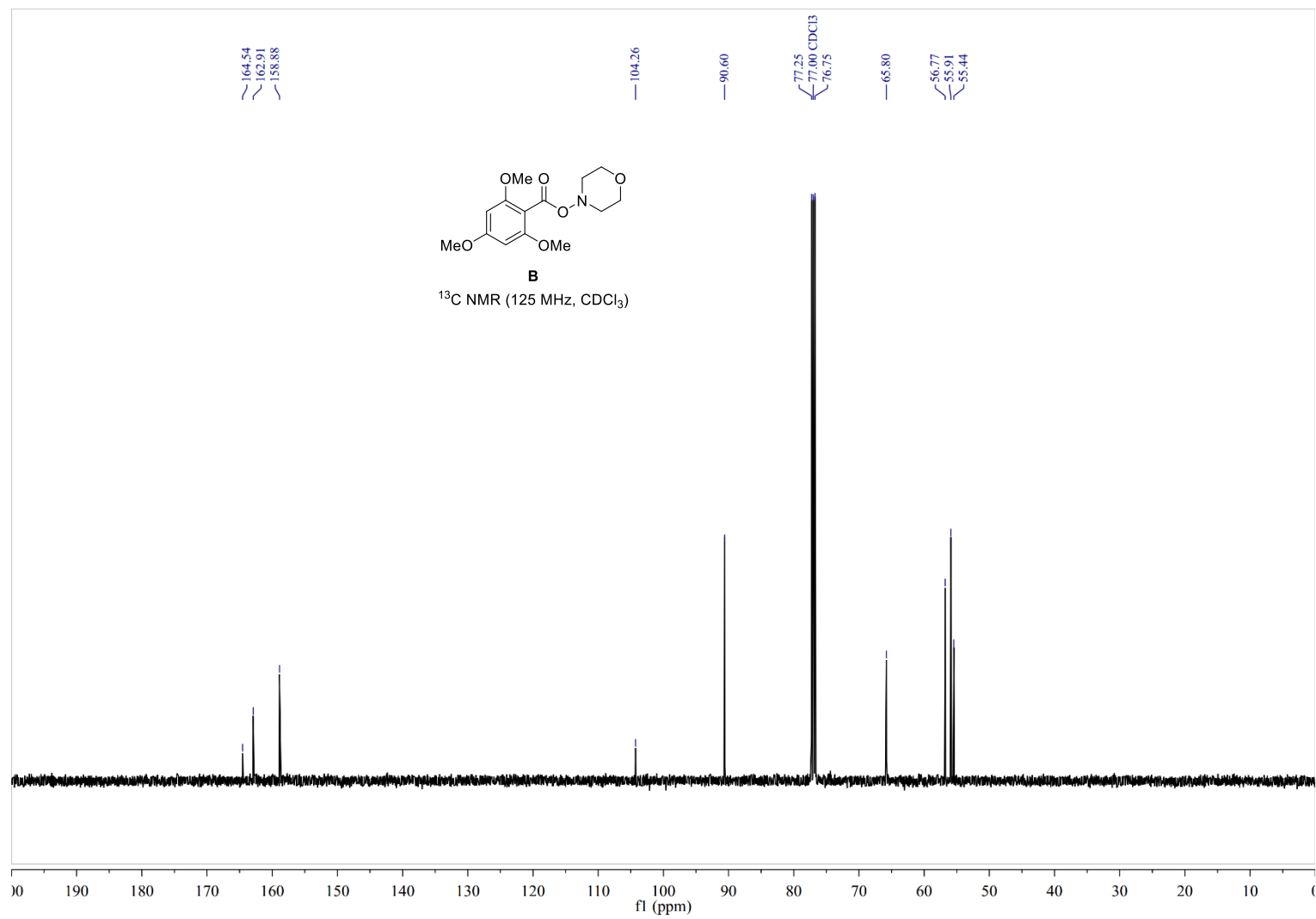
9. References

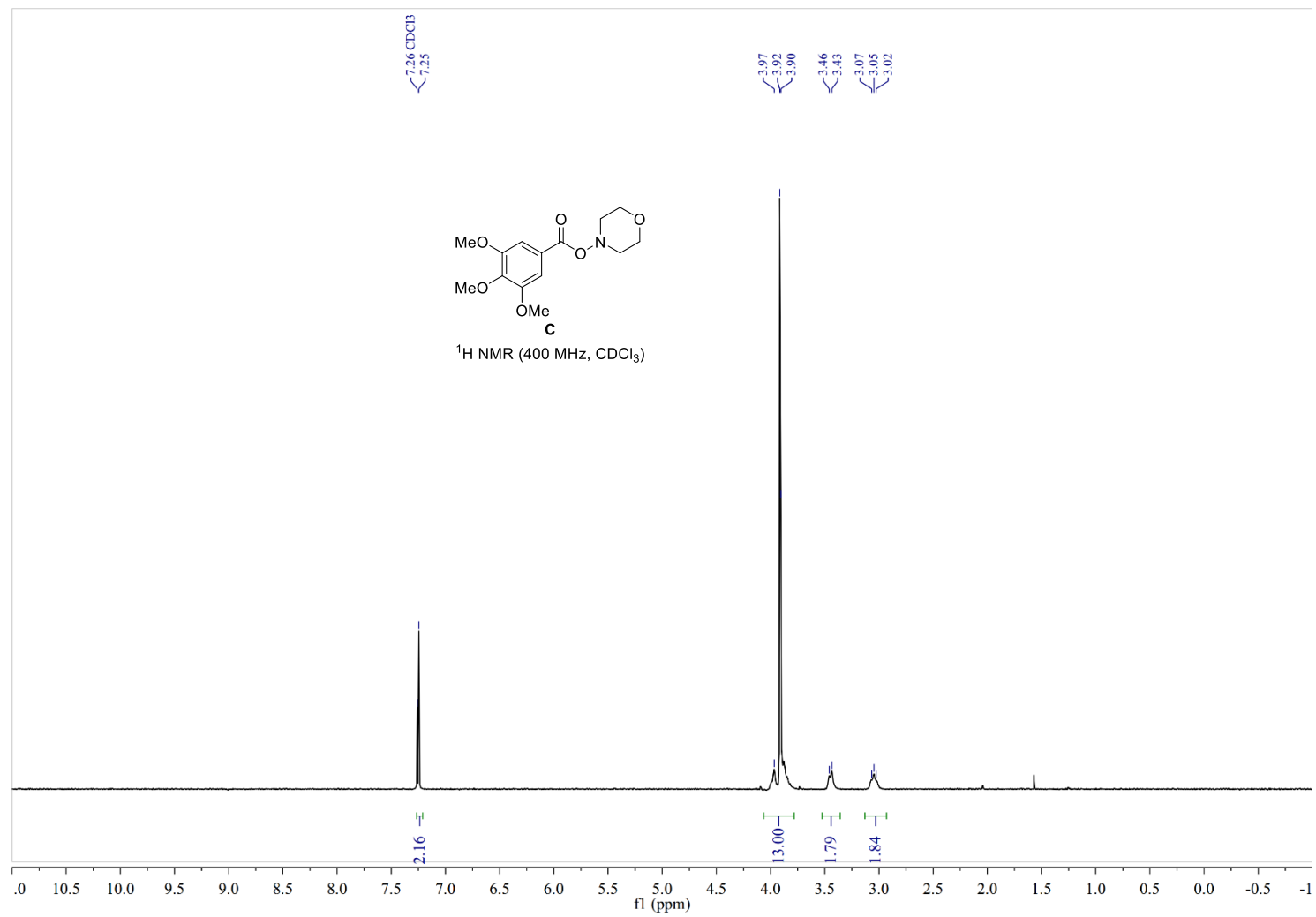
- 1 Kang, T.; Kim, N.; Cheng, P. T.; Zhang, H.; Foo, K.; Engle, K. M. Nickel-Catalyzed 1, 2-Carboamination of Alkenyl Alcohols. *J. Am. Chem. Soc.* **2021**, *143*, 13962–13970.
- 2 An, Y.; Zhang, B.-S.; Zhang, Z.; Liu, C.; Gou, X.-Y.; Ding, Y.-N.; Liang, Y.-M. A Carboxylate-Assisted Amination/Unactivated C (Sp²)-H Arylation Reaction via a Palladium/Norbornene Cooperative Catalysis. *Chem. Commun.* **2020**, *56*, 5933–5936.
- 3 Kokare, N. D.; Nagawade, R. R.; Rane, V. P.; Shinde, D. B., Design, Synthesis and Utilization of a Novel Coupling Reagent for the Preparation of *O*-Alkyl Hydroxamic Acids. *Tetrahedron Lett.* **2007**, *48*, 4437–4440.
- 4 Svejstrup, T. D.; Ruffoni, A.; Juliá, F.; Aubert, V. M.; Leonori, D., Synthesis of Arylamines via Aminium Radicals. *Angew. Chem. Int. Ed.* **2017**, *56*, 14948–14952.
- 5 (a) Biloski, A. J.; Ganem, B., Improved Oxidation of Amines with Dibenzoyl Peroxide. *Synthesis* **1983**, 537–538. (b) Berman, A. M.; Johnson, J. S. Copper-Catalyzed Electrophilic Amination of Organozinc Nucleophiles: Documentation of *O*-Benzoyl Hydroxylamines as Broadly Useful R₂N(+) and RHN(+) Synthons. *J. Org. Chem.* **2006**, *71*, 219–224. (c) Dhanju, S.; Crich, D., Synthesis of *N*, *N*, *O*-Trisubstituted Hydroxylamines by Stepwise Reduction and Substitution of *O*-Acyl *N*, *N*-Disubstituted Hydroxylamines. *Org. Lett.* **2016**, *18*, 1820–1823. (d) Shi, H.; Babinski, D. J.; Ritter, T., Modular C–H Functionalization Cascade of Aryl Iodides. *J. Am. Chem. Soc.* **2015**, *137*, 3775–3778. (e) Fan, L.; Liu, J.; Bai, L.; Wang, Y.; Luan, X., Rapid Assembly of Diversely Functionalized Spiroindenes by a Three-Component Palladium-Catalyzed C–H Amination/Phenol Dearomatization Domino Reaction. *Angew. Chem. Int. Ed.* **2017**, *56*, 14257–14261. (f) Zhu, S.; Buchwald, S. L., Enantioselective CuH-Catalyzed Anti-Markovnikov Hydroamination of 1, 1-Disubstituted Alkenes. *J. Am. Chem. Soc.* **2014**, *136*, 15913–15916.
- 6 Yu, S.; Noble, A.; Bedford, R. B.; Aggarwal, V. K. Methylene-spiro[2.3]hexanes via Nickel-Catalyzed Cyclopropanations with [1.1.1] Propellane. *J. Am. Chem. Soc.* **2019**, *141*, 20325–20334.
- 7 Kurandina, D.; Parasram, M.; Gevorgyan, V. Visible Light-Induced Room-Temperature Heck Reaction of Functionalized Alkyl Halides with Vinyl Arenes/Heteroarenes. *Angew. Chem. Int. Ed.* **2017**, *56*, 14212–14216.
- 8 Hemric, B. N.; Shen, K.; Wang, Q., Copper-Catalyzed Amino Lactonization and Amino Oxygenation of Alkenes Using *O*-Benzoylhydroxylamines. *J. Am. Chem. Soc.* **2016**, *138*, 5813–5816.

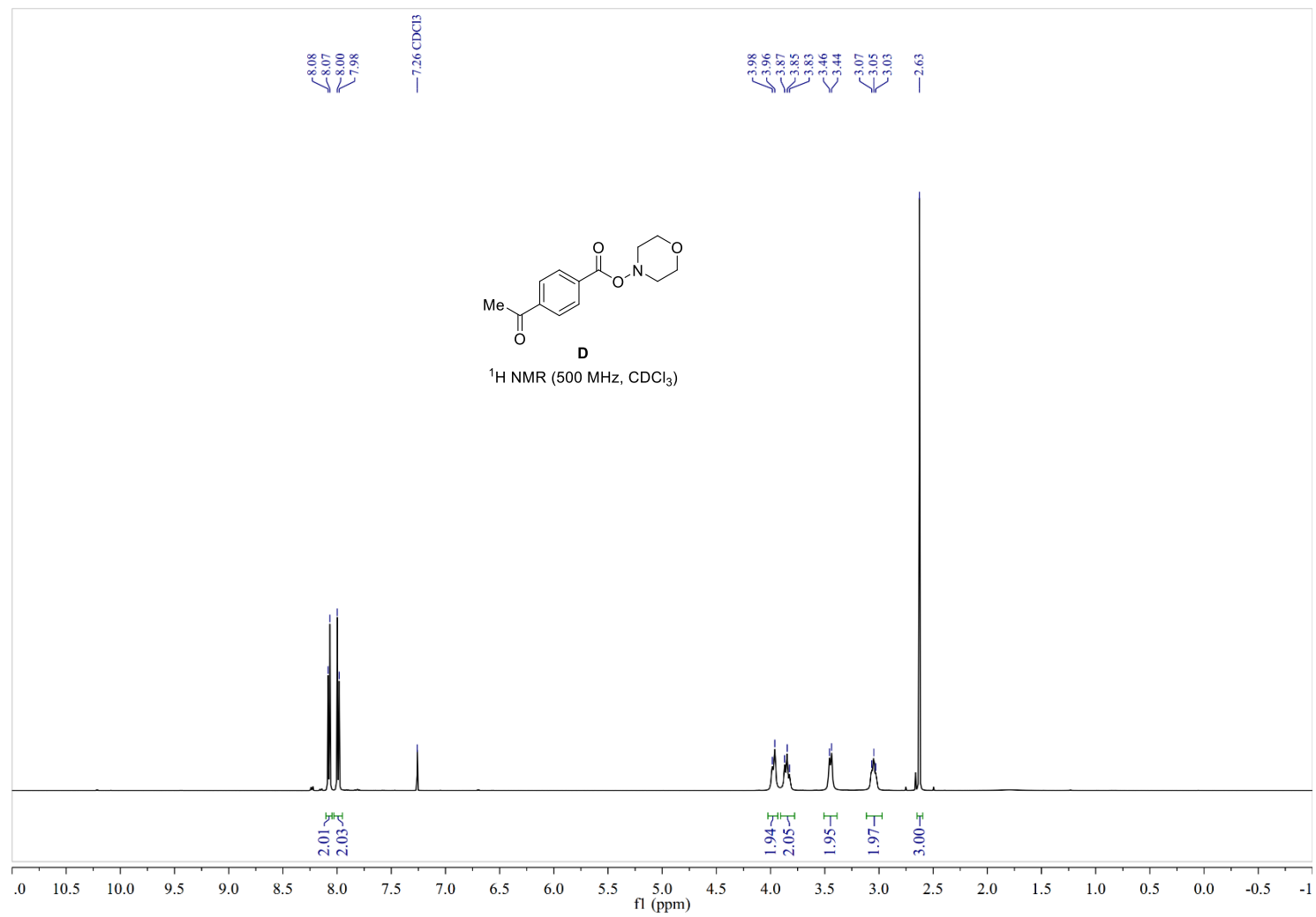
10. Copies of the NMR Spectra

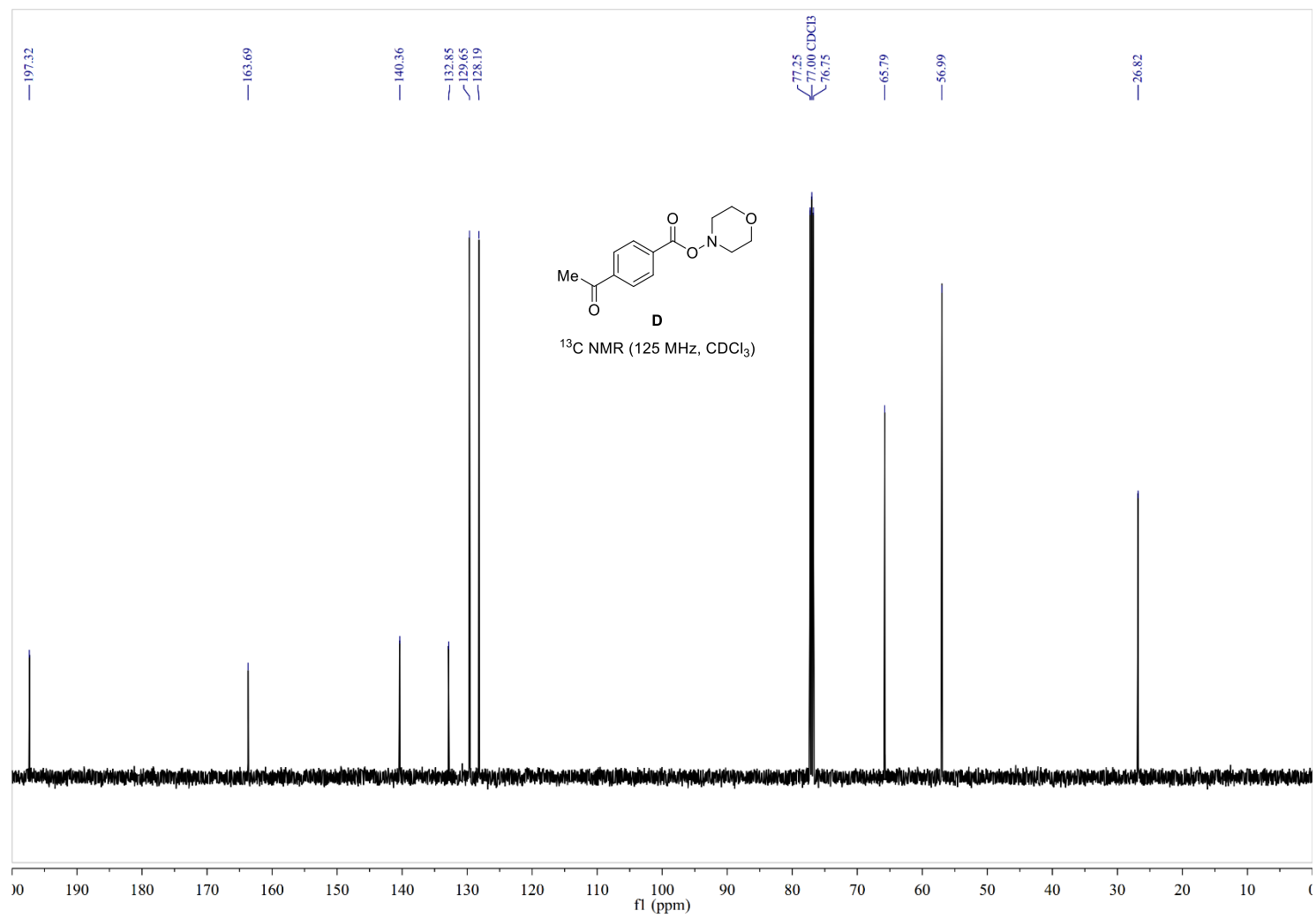


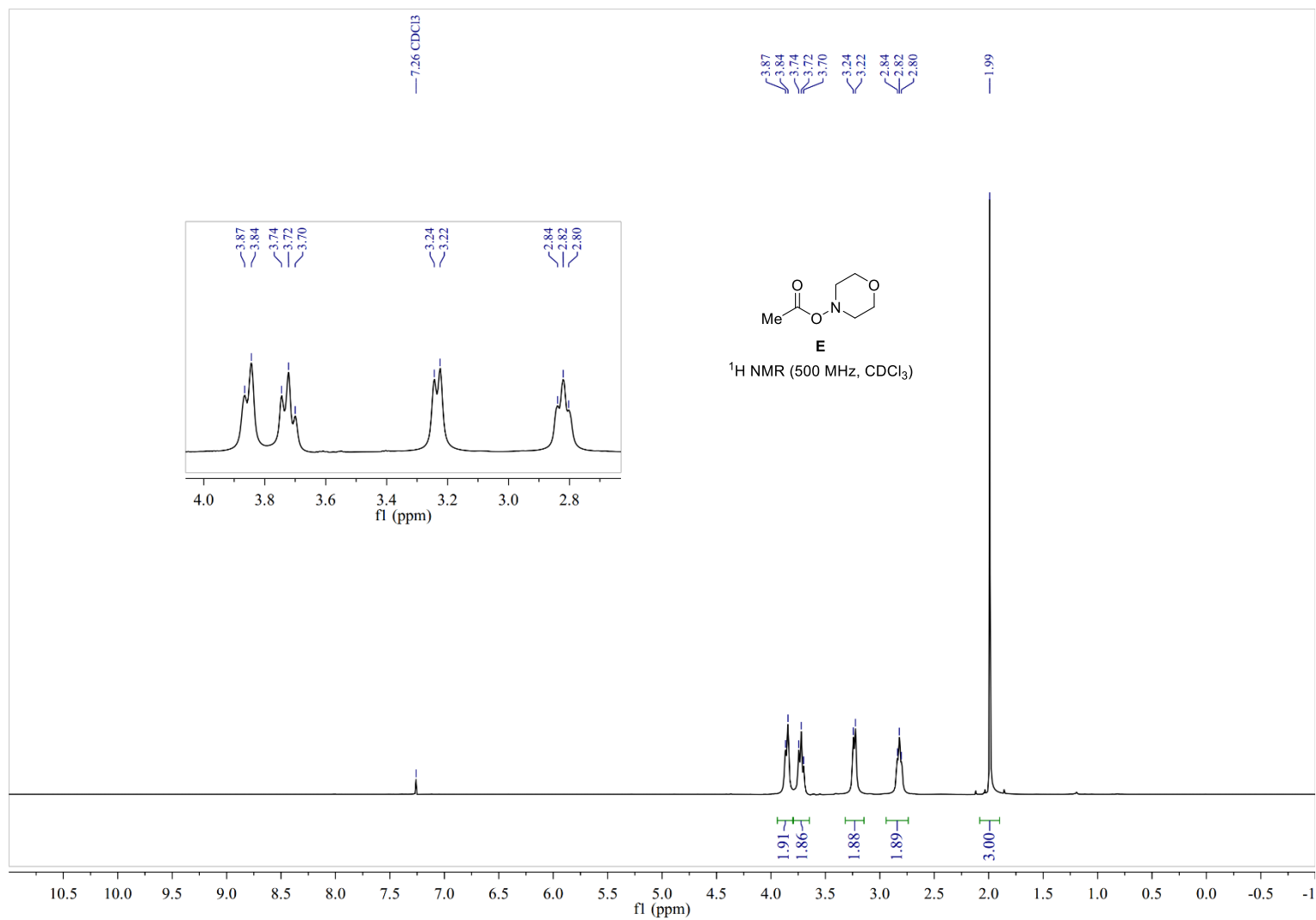


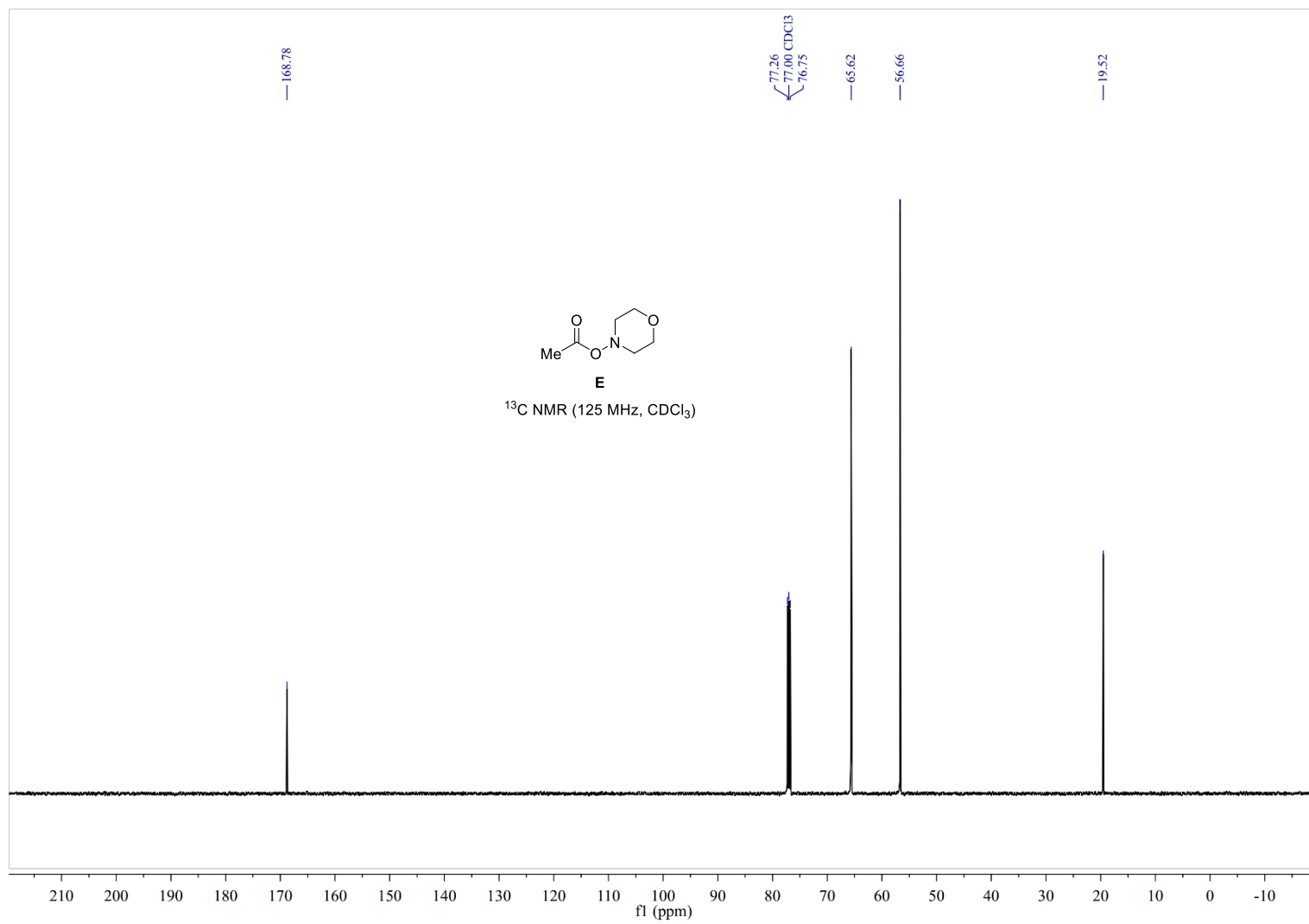


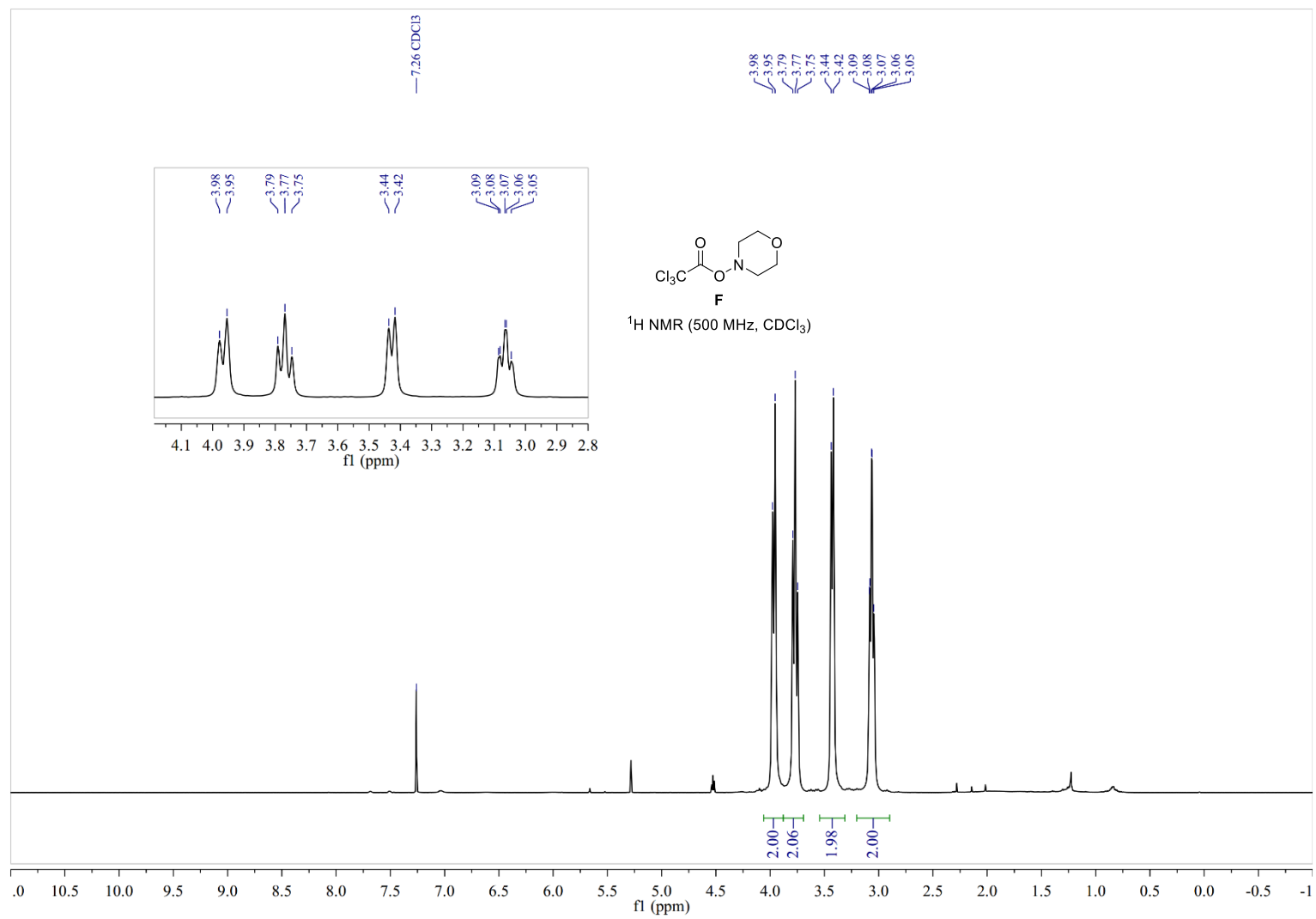


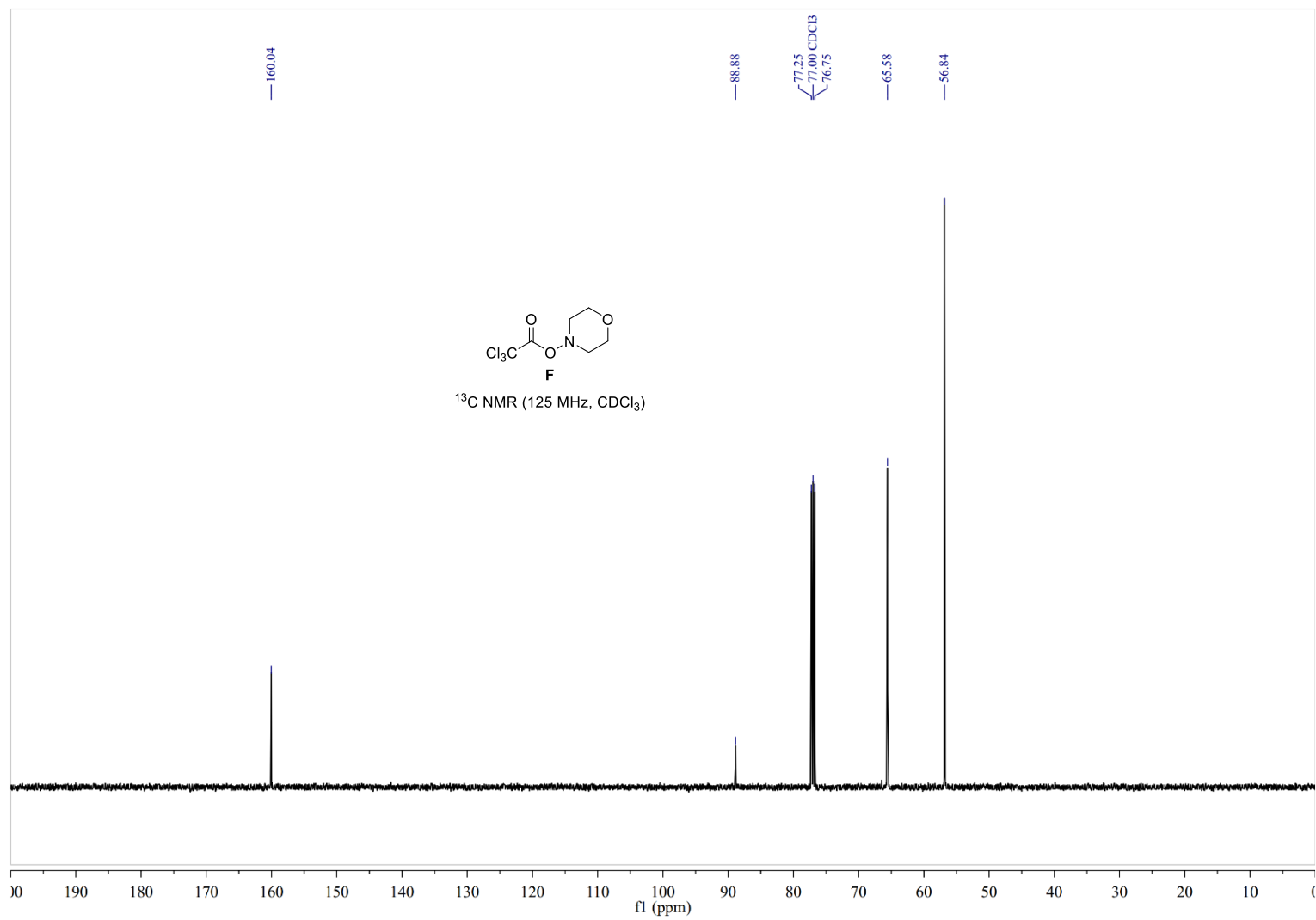


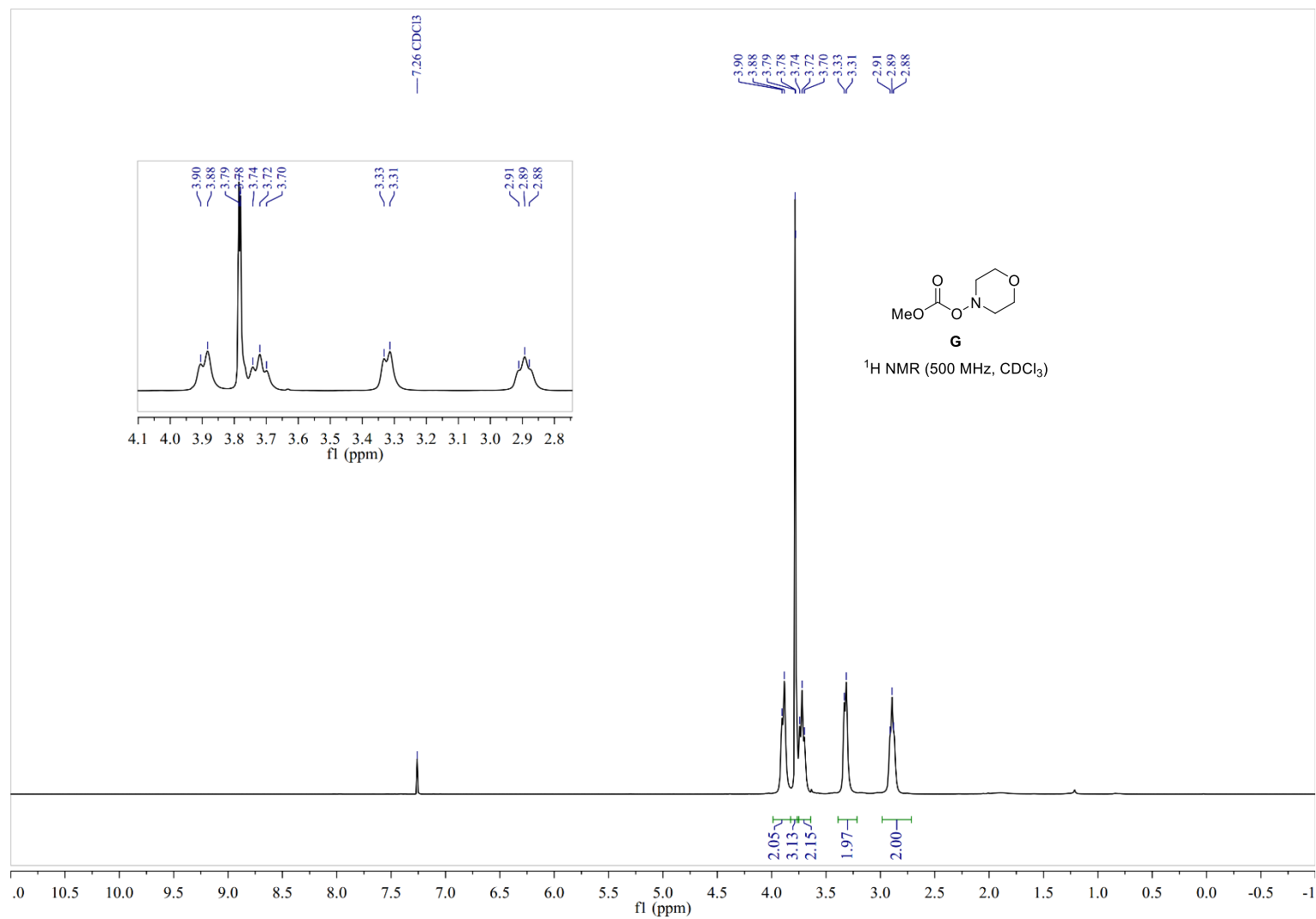


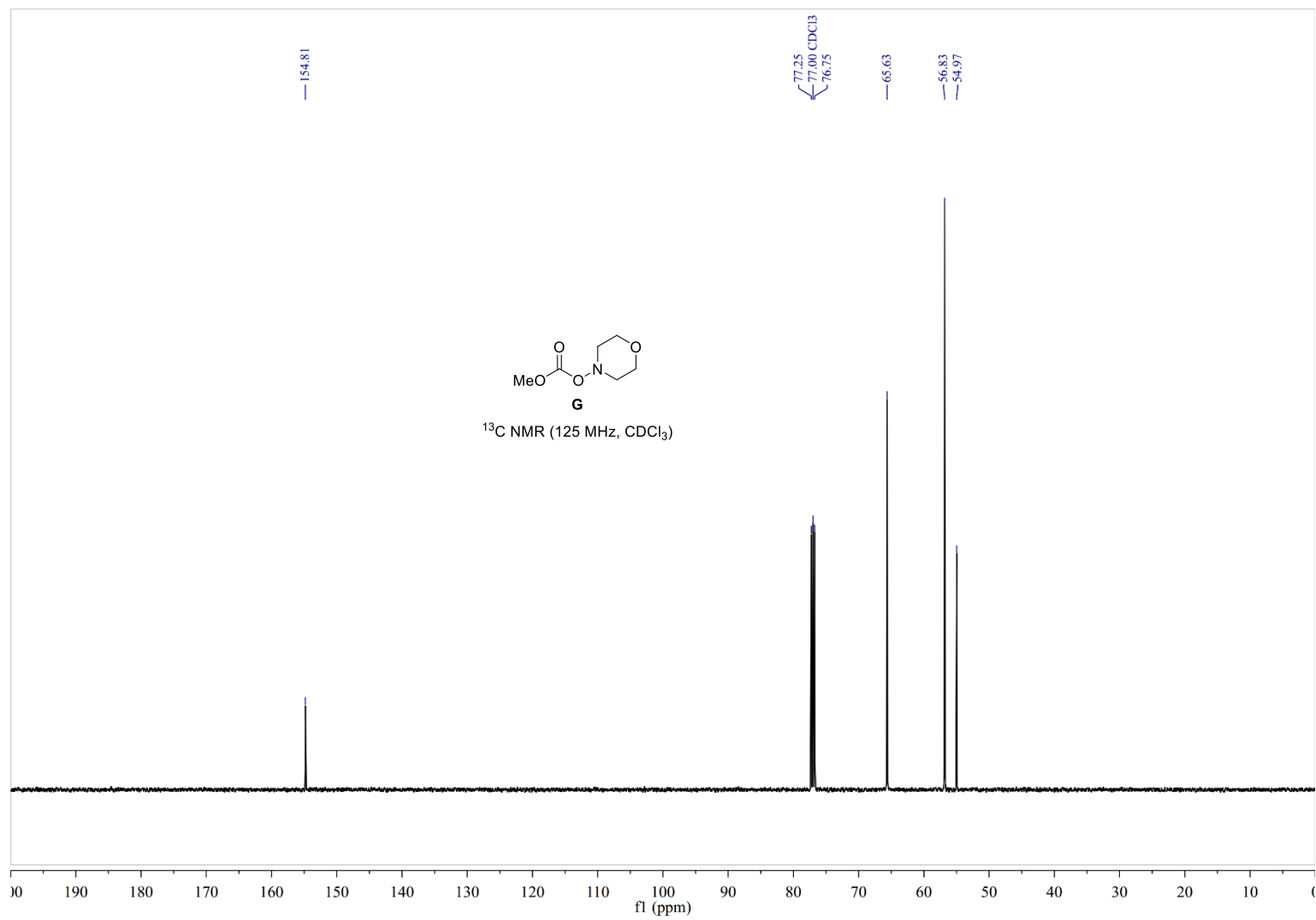


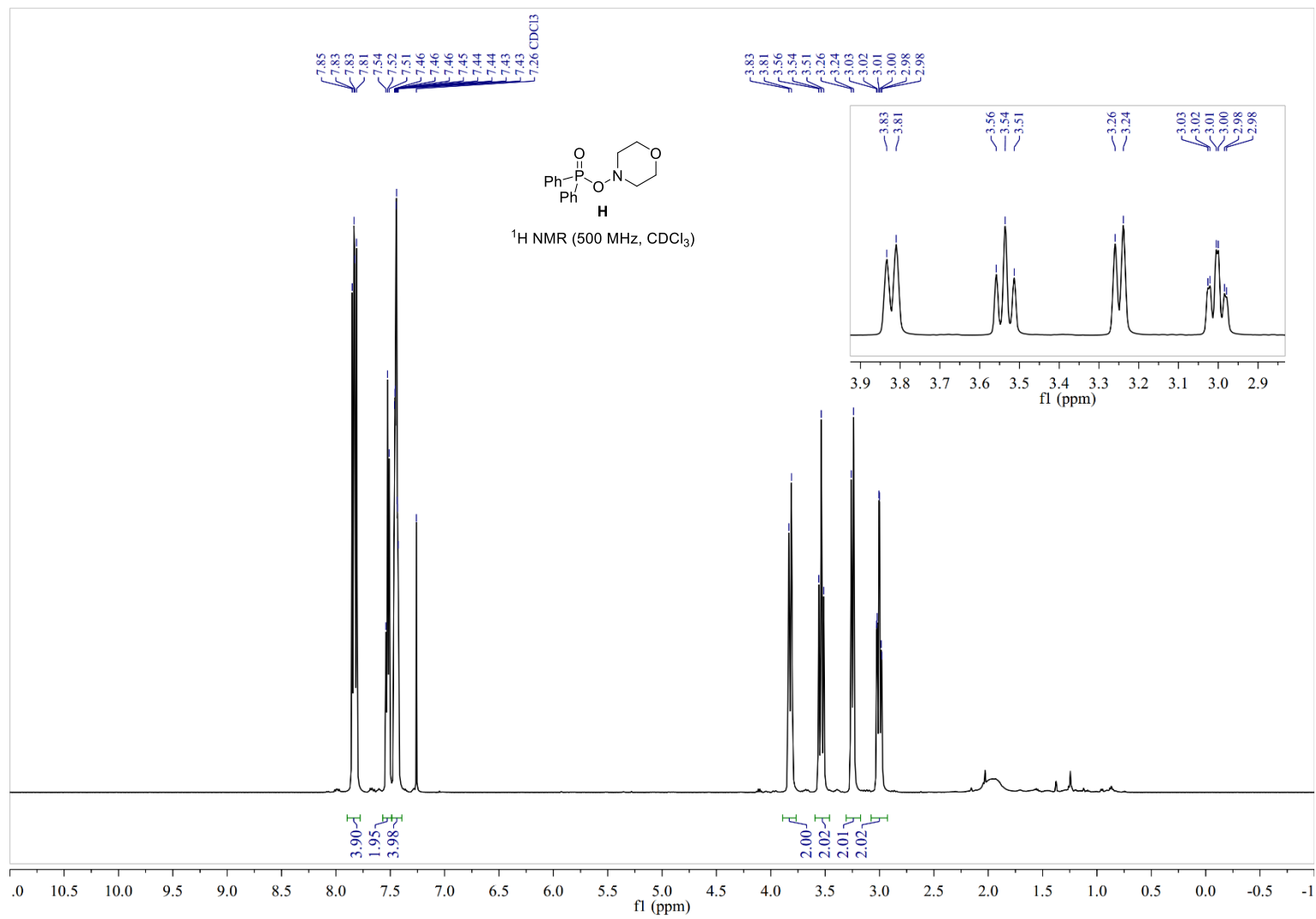


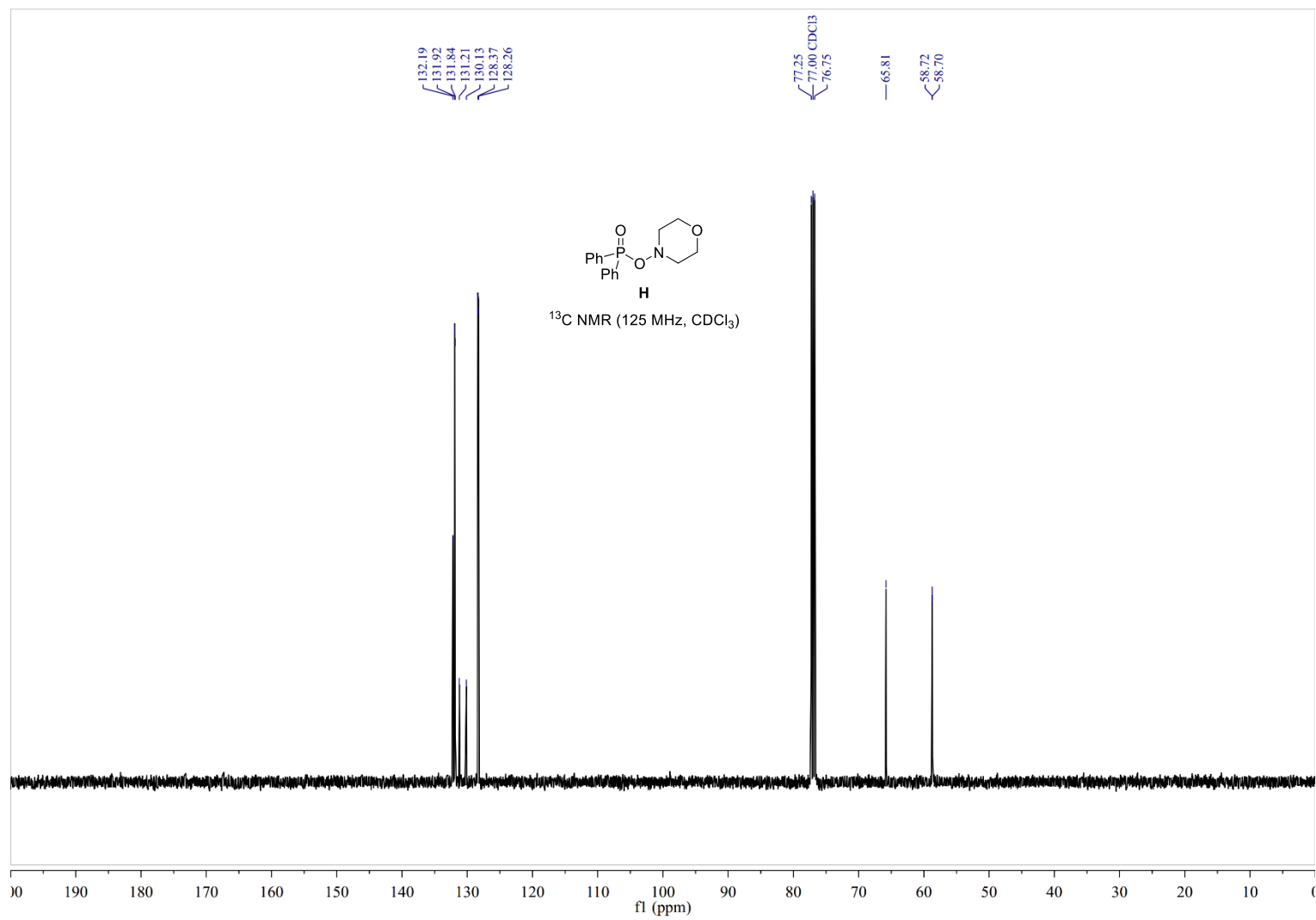


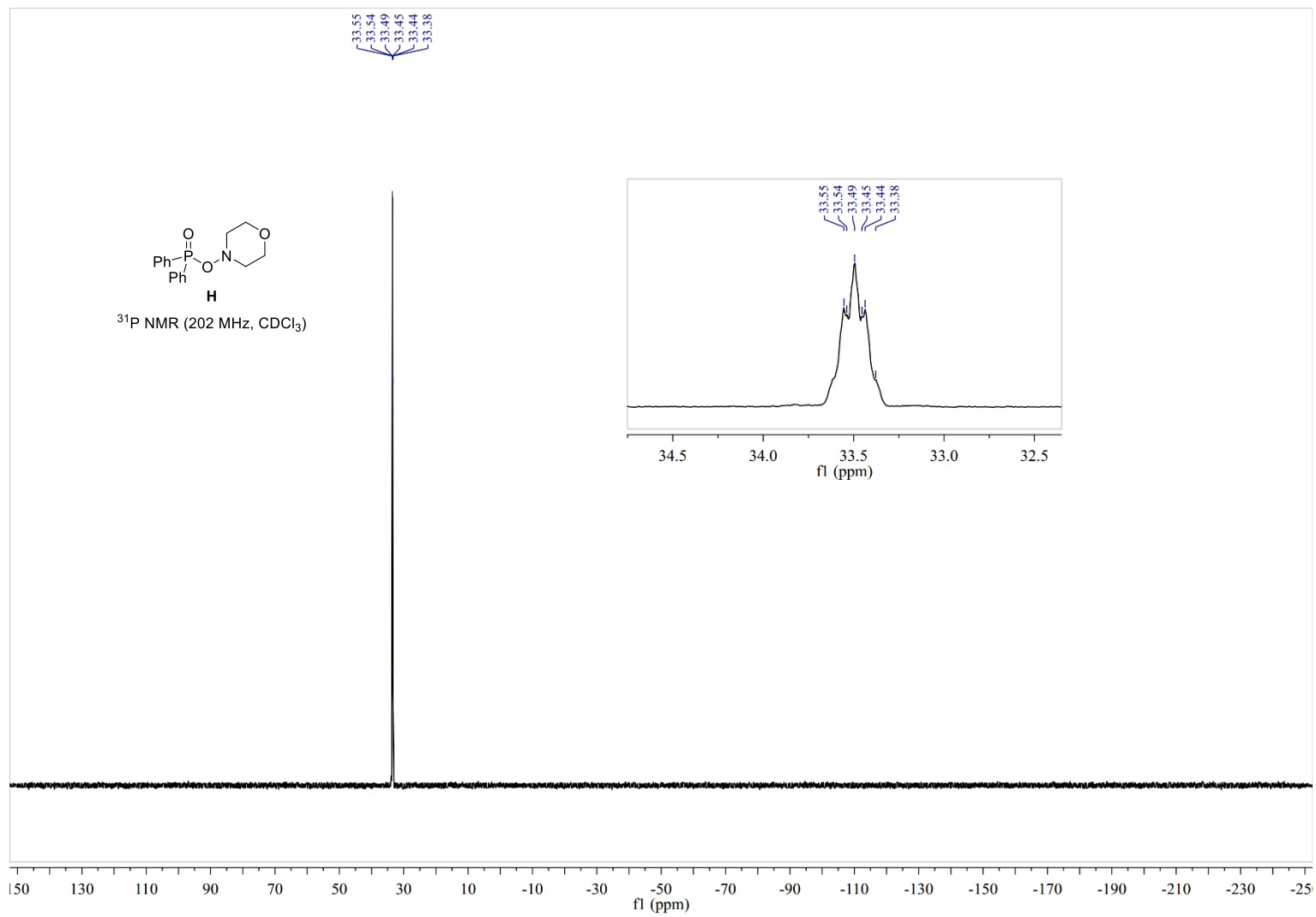






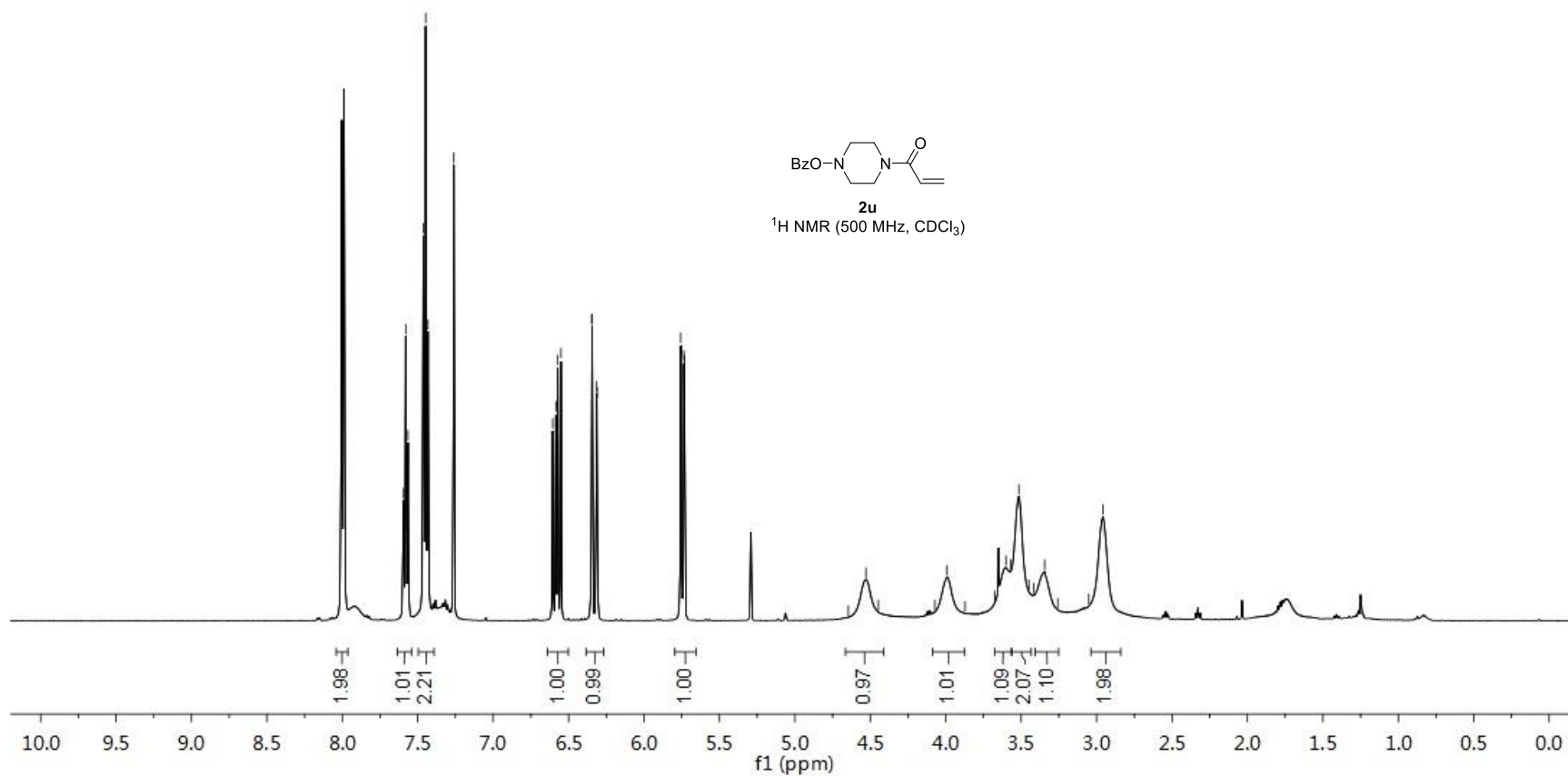


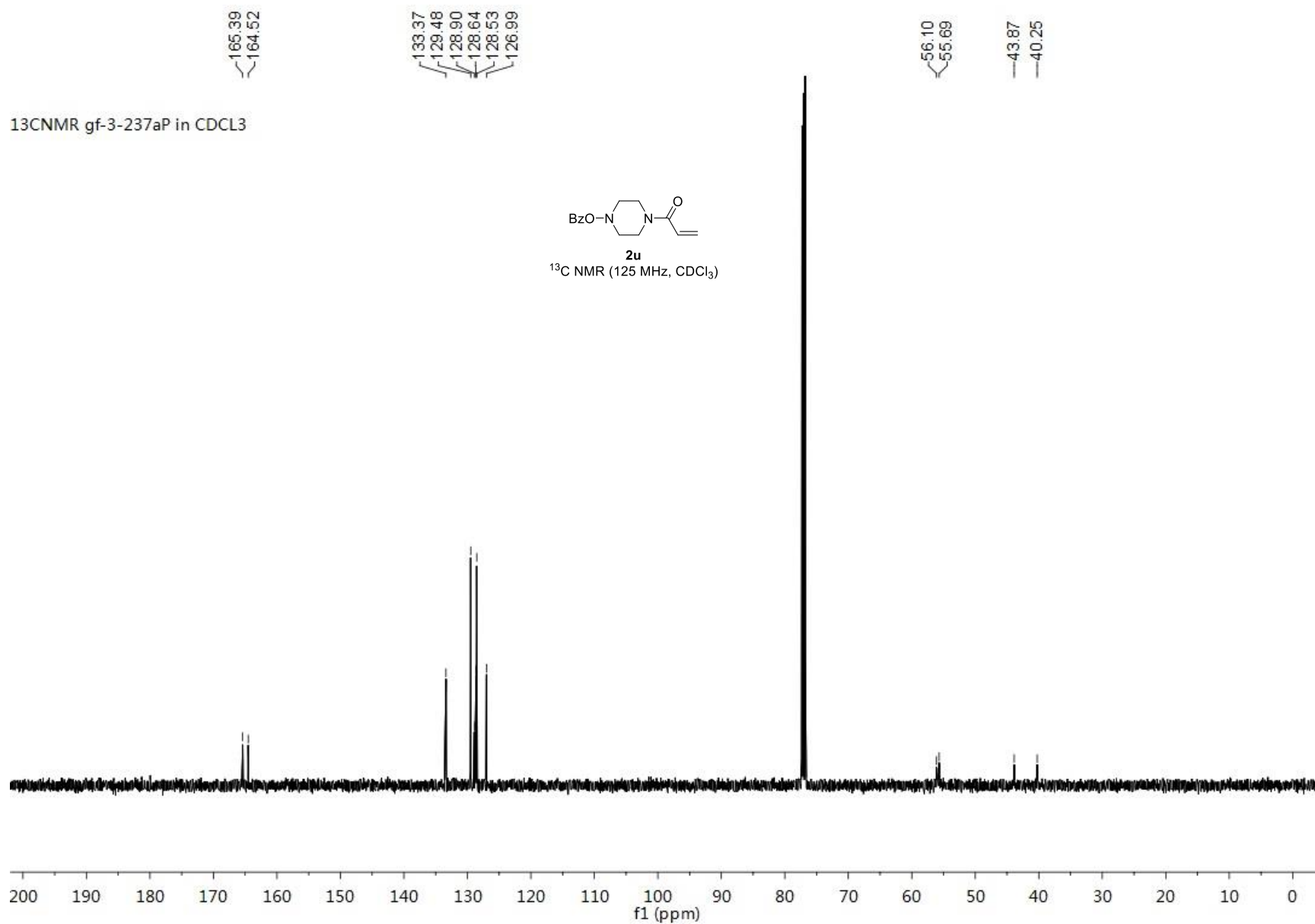


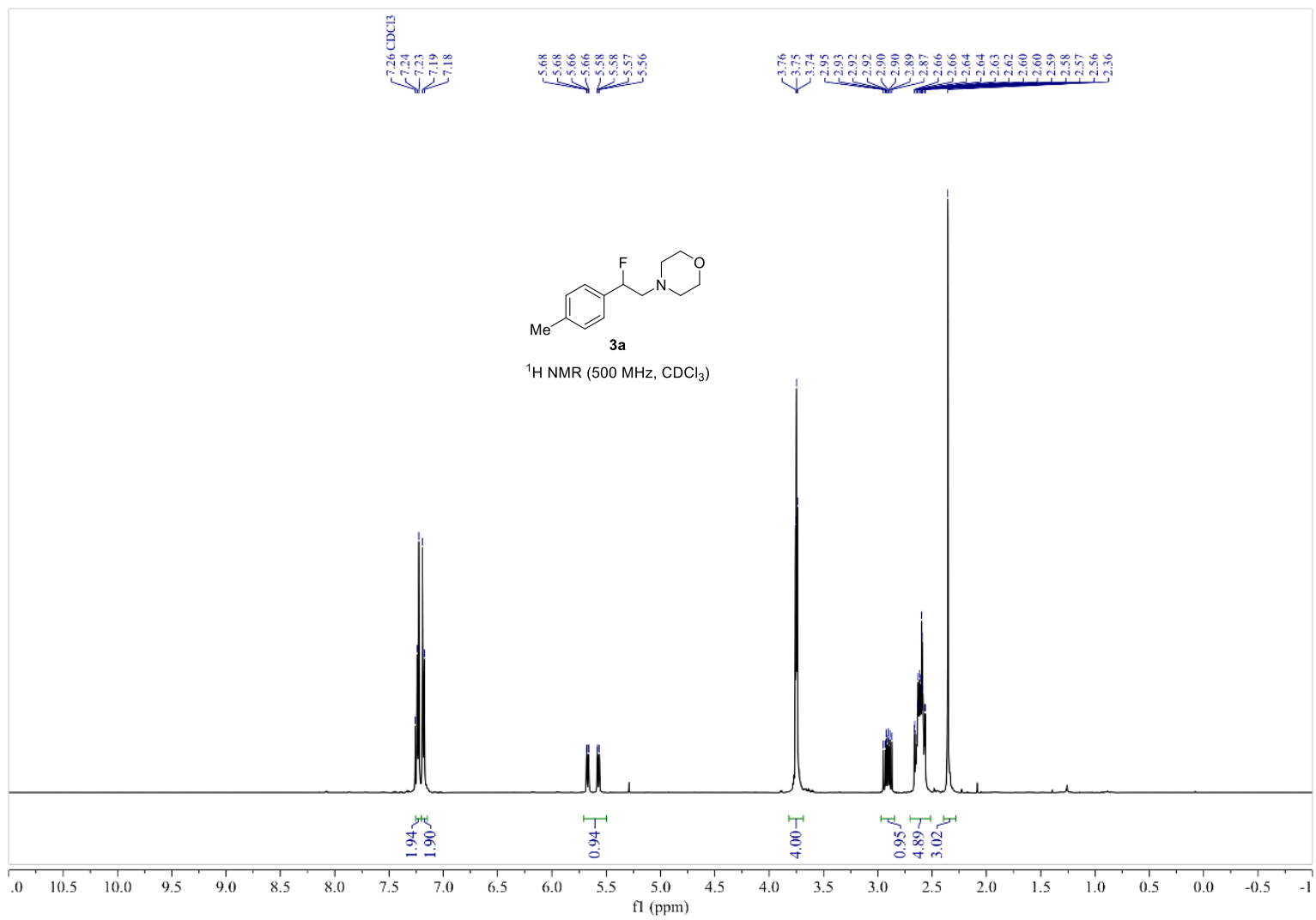


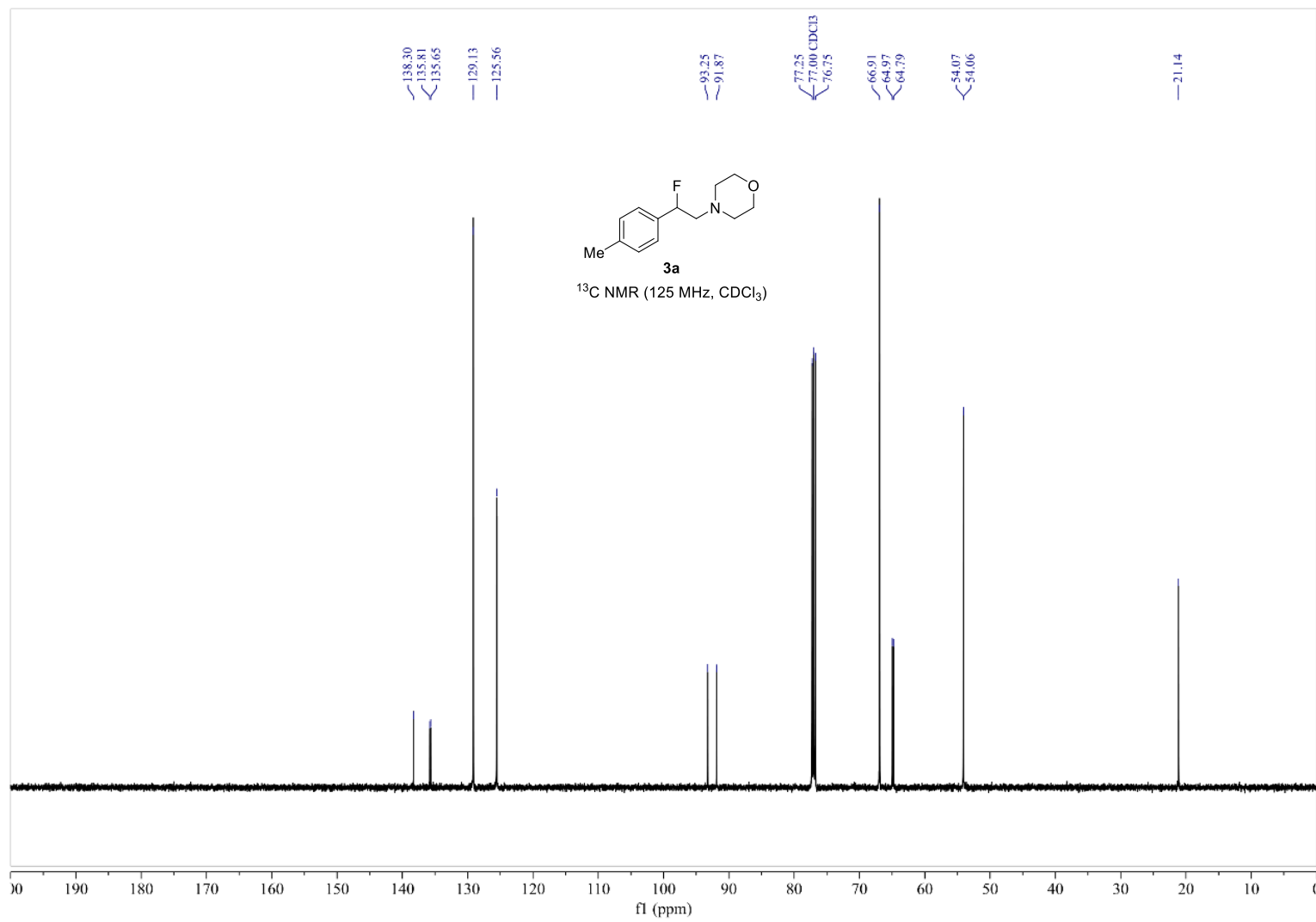
7.5942
7.5793
7.5644
7.4626
7.4470
7.4316
7.2601
6.5719
6.5507
6.3471
6.3435
6.3135
6.3089
6.3067
5.7532
5.7356
5.7321
4.6465
4.5294
4.4459
4.0723
3.9936
3.8721
3.6753
3.6008
3.5677
3.5150
3.4485
3.4169
3.3444
3.2569
3.0516
2.9578

¹H NMR of gf-3-237aP in CDCl₃



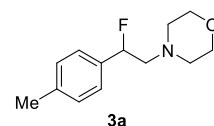




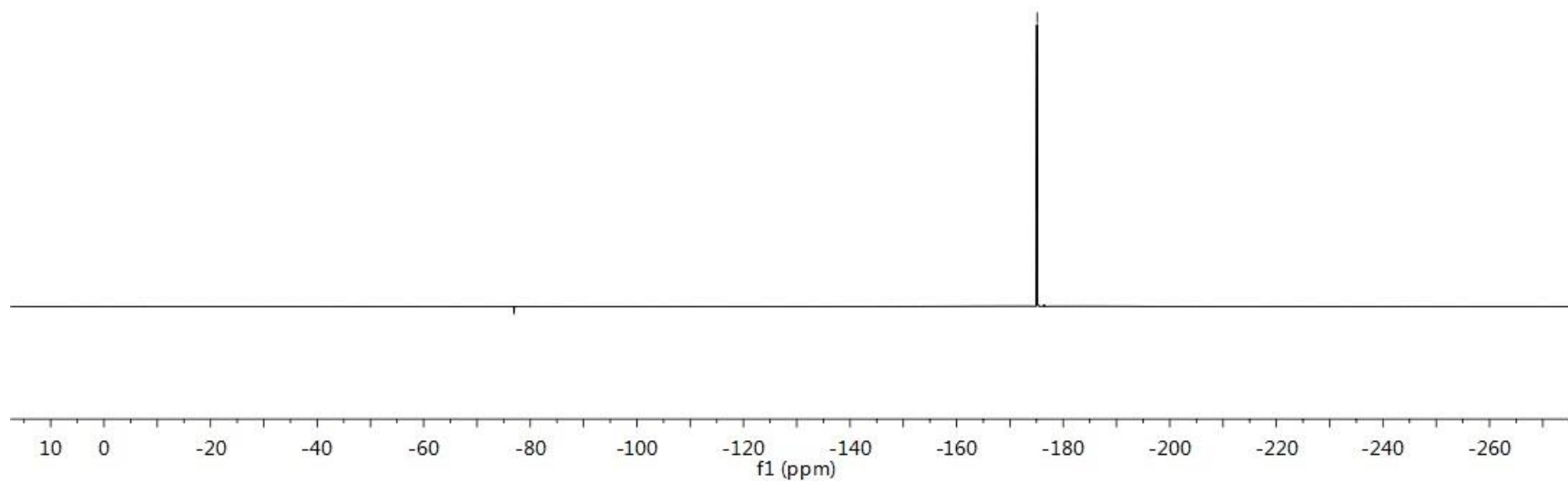


¹⁹F NMR gf-3-32a P in CDCl₃

---175.1088



¹⁹F NMR (471 MHz, CDCl₃)

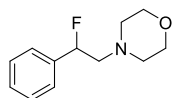


7.3240
7.3087
7.2954
7.2775
7.2631
7.2498
7.1893

5.6557
5.6512
5.6381
5.6336
5.5579
5.5534
5.5403
5.5358

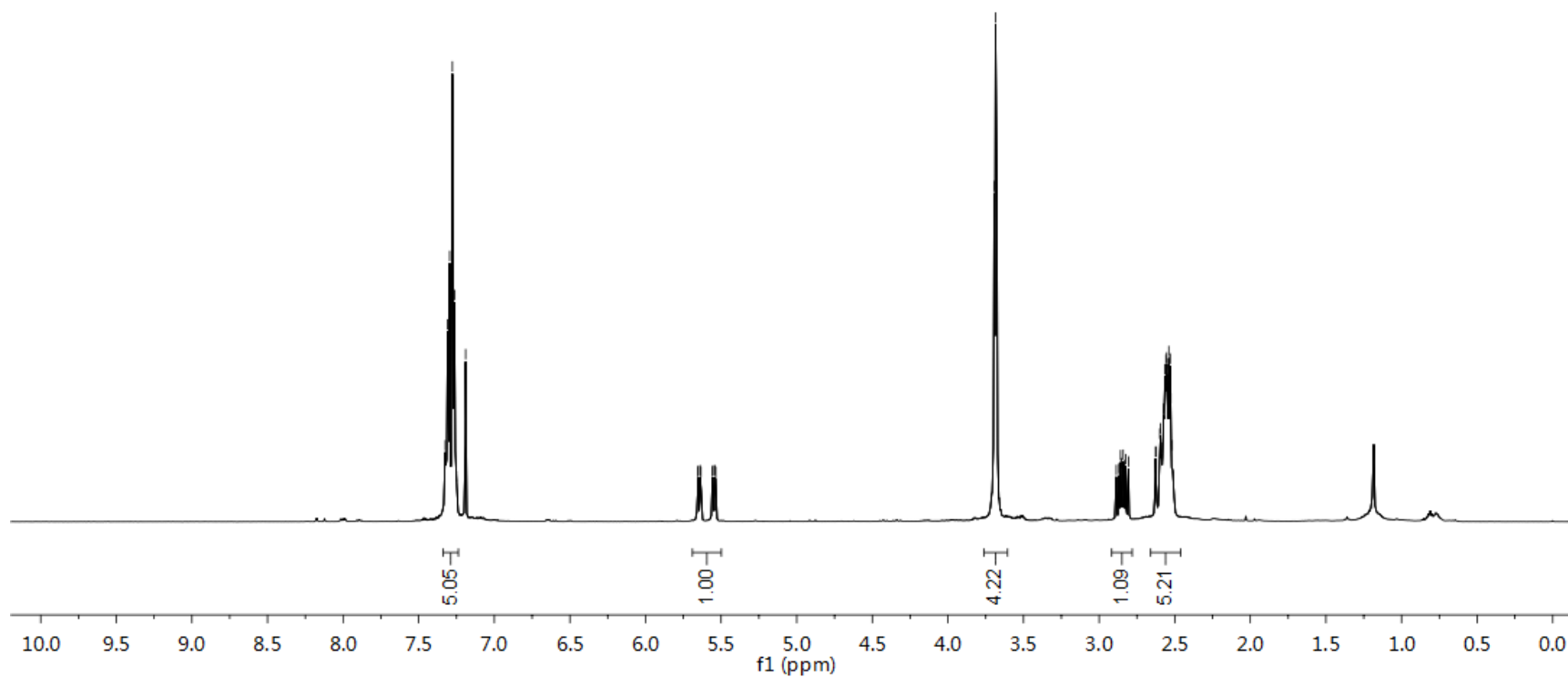
3.6945
3.6852
3.6759
2.8869
2.8693
2.8584
2.8522
2.8408
2.8345
2.8237
2.8061
2.6272
2.6223
2.5986
2.5938
2.5881
2.5736
2.5647
2.5585
2.5536
2.5492
2.5392
2.5300
2.5256
2.5165
2.5075

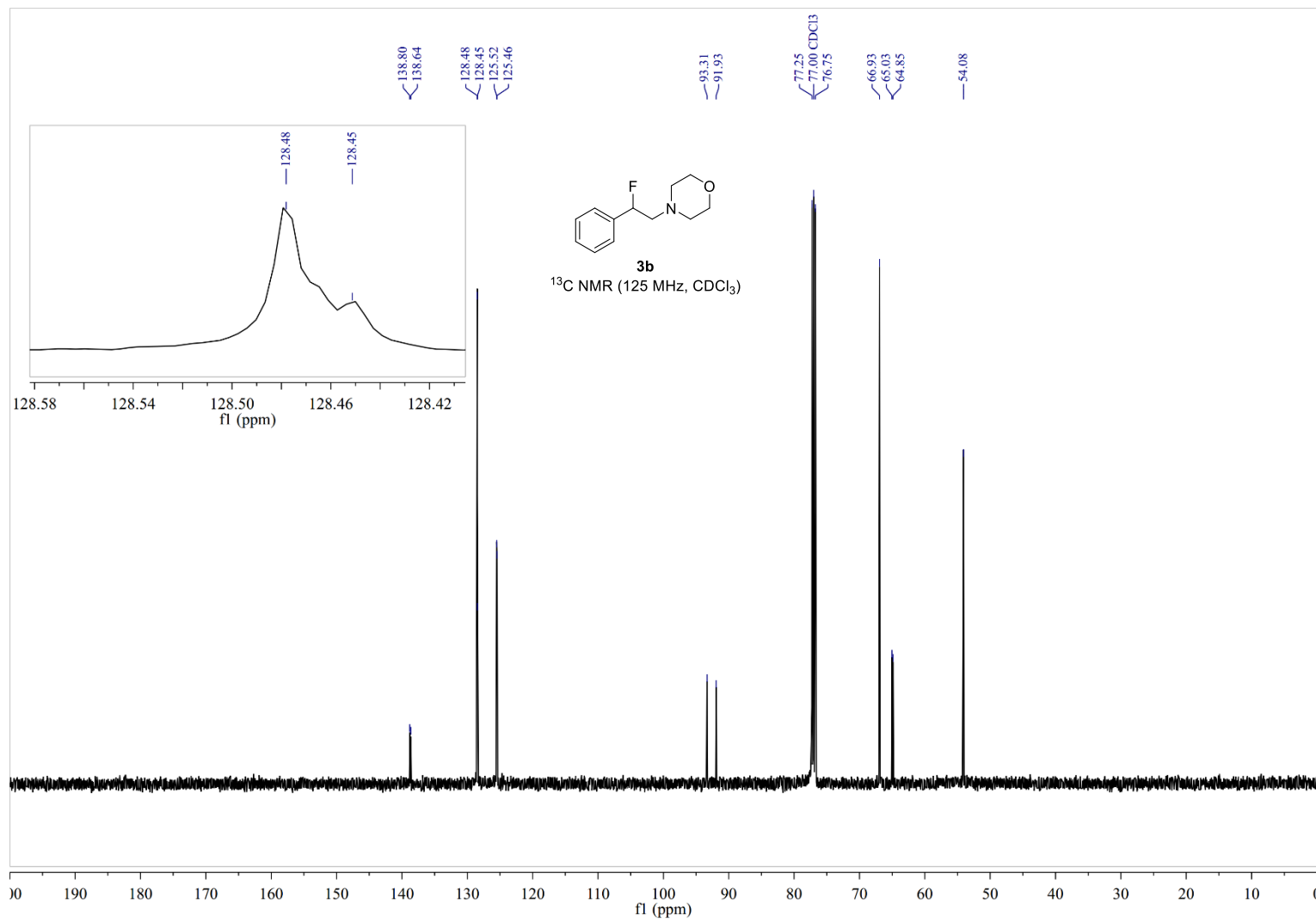
¹H NMR of gf-ck-II-56aP in CDCl₃



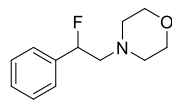
3b

¹H NMR (500 MHz, CDCl₃)





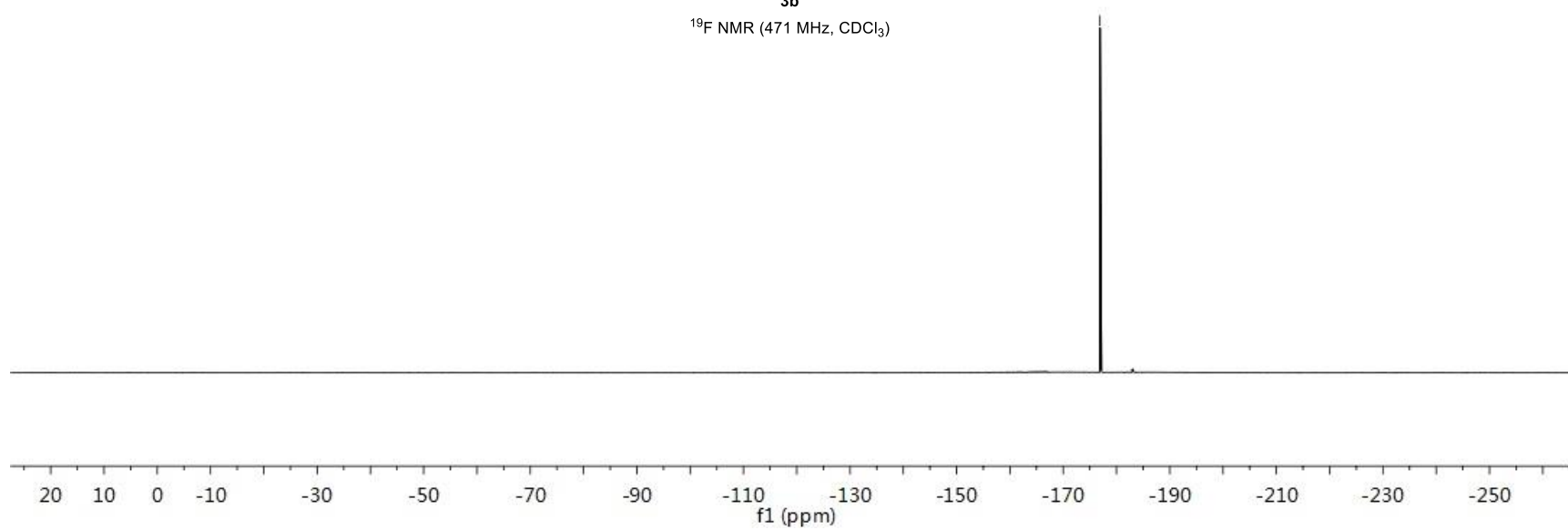
¹⁹F NMR gf-ck-II-56aP in CDCl₃



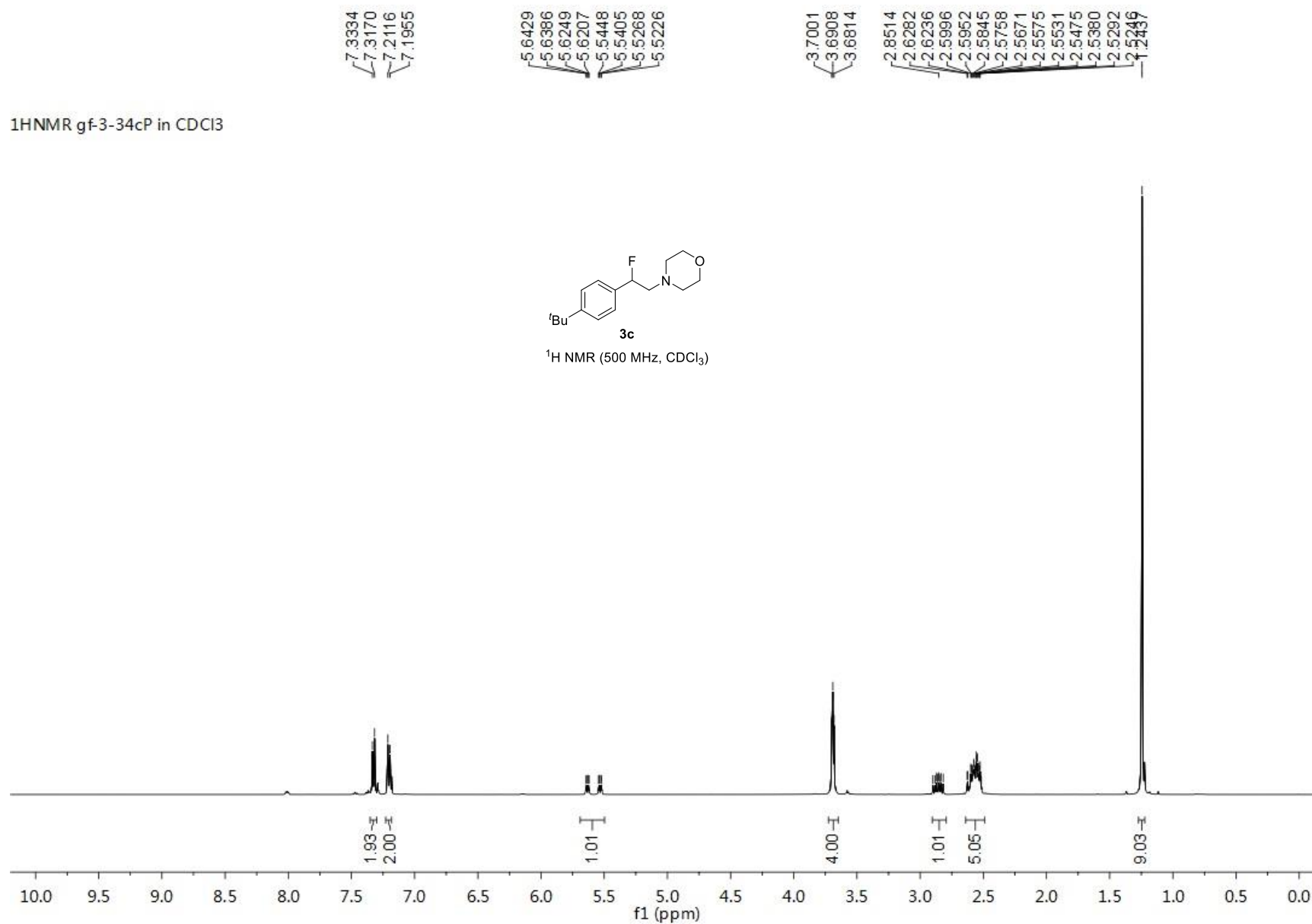
3b

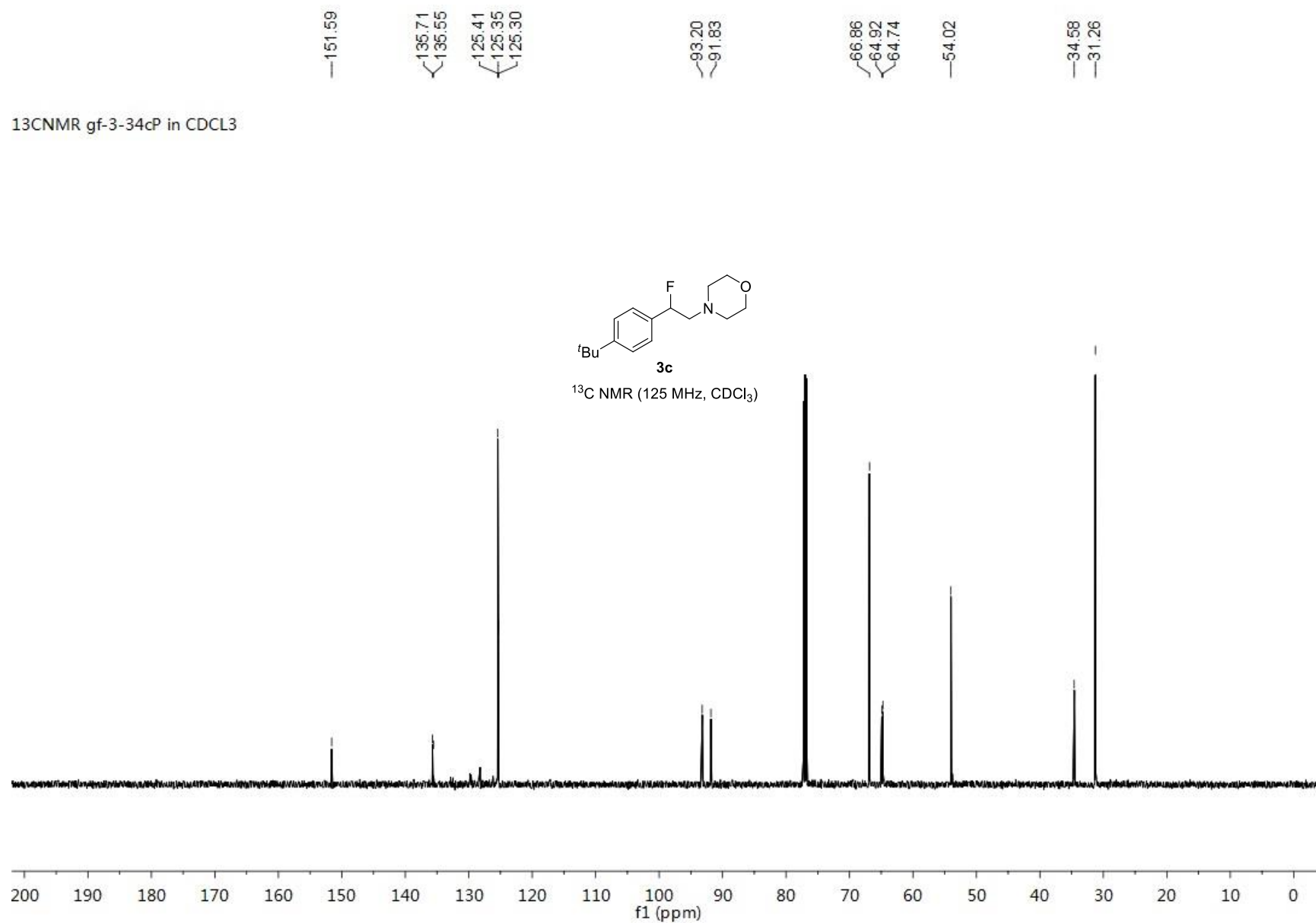
¹⁹F NMR (471 MHz, CDCl₃)

— -176.9016



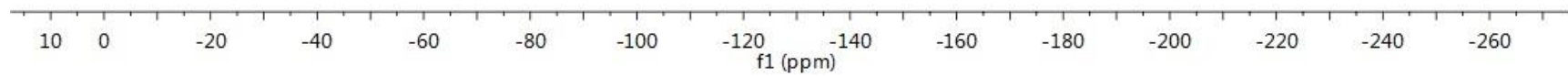
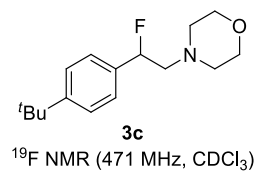
¹H NMR of gf-3-34cP in CDCl₃

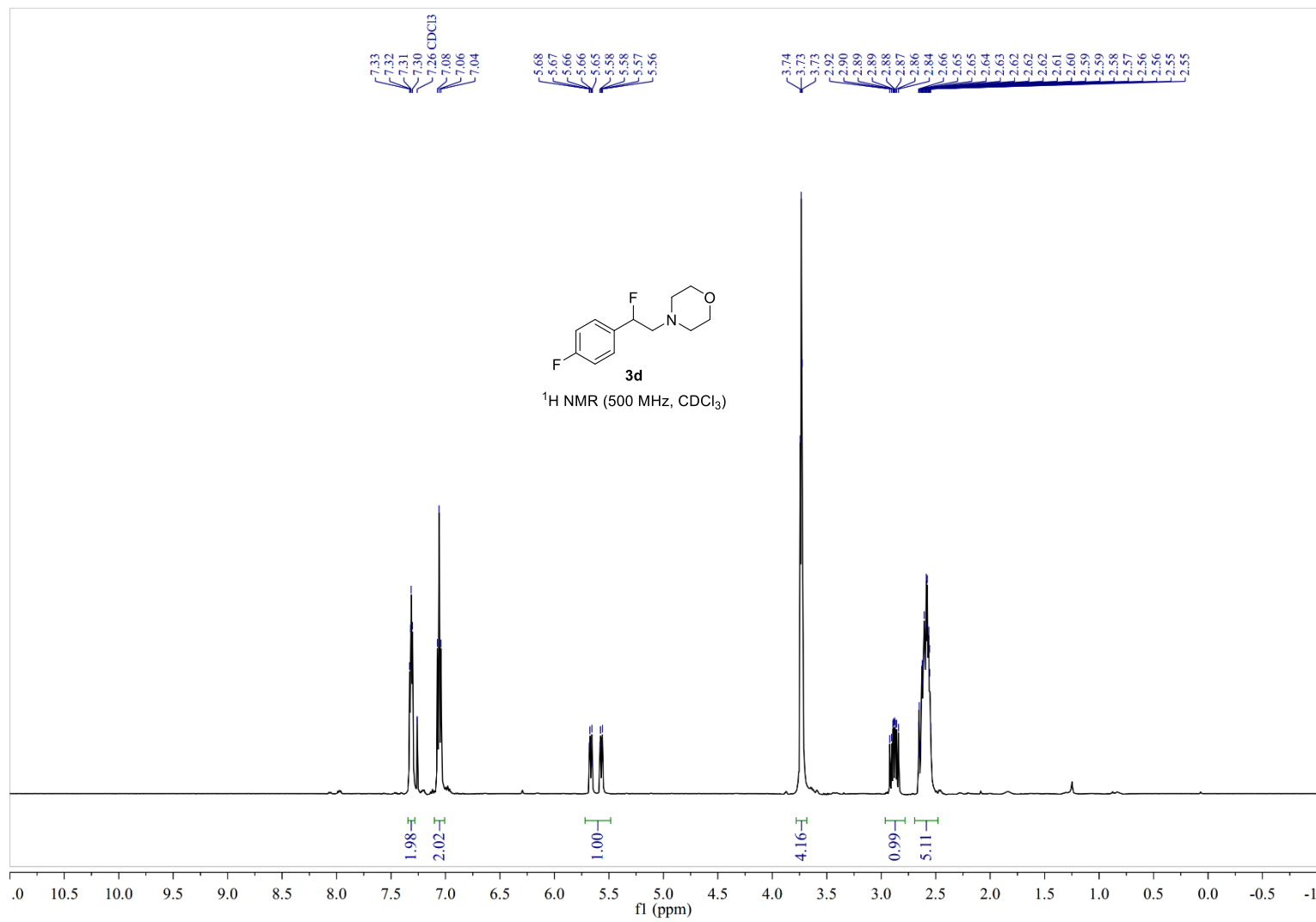


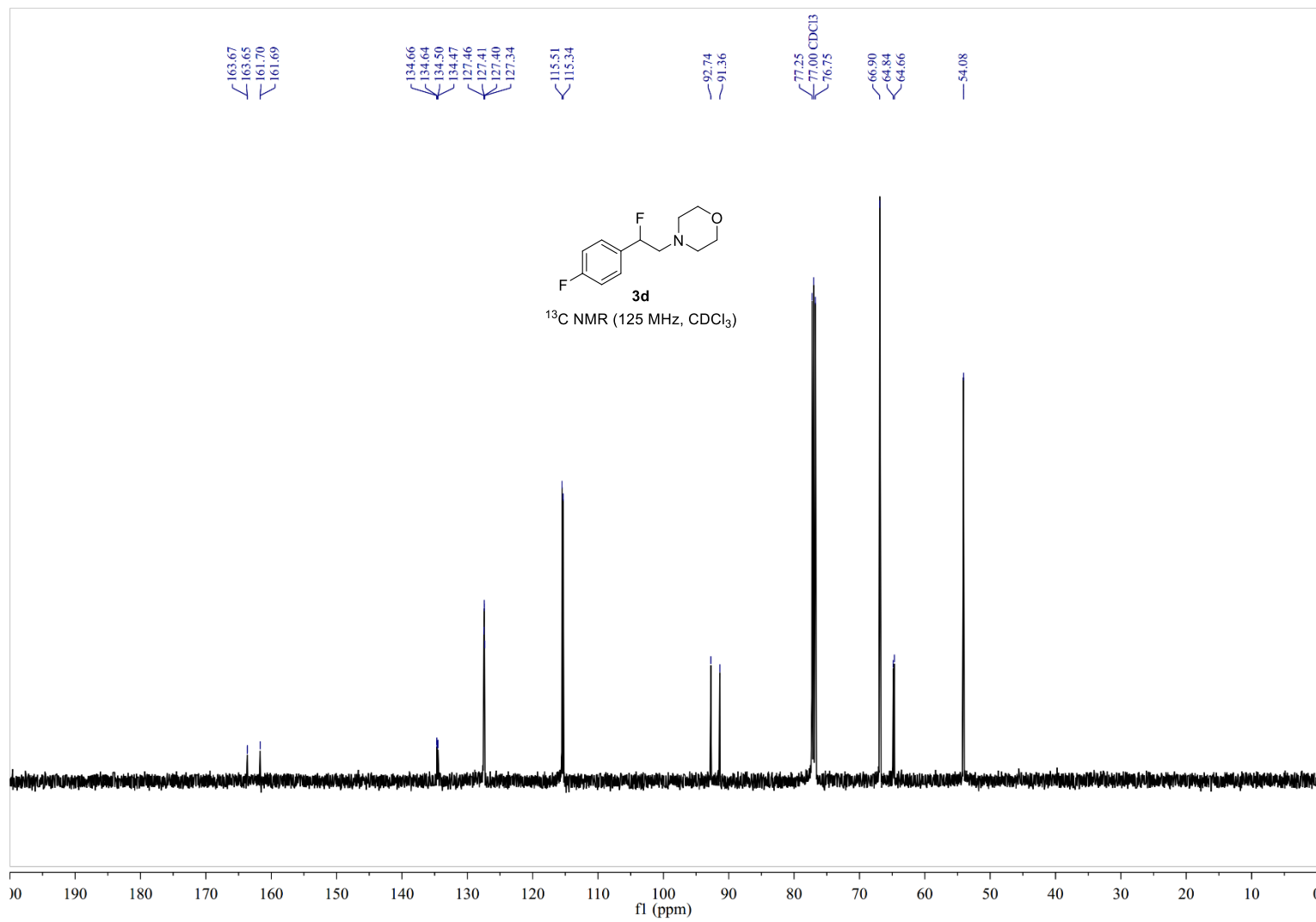


^{19}F NMR gf-3-34cP in CDCl_3

---175.1090



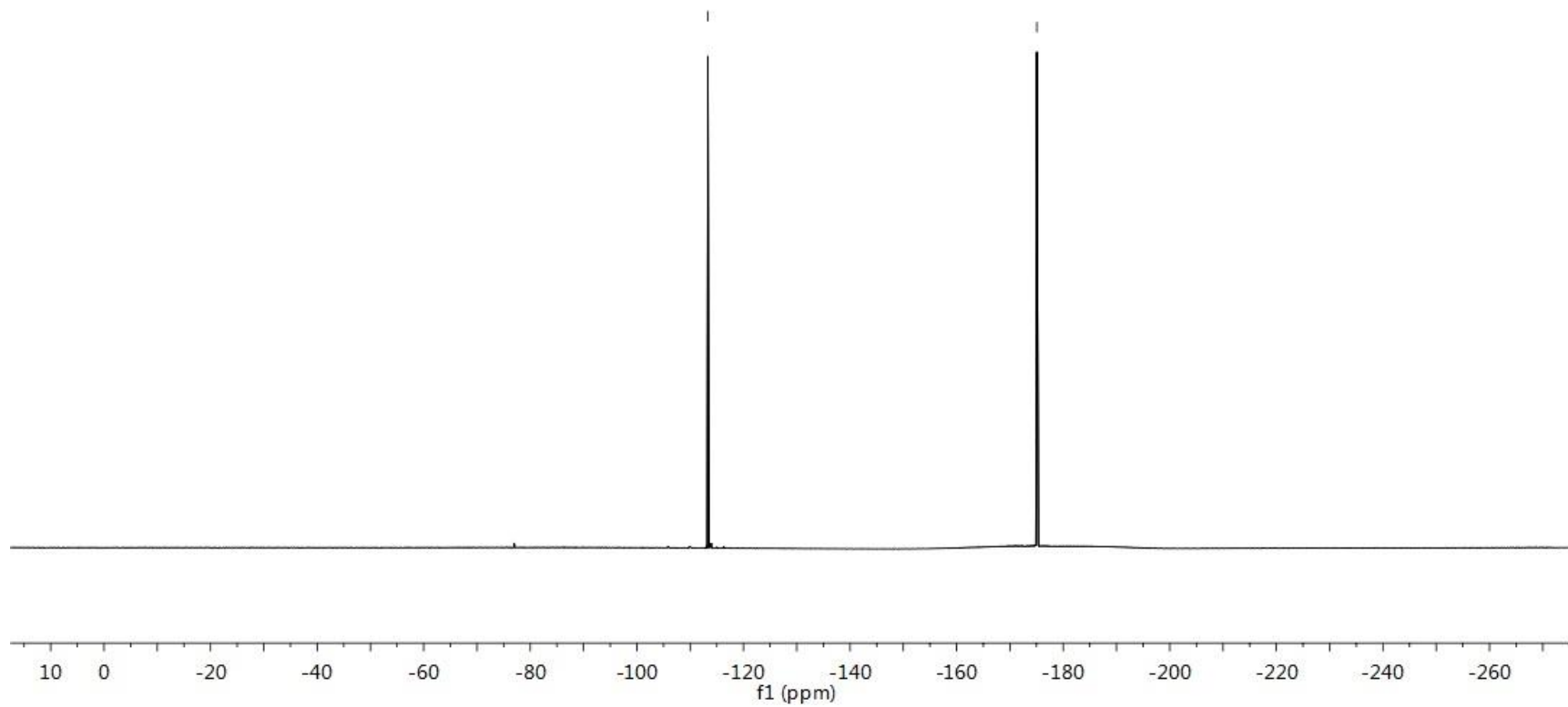
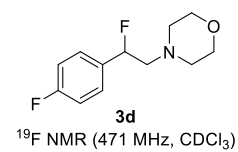


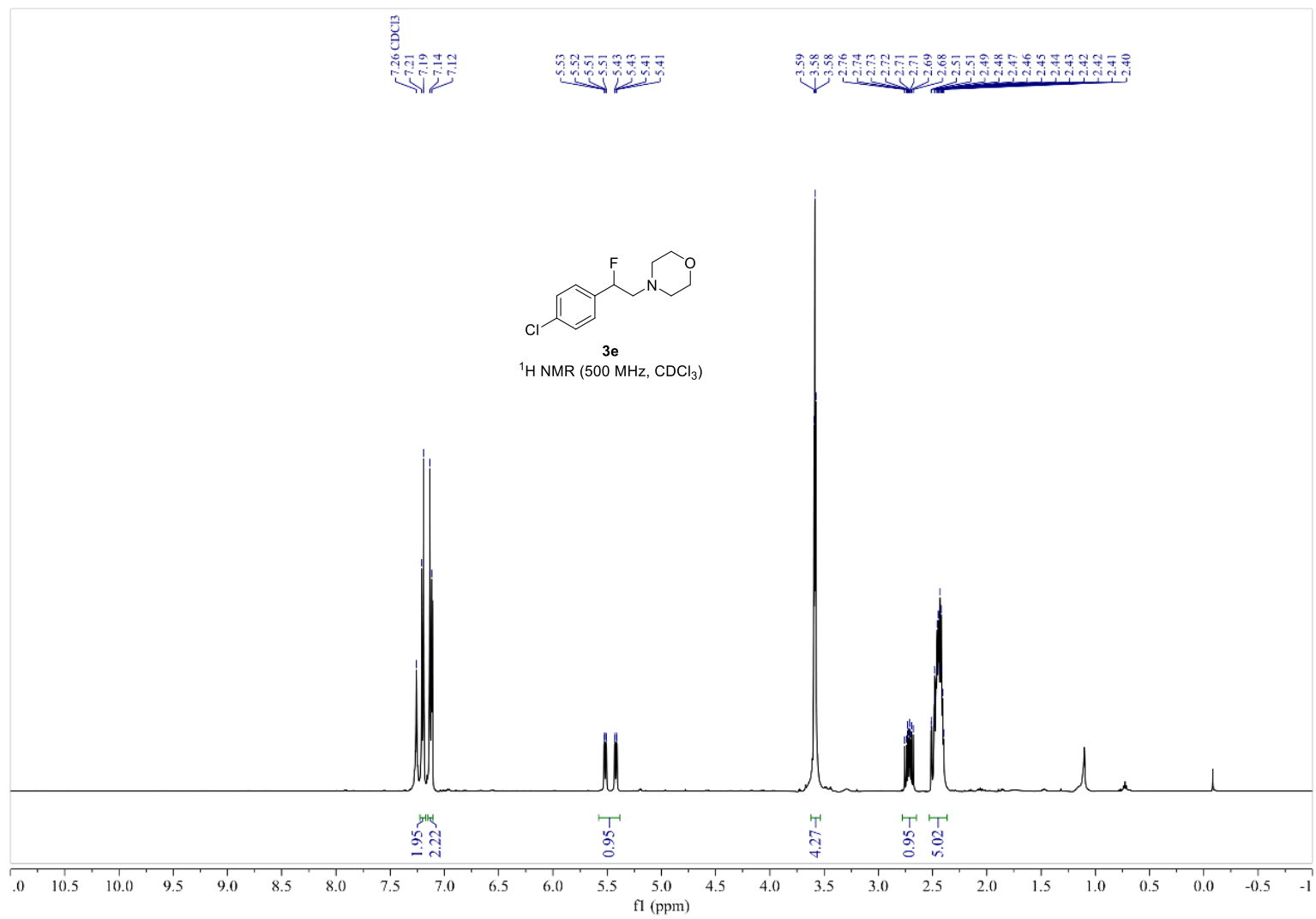


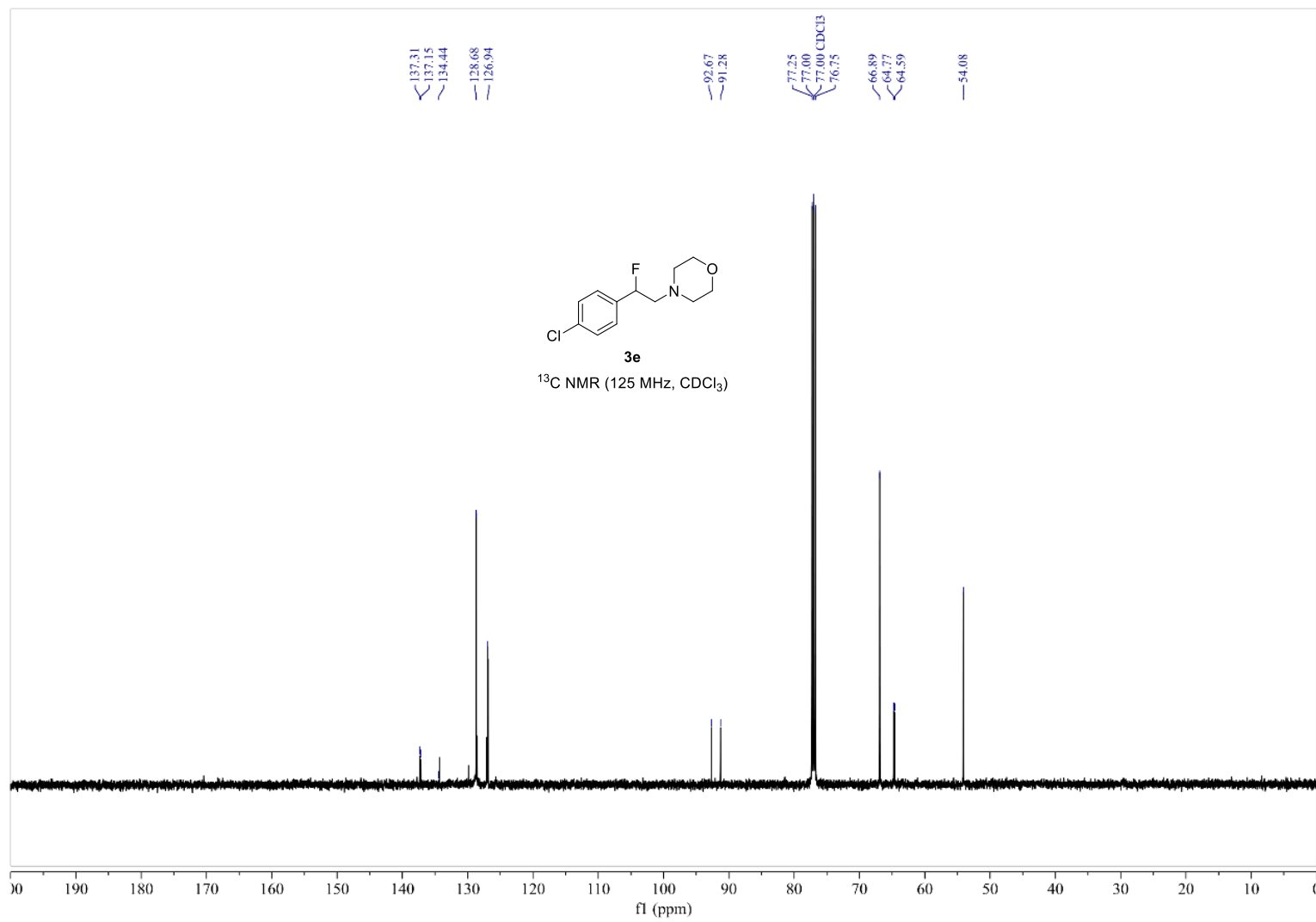
^{19}F NMR gf-3-37bP in CDCl_3

---113.3240

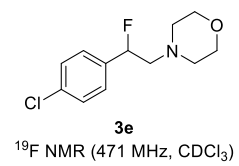
---175.0609



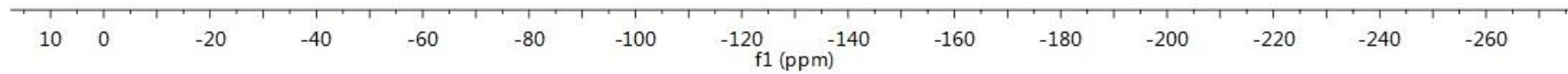


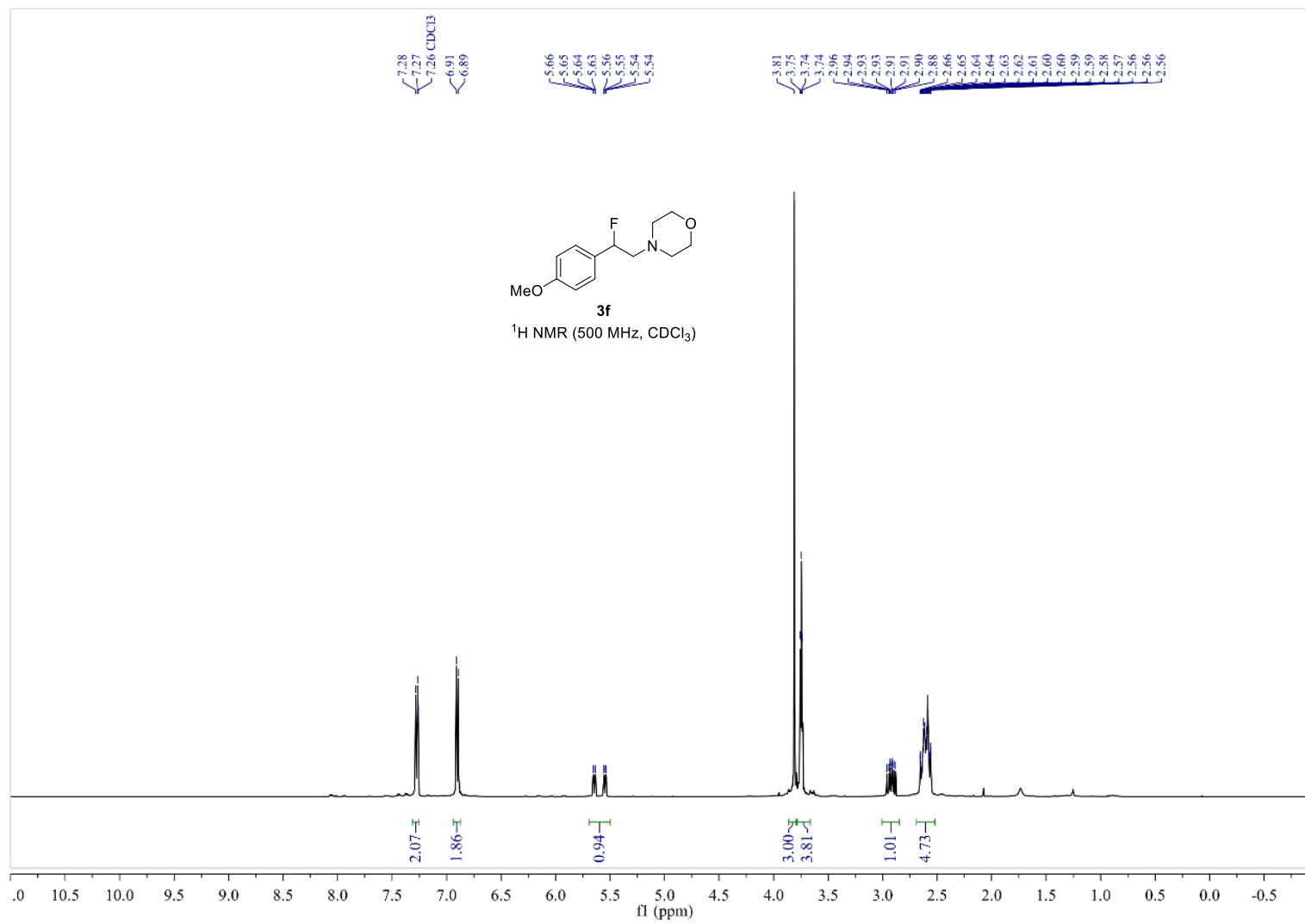


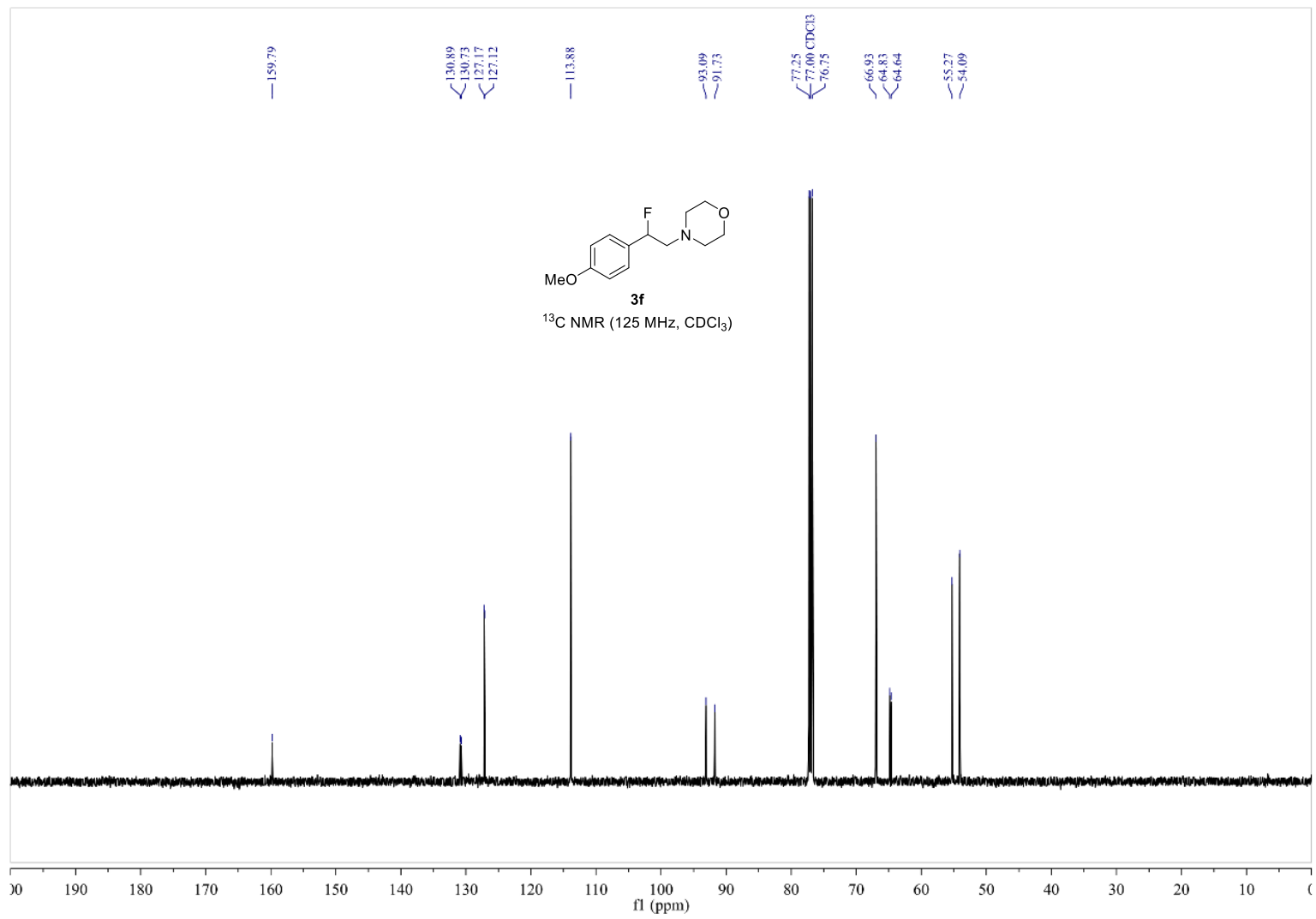
¹⁹F NMR gf-3-37cP in CDCl₃



---177.2881

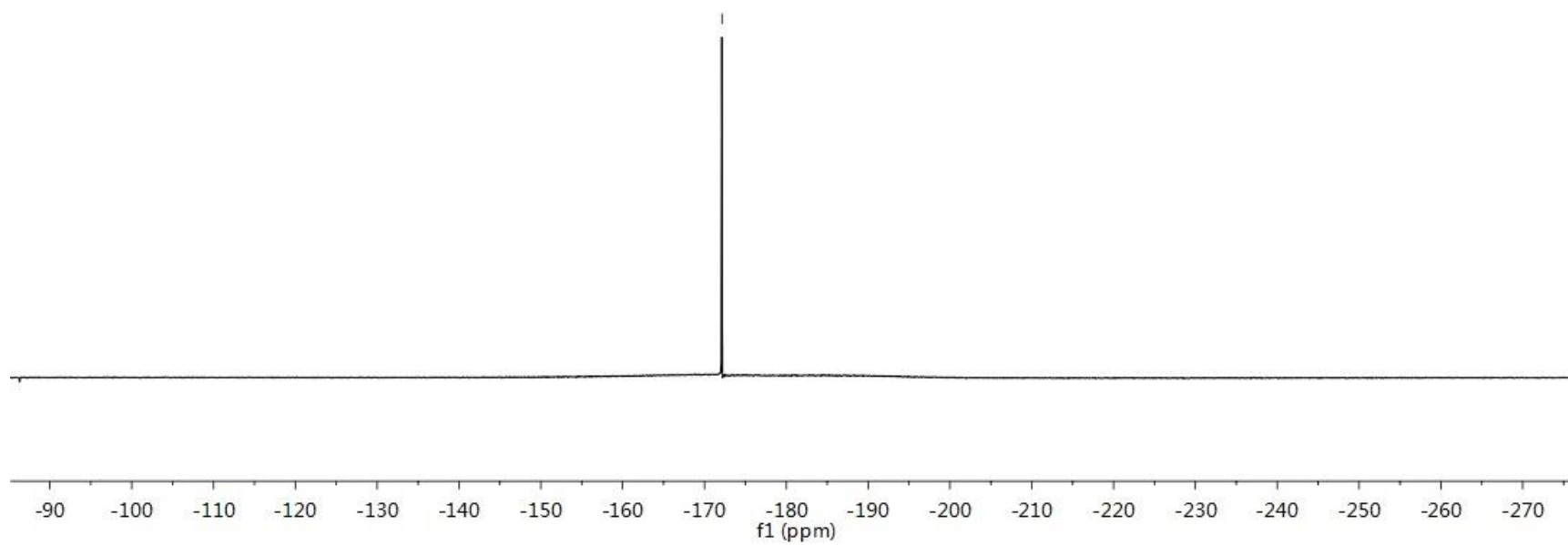
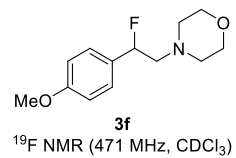


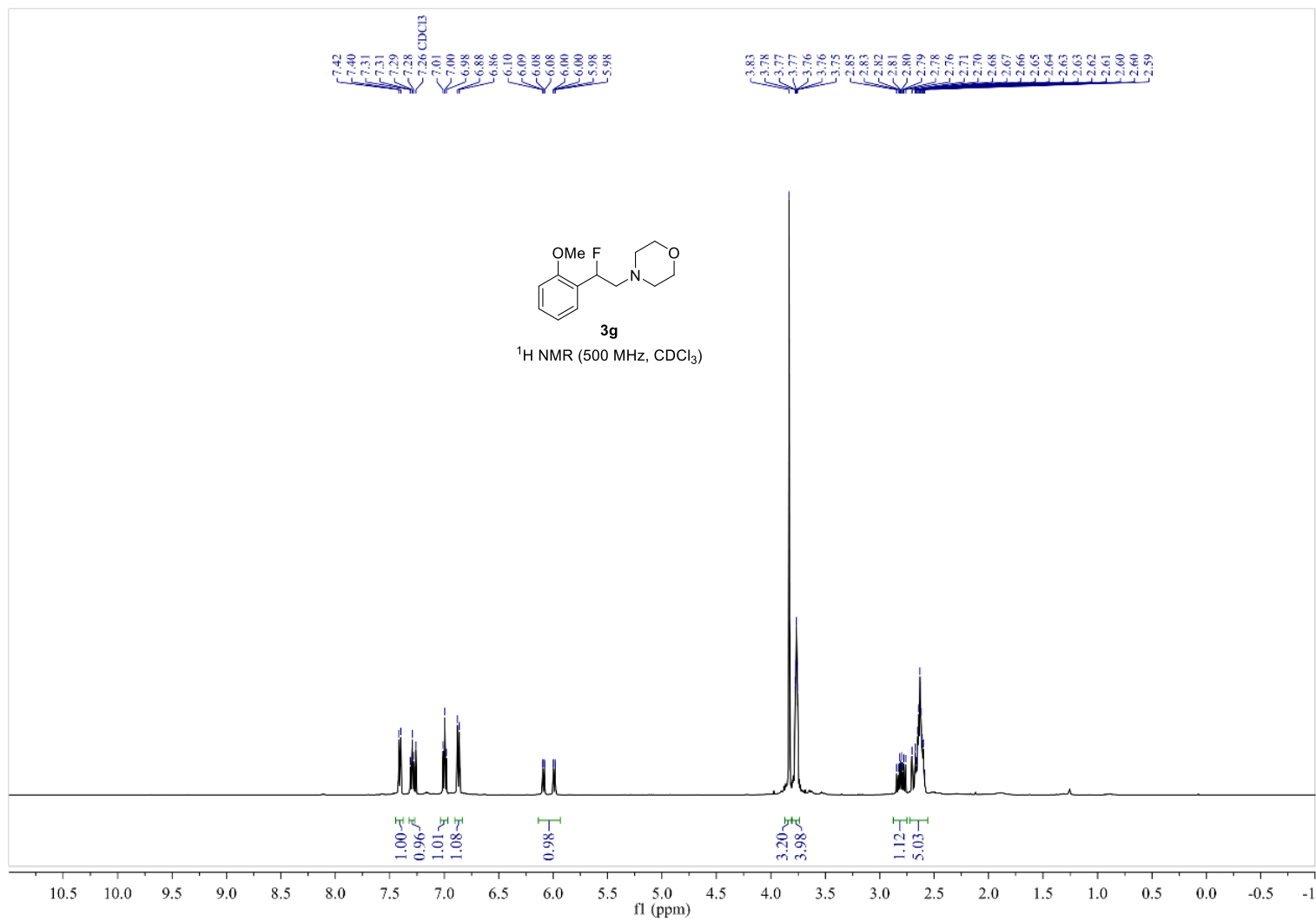


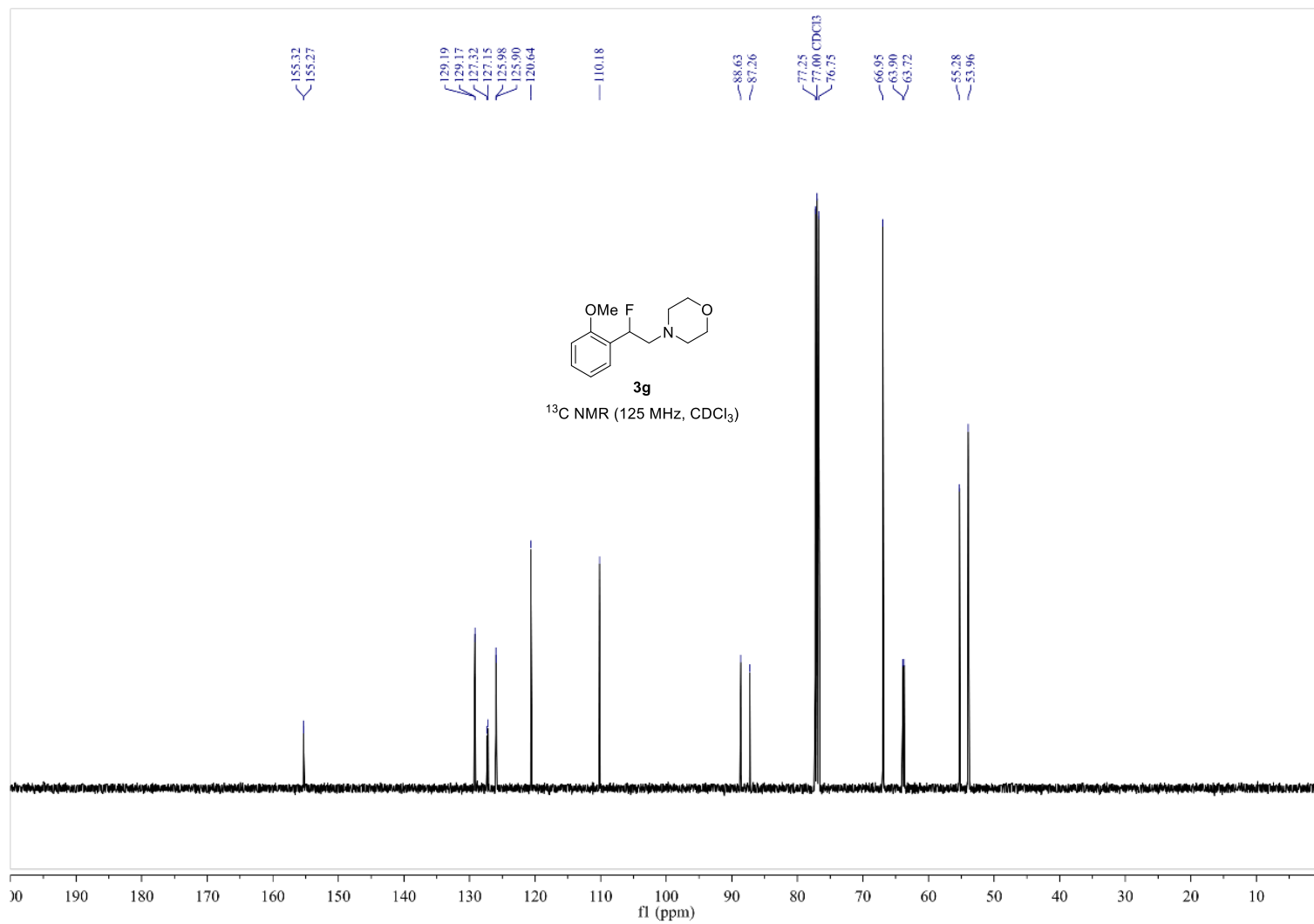


^{19}F NMR gf-3-33bP in CDCl_3

---172.1431

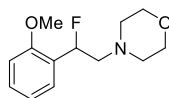






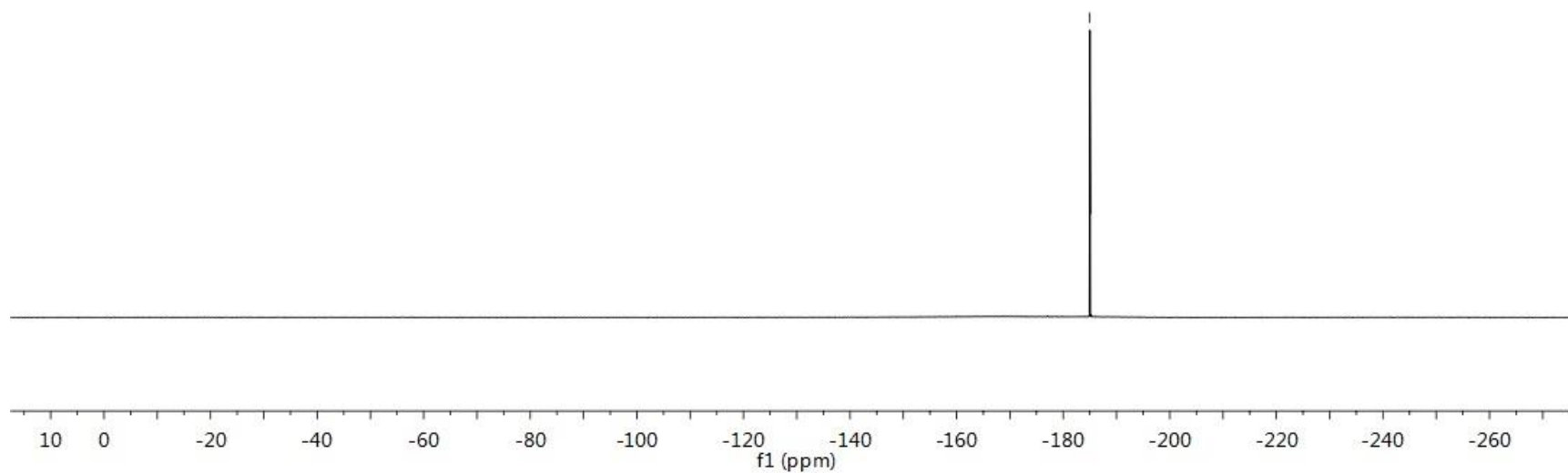
¹⁹F NMR gf-3-34bP in CDCl₃

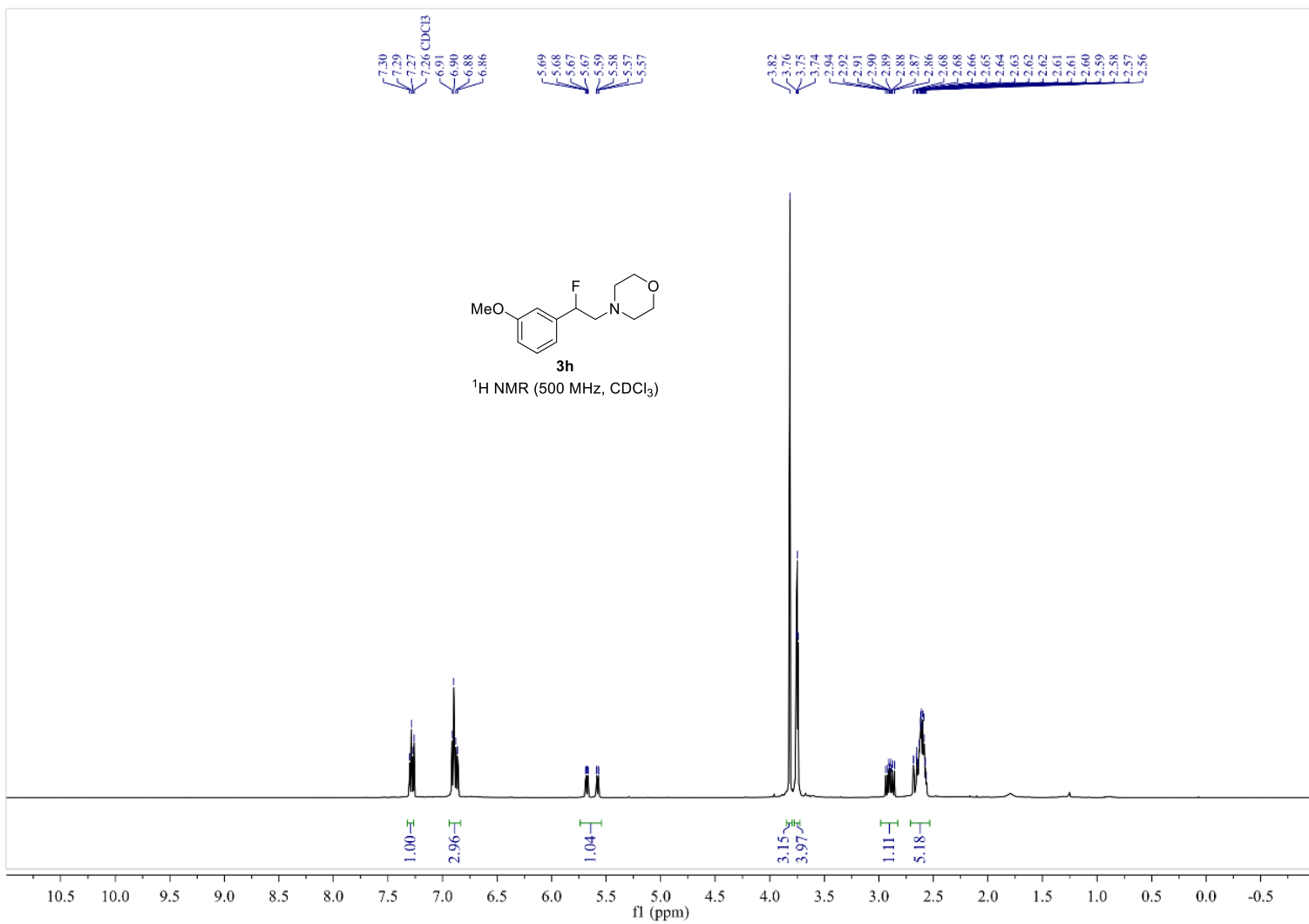
— 185.0007

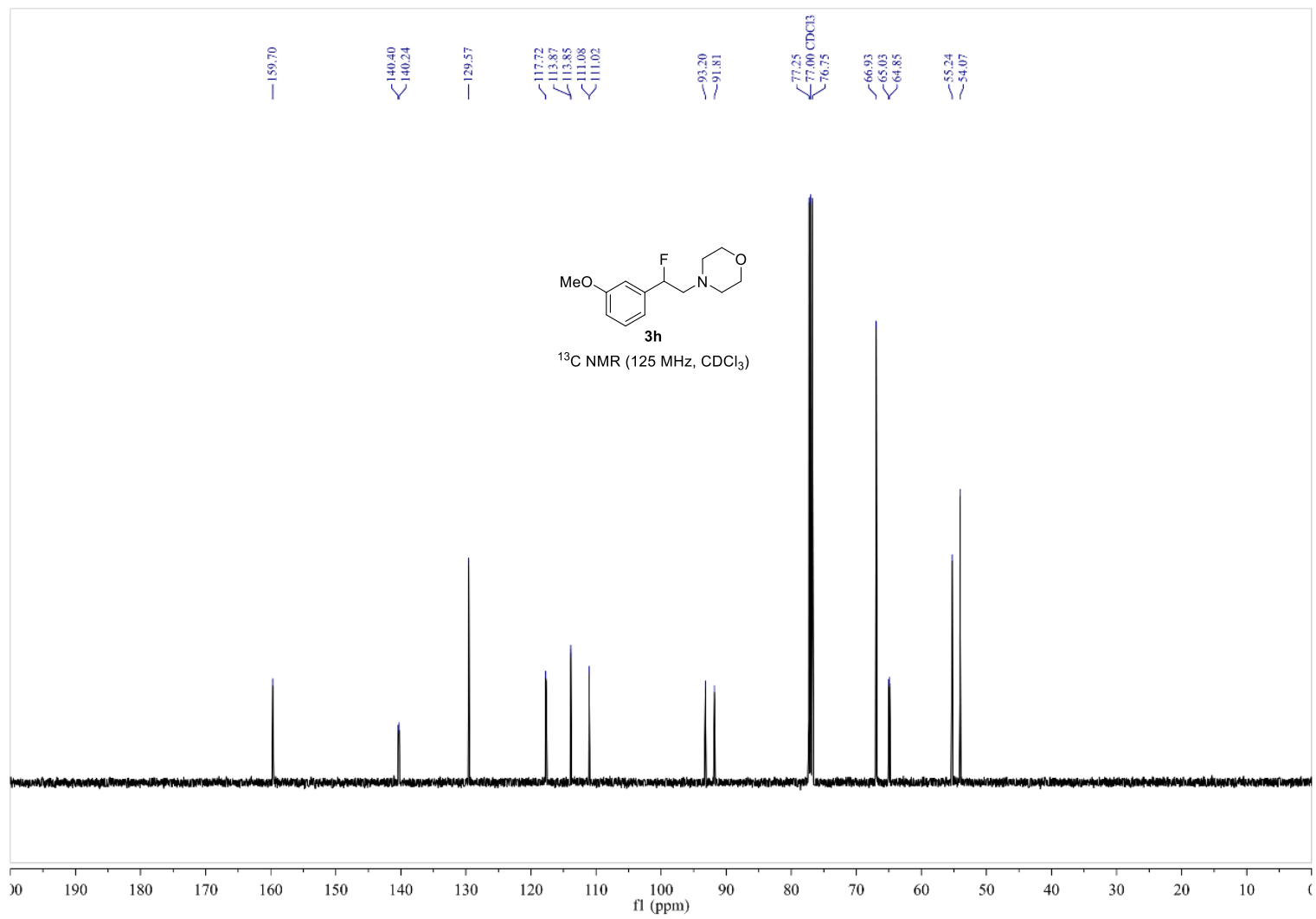


3g

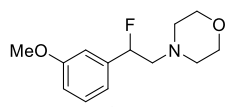
¹⁹F NMR (471 MHz, CDCl₃)





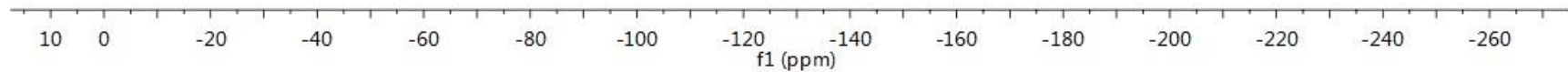


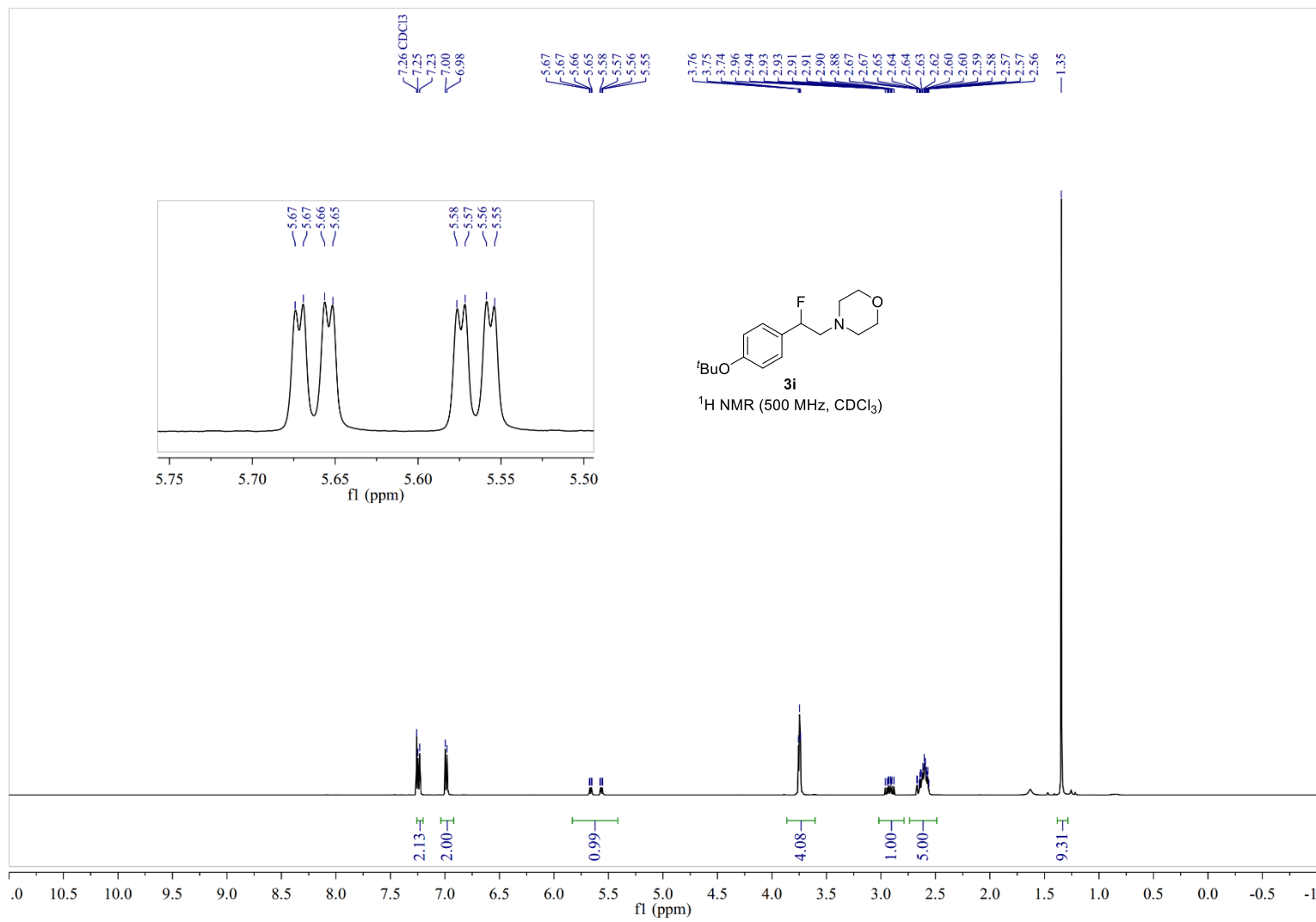
¹⁹F NMR gf-3-34aP in CDCl₃

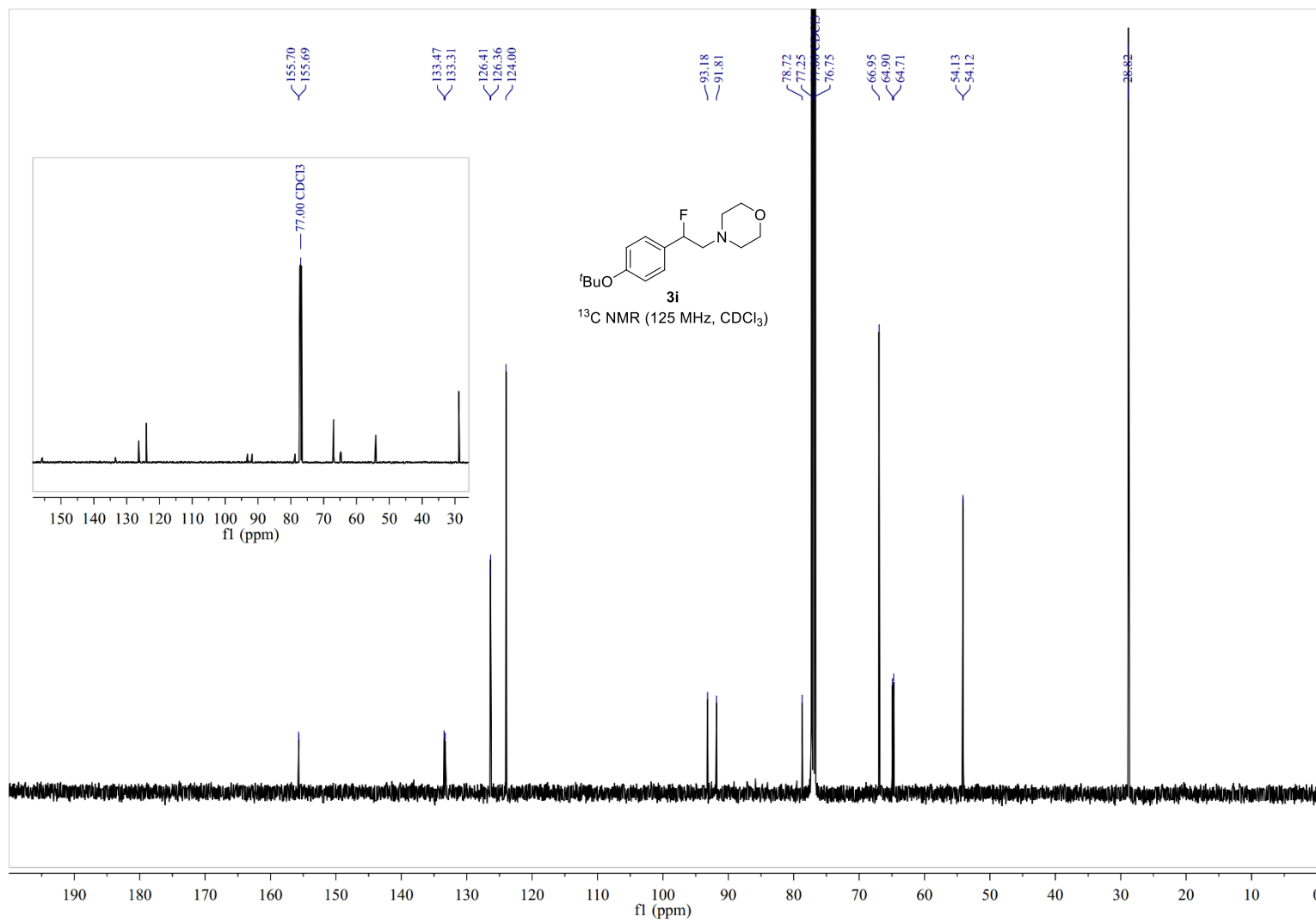


¹⁹F NMR (471 MHz, CDCl₃)

---177.0786

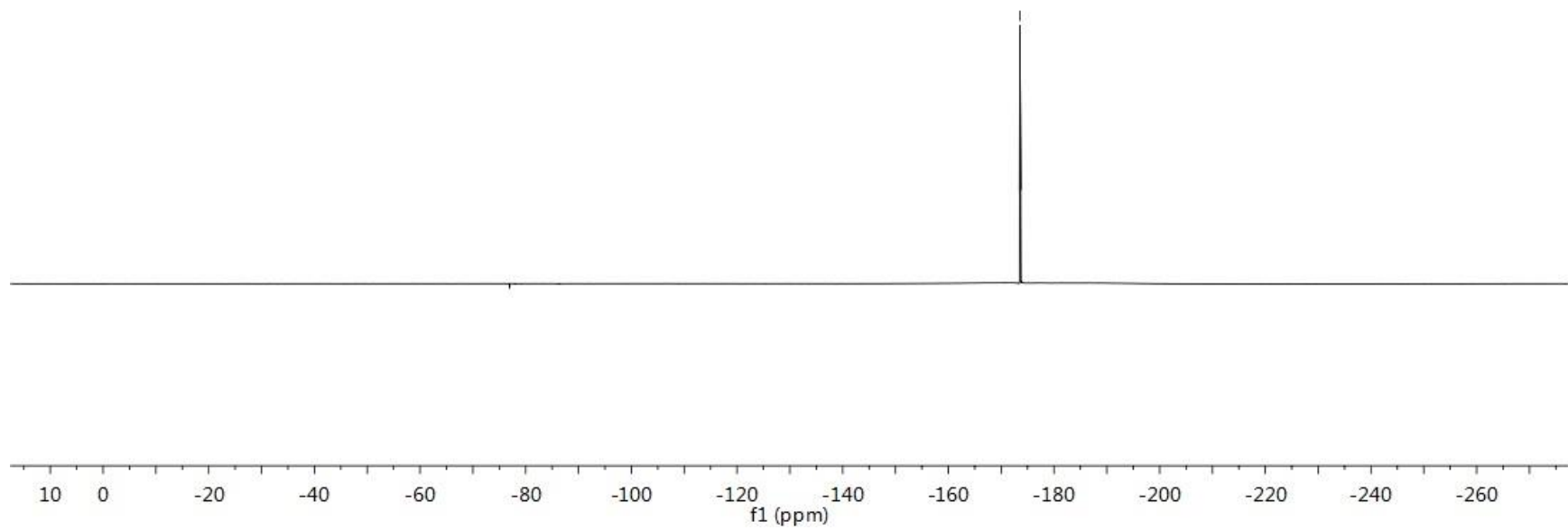
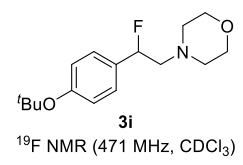




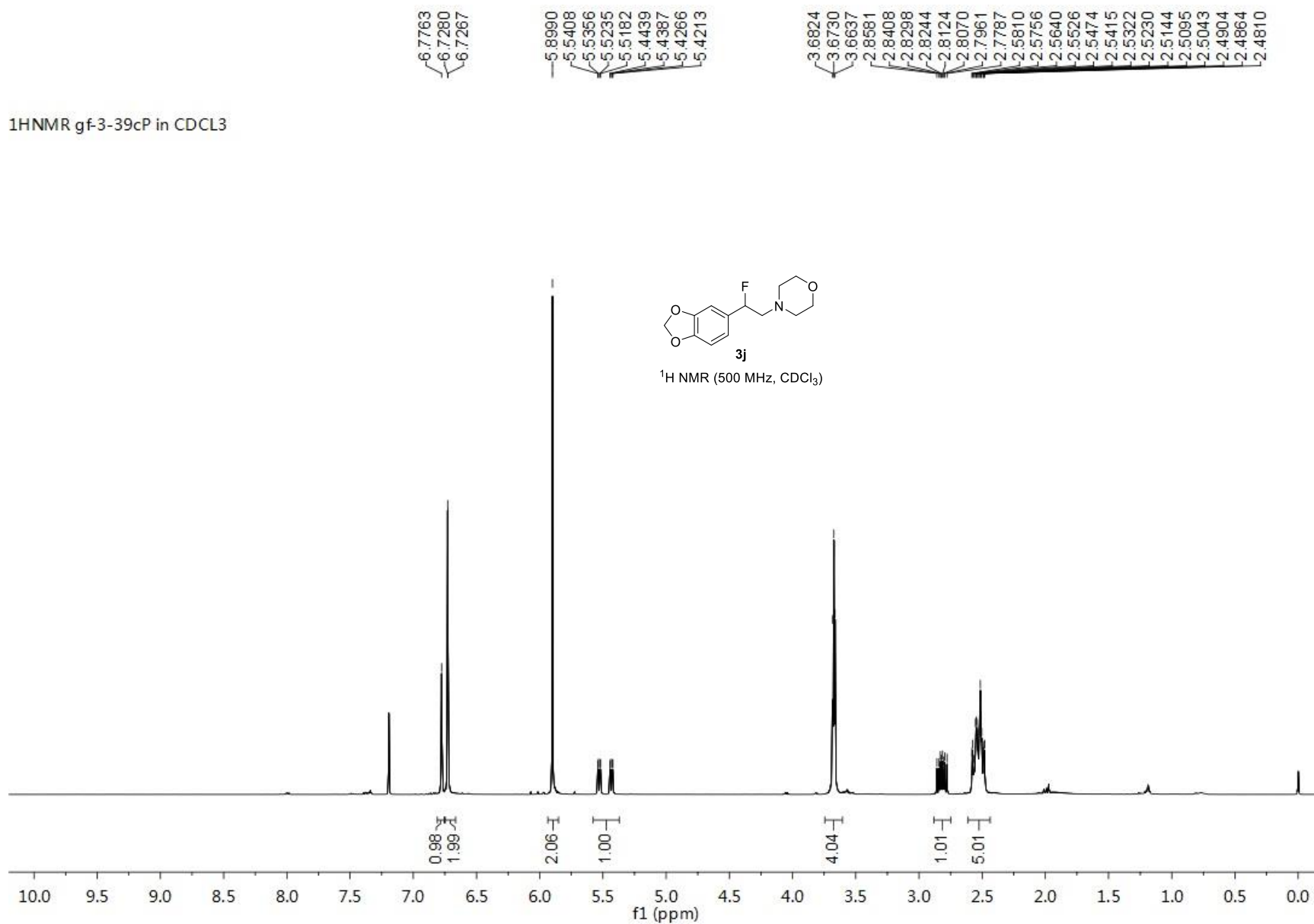


¹⁹F NMR of compound 31 in CDCl₃

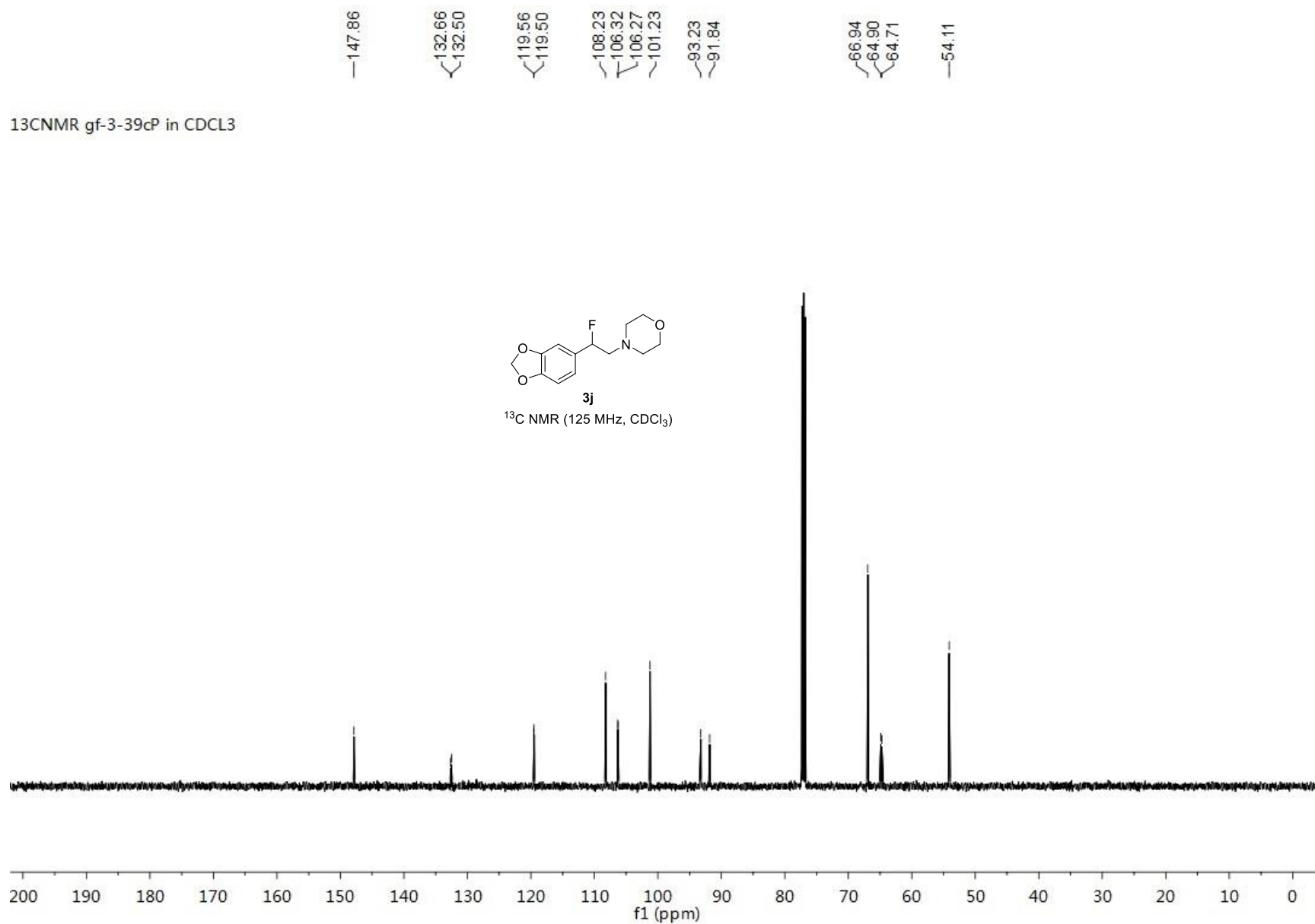
---173.5412



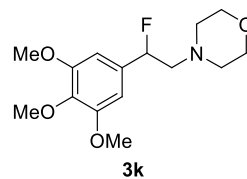
¹H NMR of **3j** in CDCl₃



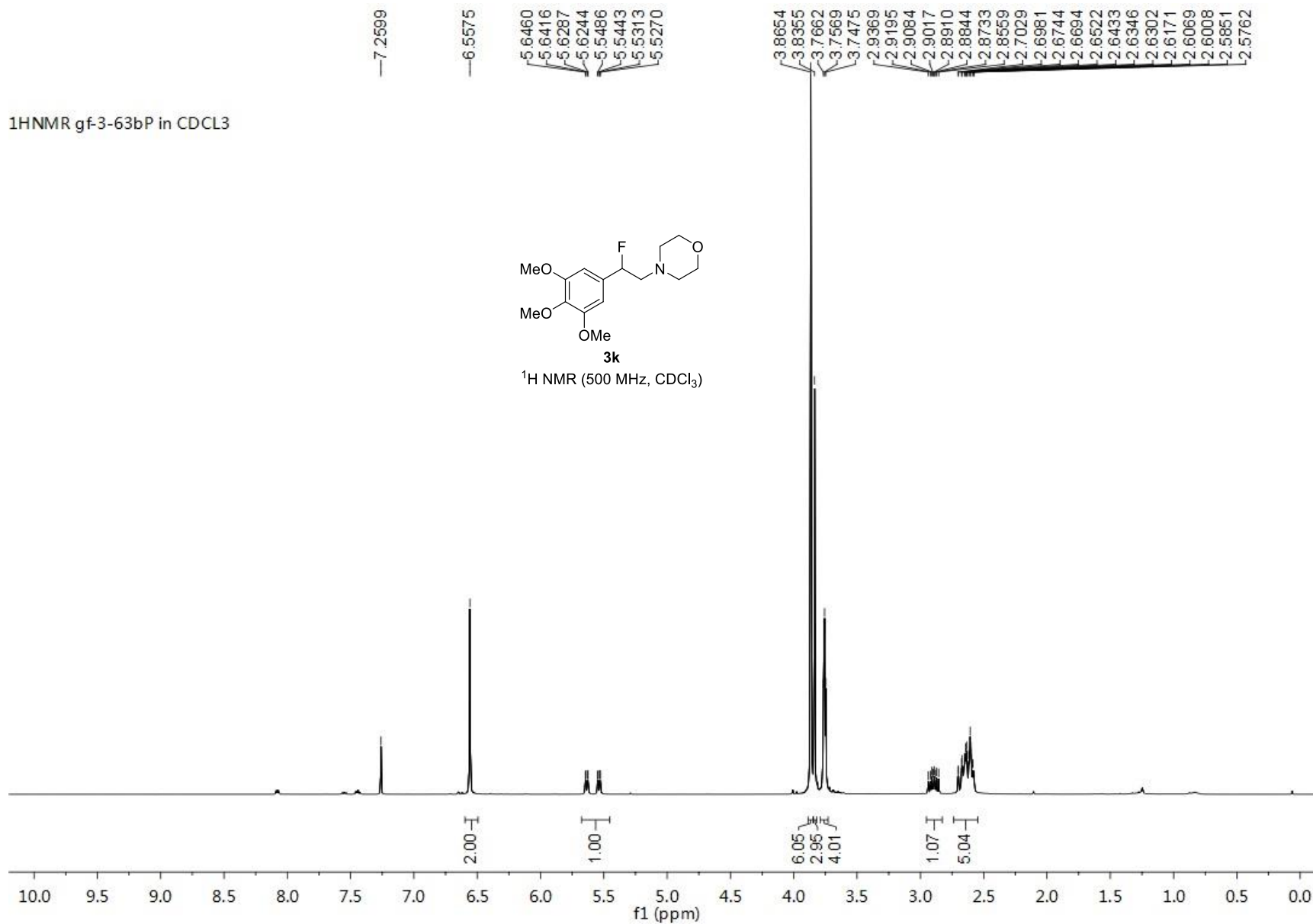
¹³CNMR gf-3-39cP in CDCl₃



¹H NMR of compound 3k in CDCl₃

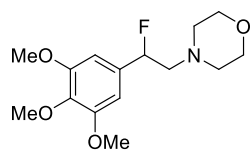


¹H NMR (500 MHz, CDCl₃)

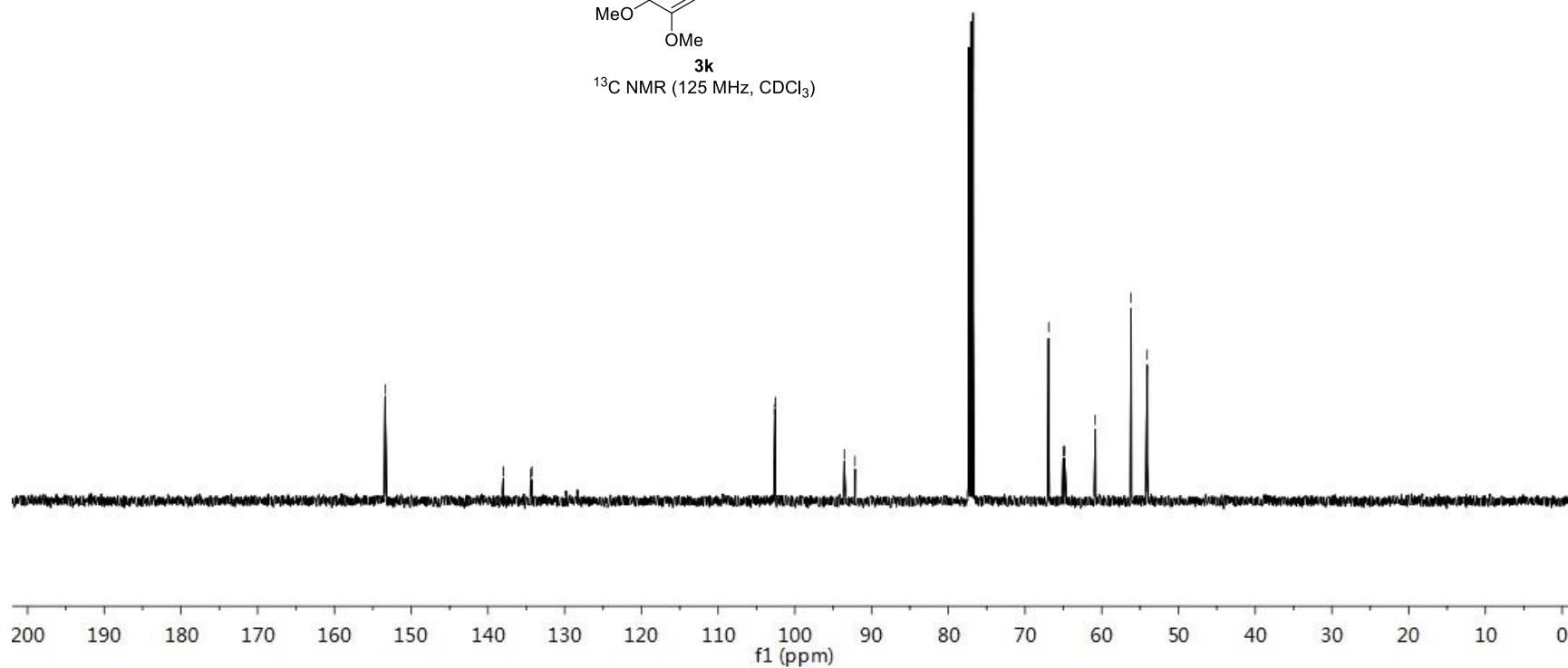


¹³CNMR gf-3-63bP in CDCl₃

153.39
138.00
134.43
134.27
102.61
102.56
93.55
92.17
66.93
65.03
64.85
60.86
56.17
54.09

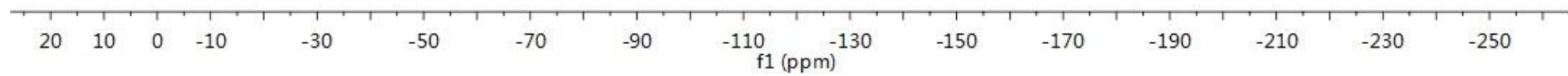
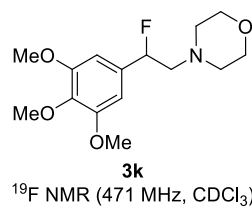


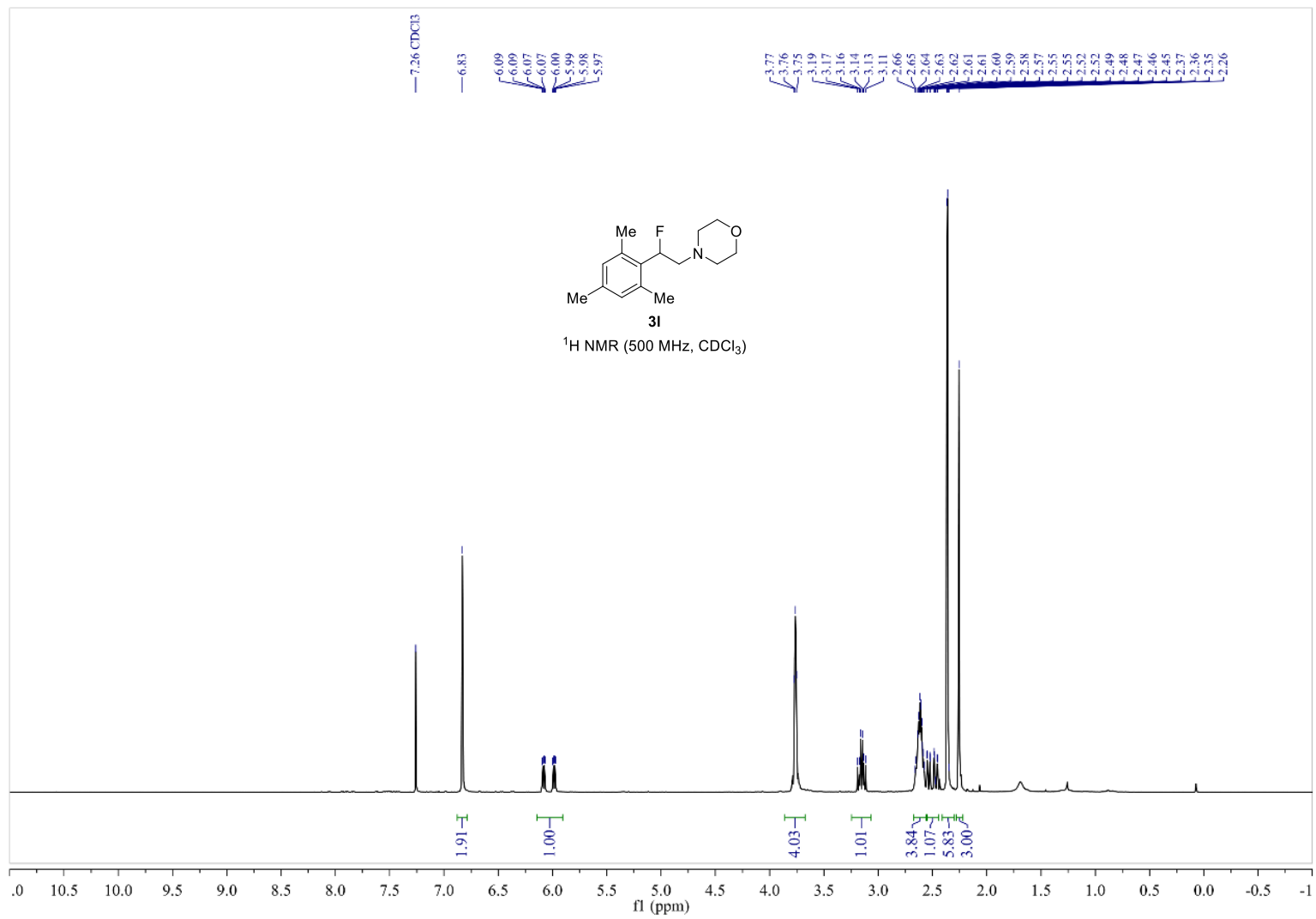
¹³C NMR (125 MHz, CDCl₃)

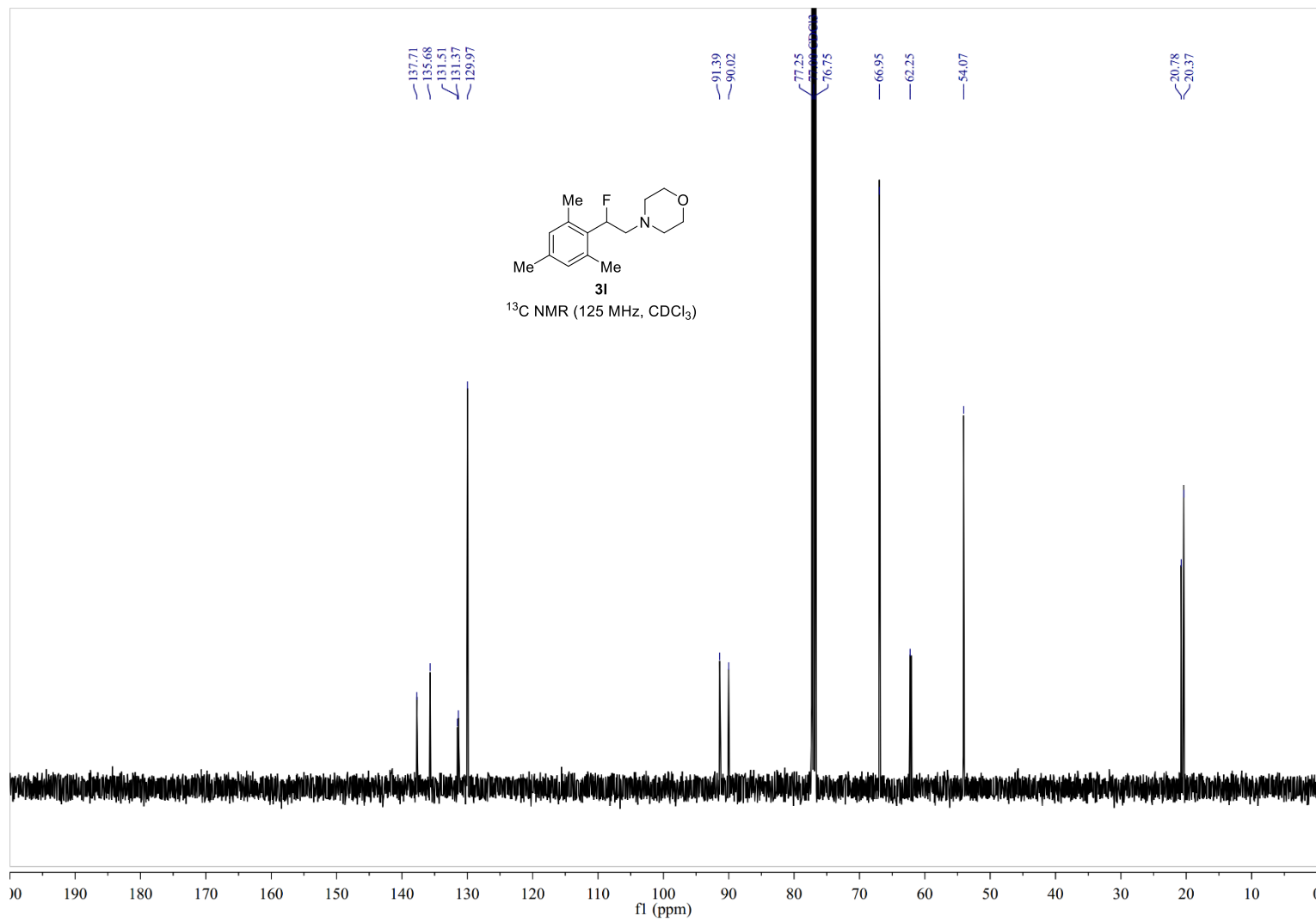


¹⁹F NMR gf-3-63bP in CDCl₃

— -174.4209

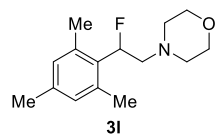




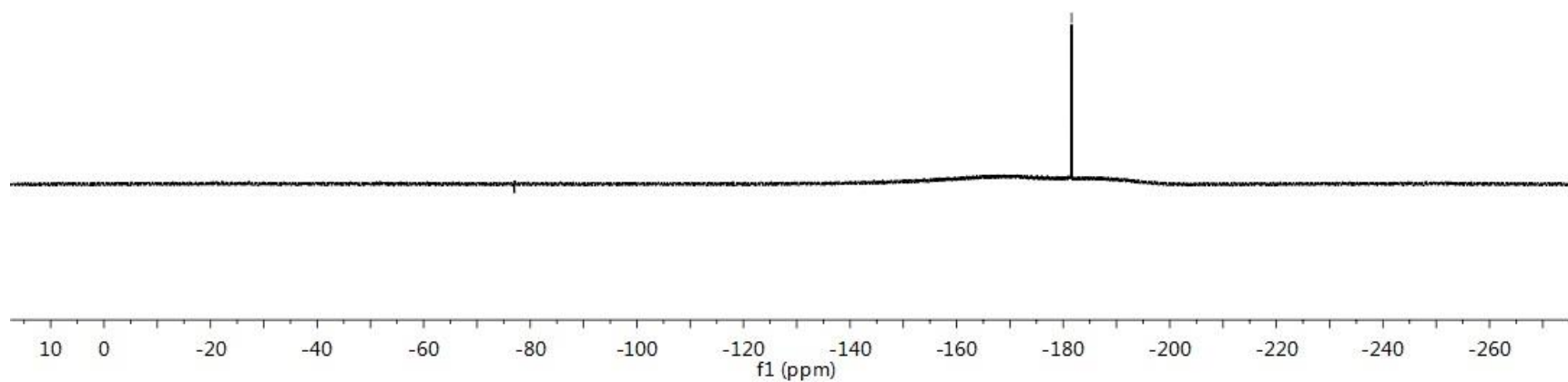


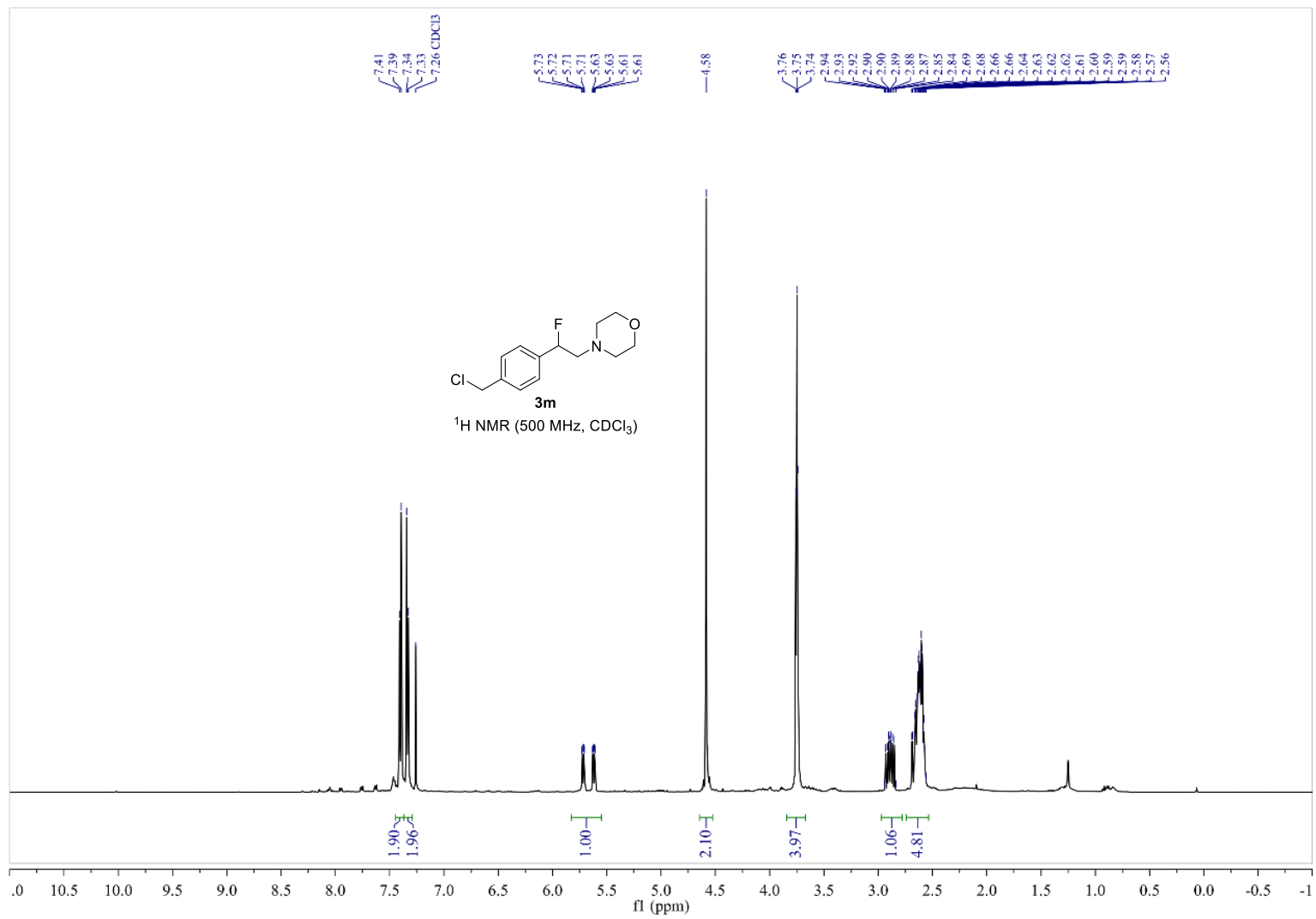
¹⁹F NMR gf-3-41aP in CDCl₃

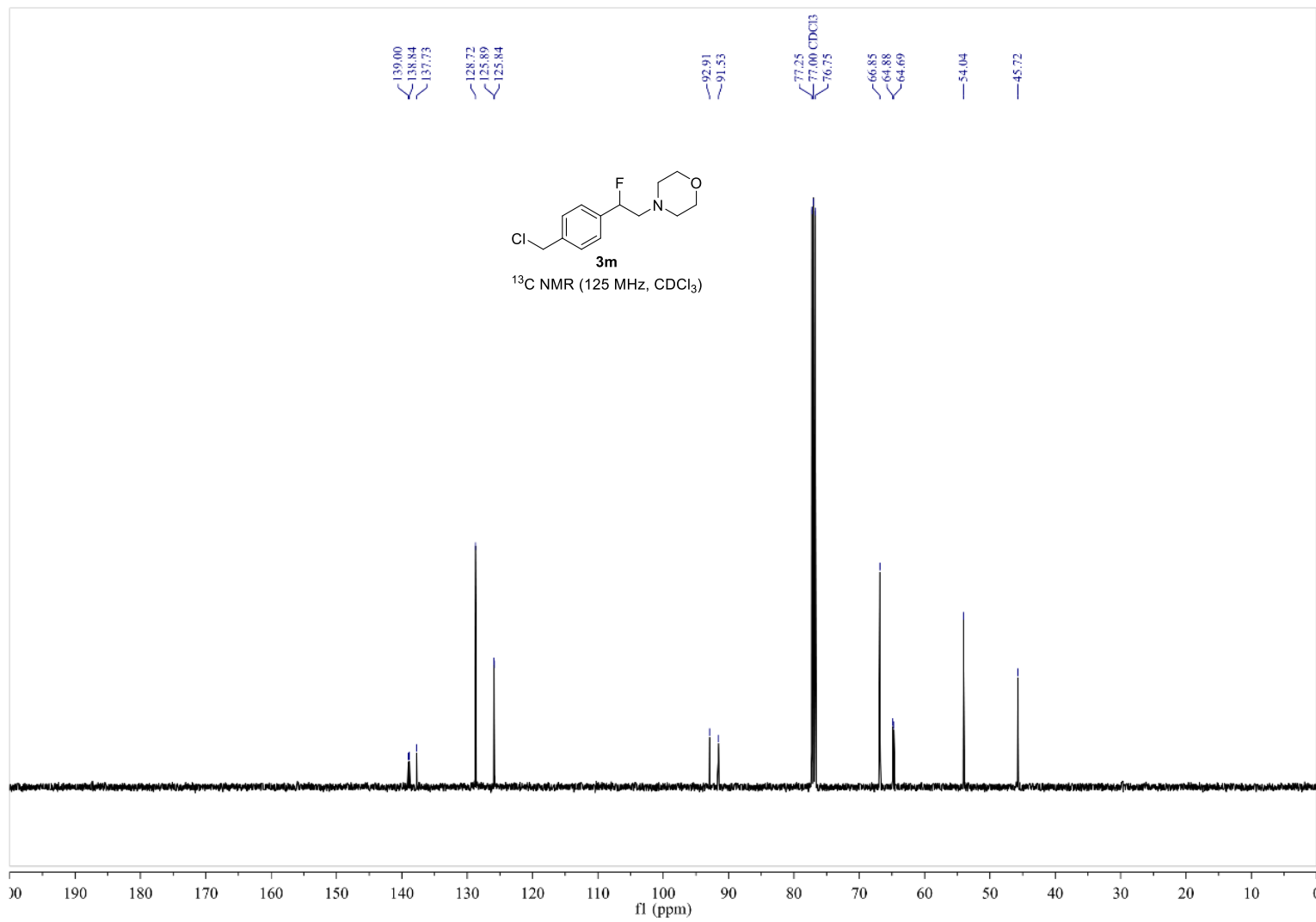
— -181.5748



¹⁹F NMR (471 MHz, CDCl₃)

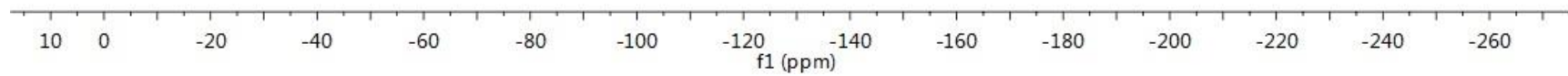
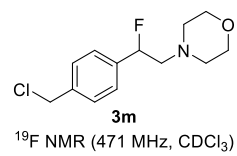


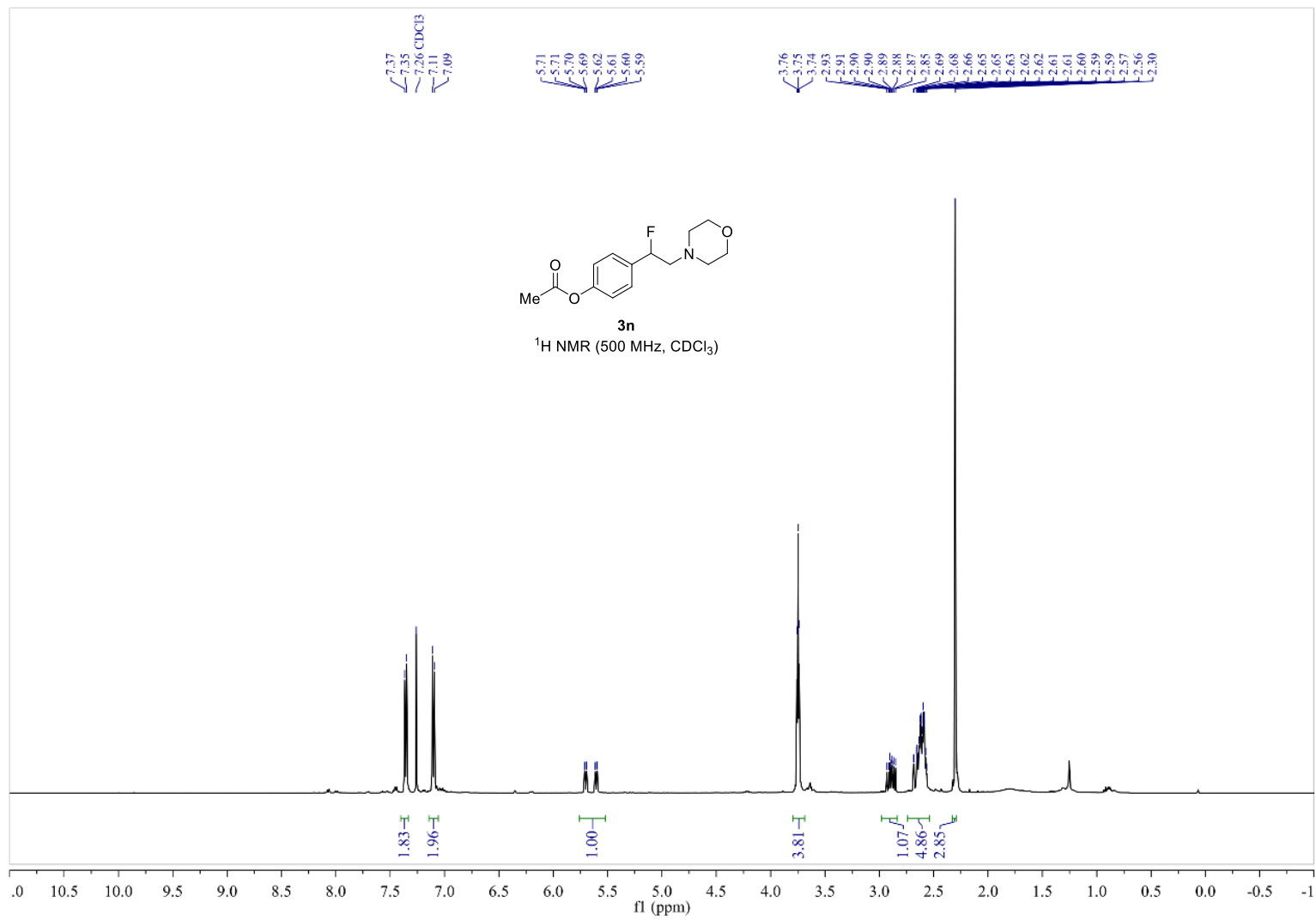


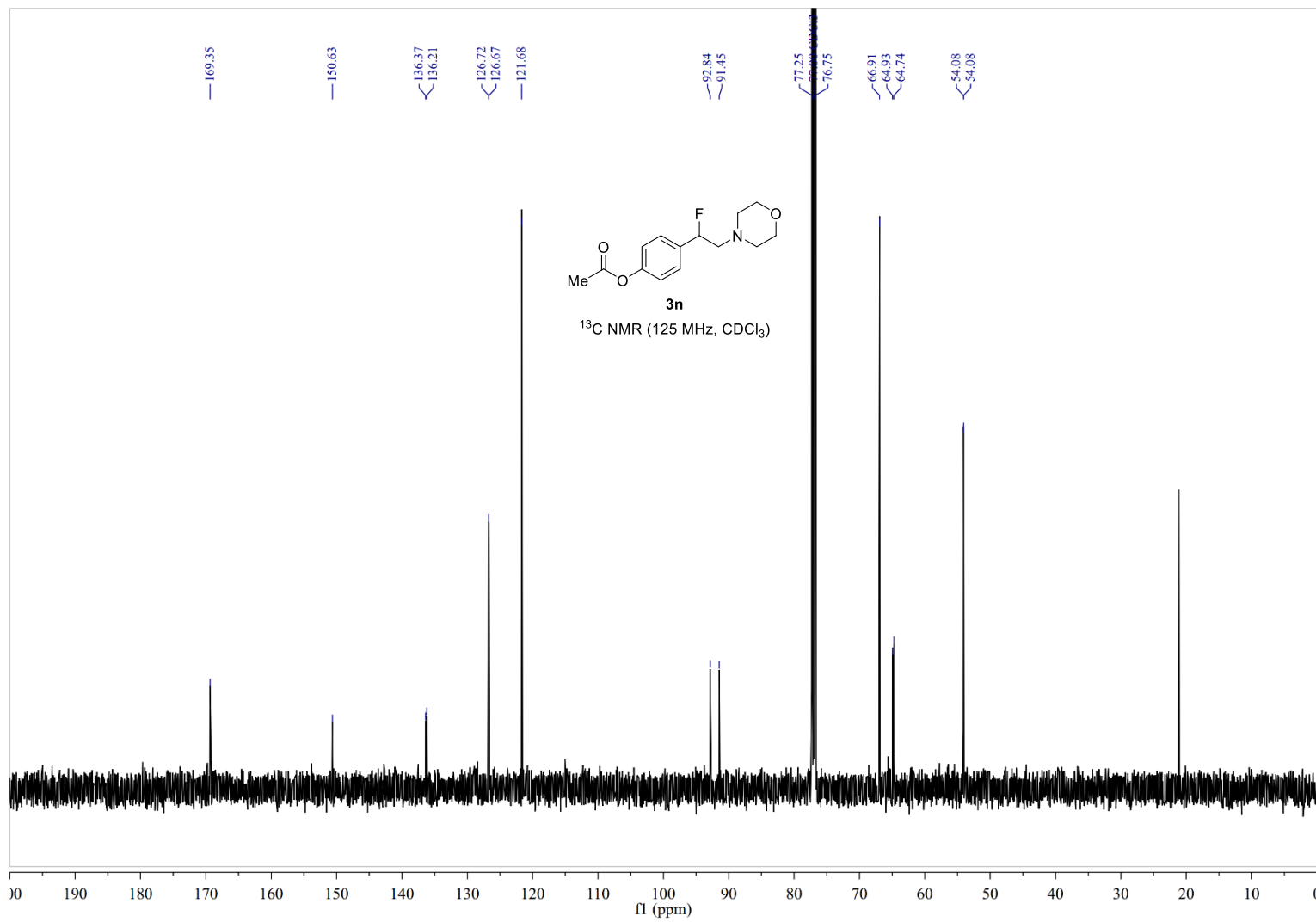


¹⁹F NMR gf-3-45aP in CDCl₃

---177.6391

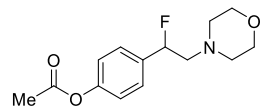




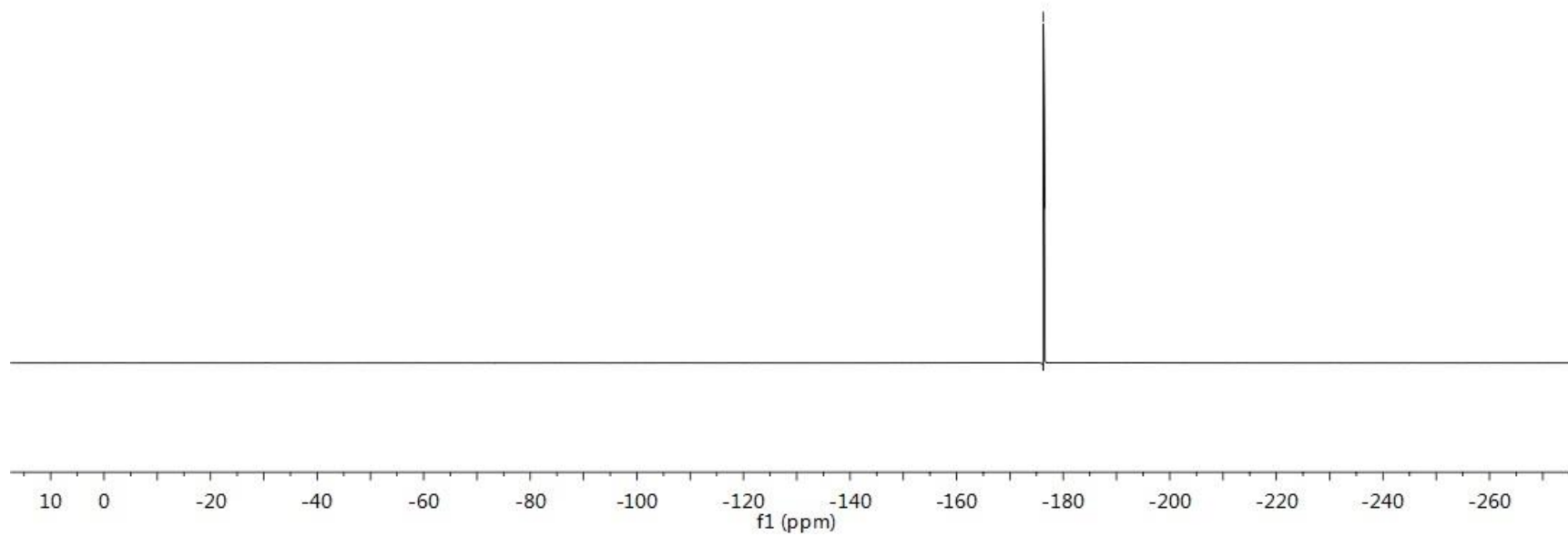


^{19}F NMR gf-3-38bP in CDCl_3

—176.2978

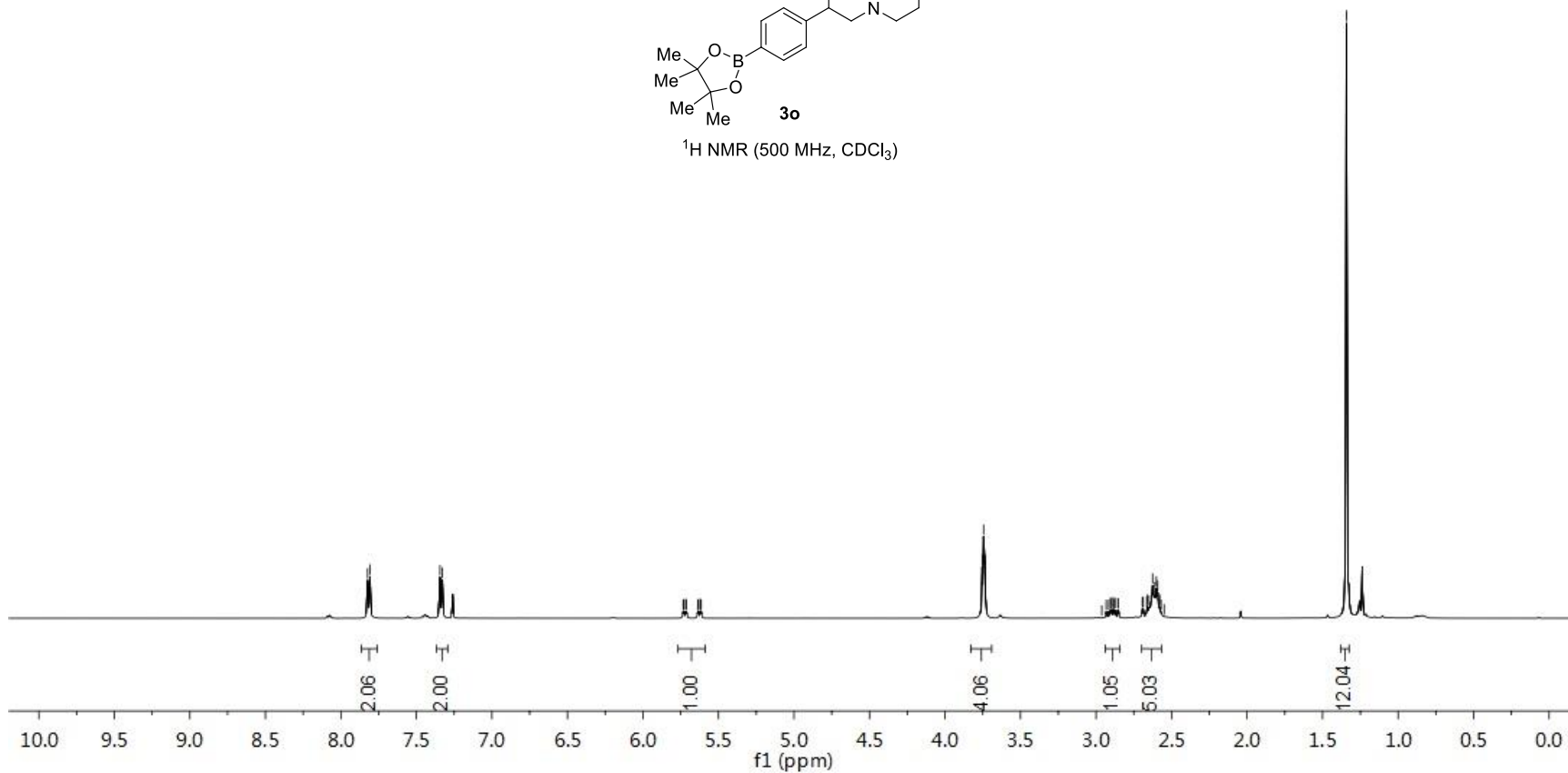
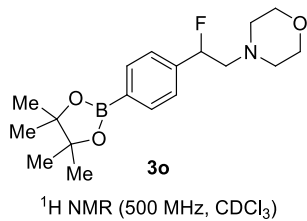


3n
 ^{19}F NMR (471 MHz, CDCl_3)



7.8232
 7.8077
 7.3442
 7.3285
 5.7340
 5.7296
 5.7168
 5.7125
 5.6361
 5.6317
 5.6189
 5.6147
 3.7537
 3.7444
 3.7351
 2.9044
 2.8975
 2.8871
 2.8690
 2.6933
 2.6883
 2.6647
 2.6596
 2.6479
 2.6254
 2.6108
 2.6006
 2.5919
 1.3418
 1.3418

¹H NMR of gf-3-66aPP in CDCl₃



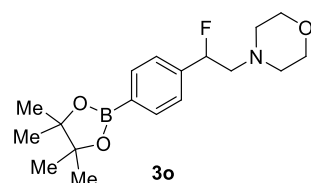
141.73
141.57
134.91
128.36
128.27
124.69
124.63

93.21
91.82
83.89

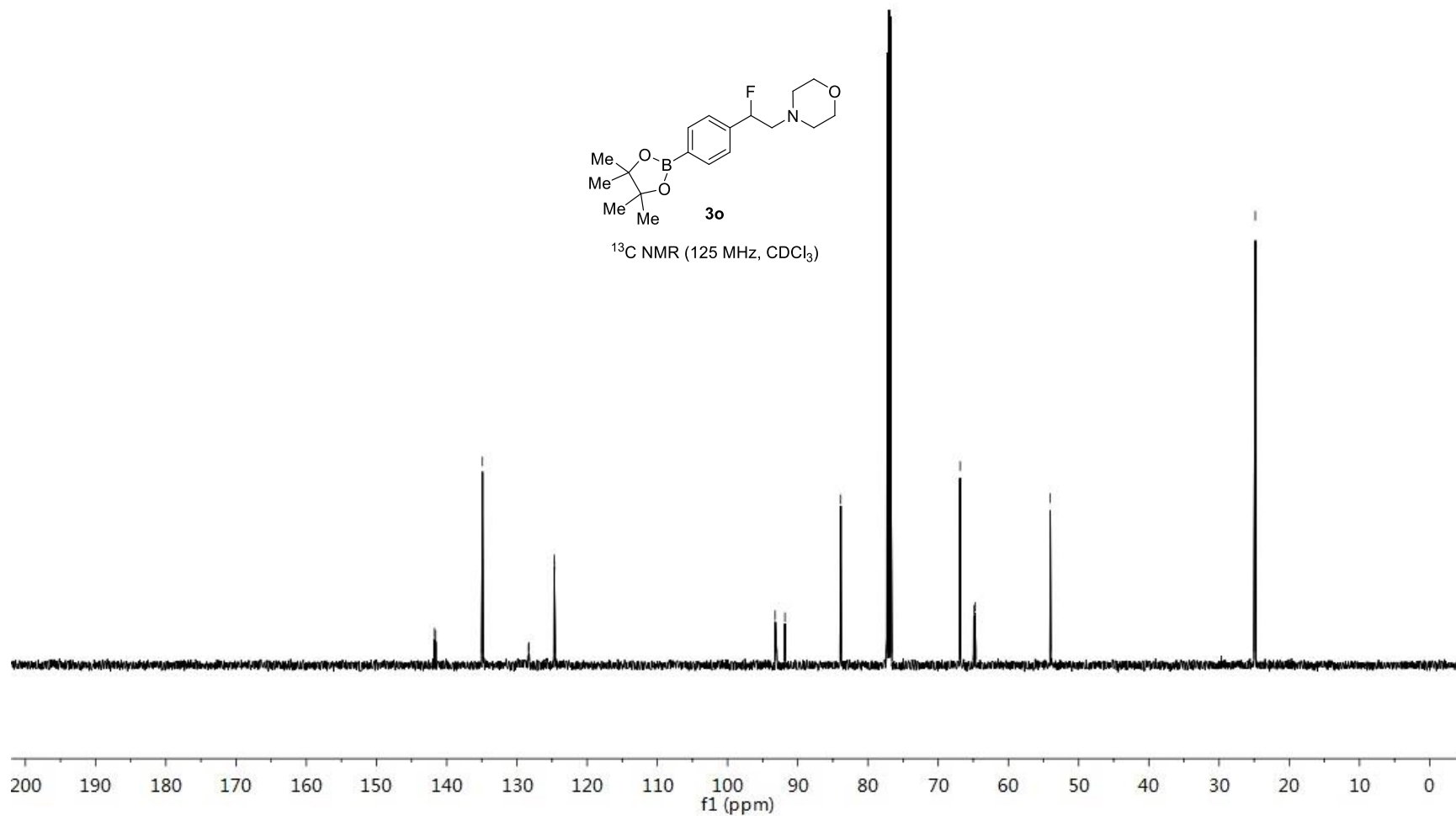
66.88
64.92
64.74
54.03

24.84

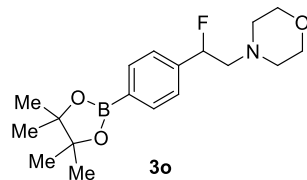
¹³CNMR gf-3-66aPP in CDCl₃



¹³C NMR (125 MHz, CDCl₃)

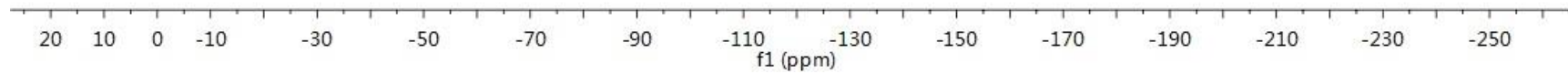


¹⁹F NMR gf-3-66aPP in CDCl₃

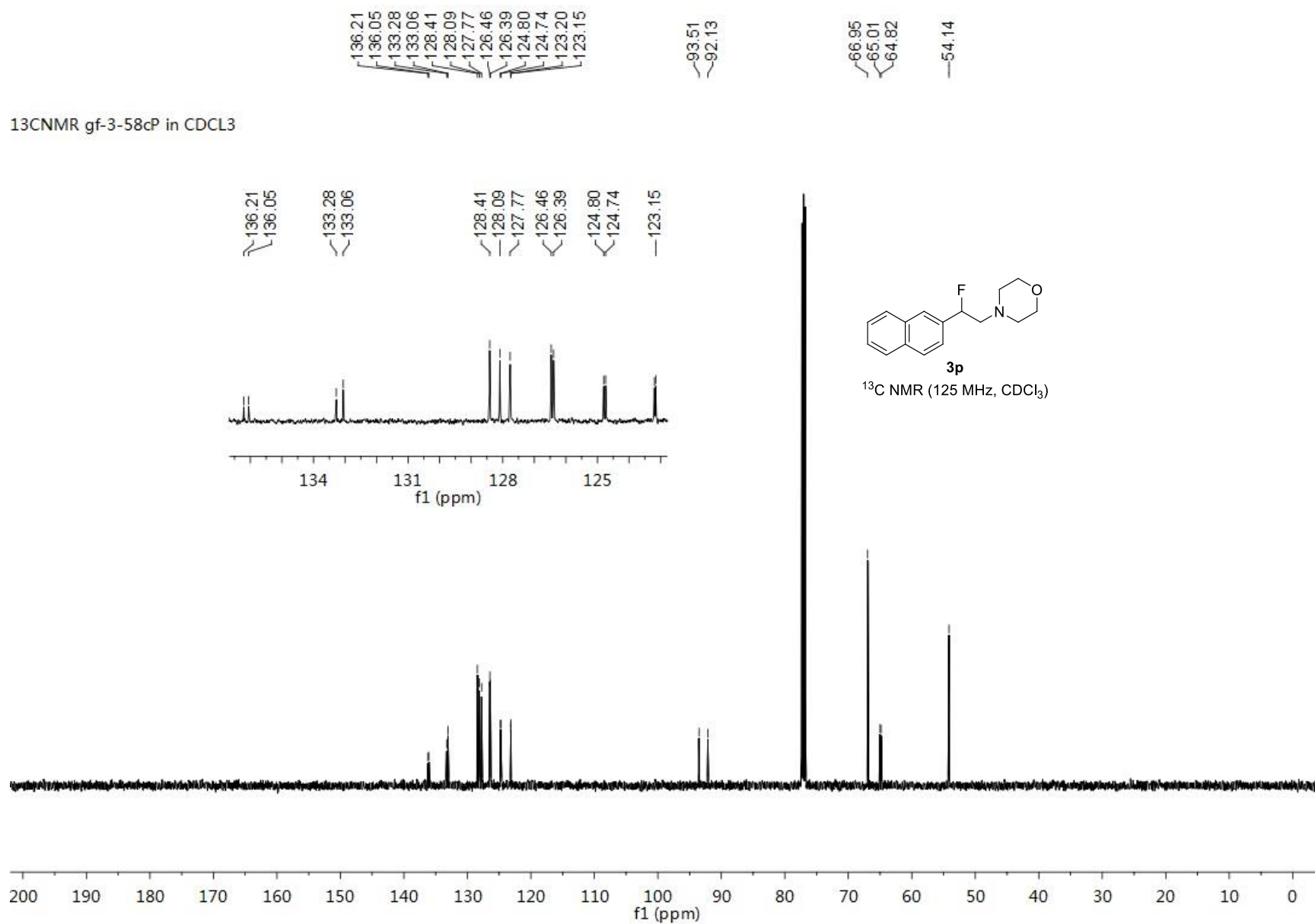


¹⁹F NMR (471 MHz, CDCl₃)

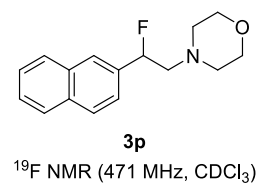
---178.8272



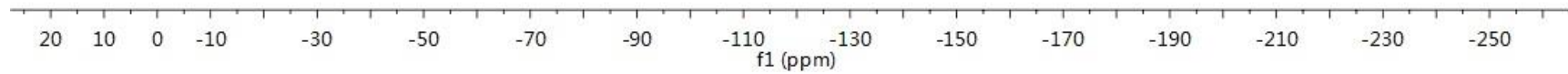
¹³CNMR gf-3-58cP in CDCl₃

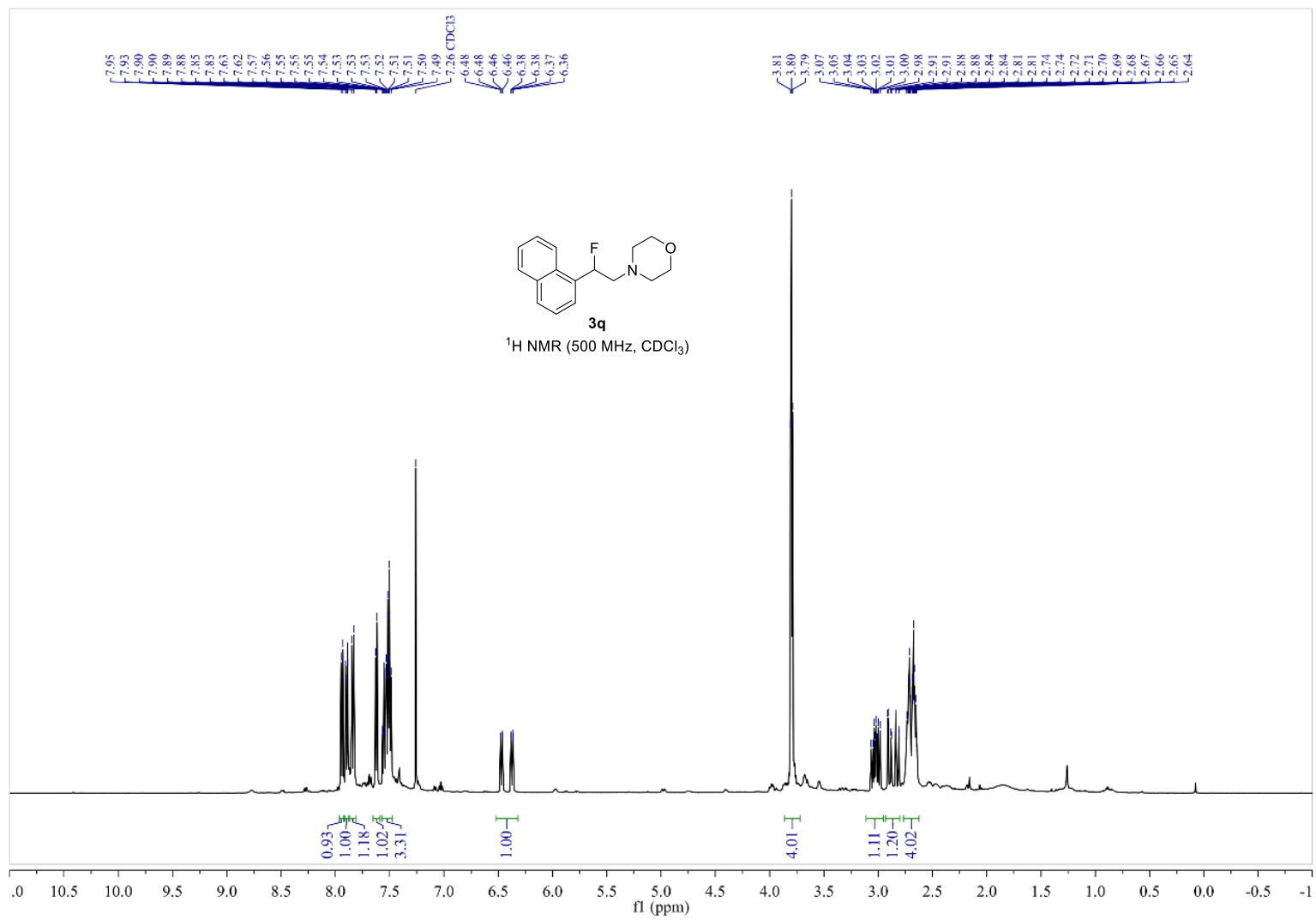


¹⁹F NMR gf-3-58cP in CDCl₃

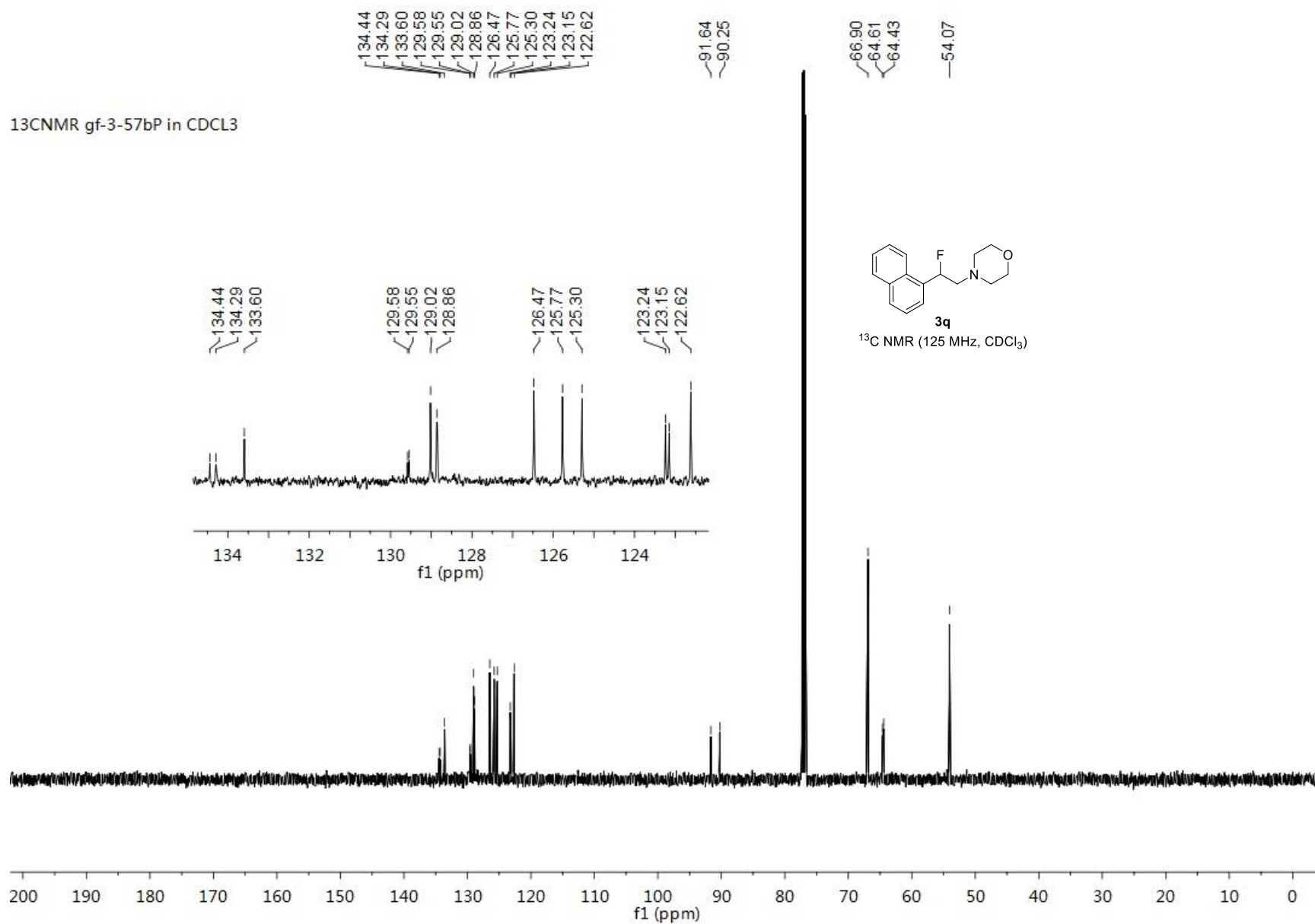


—176.5693

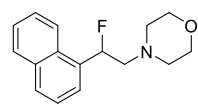




¹³CNMR gf-3-57bP in CDCl₃

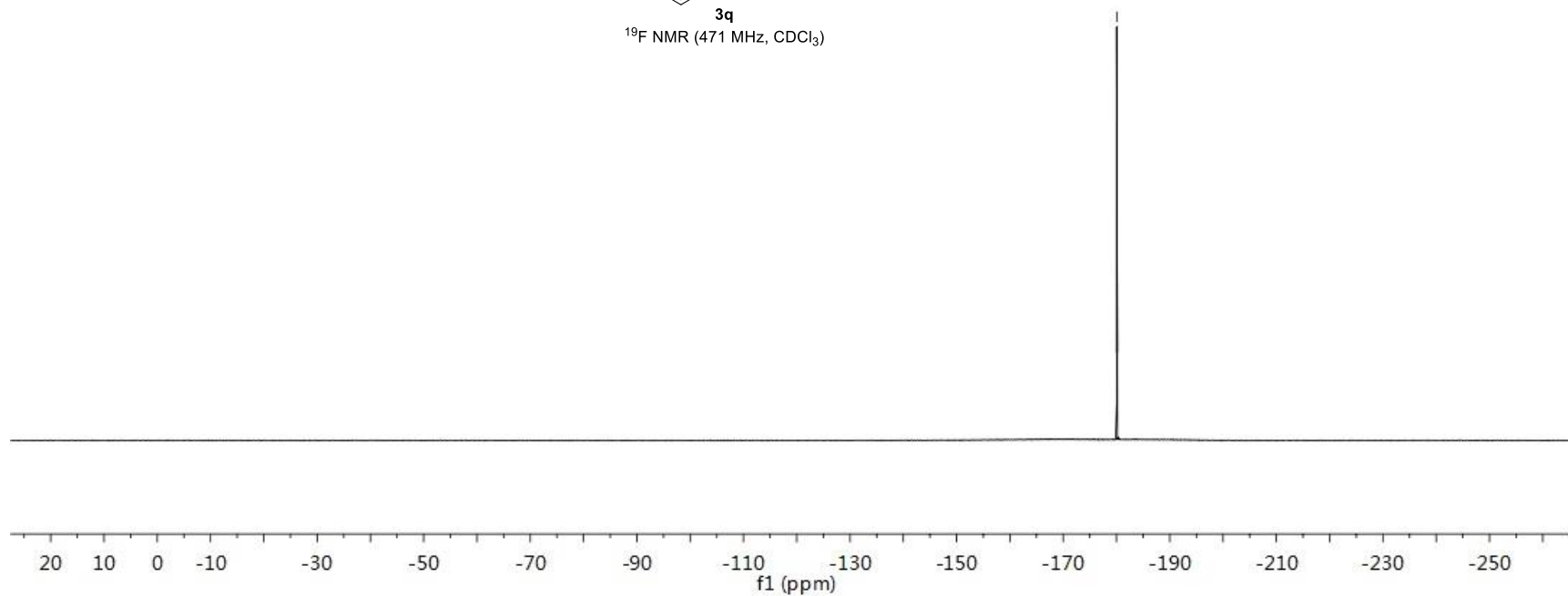


¹⁹F NMR gf-3-57bP in CDCl₃



¹⁹F NMR (471 MHz, CDCl₃)

---180.0677

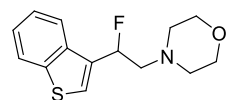


7.8872
7.8724
7.8368
7.8215
7.5024
7.4276
7.4154
7.3976
7.3816
7.3675

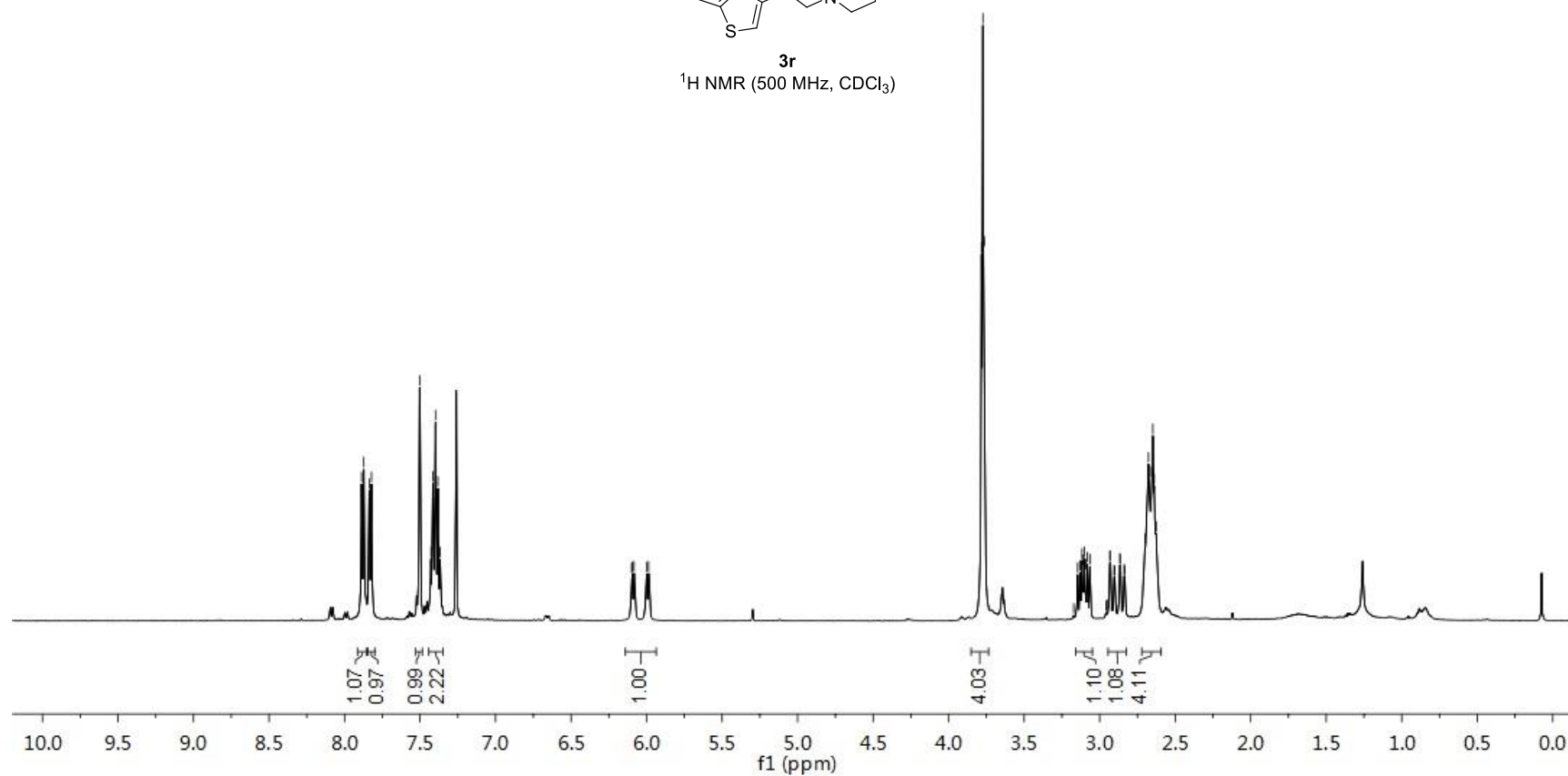
6.0990
6.0955
6.0826
5.9994
5.9864

3.7824
3.7732
3.7640
3.1721
3.1459
3.1291
3.1174
3.1095
3.1006
3.0927
3.0809
3.0642
2.9582
2.9500
2.9328
2.9283
2.9044
2.8998
2.8668
2.8622
2.8382
2.8336
2.6987
2.6769
2.6573
2.6482
2.6391
2.6256

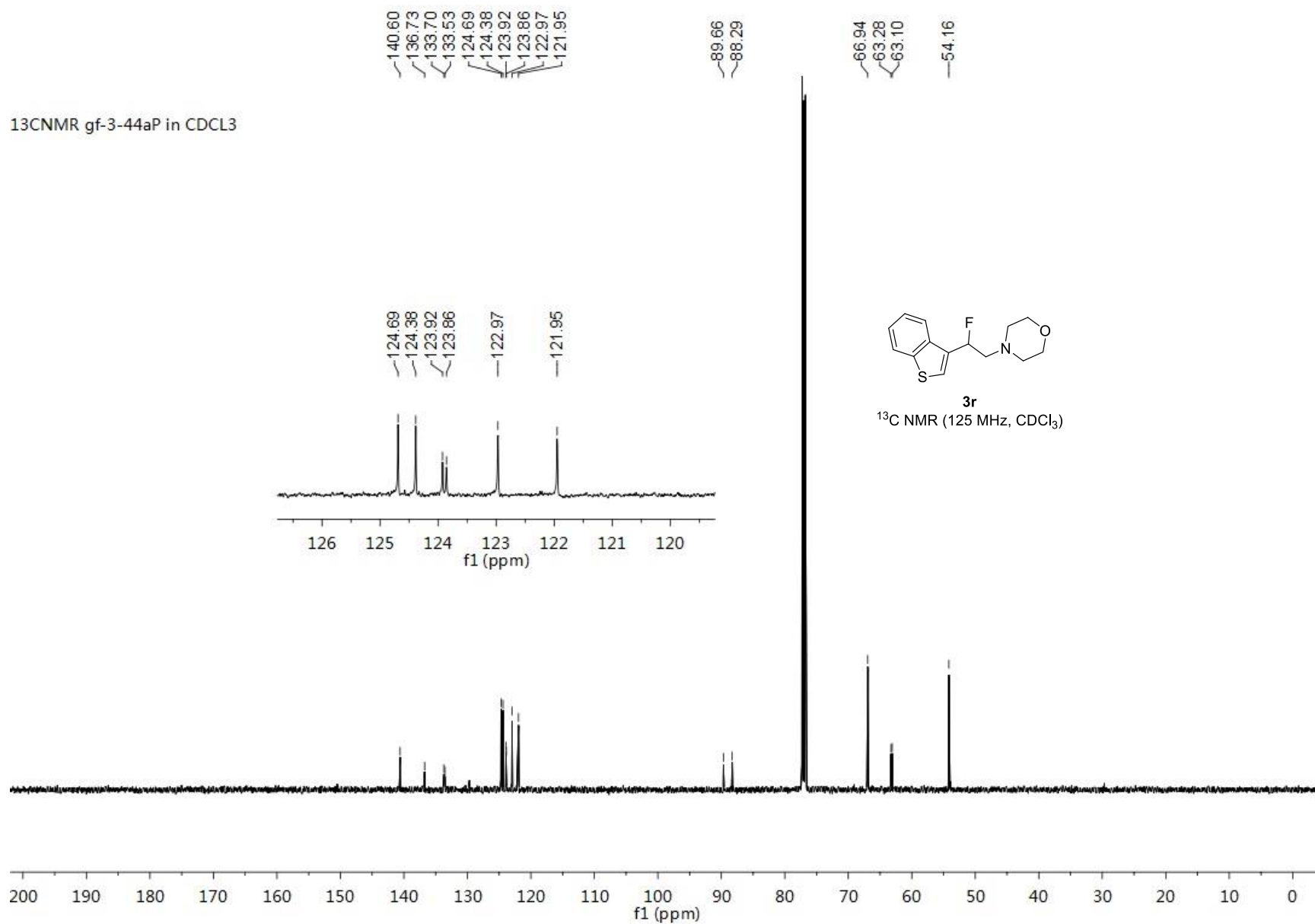
¹HMR gf-3-44aP in CDCl₃



3r
¹H NMR (500 MHz, CDCl₃)

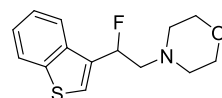


¹³CNMR gf-3-44aP in CDCl₃



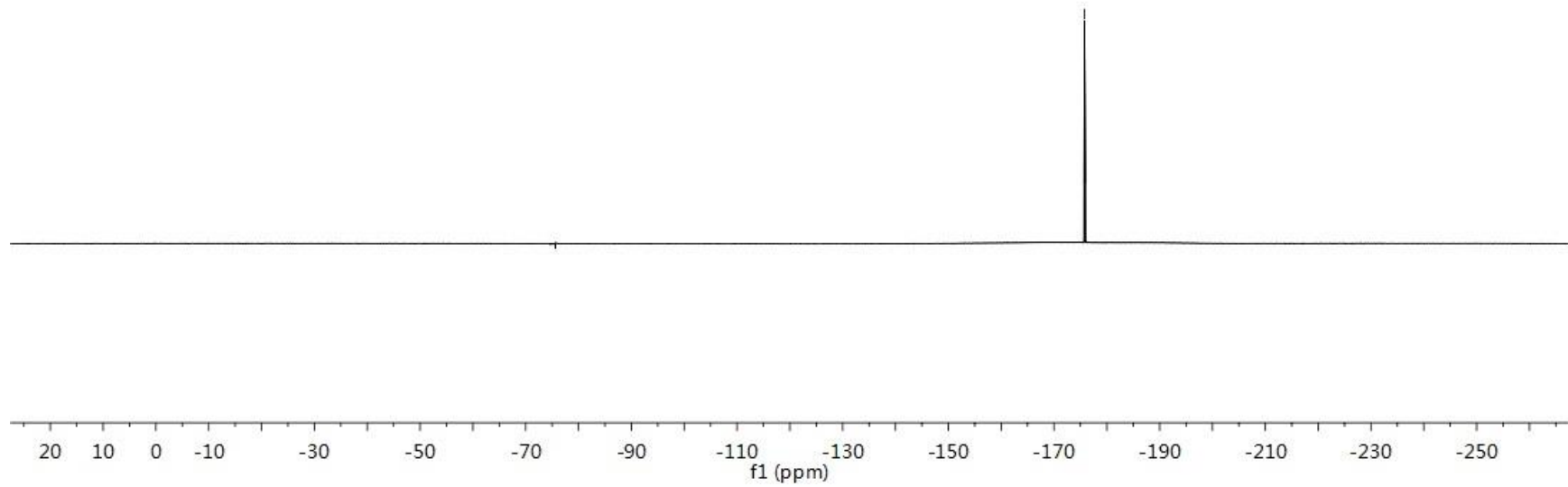
¹⁹F NMR gf-3-44aP in CDCl₃

—175.7704



3r

¹⁹F NMR (471 MHz, CDCl₃)

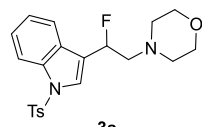


7.9892
7.9725
7.7845
7.7679
7.6359
7.6311
7.6079
7.5922
7.3576
7.3425
7.3273
7.2722
7.2599
7.2440
7.2281

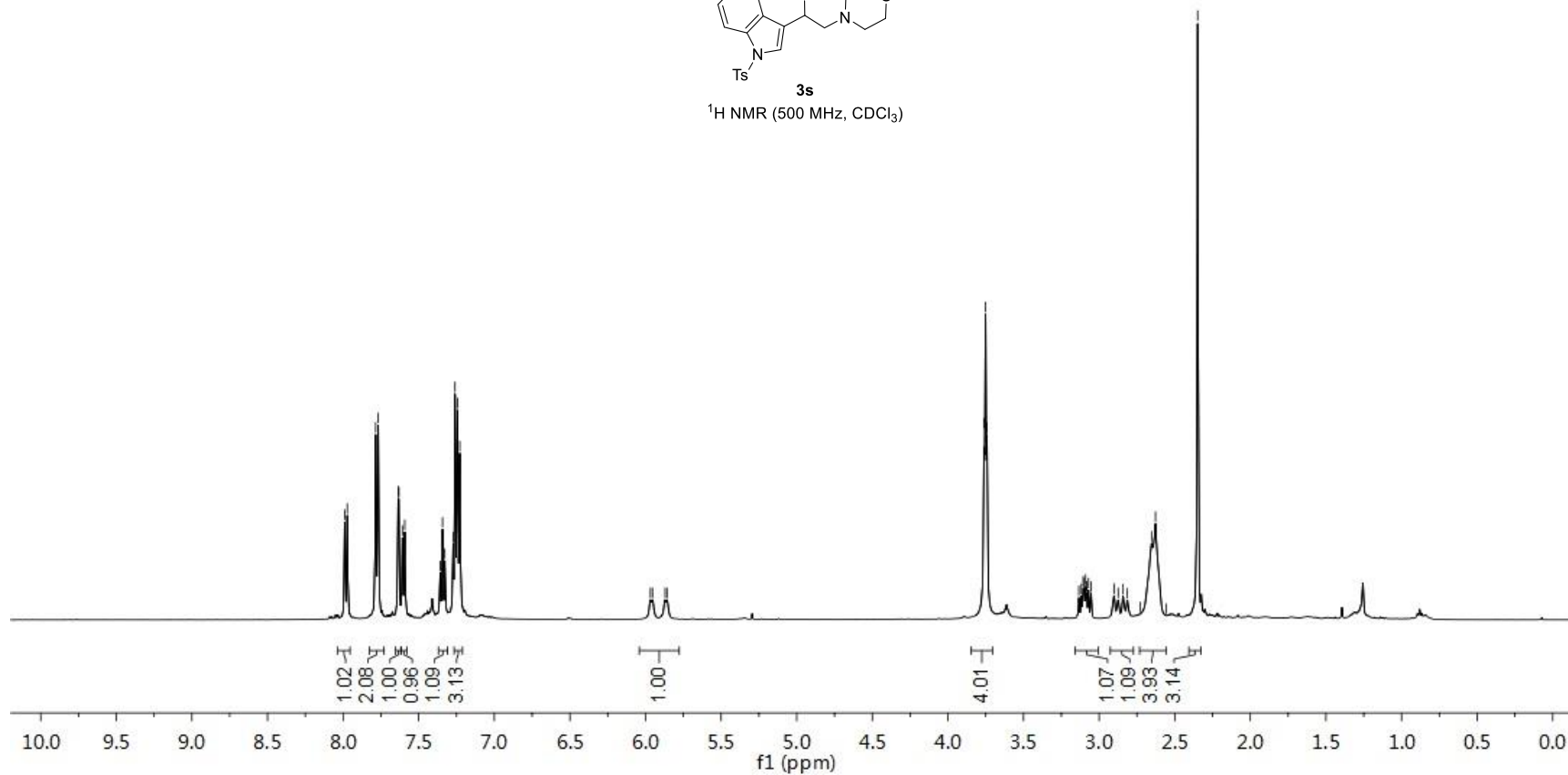
5.9673
5.9539
5.8703
5.8575

3.7599
3.7510
3.7420
3.1341
3.1177
3.1058
3.0983
3.0895
3.0821
3.0701
3.0537
2.9016
2.8742
2.8418
2.8132
2.7297
2.6531
2.6274
2.5564
2.3484

¹H NMR of 3-43cP in CDCl₃



¹H NMR (500 MHz, CDCl₃)



¹³C NMR gf-3-43cP in CDCl₃

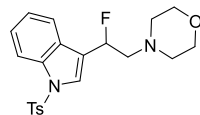
145.27
135.15
135.09
130.01
126.92
125.17
123.78
123.70
123.49
119.76

87.86
86.51

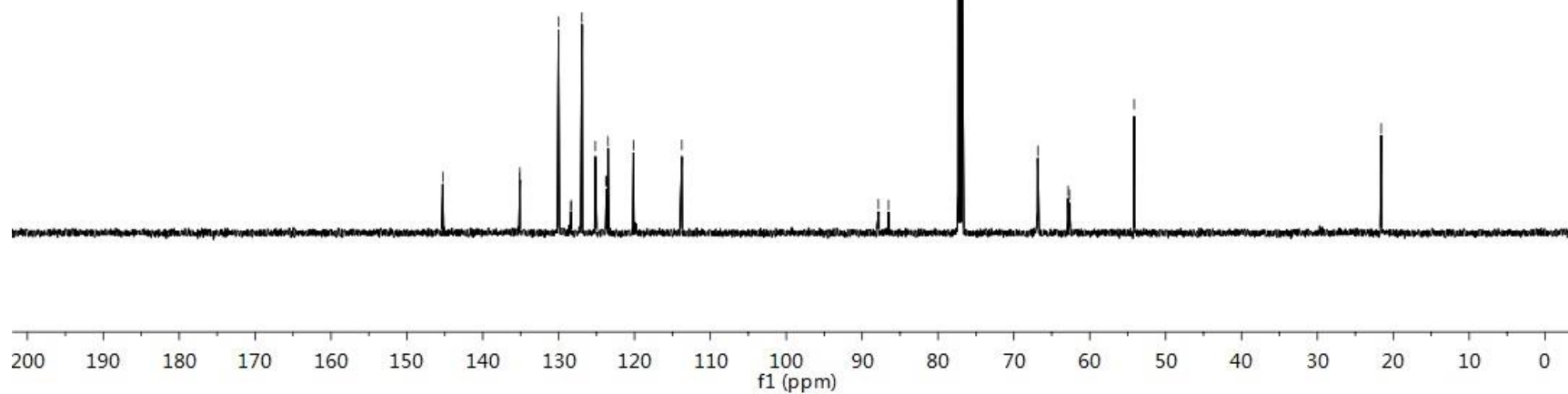
66.85
62.87
62.69

54.14

21.60

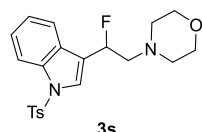


¹³C NMR (125 MHz, CDCl₃)

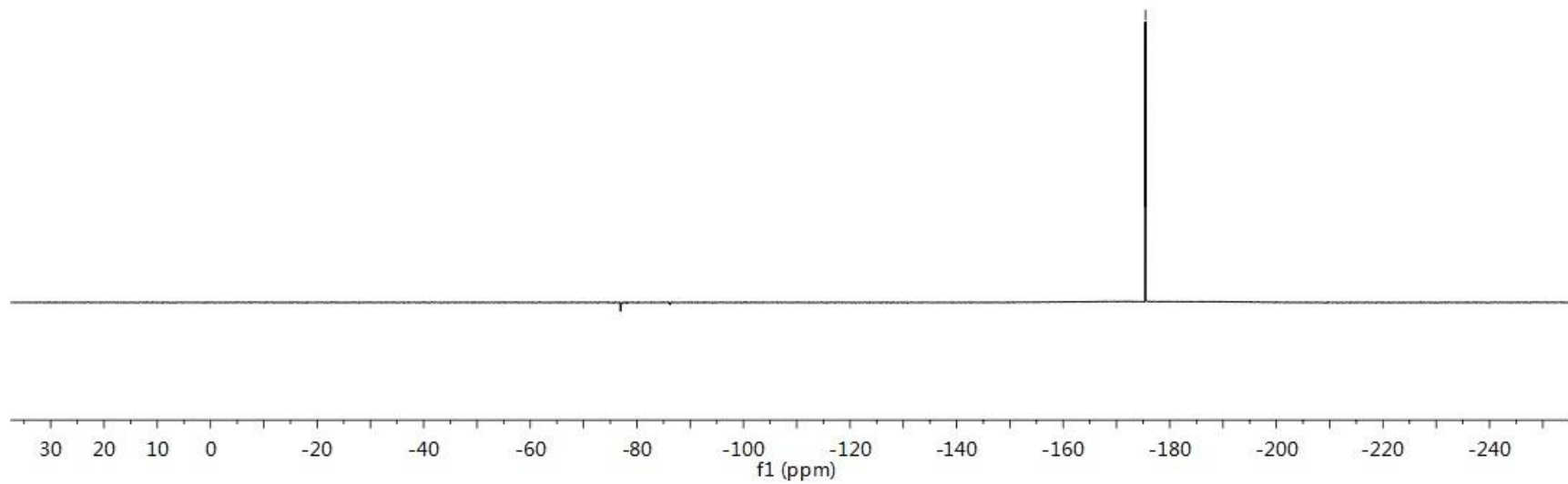


¹⁹F NMR gf-3-43cP in CDCl₃

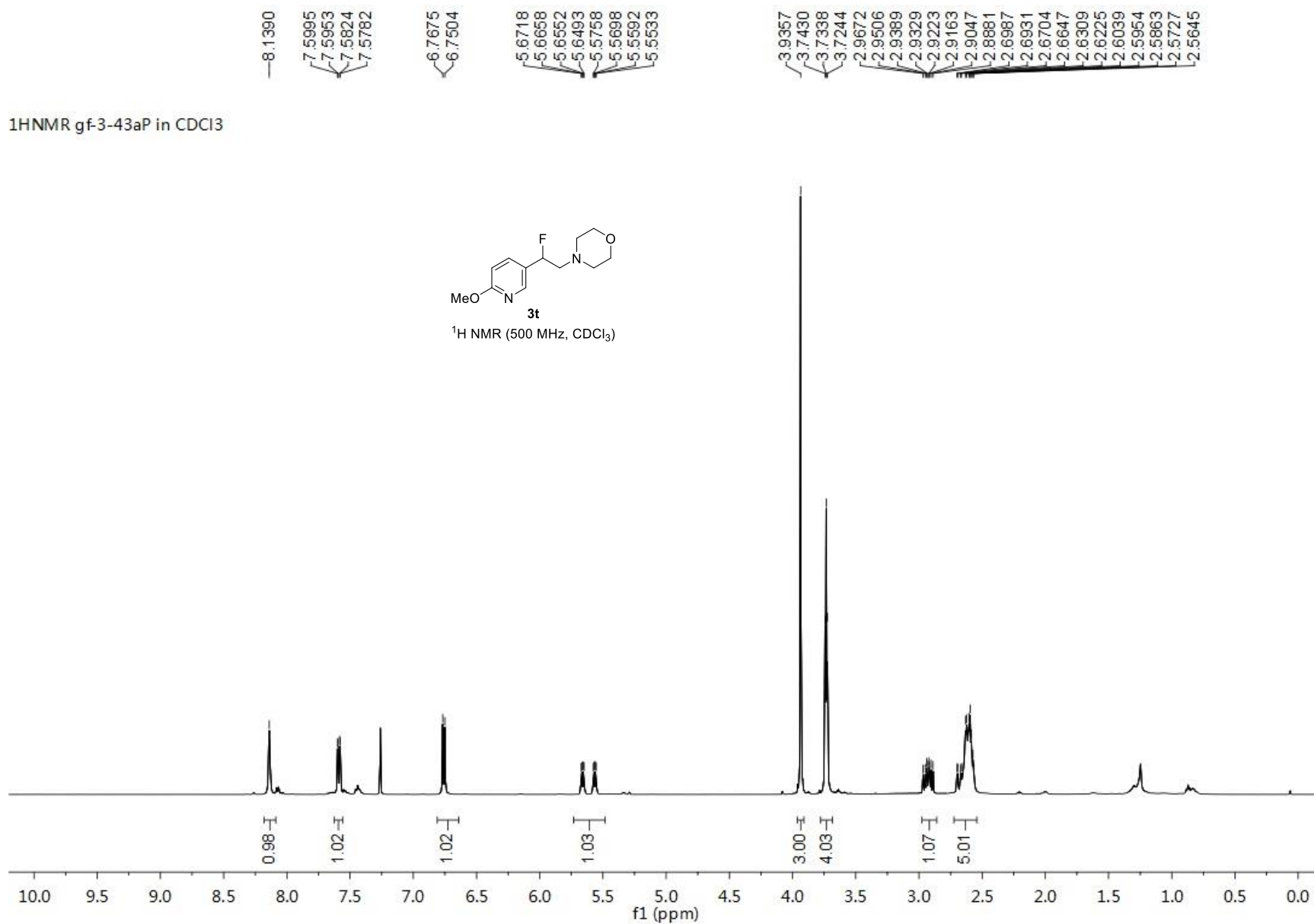
---175.4343



¹⁹F NMR (471 MHz, CDCl₃)

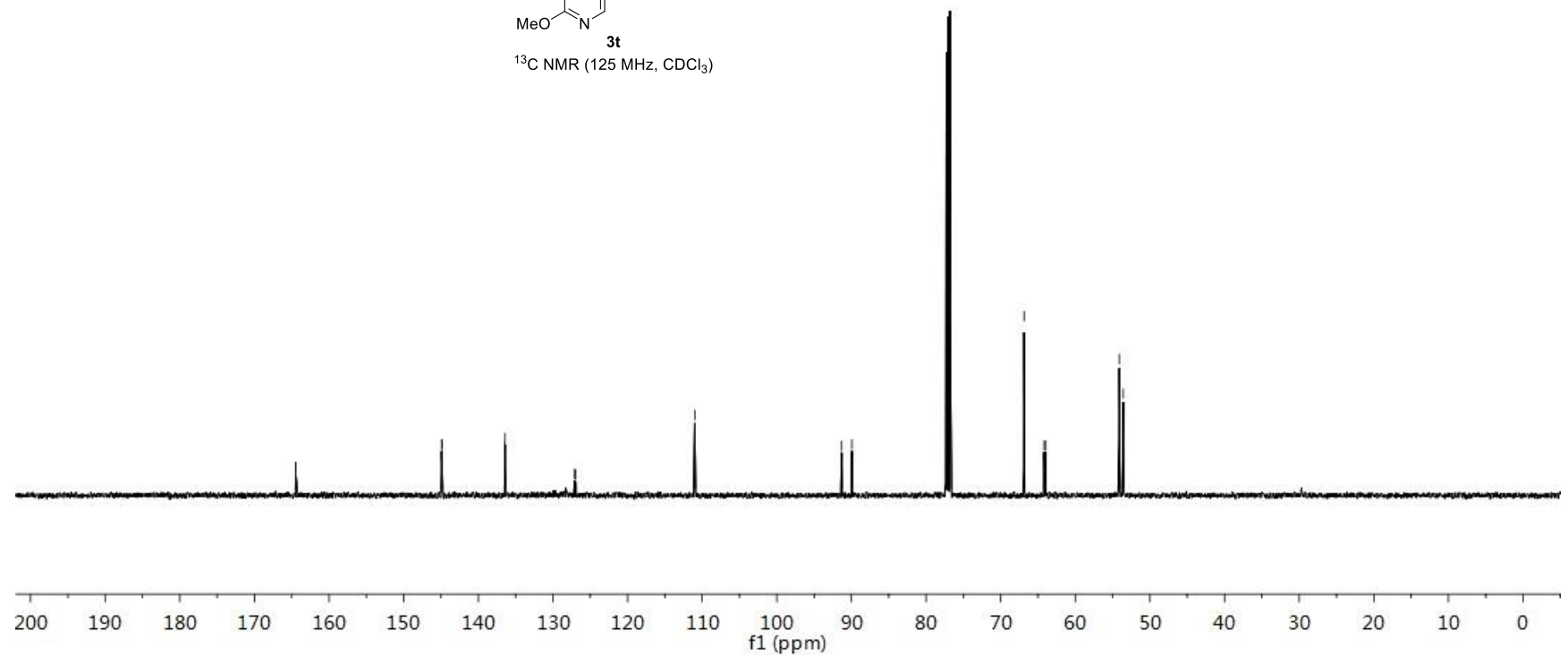
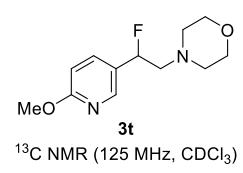


S115

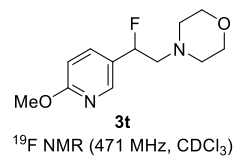


164.48
164.47
144.92
144.86
136.46
136.42
127.10
126.93
110.99
91.31
89.94
66.86
64.20
64.01
54.11
53.59

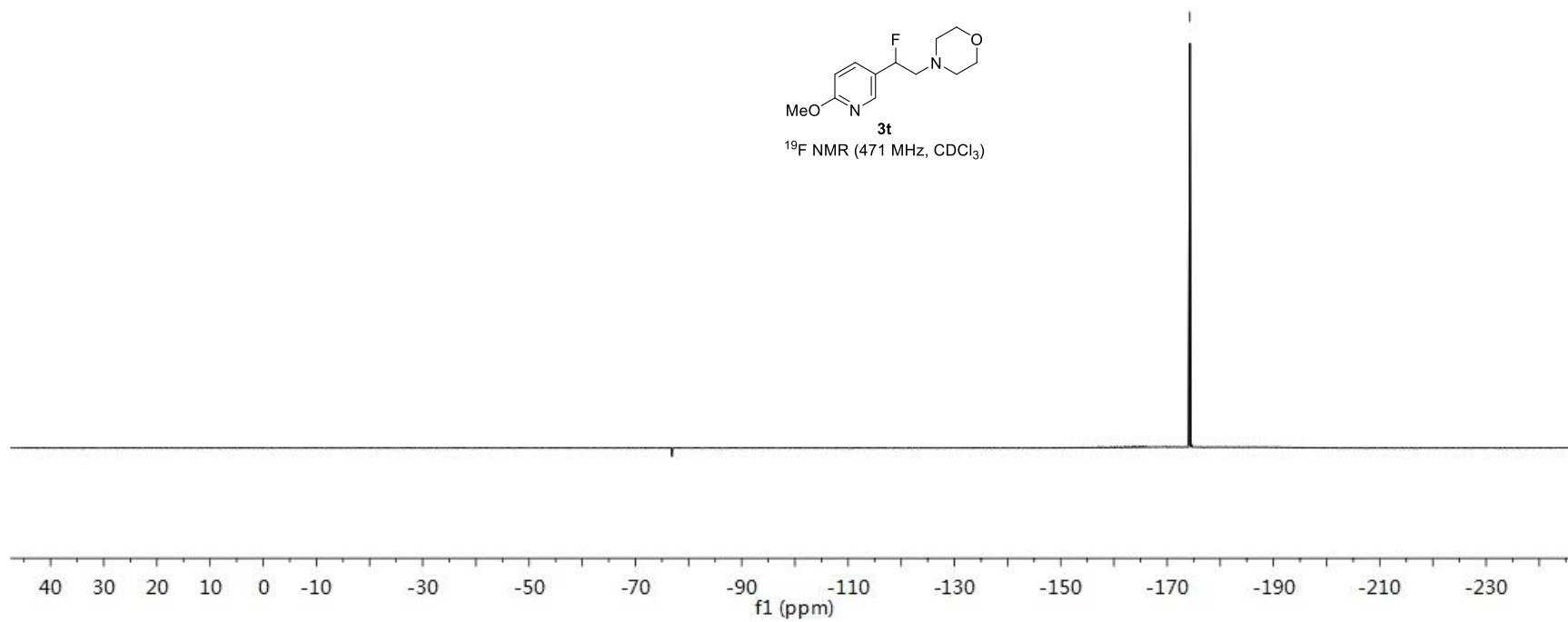
¹³CNMR gf-3-43aP in CDCl₃



¹⁹F NMR gf-3-43aP in CDCl₃

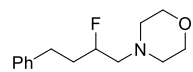


—174.2810

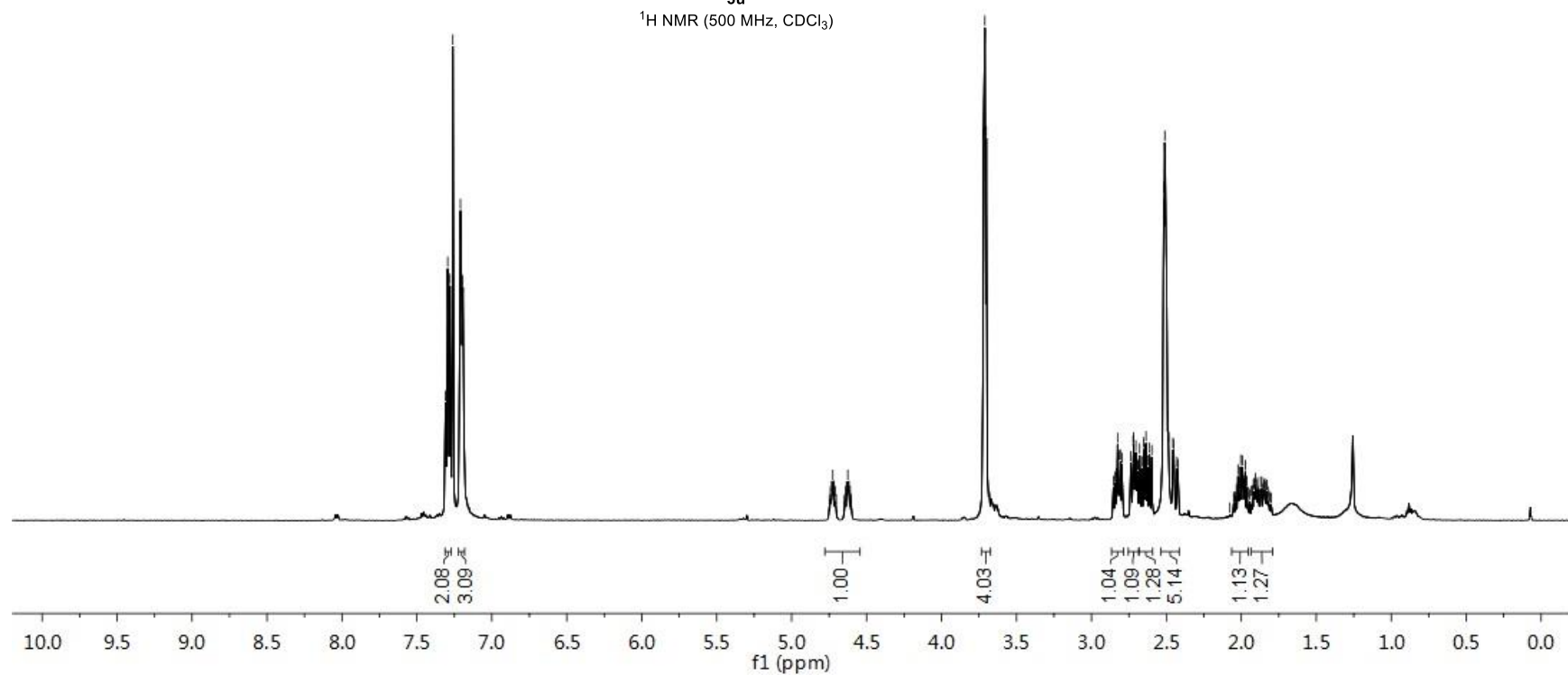


7.3092
7.2941
7.2791
7.2602
7.2092
7.2009
7.1933
4.7336
4.7255
4.7173
4.6339
4.6255
4.6174
3.7204
3.7111
3.7017
3.7017
2.8538
2.8433
2.8345
2.8254
2.8157
2.8069
2.7964
2.7369
2.7222
2.7184
2.7090
2.7039
2.6946
2.6909
2.6784
2.6638
2.6510
2.6399
2.6361
2.6251
2.6123
2.5973
2.5173
2.5107
2.5006
2.4897
2.4839
2.4561
2.4505
2.4283
2.4227
2.0331
2.0225
2.0120
2.0044
1.9917
1.9866
1.9810
1.9734
1.9629
1.9255
1.9187
1.9136
1.9113
1.9059
1.8994
1.8913
1.8706
1.8631
1.8560
1.8487
1.8432
1.8366
1.8287

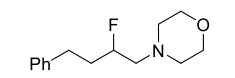
¹H NMR of 3-139dP in CDCl₃



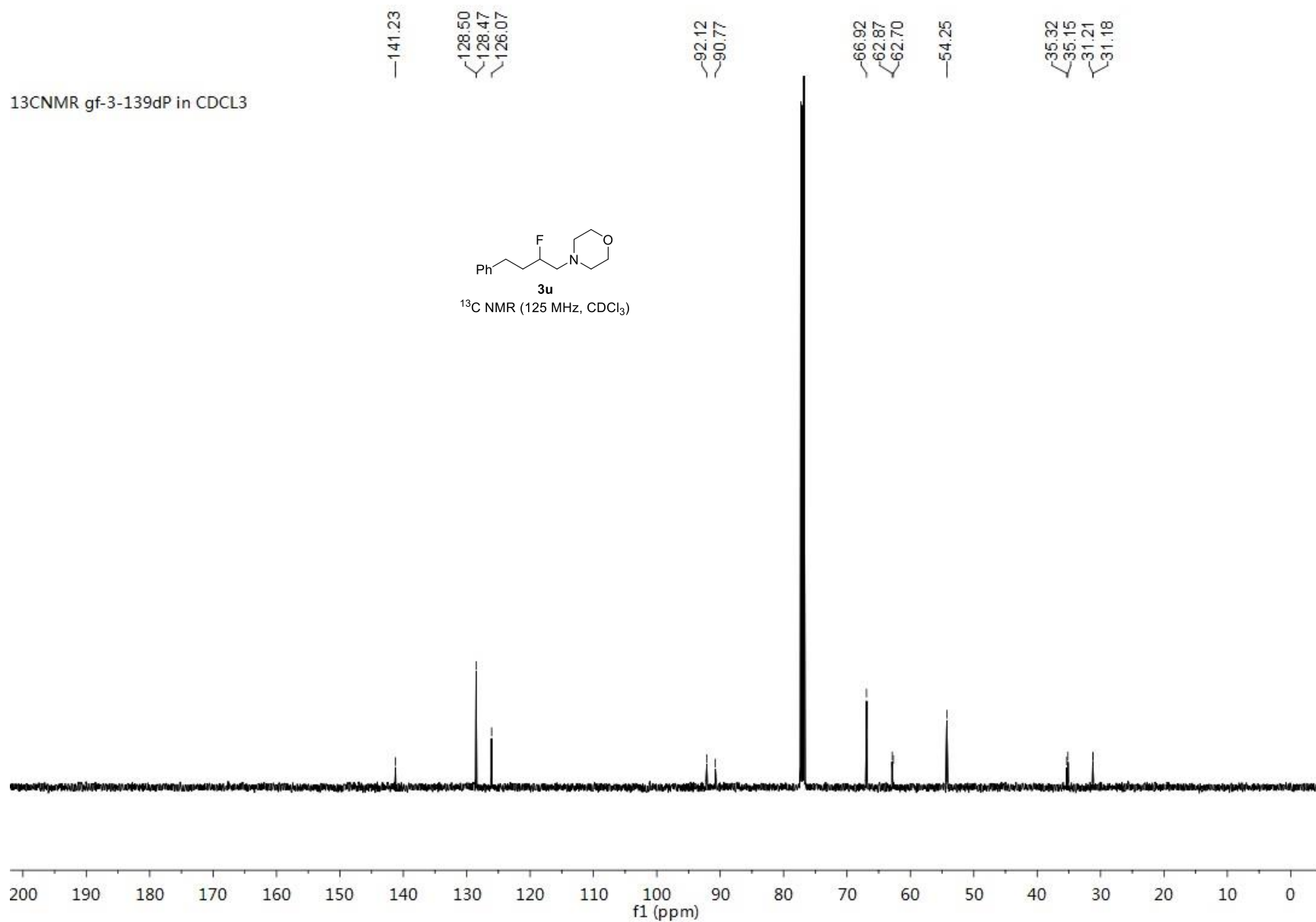
3u
¹H NMR (500 MHz, CDCl₃)



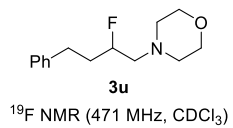
¹³CNMR gf-3-139dP in CDCl₃



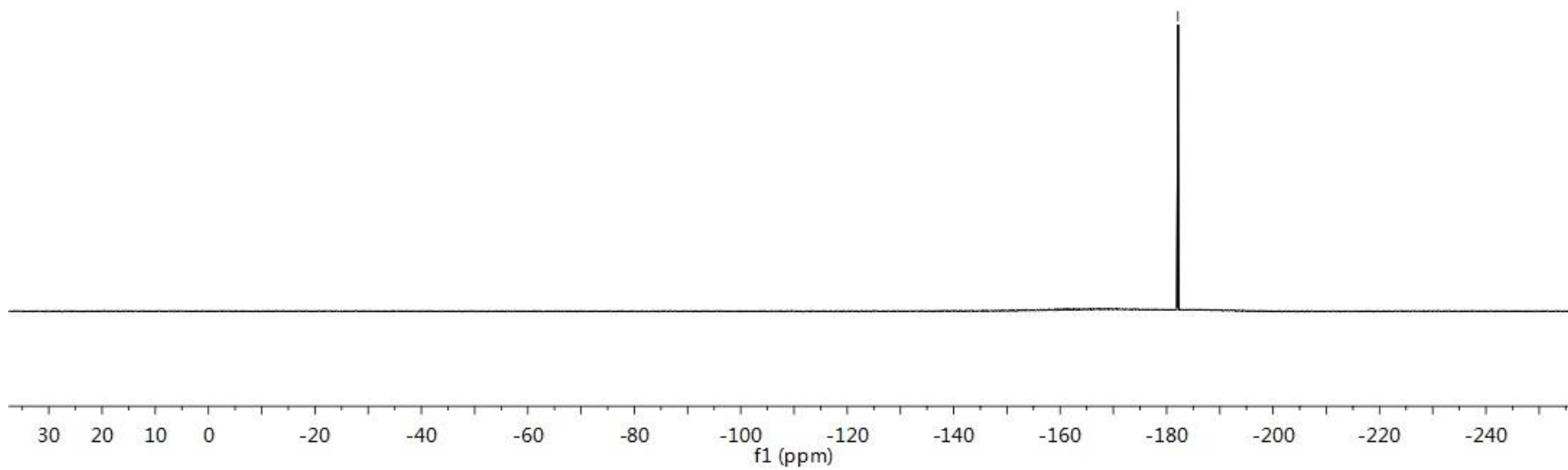
¹³C NMR (125 MHz, CDCl₃)



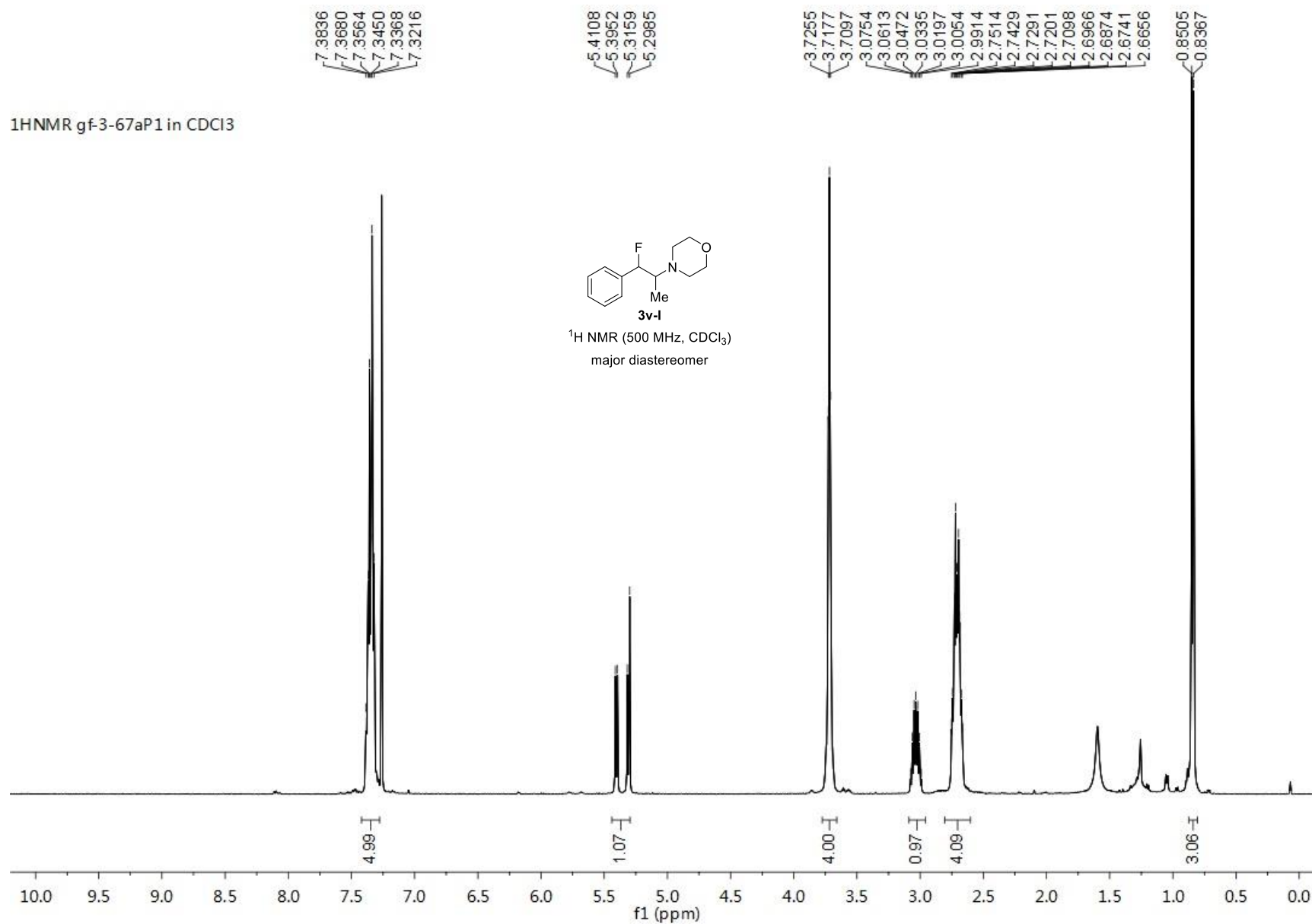
¹⁹F NMR gf-3-139dP in CDCl₃



— -182.1342



¹H NMR of **3v-I** in CDCl₃



¹³C NMR gf-4-66cP1 in CDCl₃

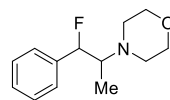
138.62
138.46
128.52
128.50
128.33
126.70
126.65

96.46
95.06

67.59
63.56
63.39

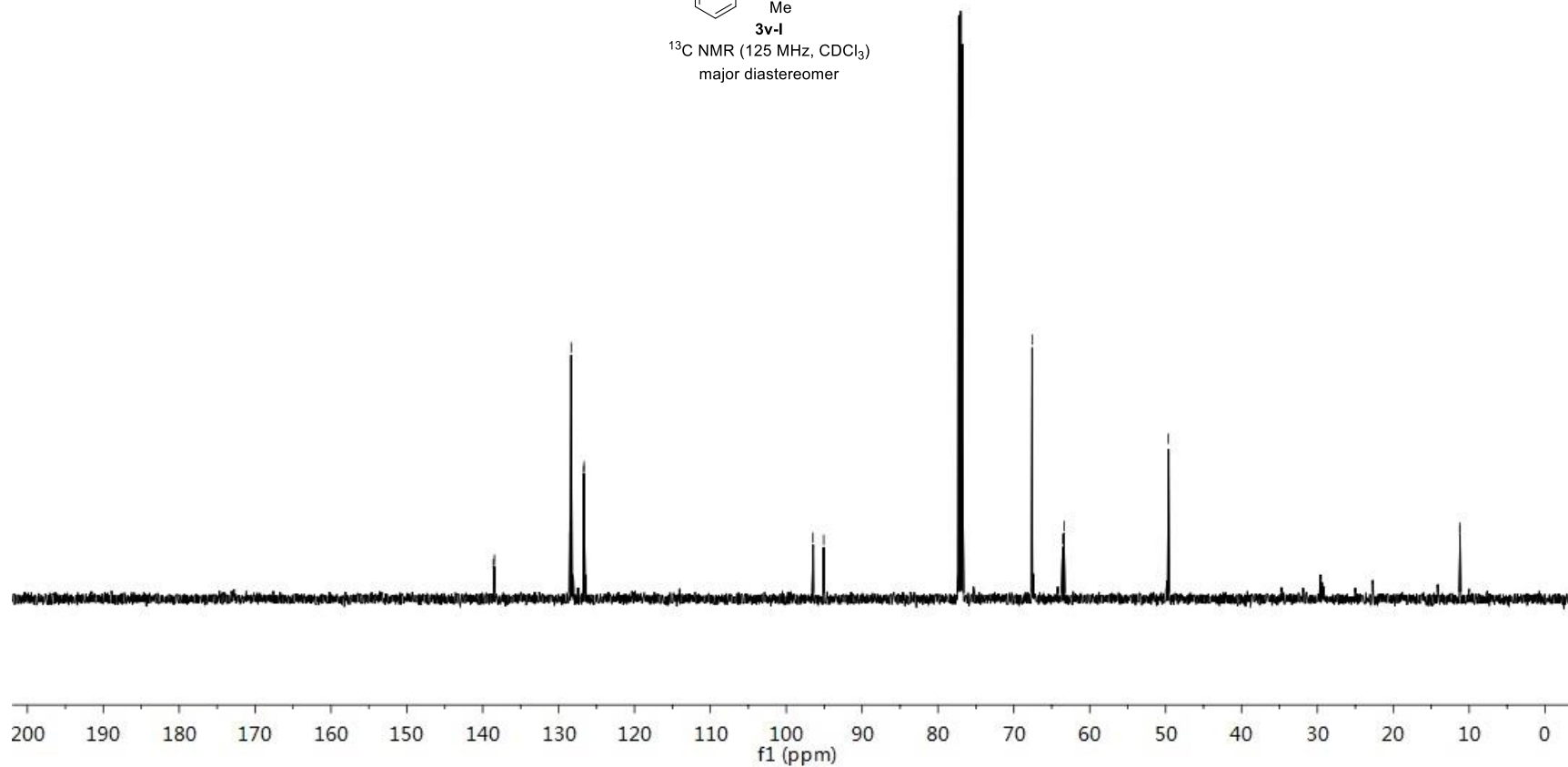
49.64

11.25
11.20

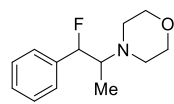


3v-I

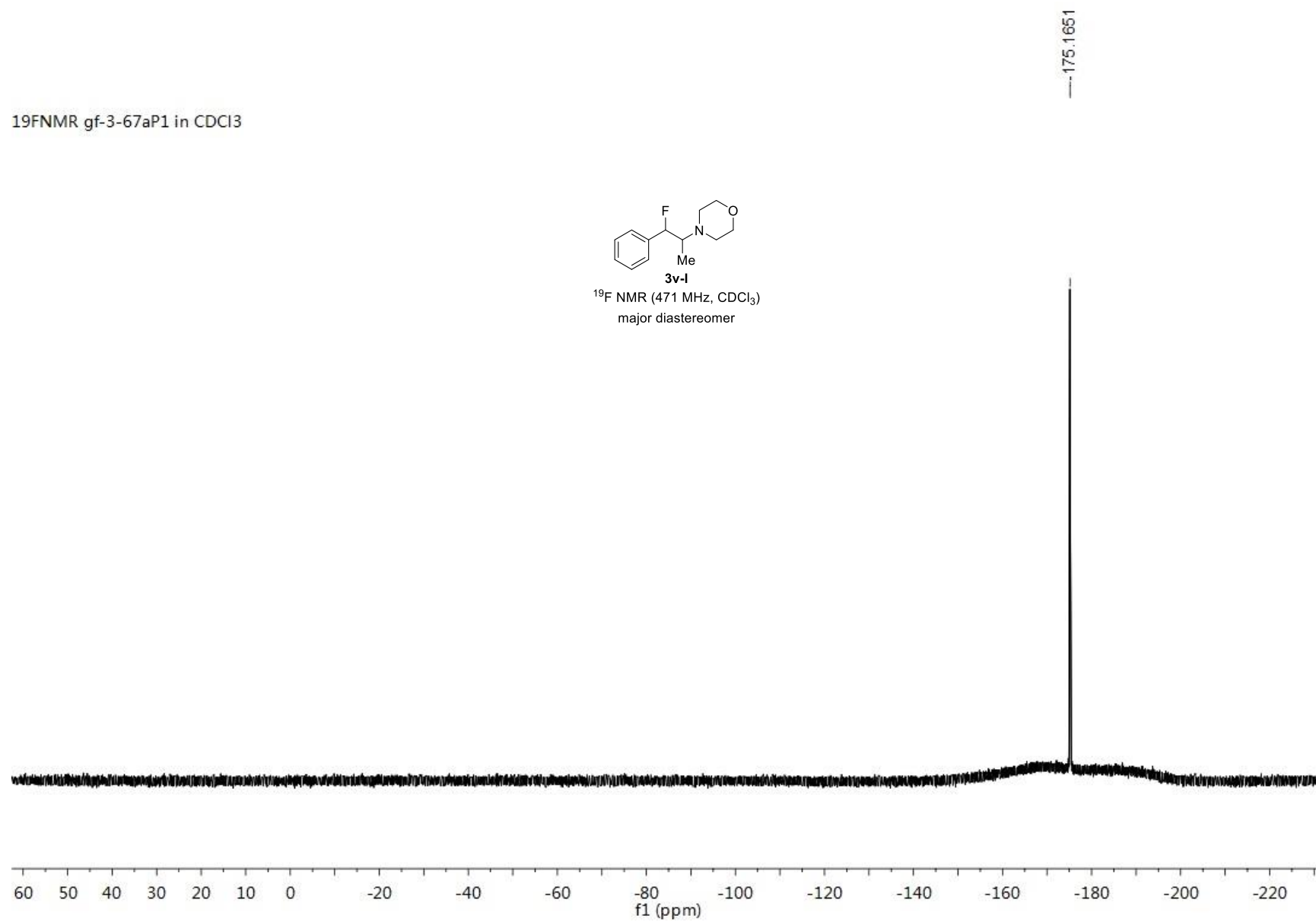
¹³C NMR (125 MHz, CDCl₃)
major diastereomer



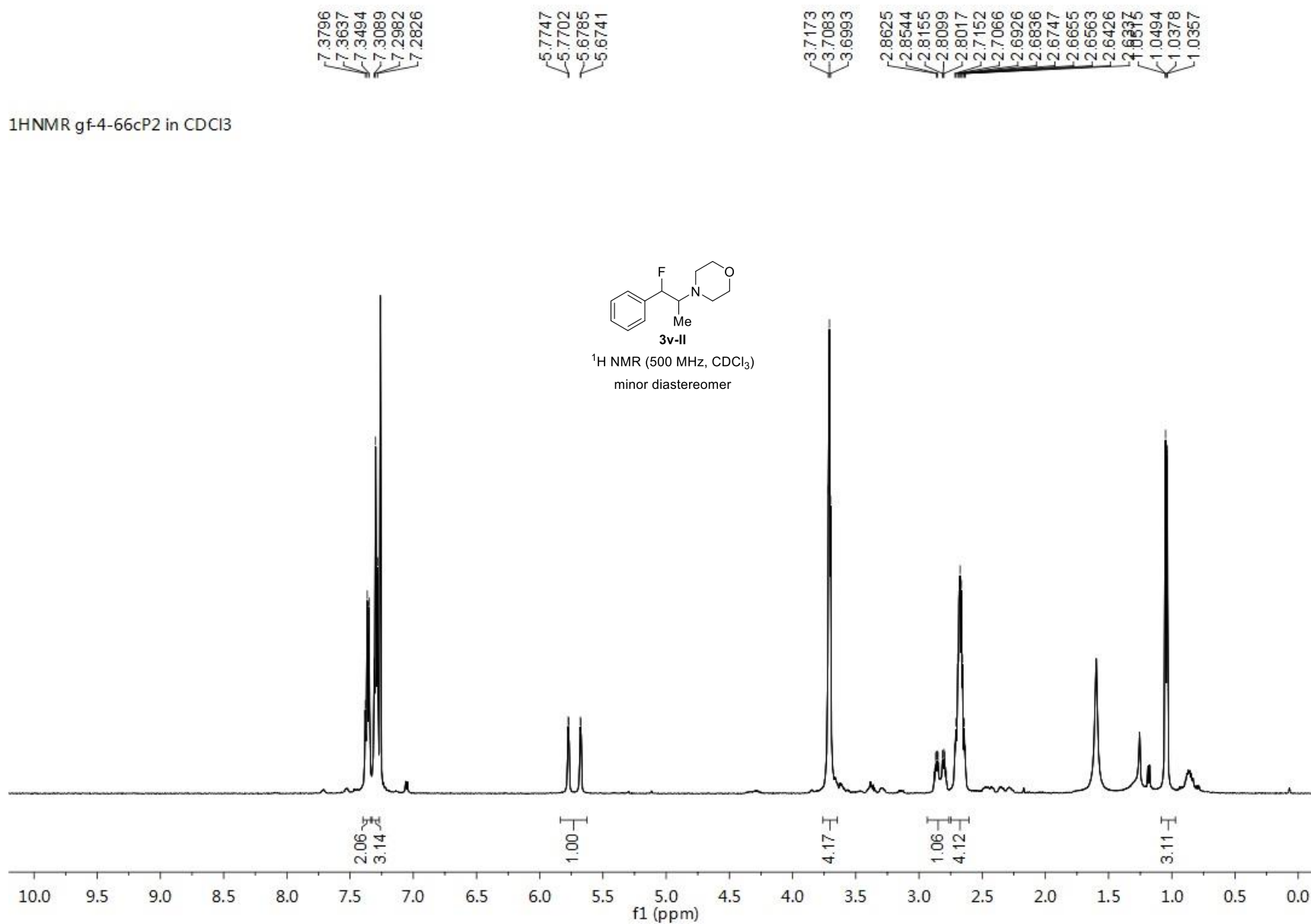
¹⁹F NMR gf-3-67aP1 in CDCl₃



¹⁹F NMR (471 MHz, CDCl₃)
major diastereomer

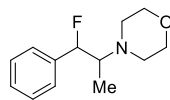


¹H NMR of gf-4-66cP2 in CDCl₃



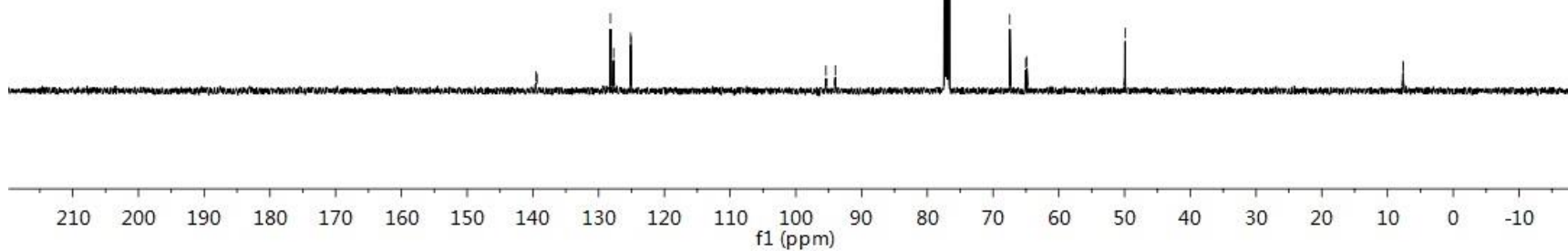
¹³CNMR gf-4-66cP2 in CDCl₃

139.54
139.37
128.21
127.72
125.13
125.07
95.41
93.99
67.47
65.02
64.85
49.93
7.65
7.60

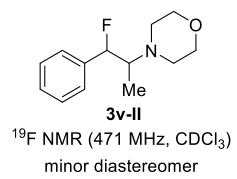


3v-II

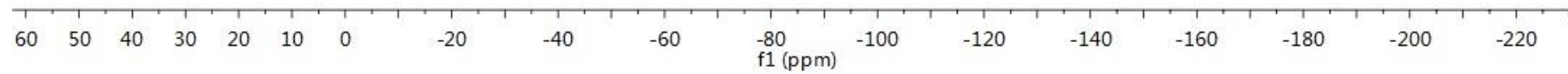
¹³C NMR (125 MHz, CDCl₃)
minor diastereomer



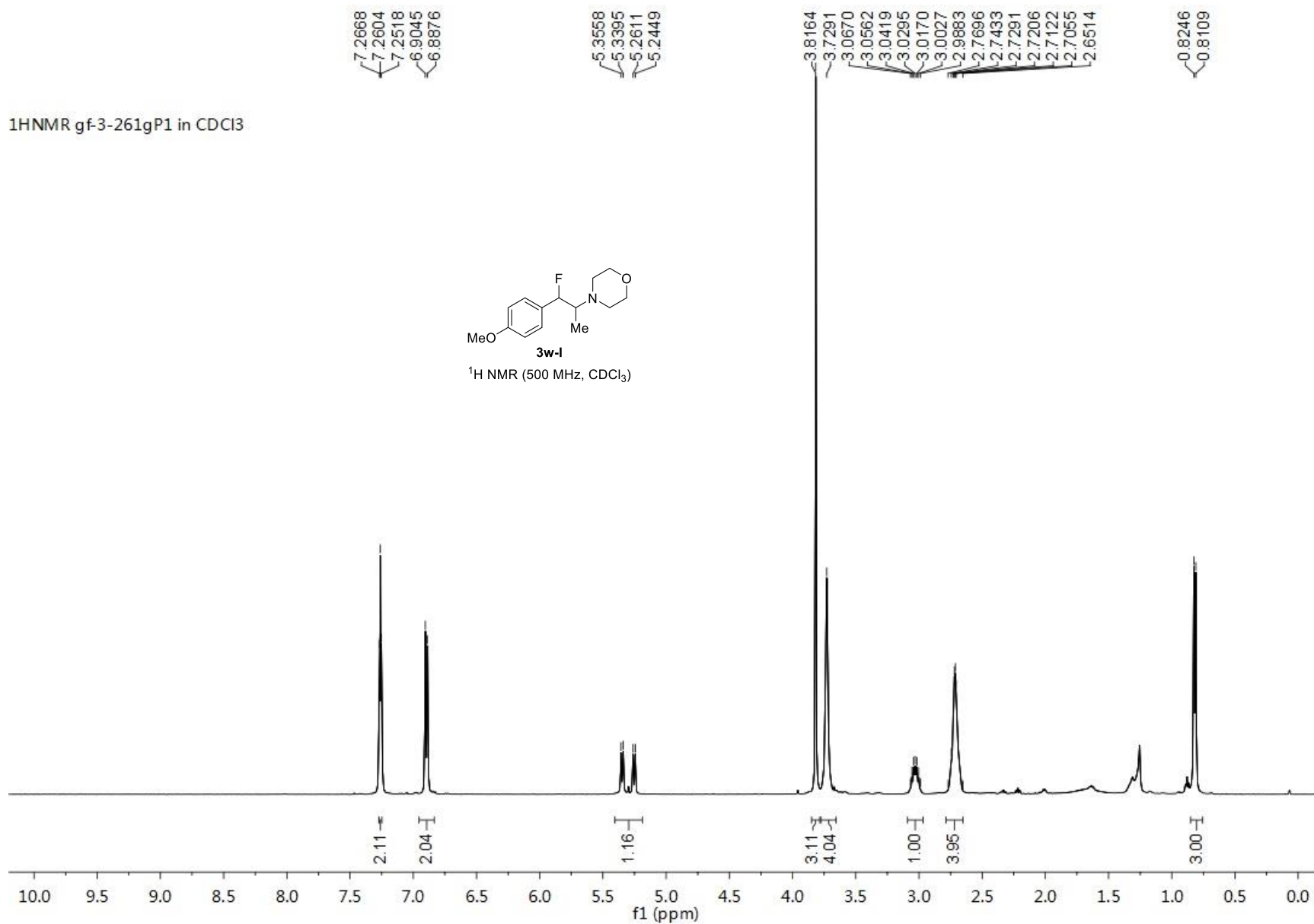
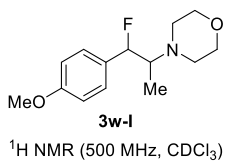
¹⁹F NMR gf-4-66cP2 in CDCl₃

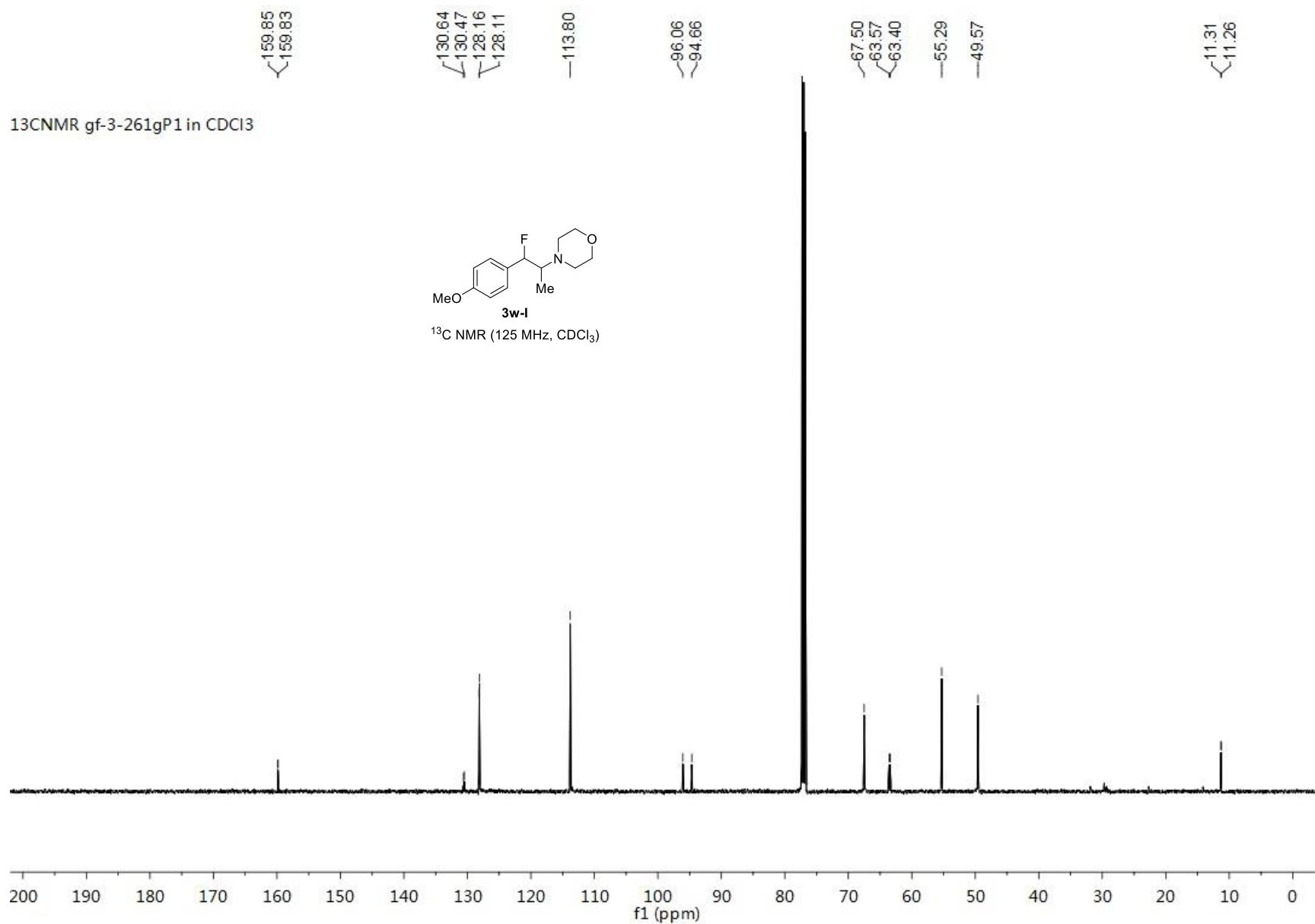


---193.6840

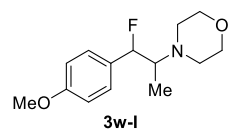


¹H NMR of compound 3w-I in CDCl₃



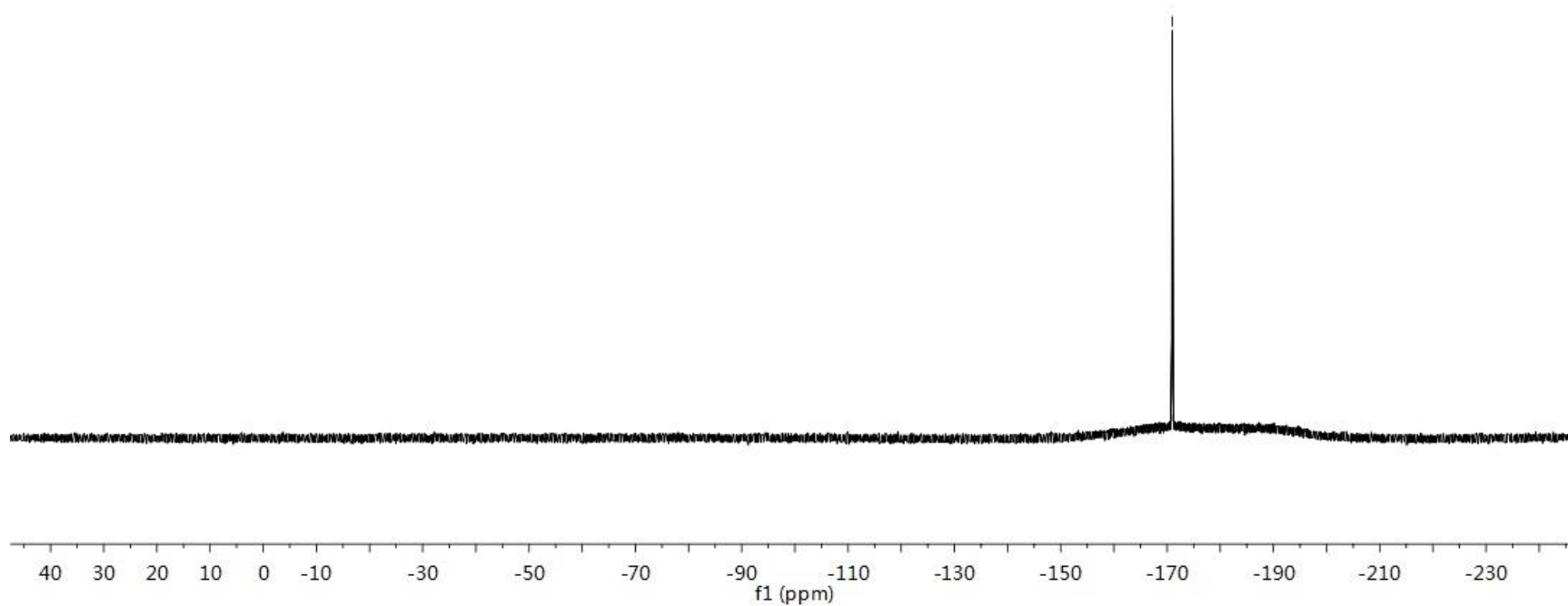


¹⁹F NMR gf-3-261gP1 in CDCl₃



¹⁹F NMR (471 MHz, CDCl₃)

---170.9617



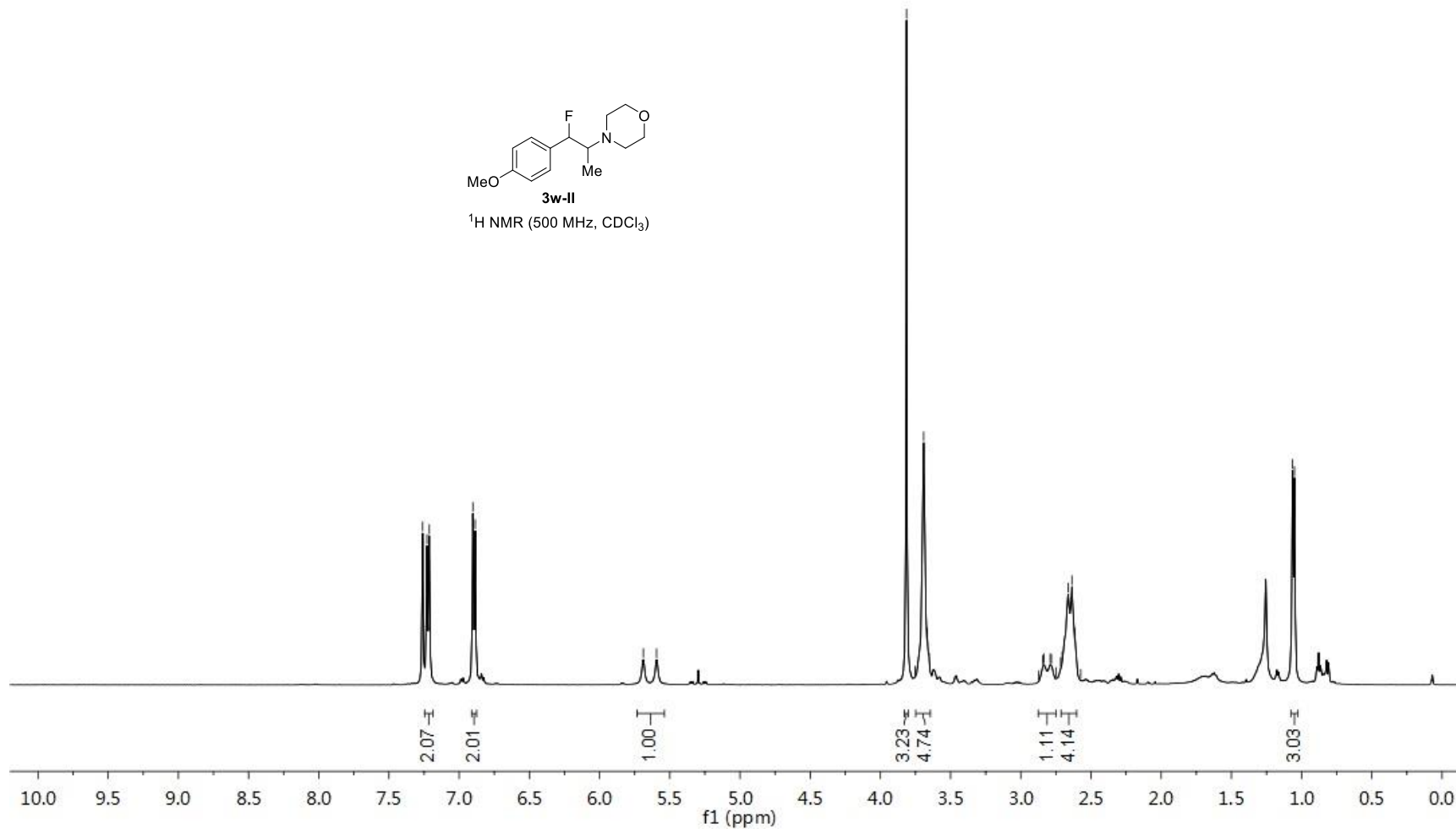
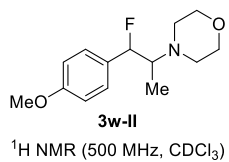
7.2603
7.2297
7.2128
6.9023
6.8853

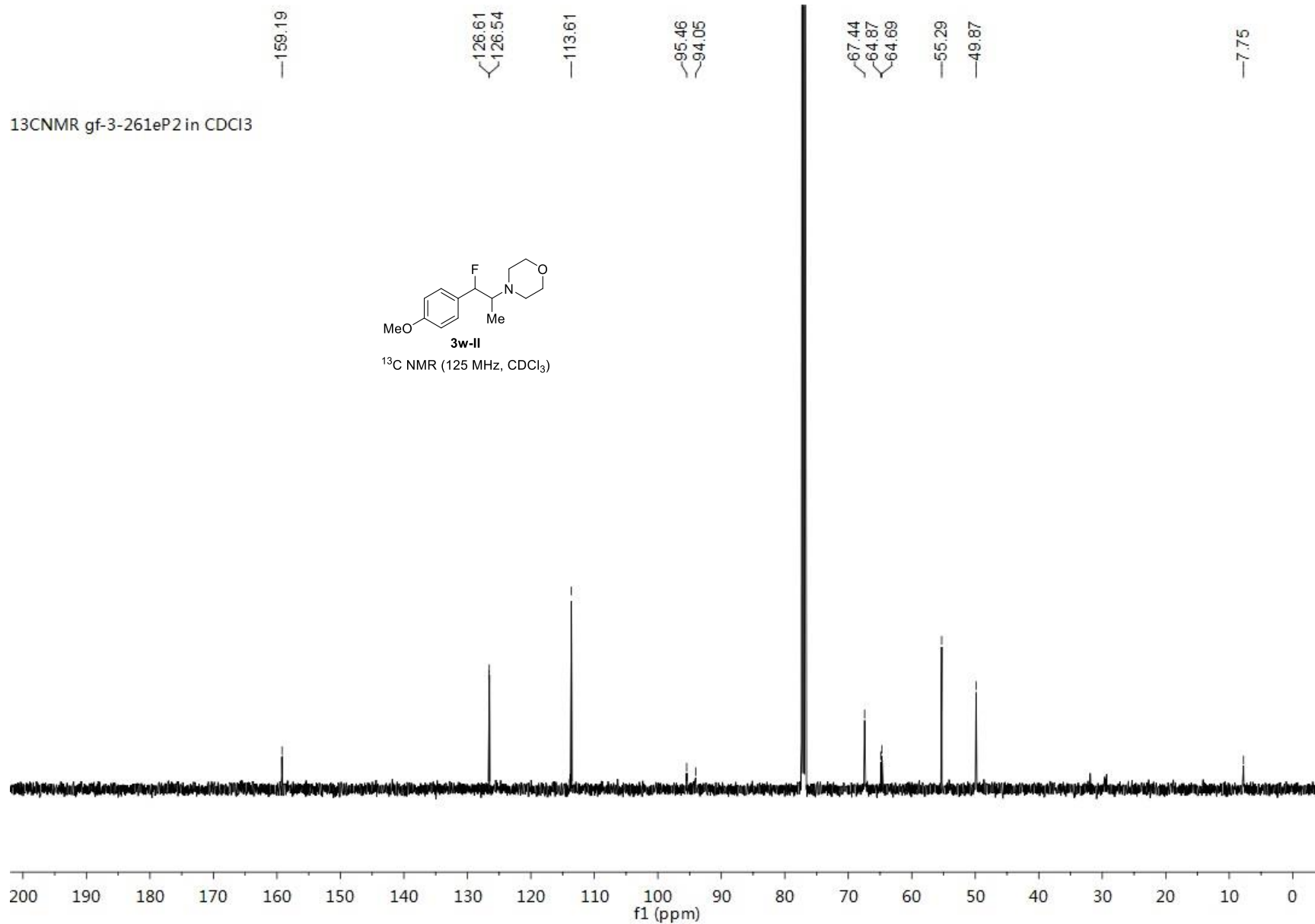
5.6887
5.5942

3.8136
3.7532
3.6919
3.6662
2.8737
2.8432
2.8343
2.7910
2.7821
2.7496
2.7161
2.7050
2.6632
2.6359
2.6142
2.5729

1.0643
1.0511

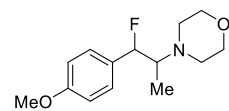
¹H NMR of **3w-II** in CDCl₃



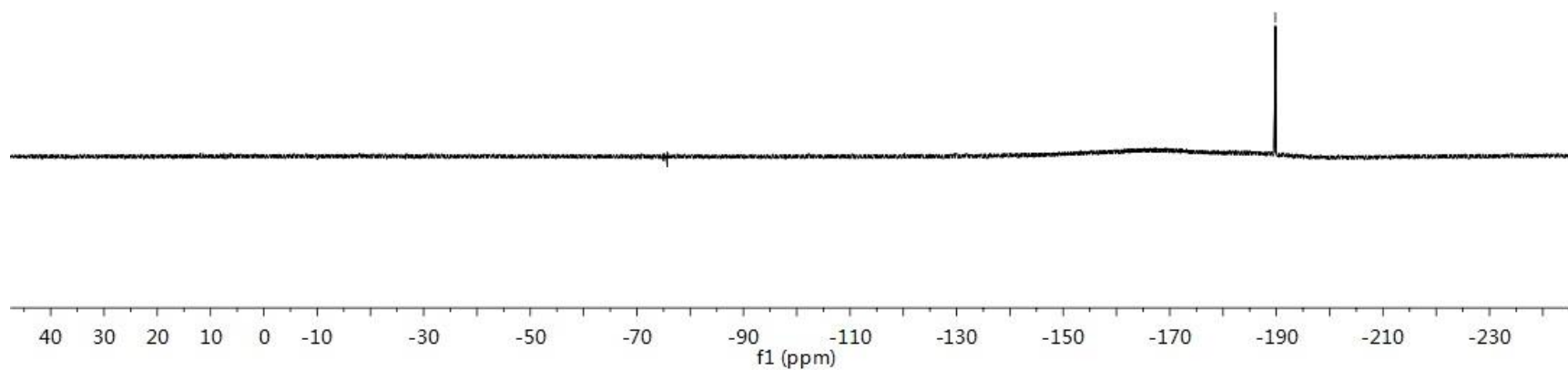


¹⁹F NMR of compound 3w-II in CDCl₃

— 189.8105

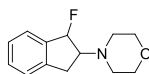


3w-II
¹⁹F NMR (471 MHz, CDCl₃)

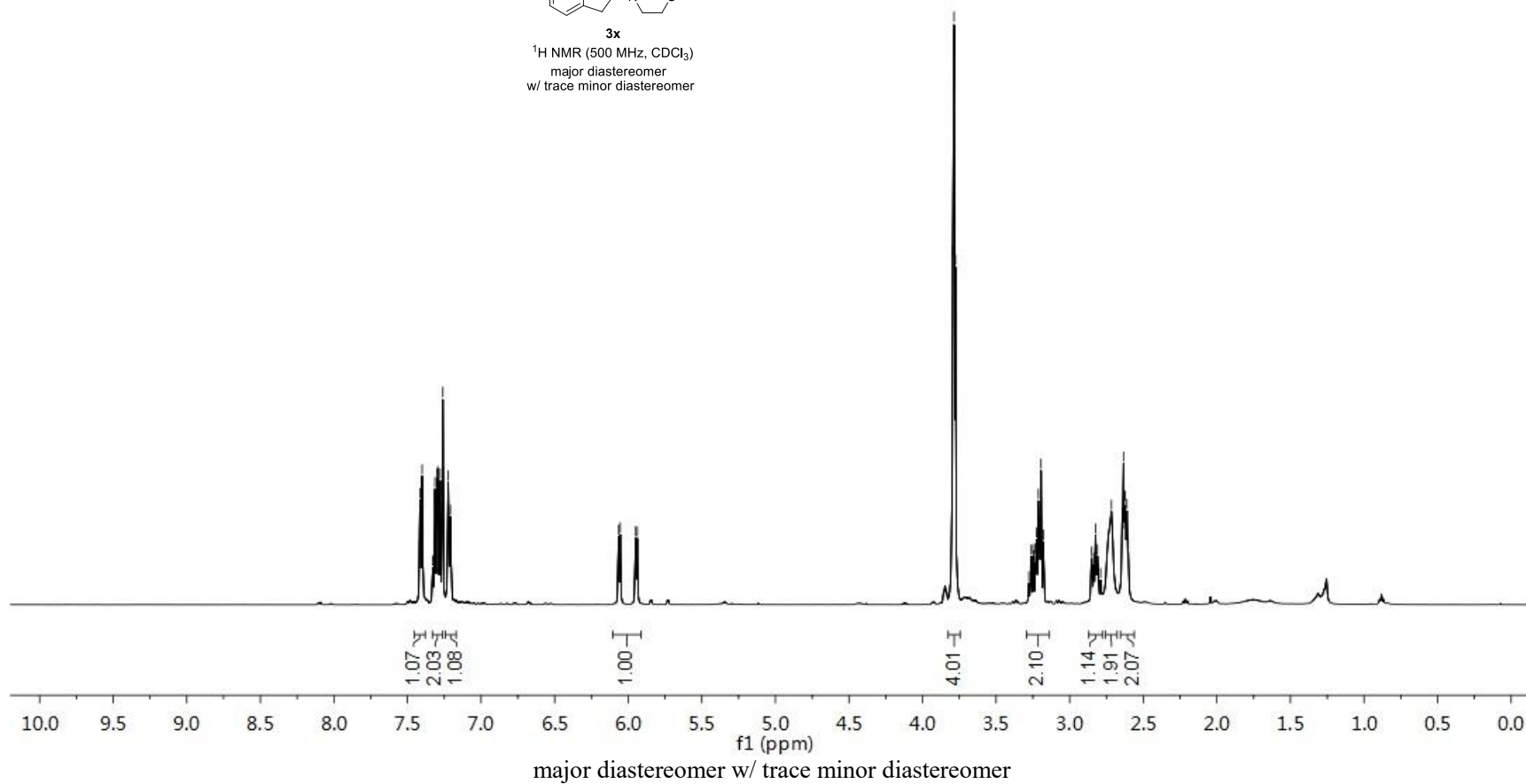


7.4147
7.4006
7.3279
7.3134
7.2989
7.2949
7.2920
7.2777
7.2598
7.2231
7.2087
6.0649
6.0536
5.9494
5.9386
3.7951
3.7858
3.7763
3.2133
3.2072
3.1959
2.8363
2.8243
2.8116
2.7896
2.7162
2.6431
2.6341
2.6240
2.6113
2.6022

¹H NMR gf-3-68cP col in CDCl₃



3x
¹H NMR (500 MHz, CDCl₃)
major diastereomer
w/ trace minor diastereomer



¹³CNMR gf-3-68cP col in CDCL₃

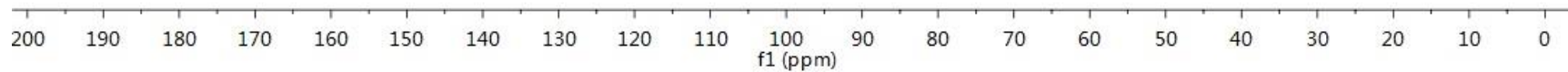
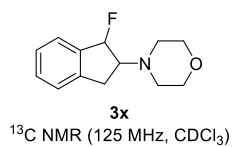
139.53
139.49
139.11
138.94
129.43
129.41
127.28
127.27
124.82
124.64

99.71
98.28

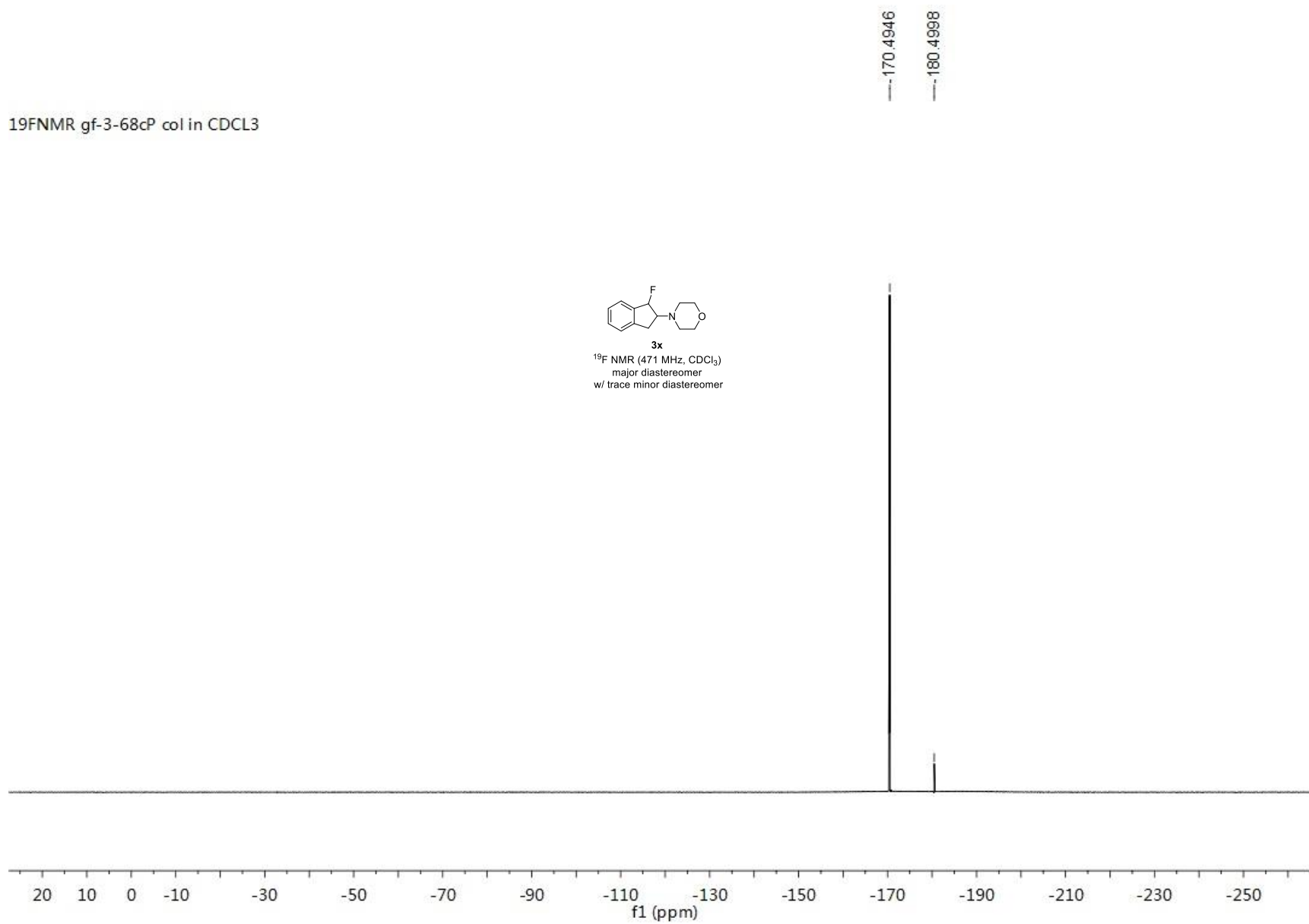
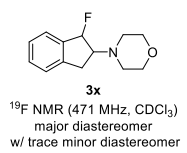
73.36
73.22
66.89

52.42

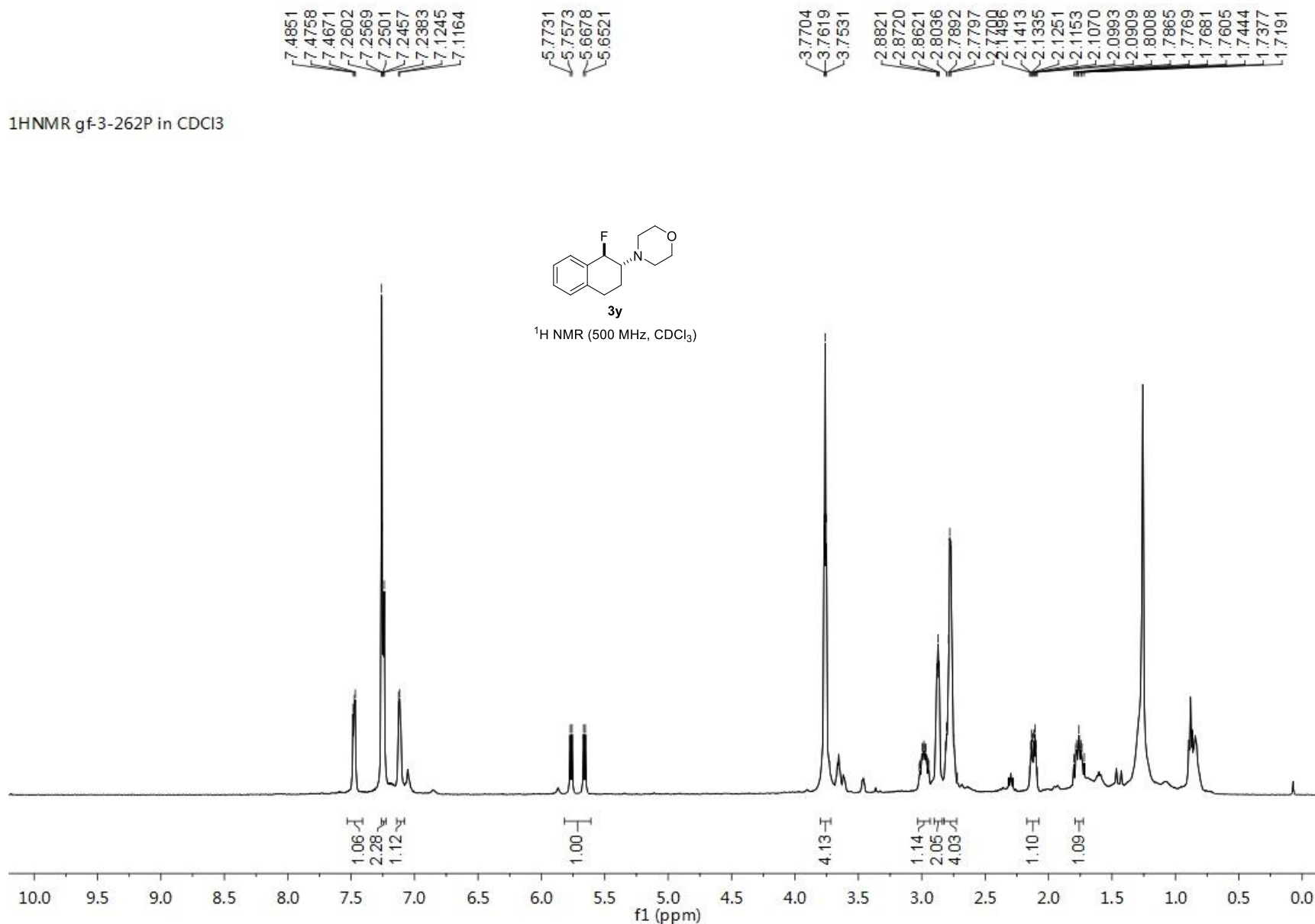
34.21
34.17



¹⁹F NMR gf-3-68cP col in CDCl₃



¹H NMR of compound 3y in CDCl₃



¹³C NMR gf-3-262PP in CDCl₃

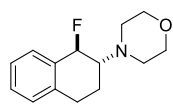
137.02
136.99
134.50
134.36
128.44
128.43
128.40
128.19
126.45

90.46
89.08

67.31
65.15
65.01

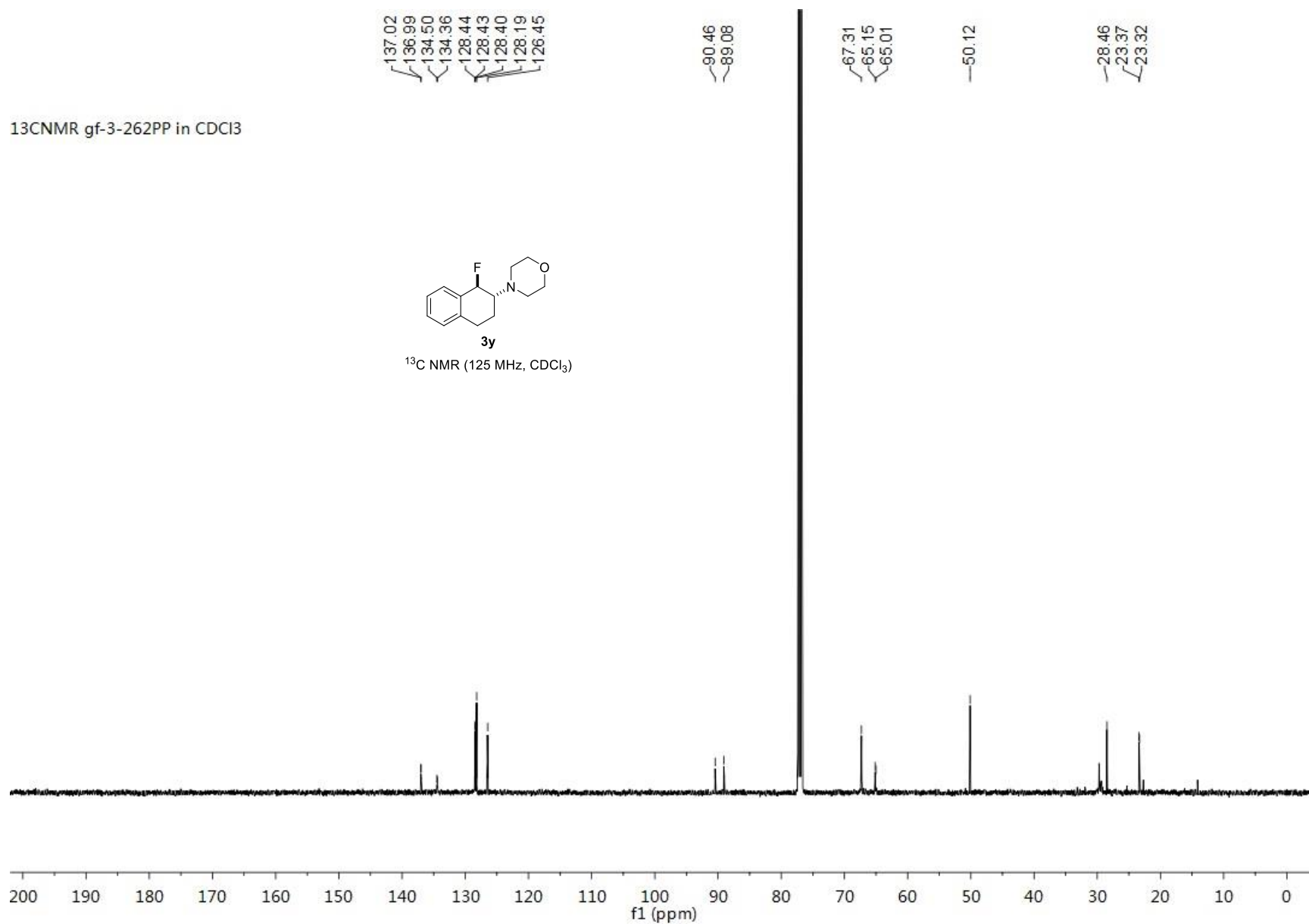
50.12

28.46
23.37
23.32

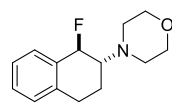


3y

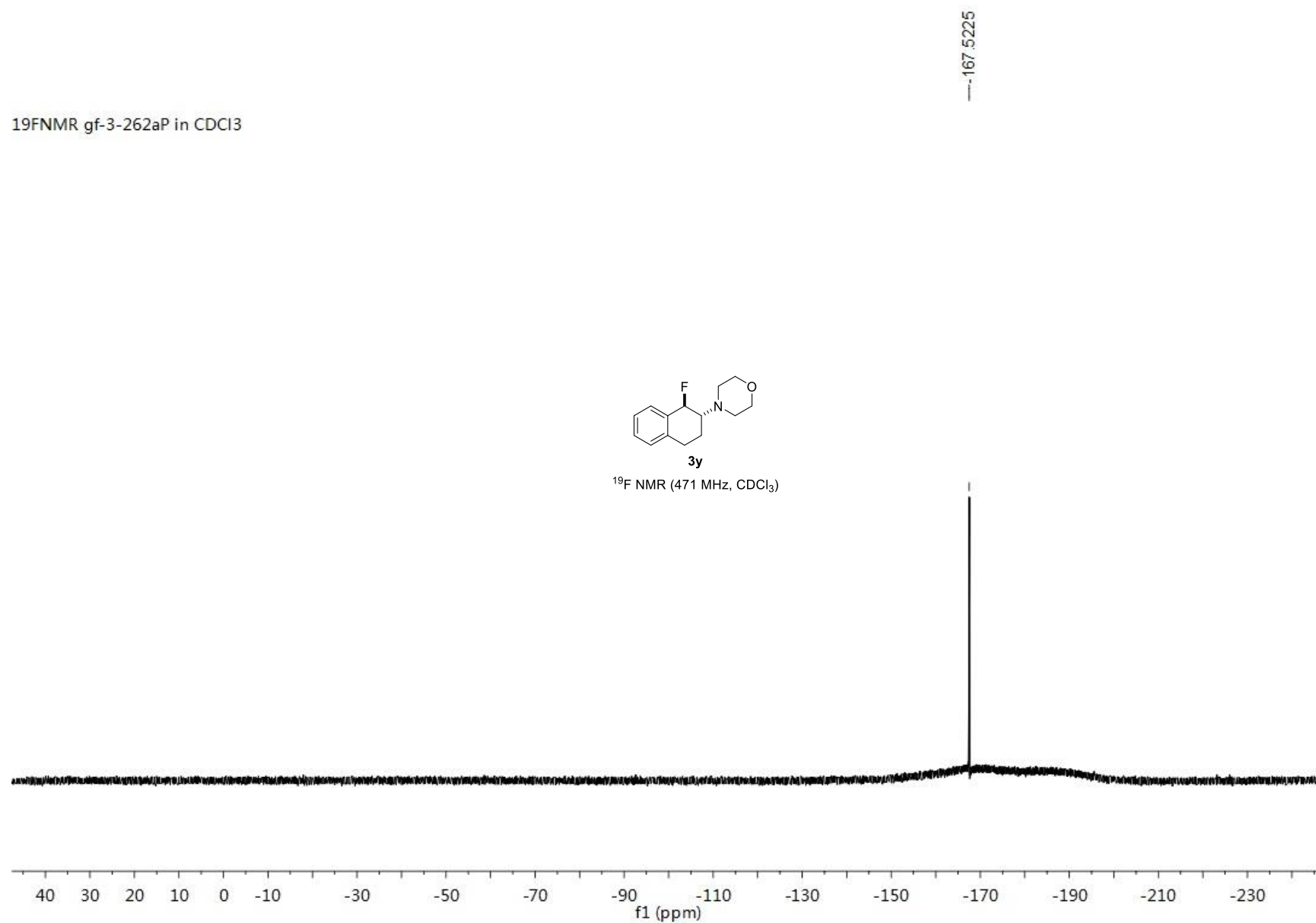
¹³C NMR (125 MHz, CDCl₃)

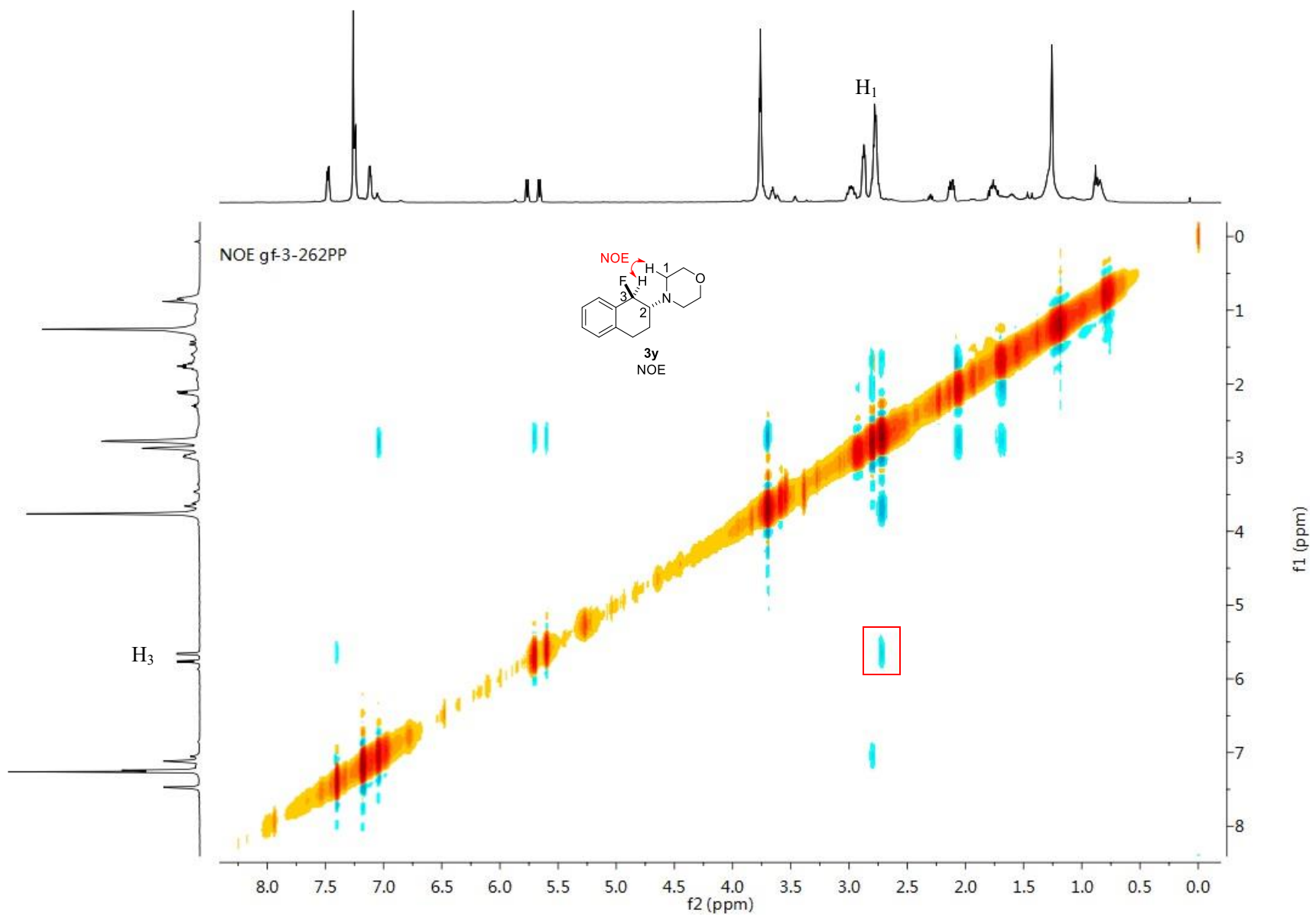


¹⁹F NMR gf-3-262aP in CDCl₃

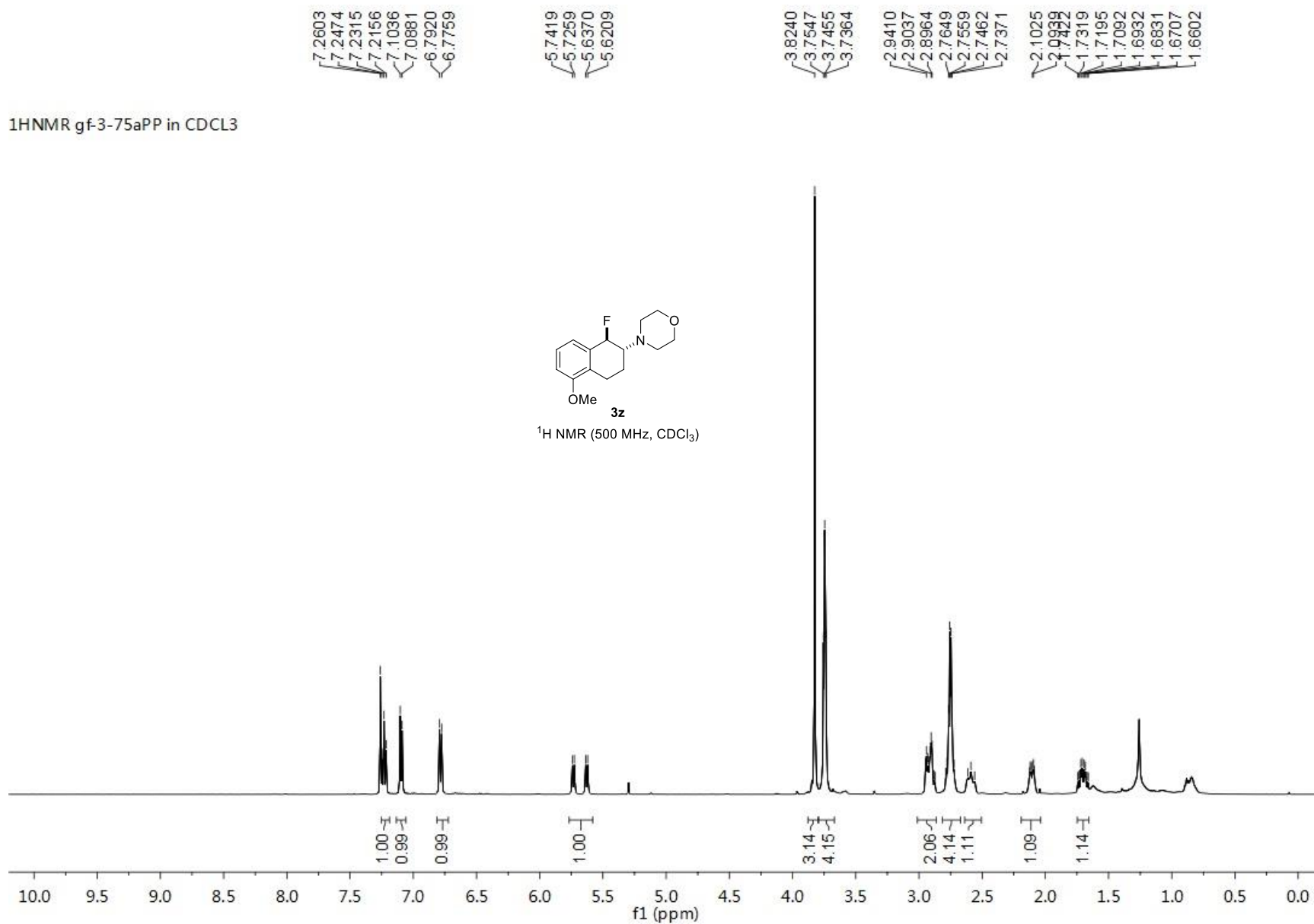


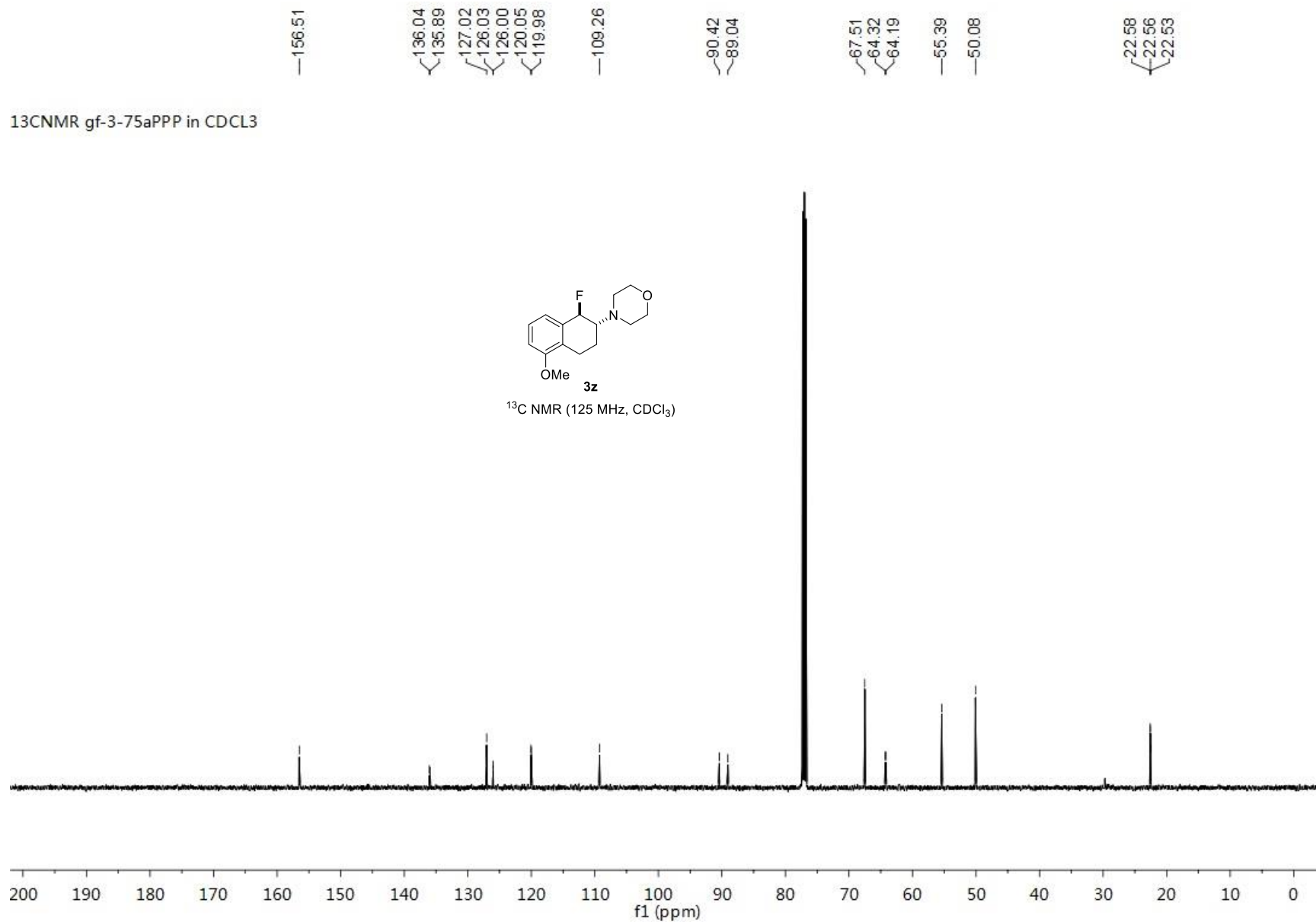
¹⁹F NMR (471 MHz, CDCl₃)





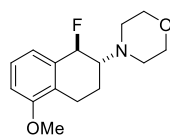
¹H NMR of compound 3z in CDCl₃



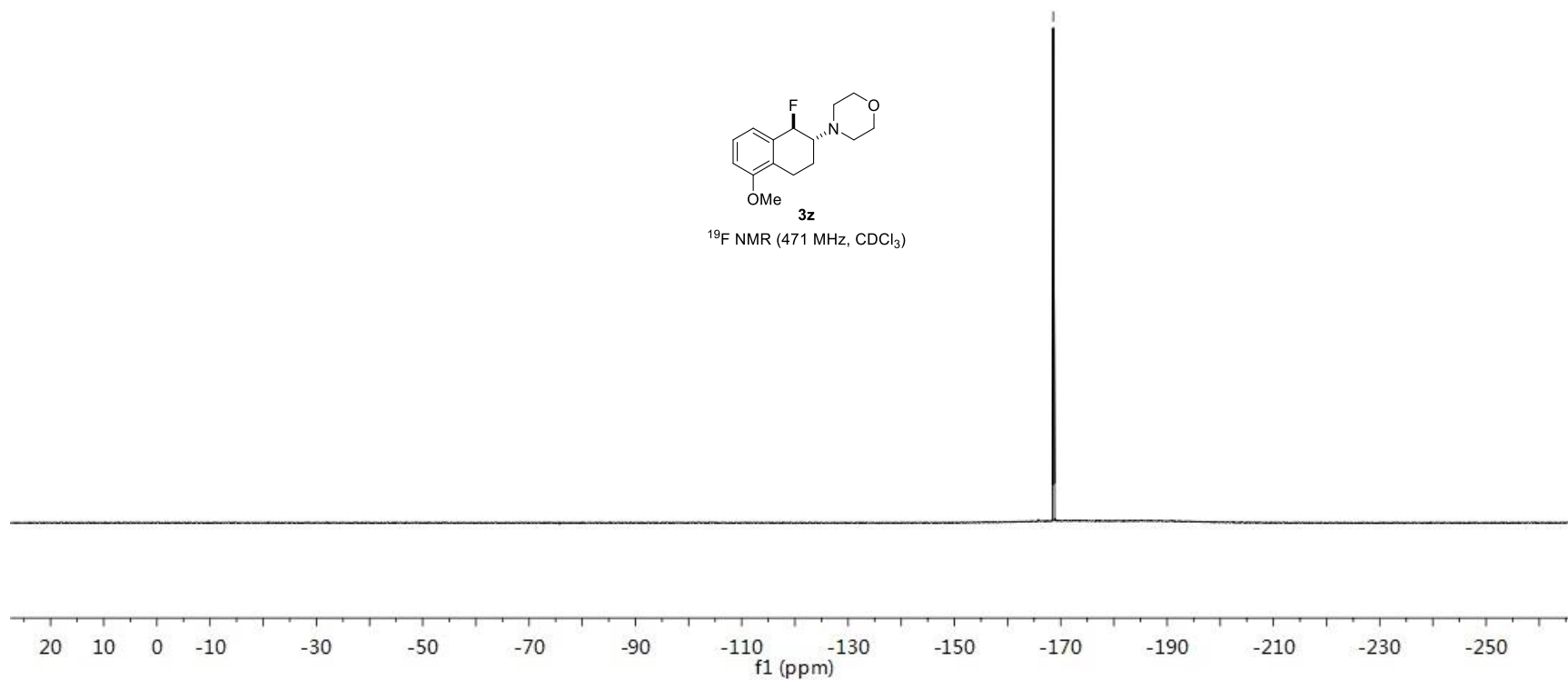


¹⁹F NMR gf-3-75aPPP in CDCl₃

168.6082



¹⁹F NMR (471 MHz, CDCl₃)

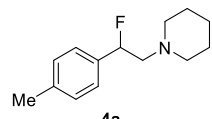


7.2384
7.2226
7.1819
7.1660

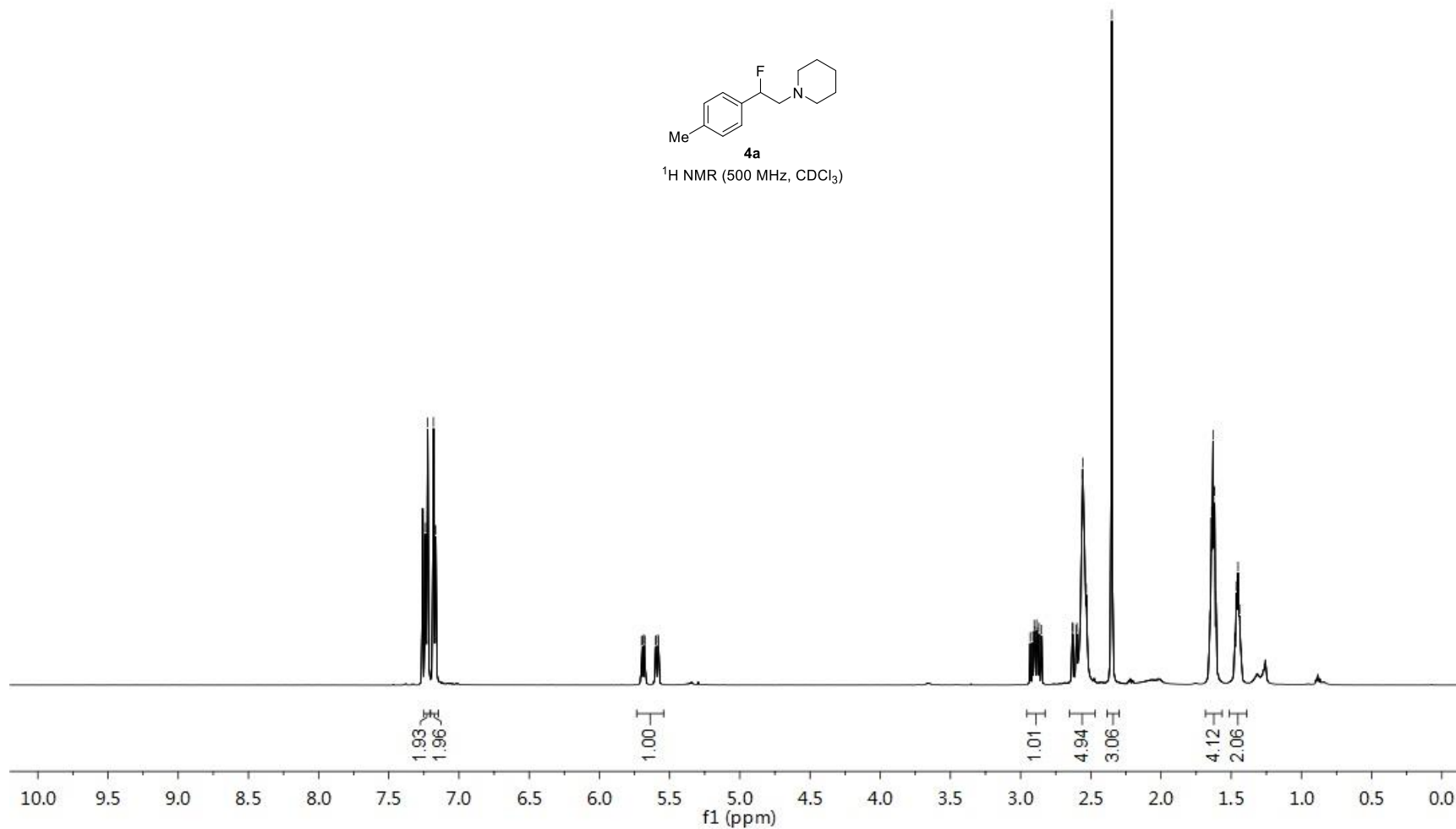
5.6985
5.6944
5.6807
5.6766
5.6005
5.5964
5.5826
5.5786

2.9319
2.9140
2.9033
2.8972
2.8854
2.8793
2.8686
2.8507
2.8320
2.8273
2.6033
2.5987
2.5612
2.5566
2.5330
2.5283
2.3507
1.6520
1.6401
1.6296
1.6192
1.6079
1.4745
1.4627
1.4514
1.4401
1.4288

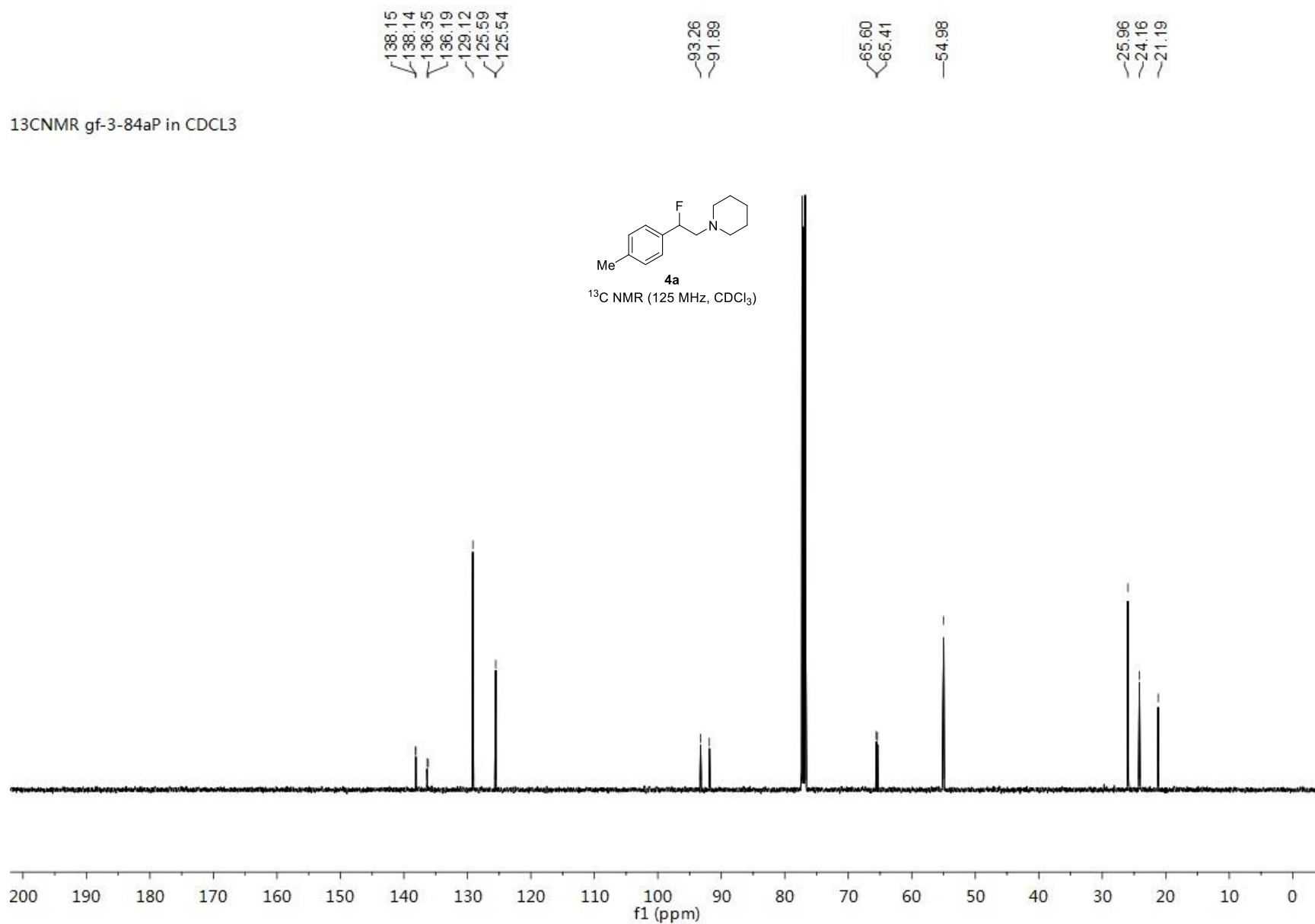
¹H NMR of gf-3-84aP in CDCl₃



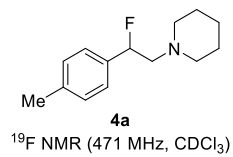
¹H NMR (500 MHz, CDCl₃)



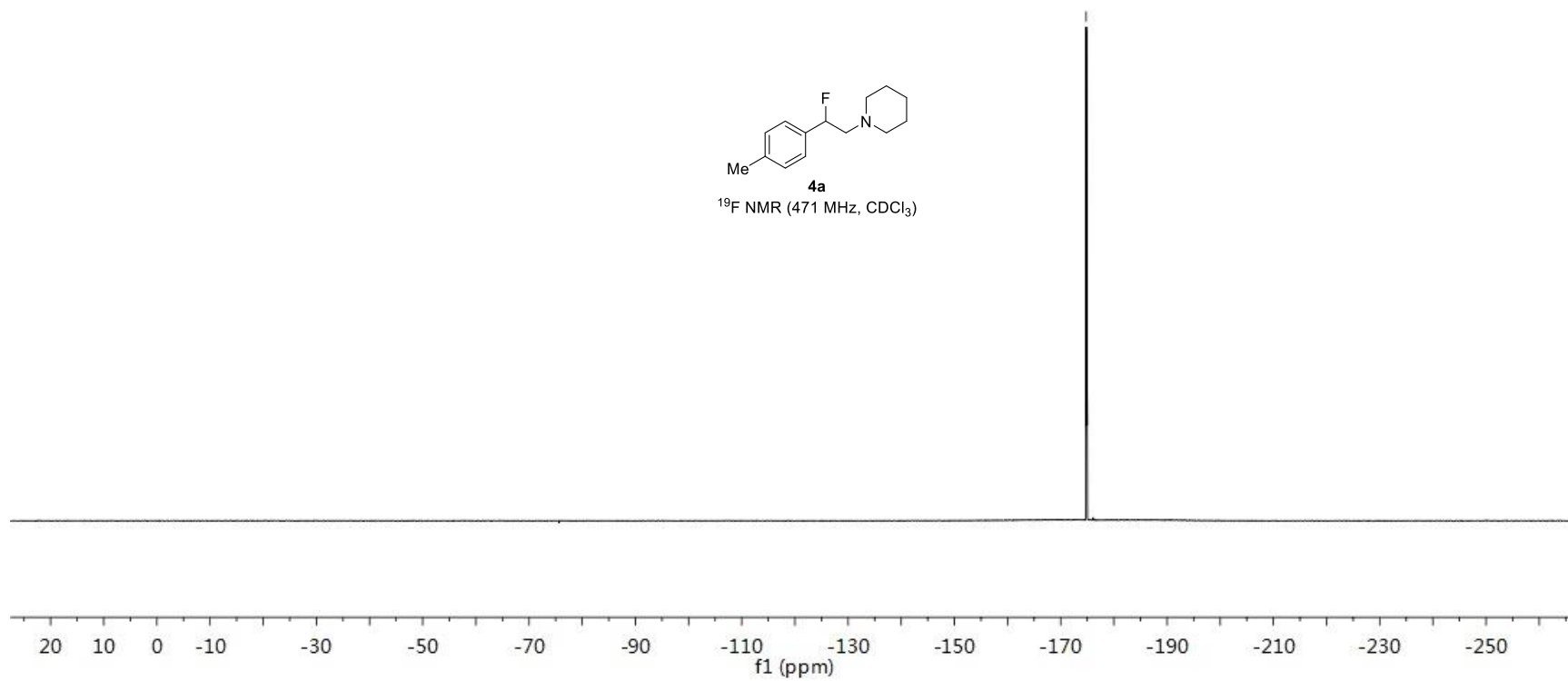
¹³CNMR gf-3-84aP in CDCl₃

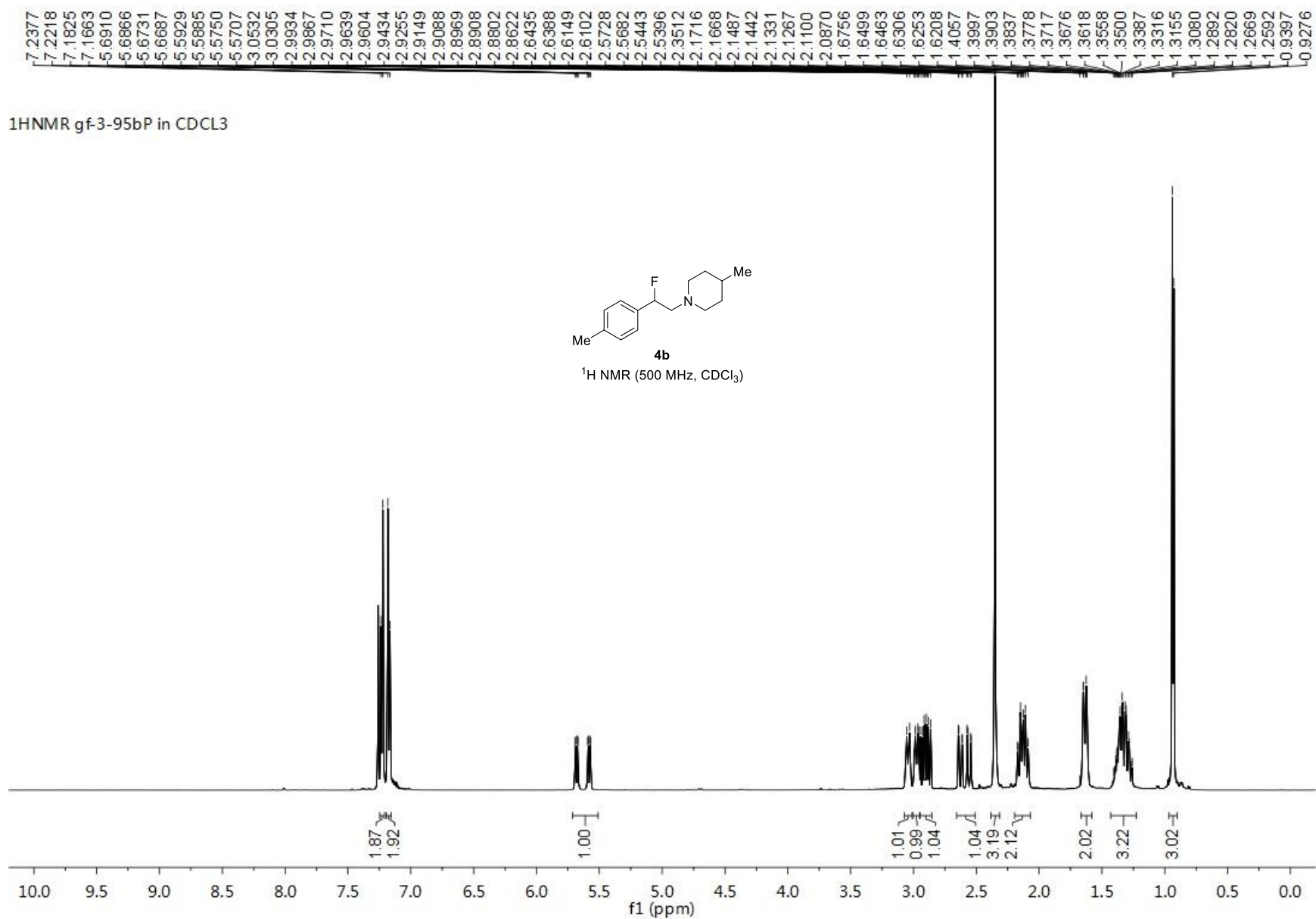


¹⁹F NMR gf-3-84aP in CDCl₃

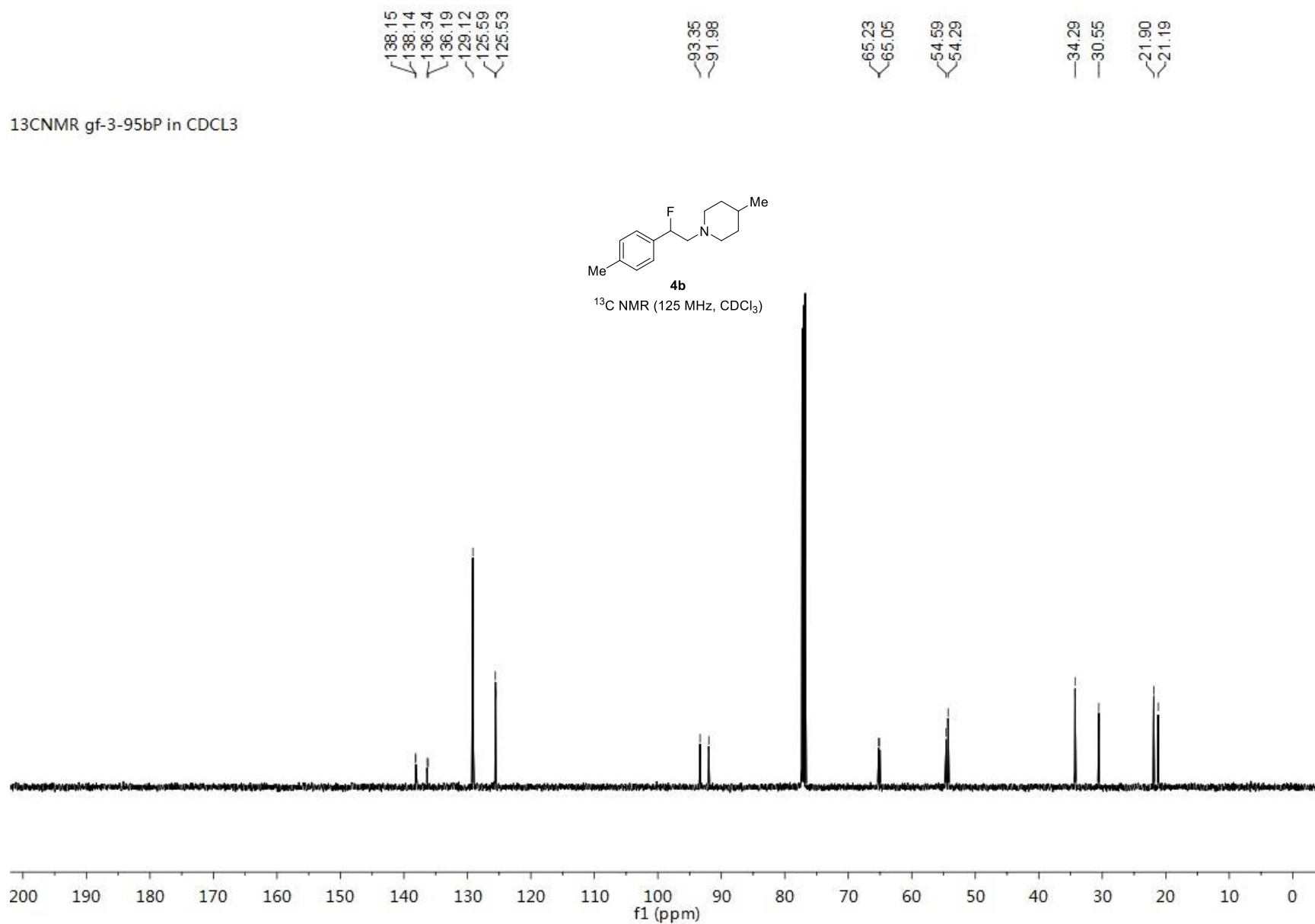
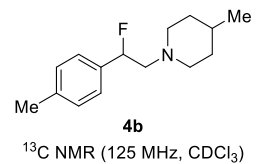


-174.7714



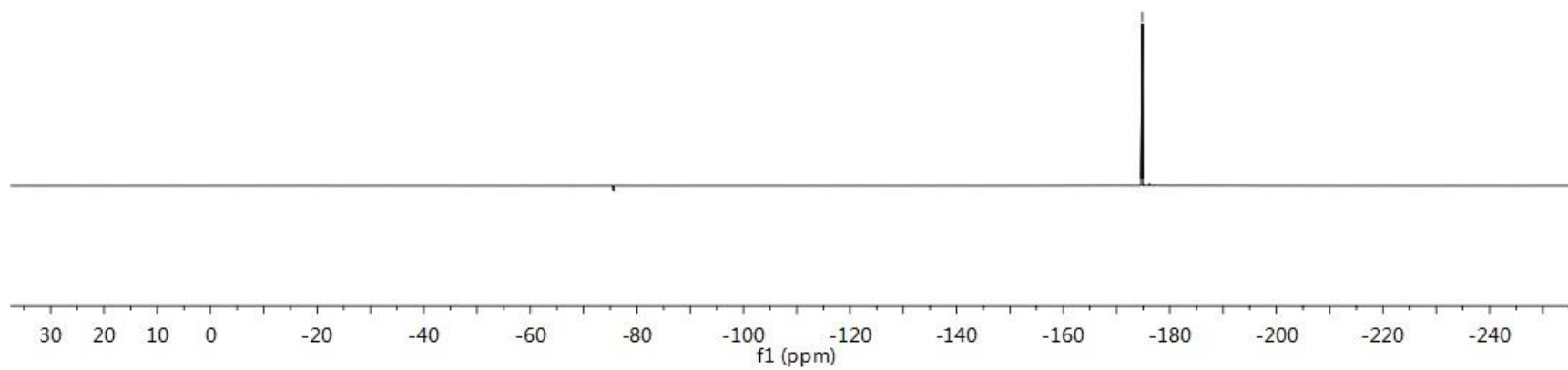
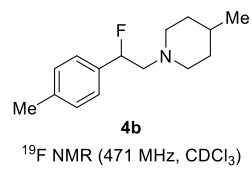


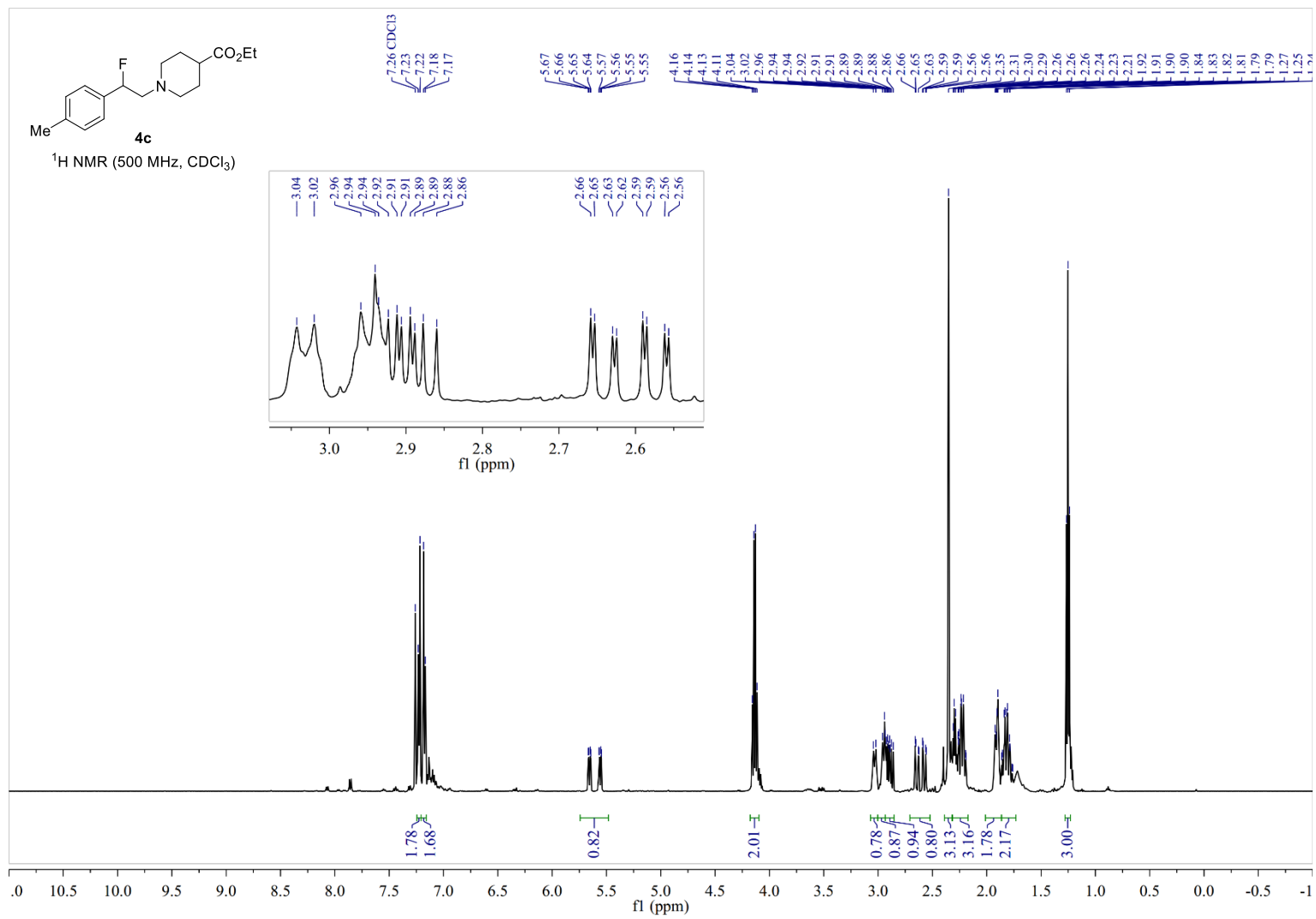
¹³C NMR gf-3-95bP in CDCl₃

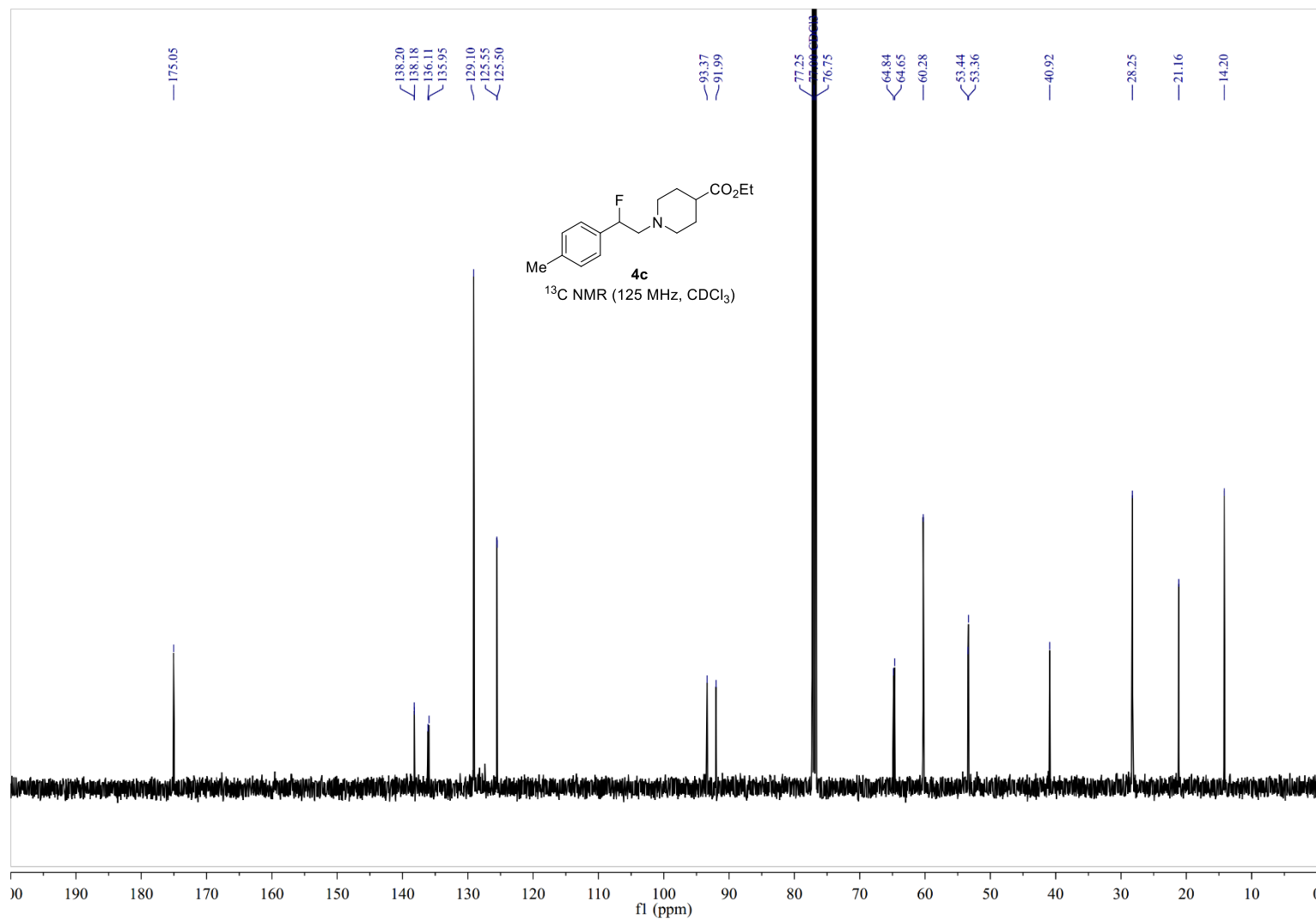


^{19}F NMR gf-3-95bP in CDCl_3

---174.8311

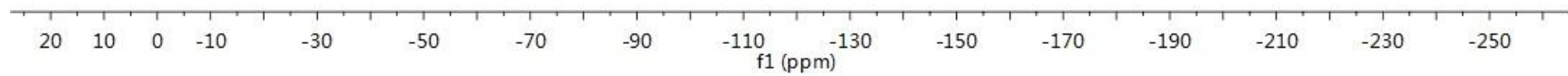
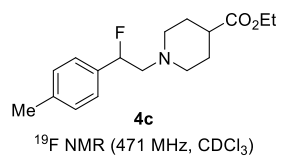


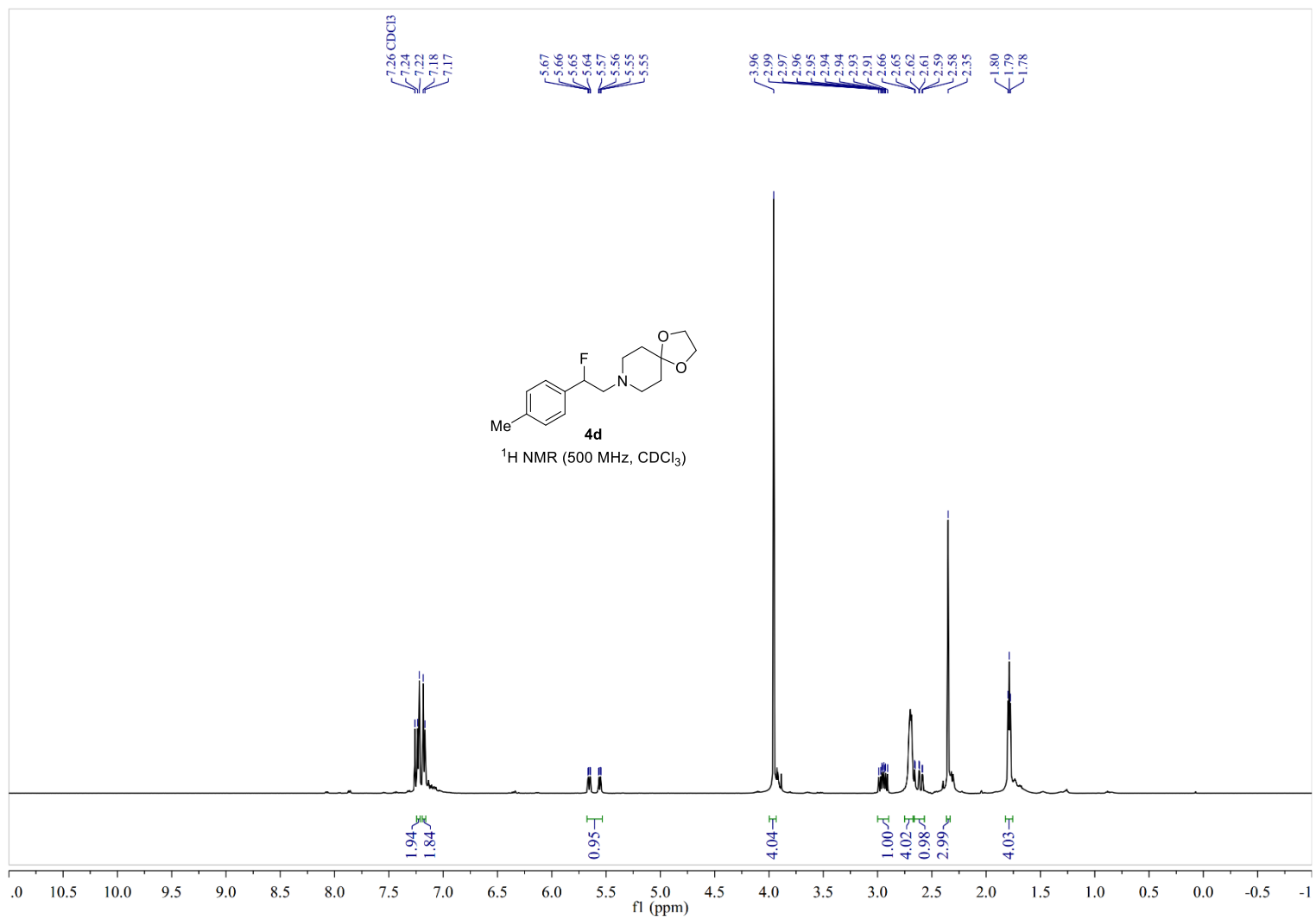


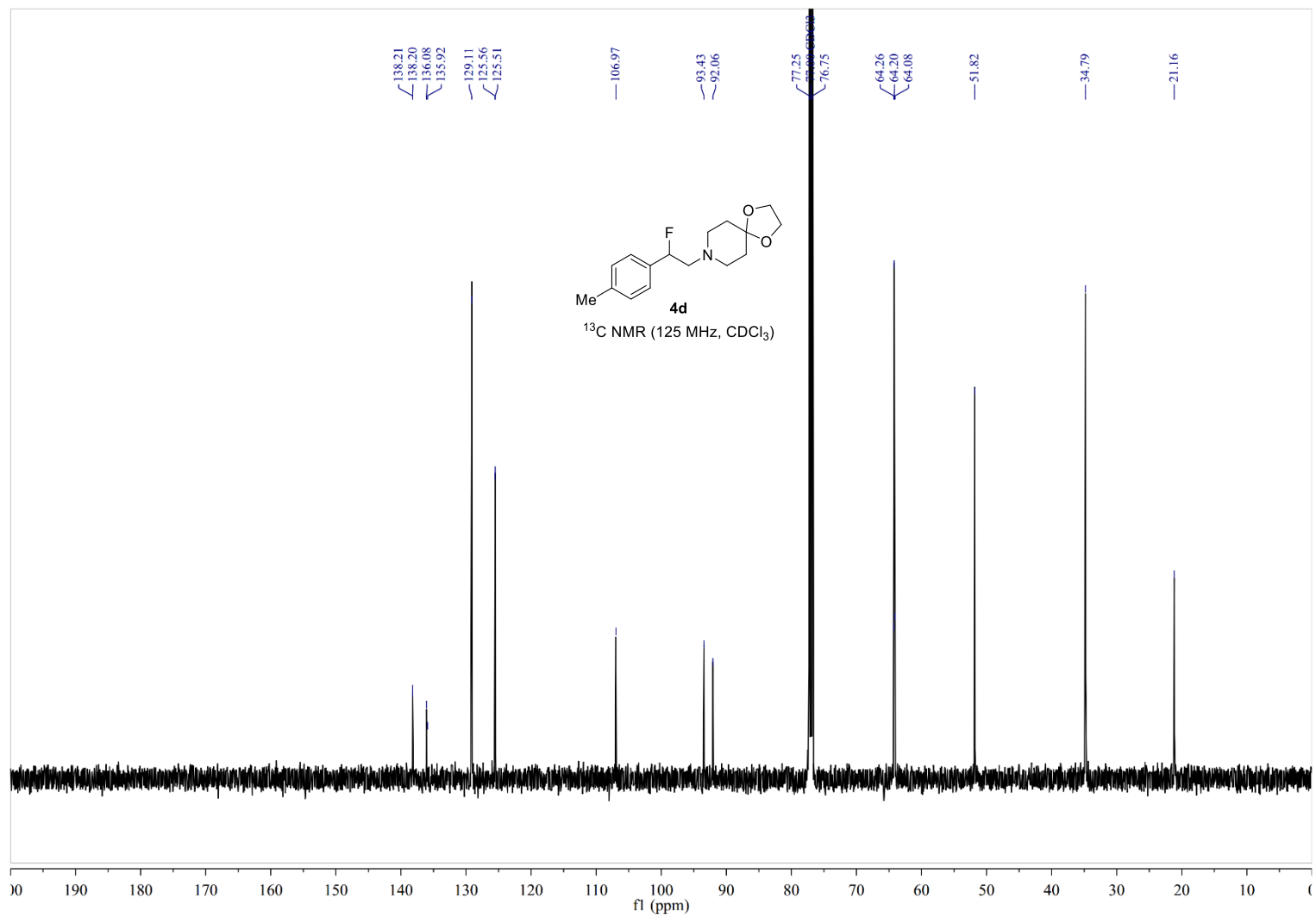


^{19}F NMR gf-3-84cP in CDCl_3

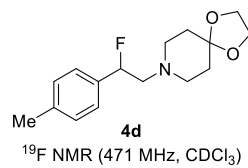
—174.9926



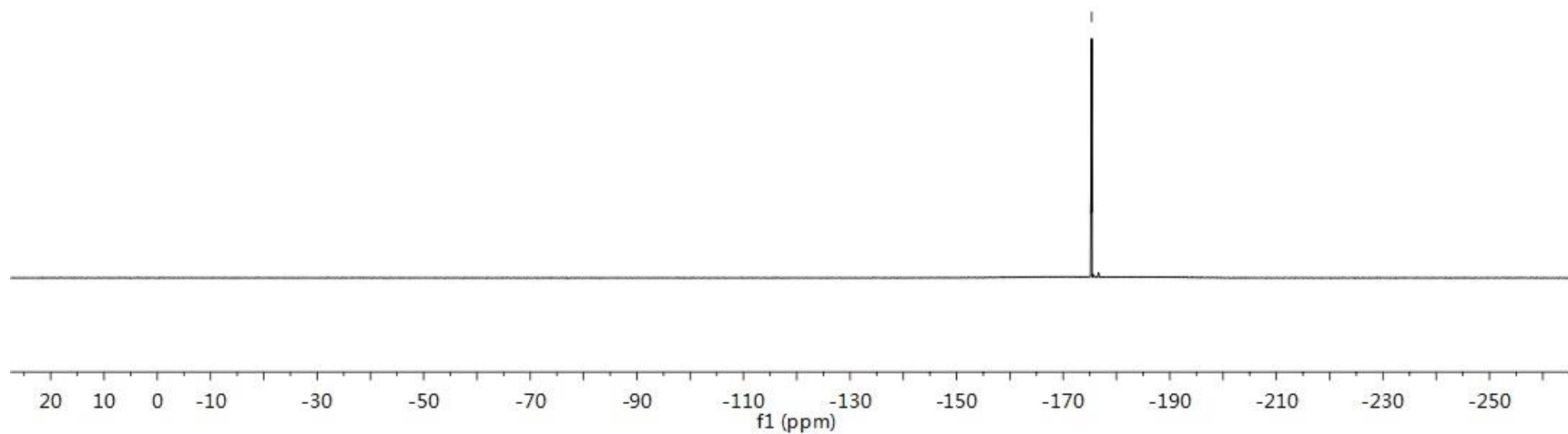




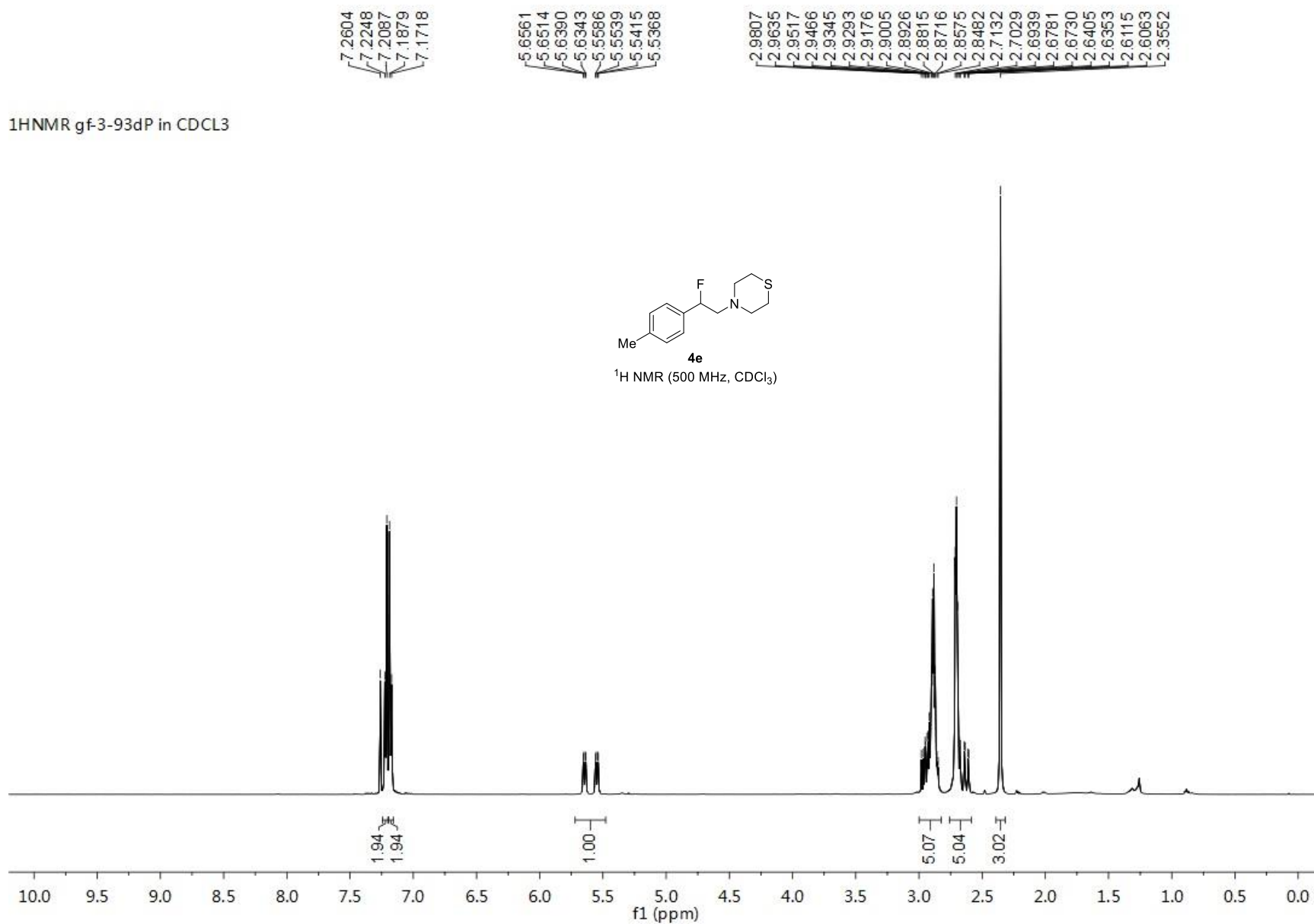
¹⁹F NMR of compound 4d in CDCl₃



-175.3455



¹H NMR of gf-3-93dP in CDCl₃



138.33
138.32
135.91
135.75
129.17
125.59
125.54

93.24
91.86

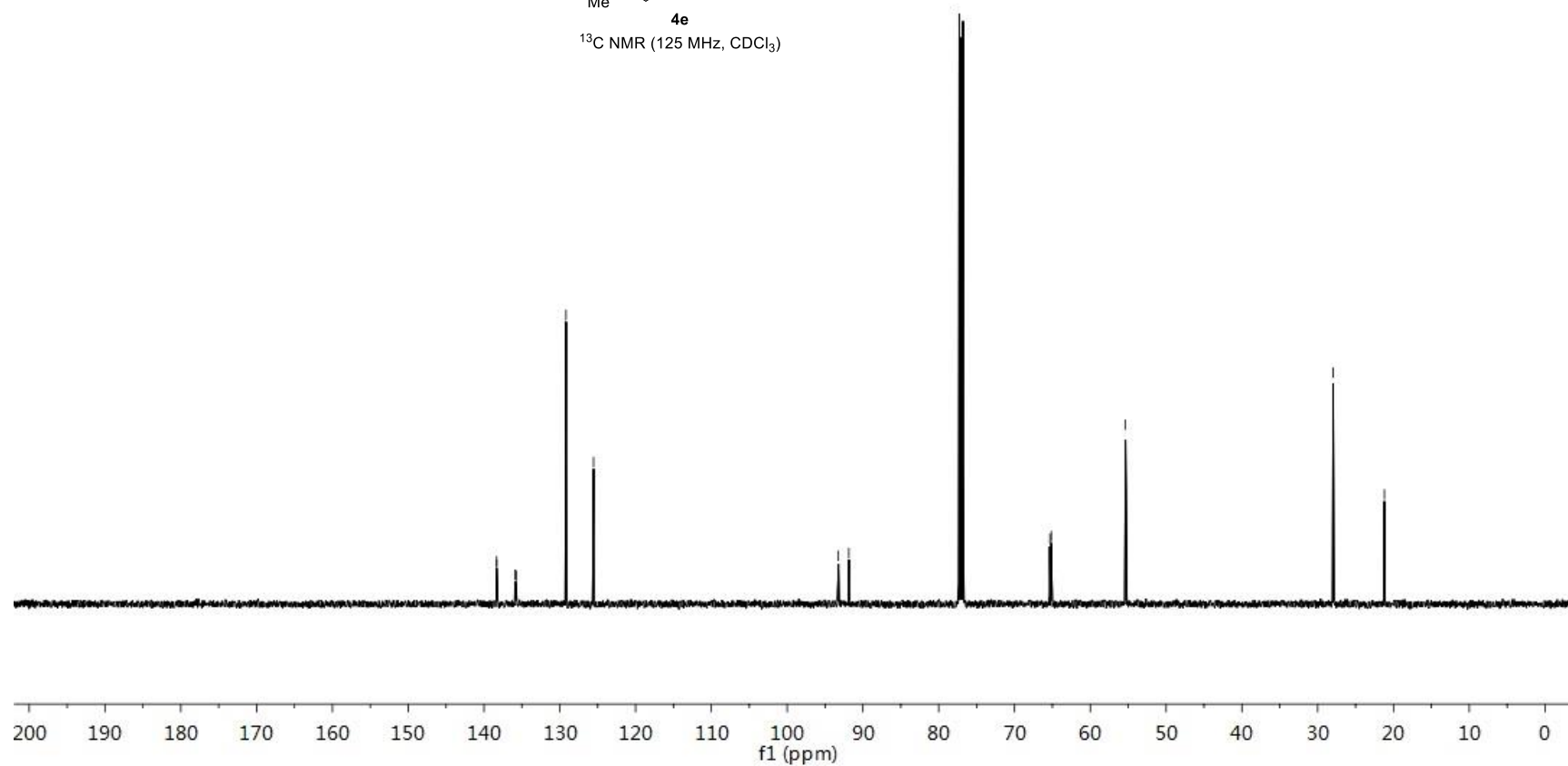
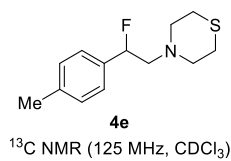
65.34
65.15

55.36

27.96

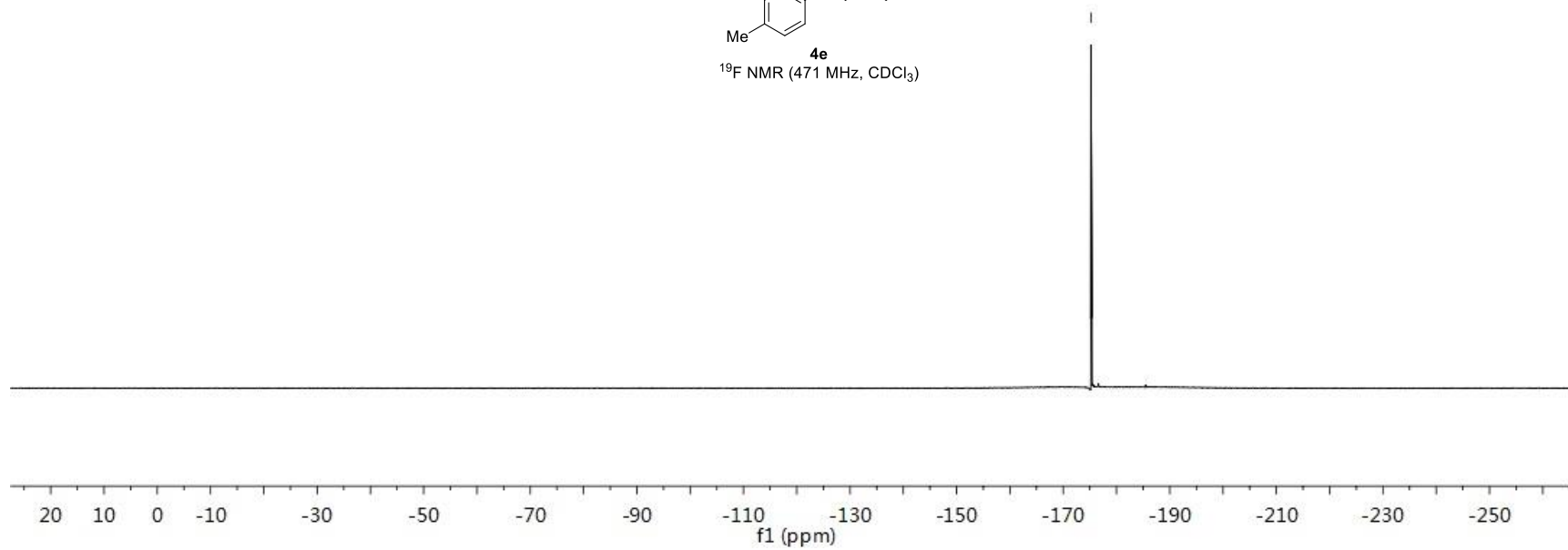
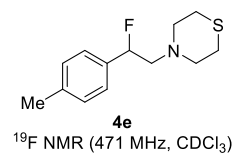
21.20

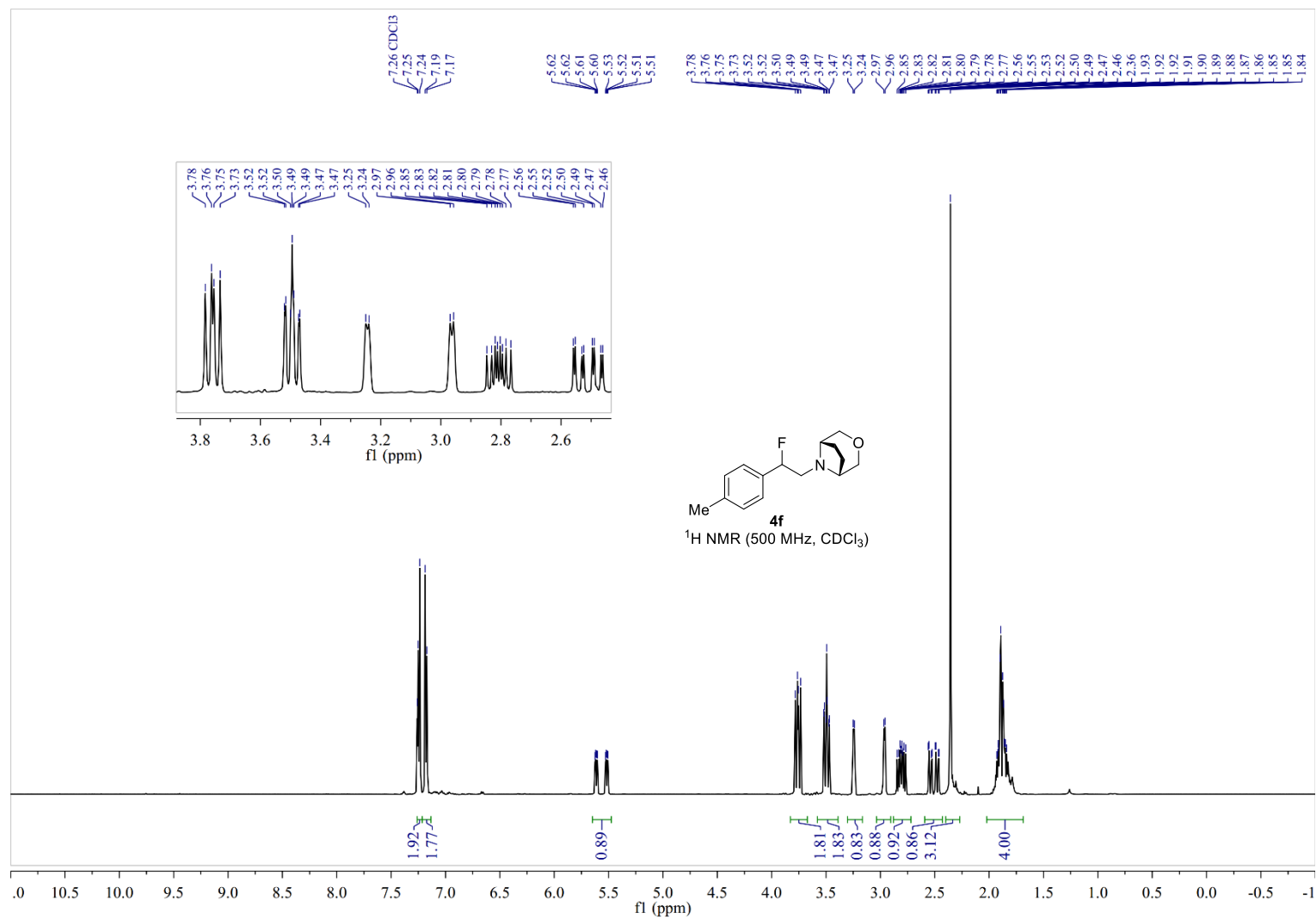
¹³C NMR gf-3-93dP in CDCl₃

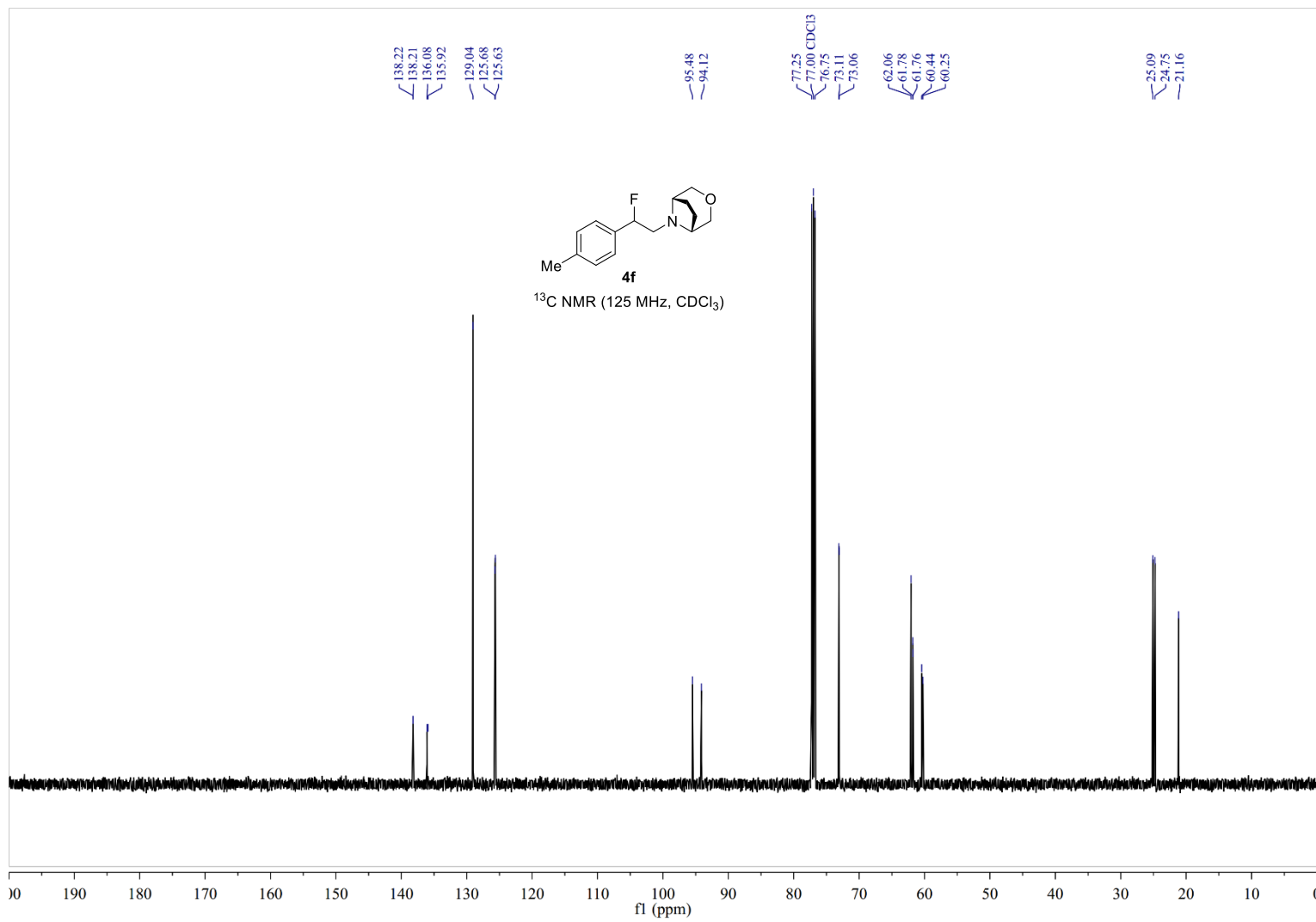


¹⁹F NMR gf-3-93dP in CDCl₃

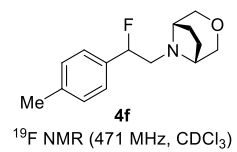
—175.1931



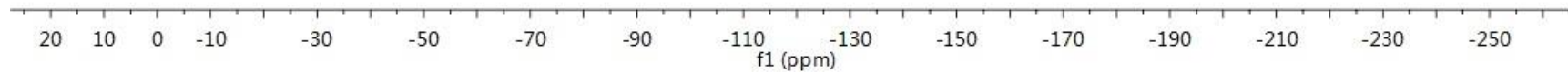




¹⁹F NMR gf-3-86bP in CDCl₃



→ -176.7306



7.2604
7.2356
7.2197
7.1912
7.1752

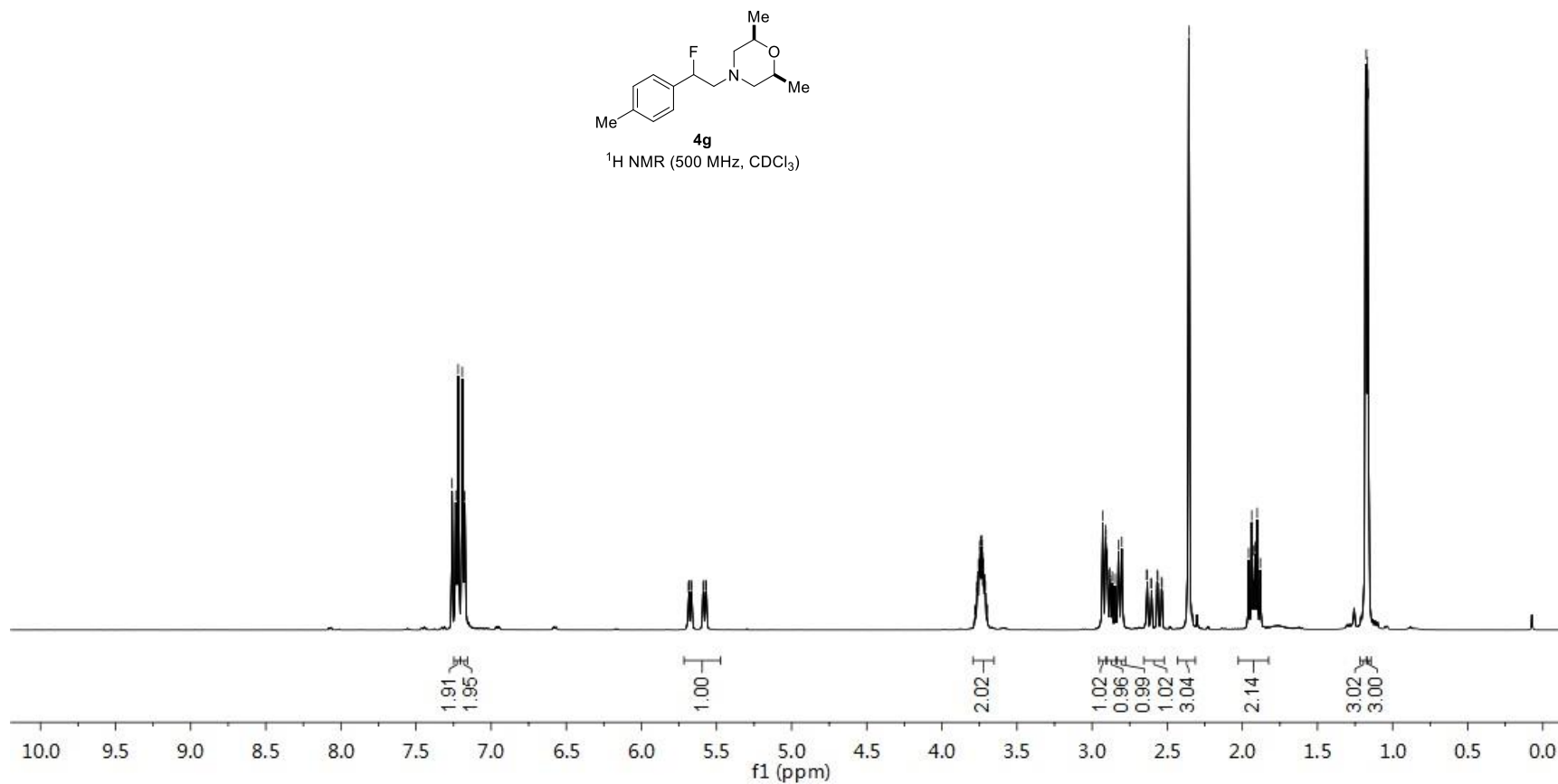
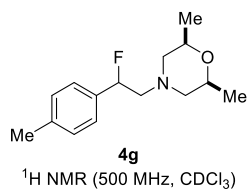
5.6879
5.6837
5.6700
5.6659
5.5900
5.5858
5.5721
5.5680

3.7636
3.7555
3.7507
3.7476
3.7431
3.7350
3.7305
3.7274
3.7225
3.7179
3.7145

2.9287
2.9102
2.9075
2.9003
2.8241
2.8021
2.3565

1.9375
1.9227
1.9162
1.9012
1.1749
1.1677
1.1623

¹H NMR of **4g** in CDCl₃



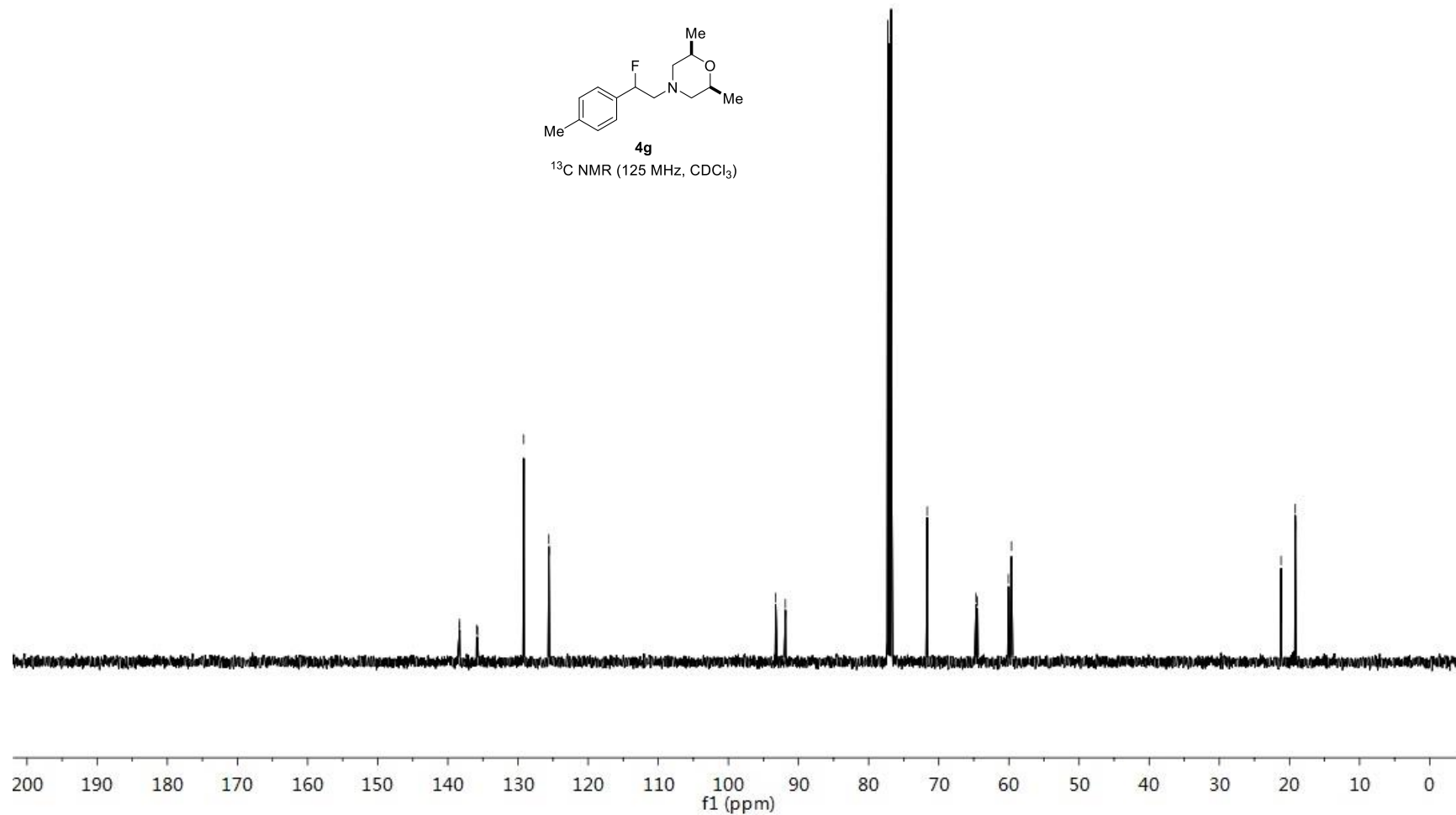
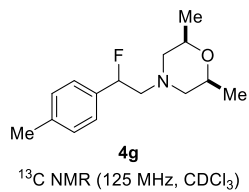
138.35
138.33
135.92
135.76
129.18
125.59
125.54

93.25
91.87

71.67
71.65
64.68
64.50
60.07
59.64

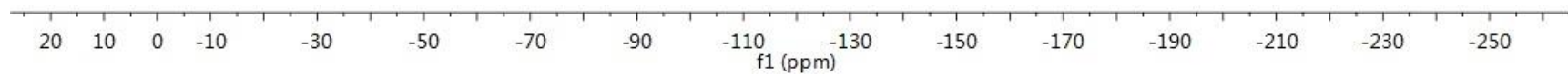
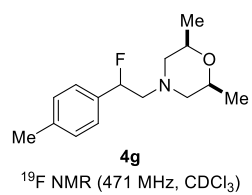
21.20
19.17
19.14

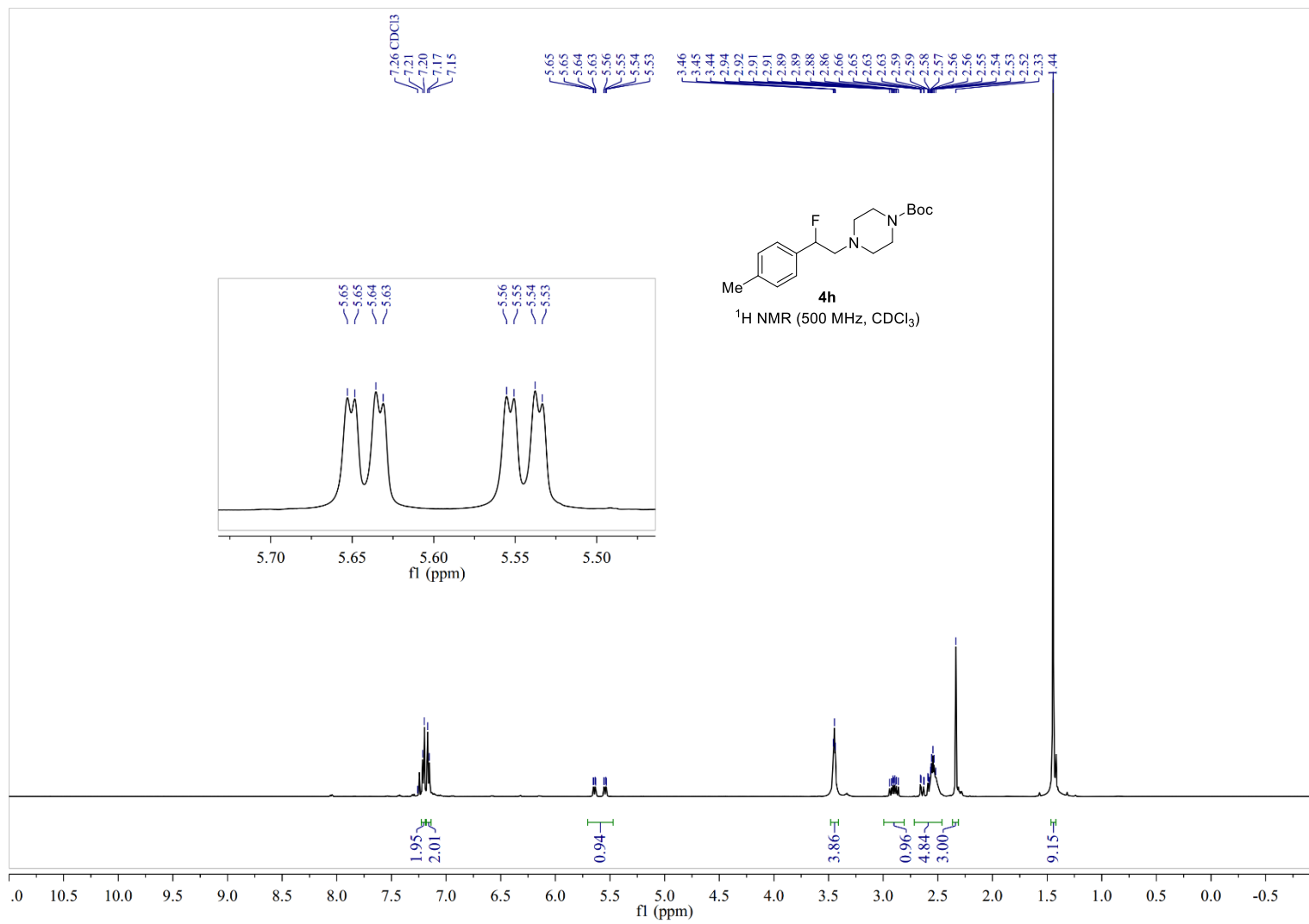
¹³C NMR gf-3-95aP in CDCl₃

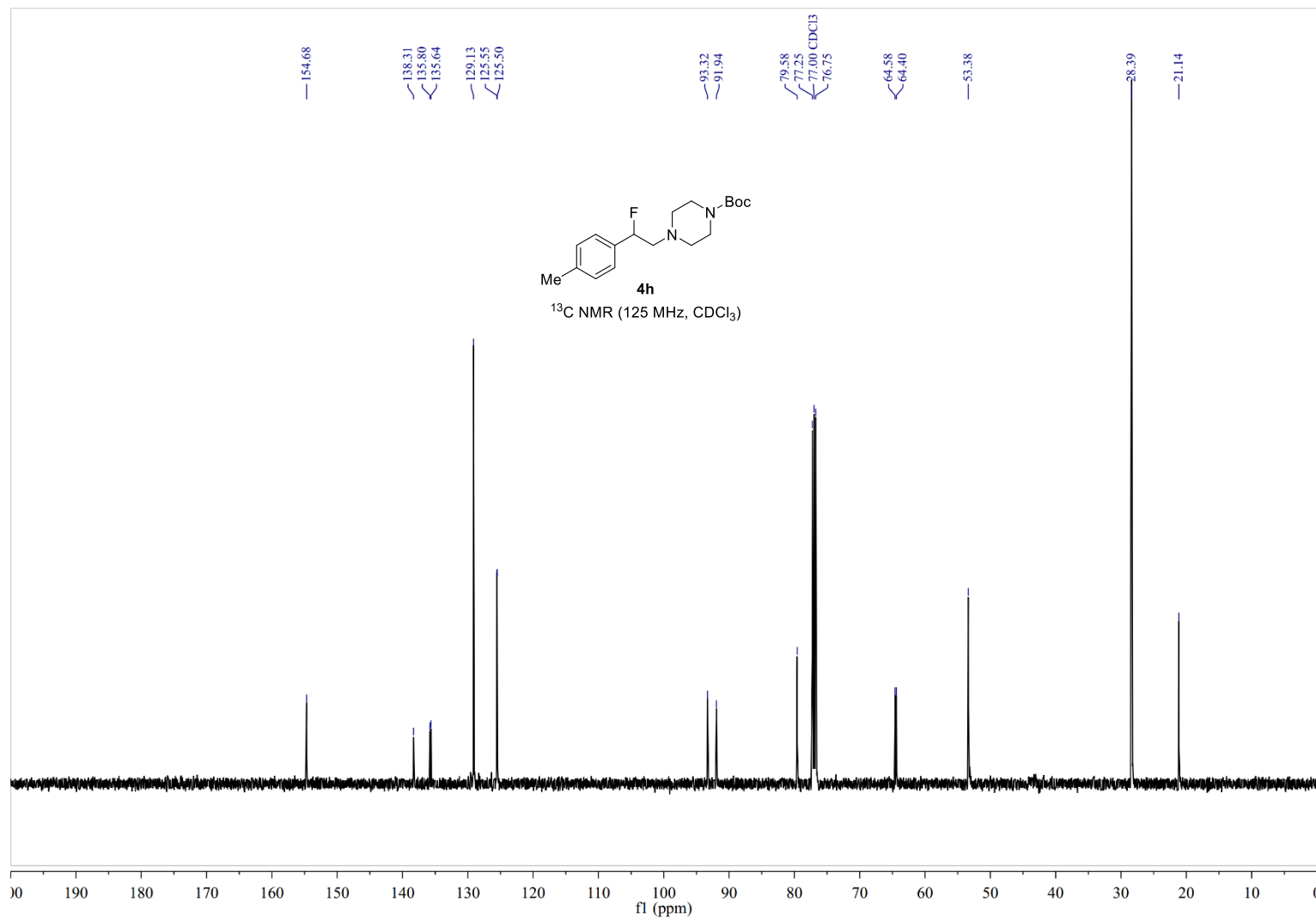


¹⁹F NMR gf-3-95aP in CDCl₃

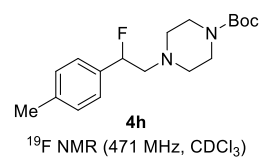
—175.1683



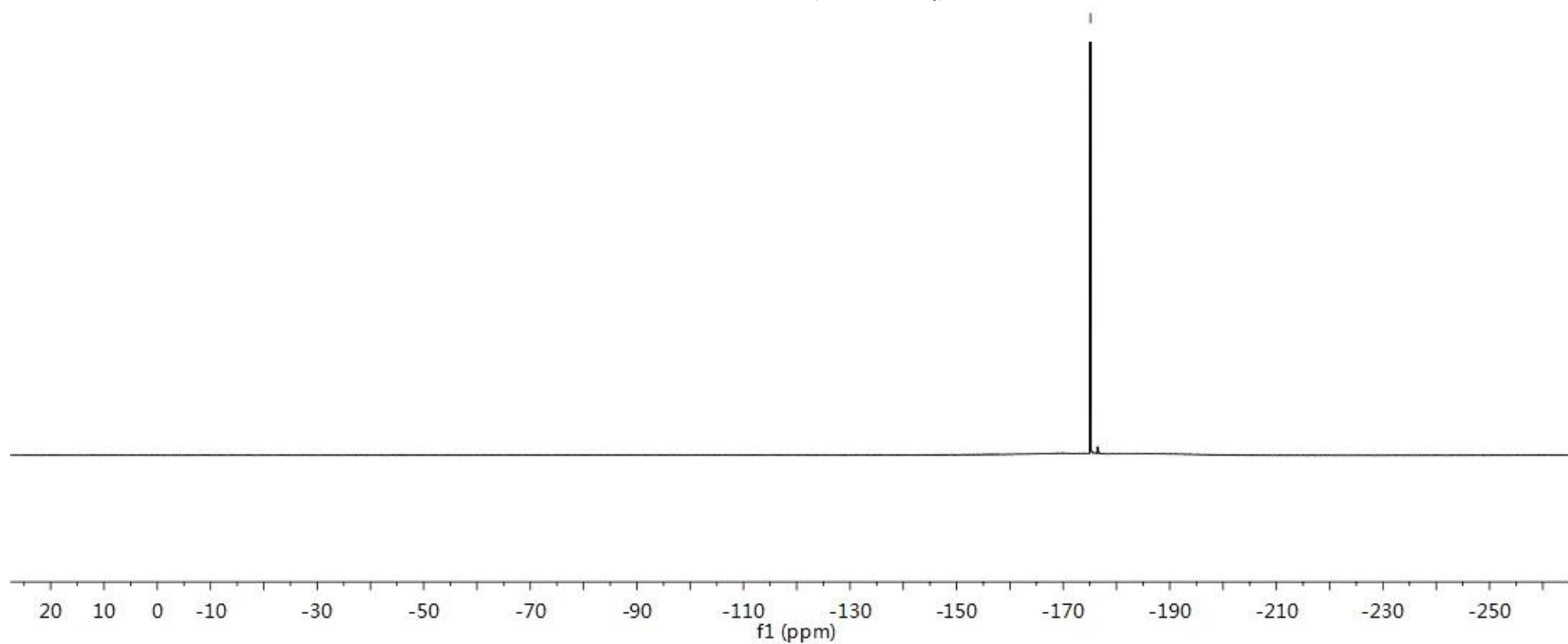


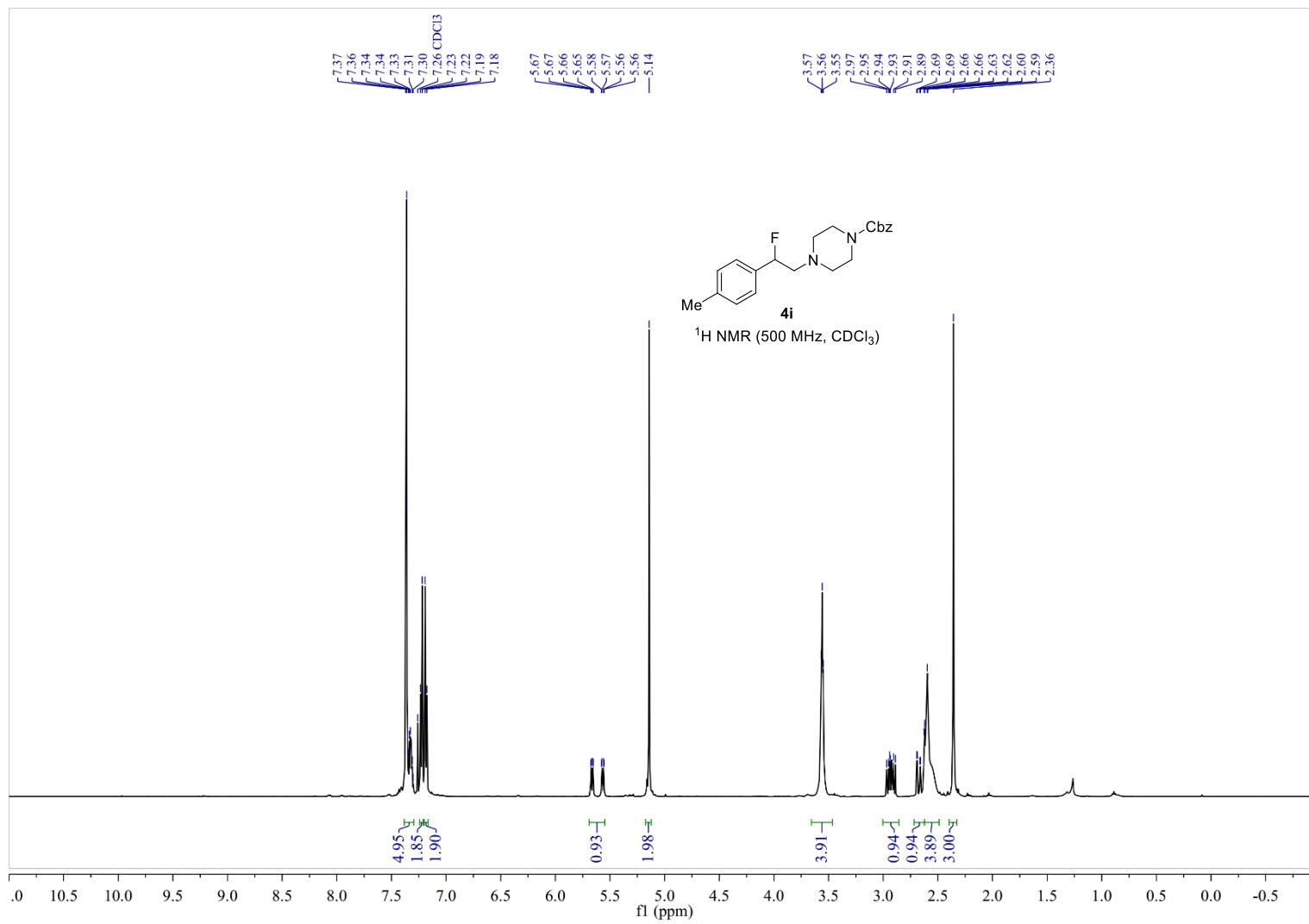


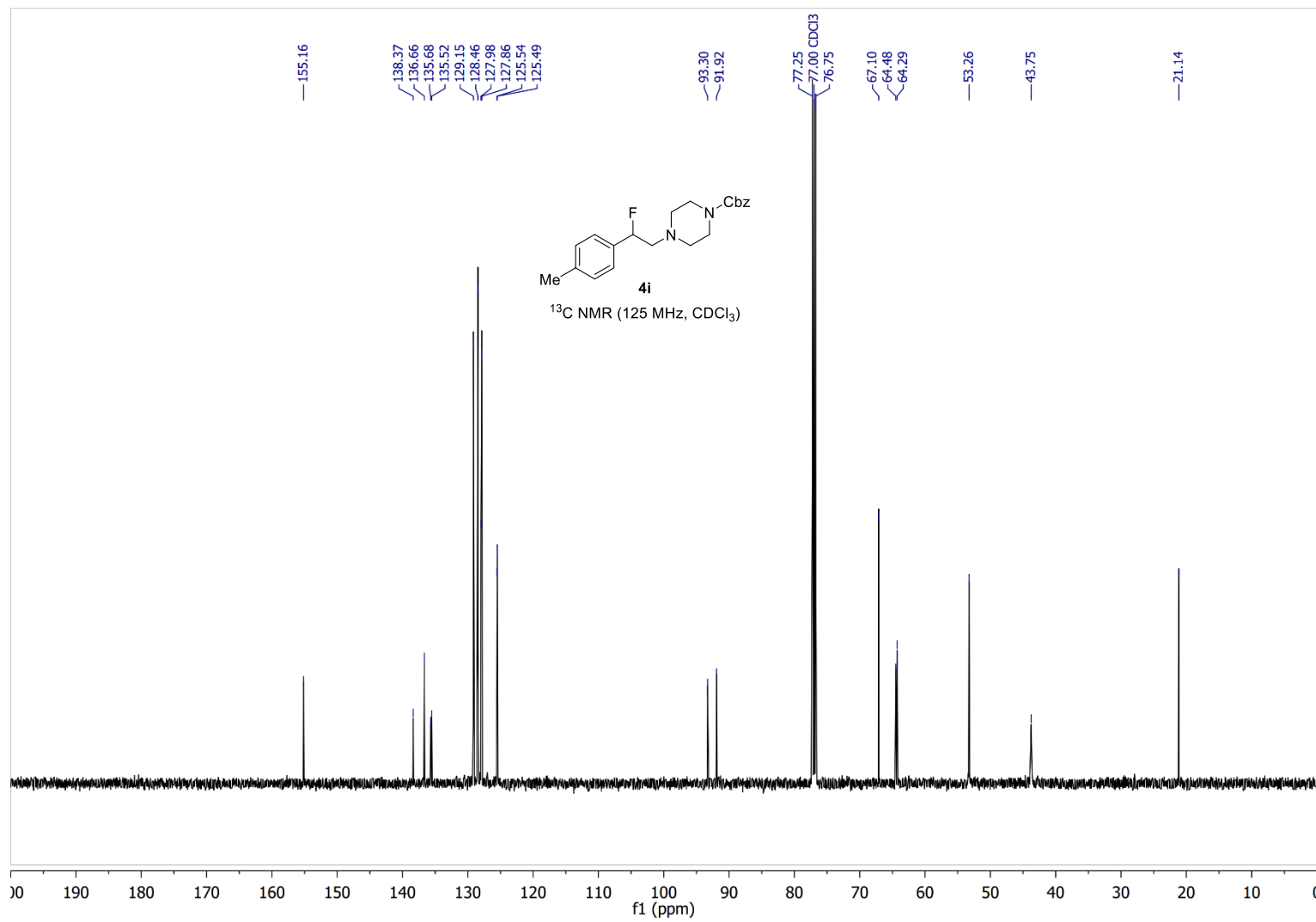
¹⁹F NMR of compound 4h in CDCl₃



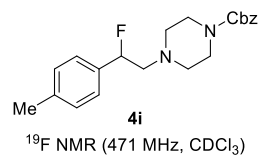
175.1202



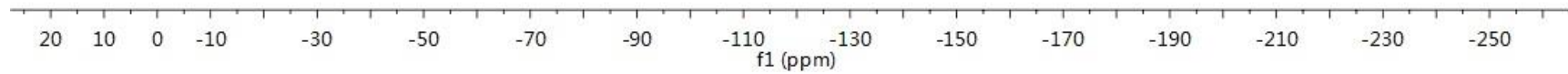


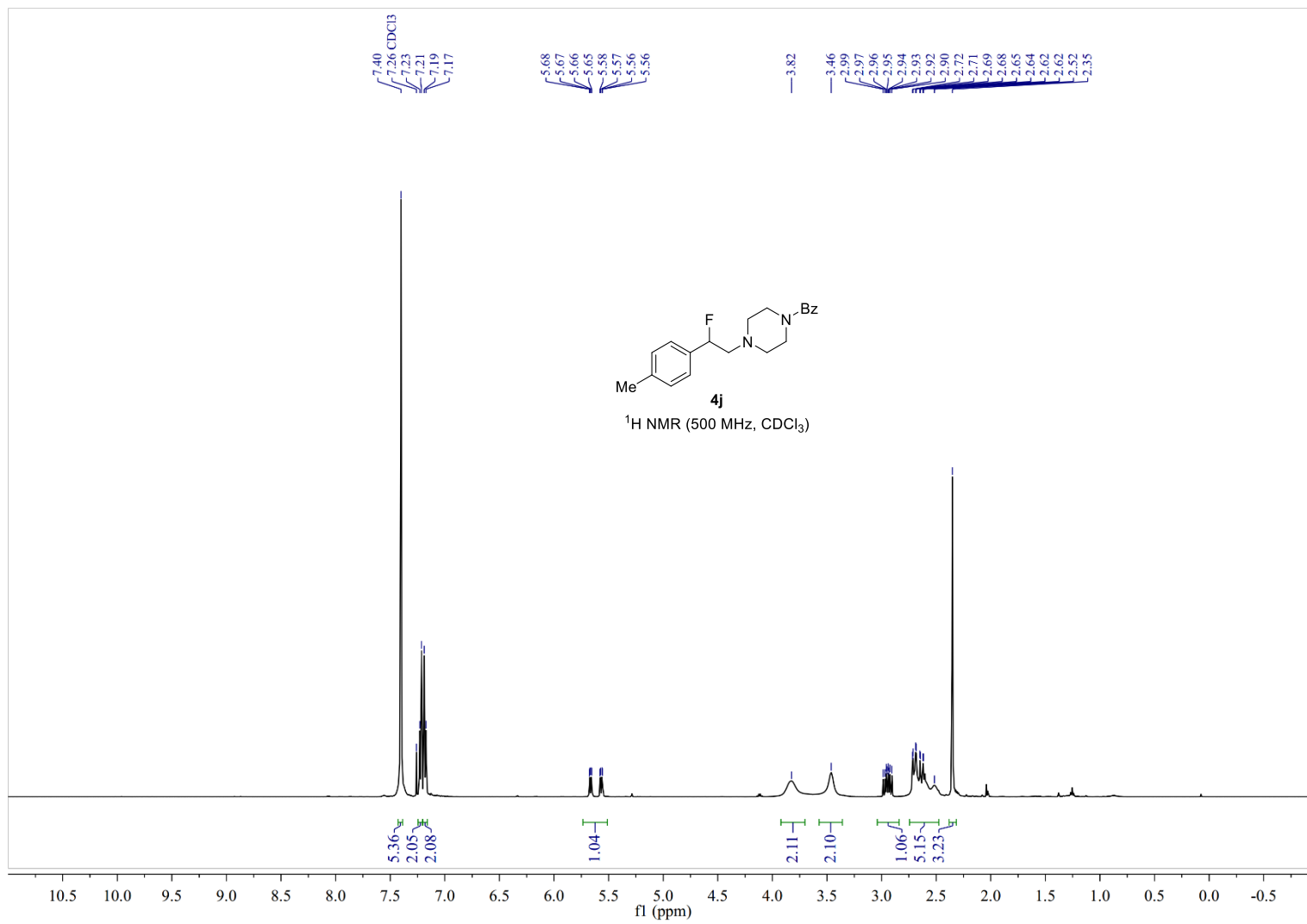


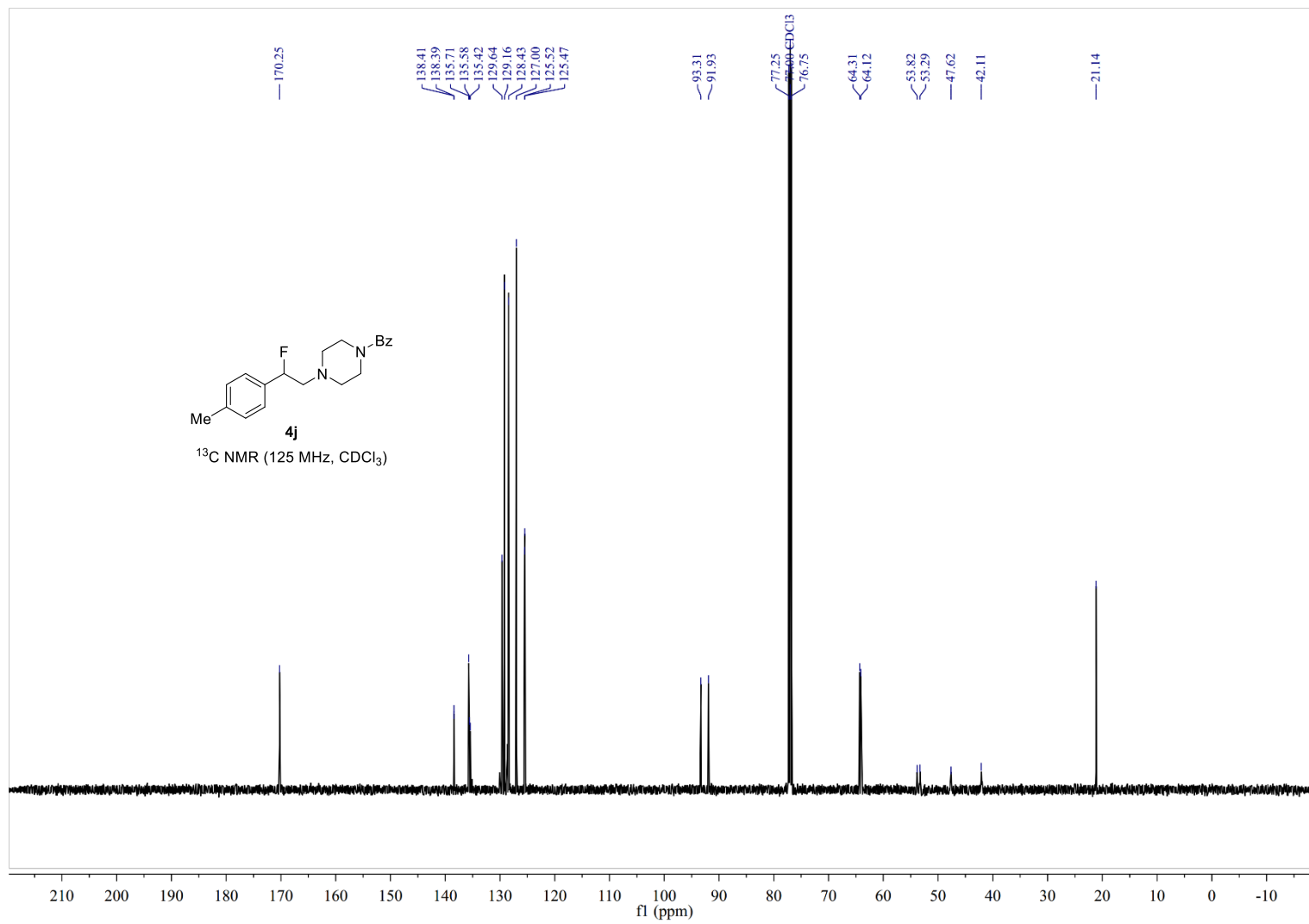
^{19}F NMR gf-3-84bPP in CDCl_3



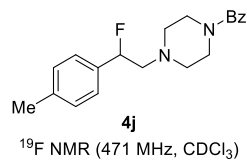
-175.1542



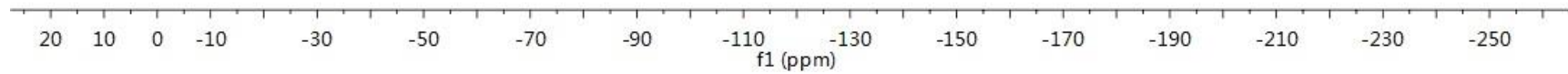




¹⁹F NMR gf-3-96dP in CDCl₃

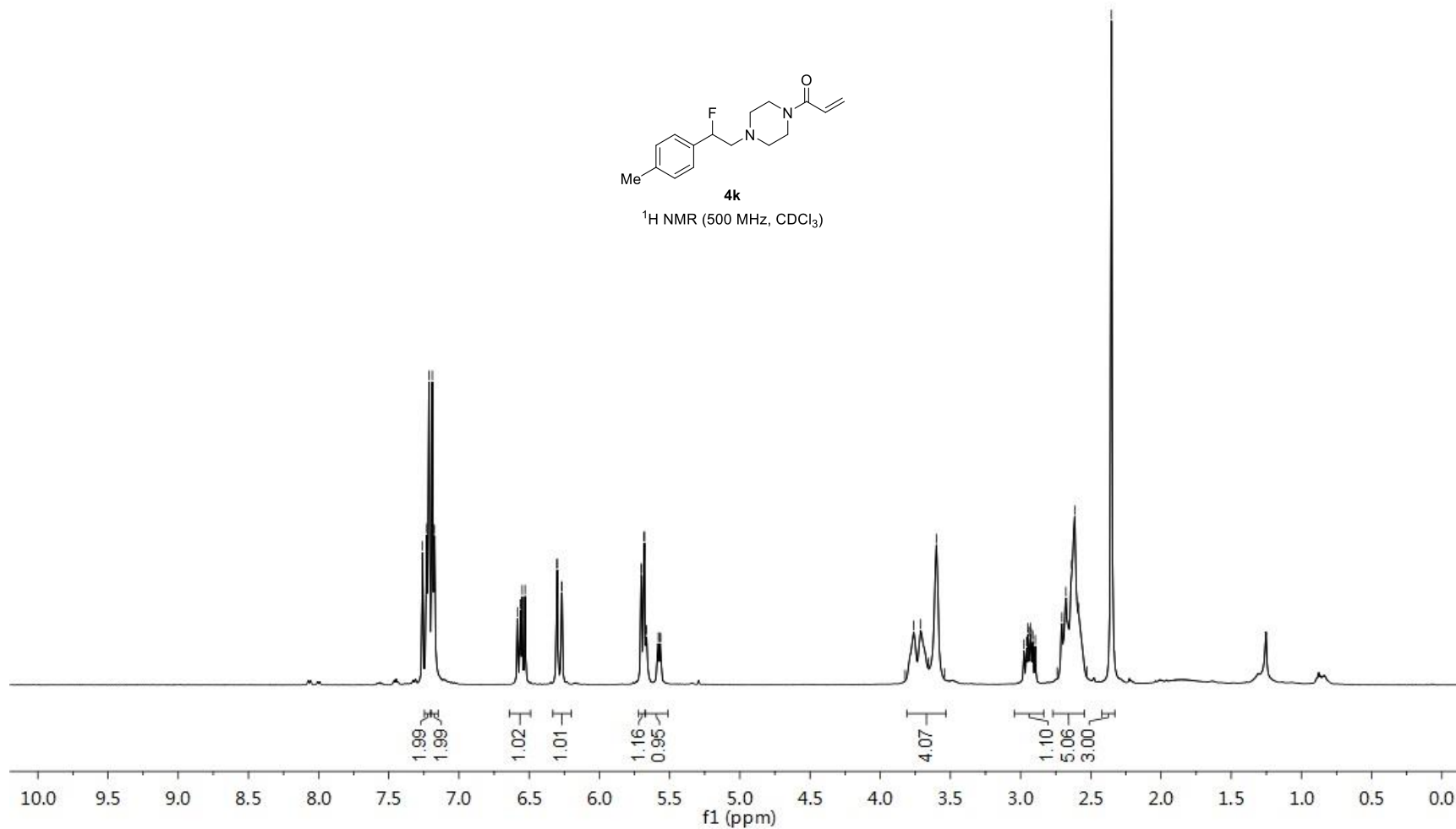
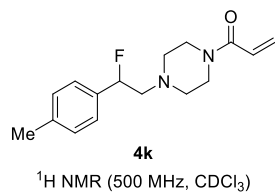


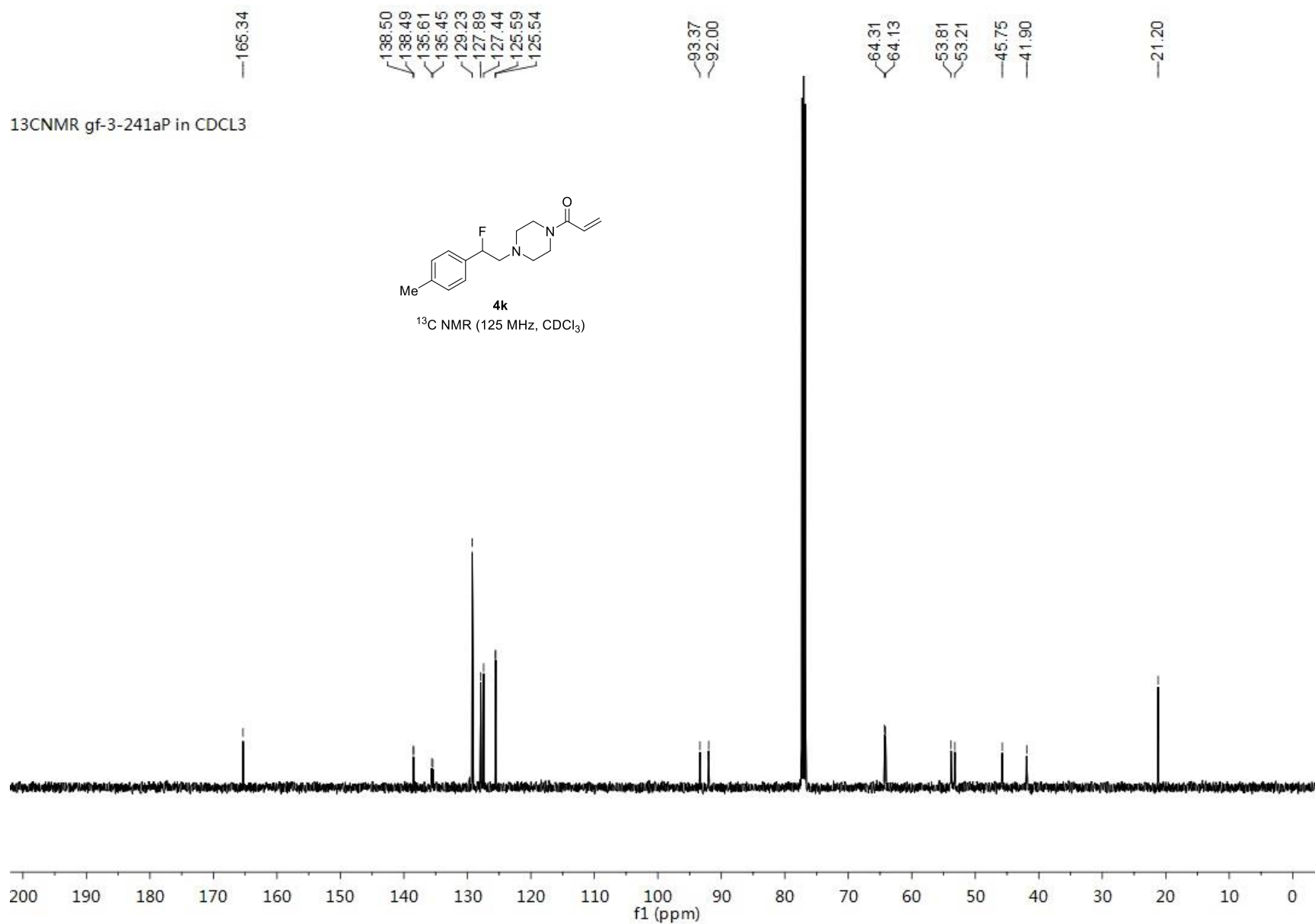
175.1636



7.2603
7.2301
7.2144
7.1900
7.1742
6.5505
6.5293
6.3029
6.3000
6.2693
6.2663
6.2658
5.7000
5.6818
5.6789
5.6650
5.5815
5.5668
5.5651
3.8236
3.7605
3.7116
3.6570
3.6000
3.5419
2.9756
2.9581
2.9468
2.9418
2.9294
2.9243
2.9130
2.8955
2.7402
2.7060
2.6774
2.6375
2.6151
2.5876
2.5290
2.3531

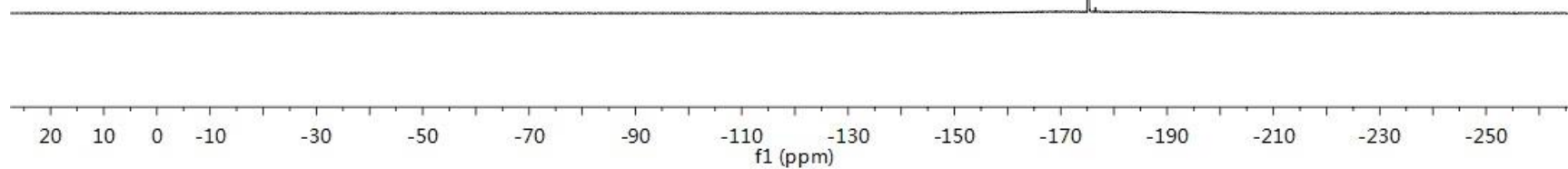
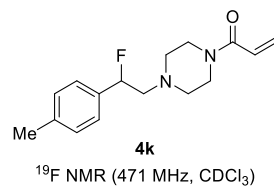
¹H NMR of gf-3-241aP in CDCl₃





¹⁹F NMR gf-3-241aP in CDCl₃

---175.1631



8.3103
8.3008

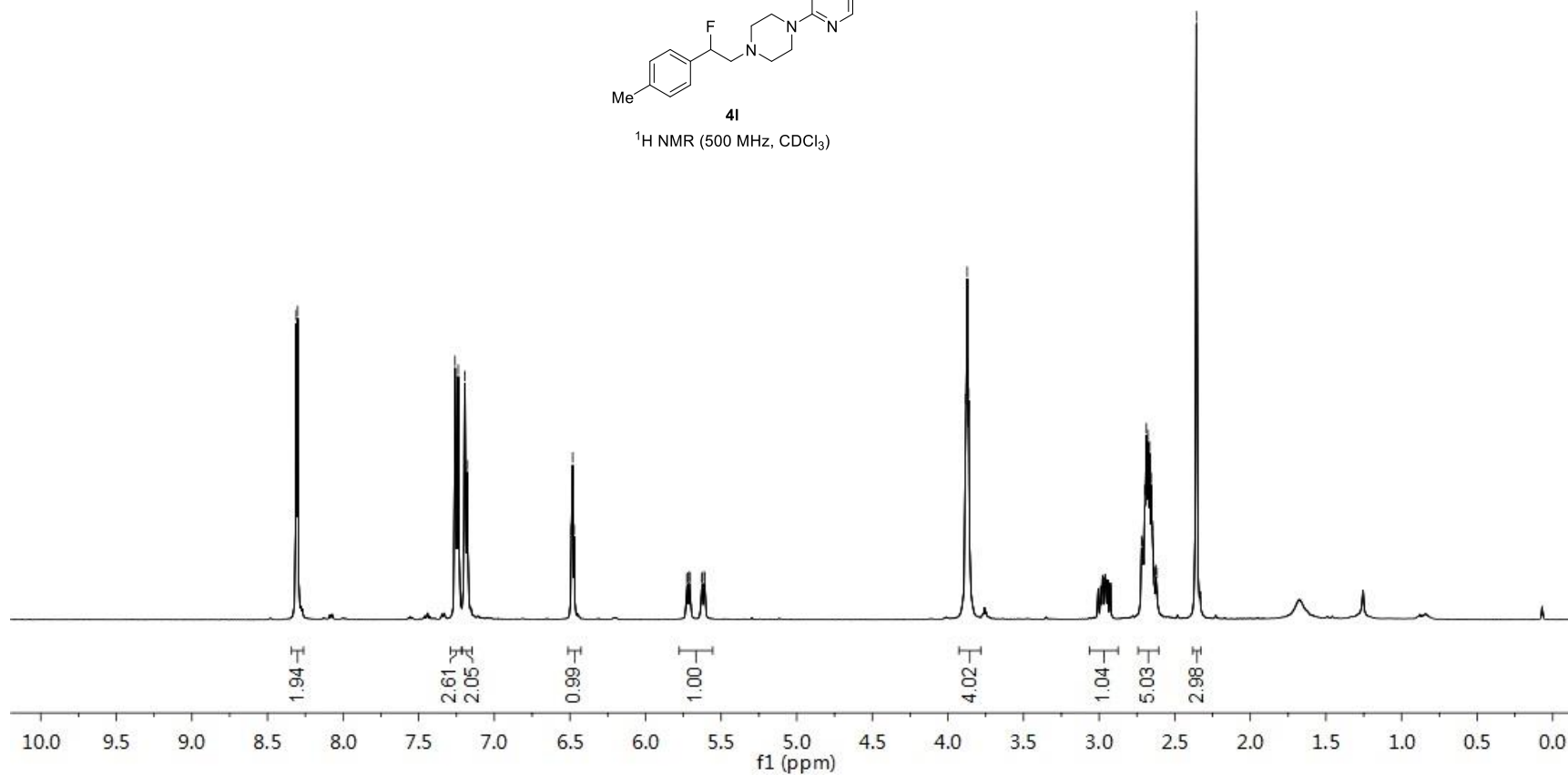
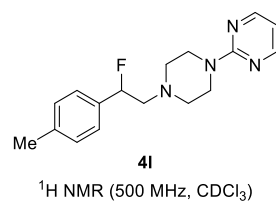
7.2597
7.2546
7.2385
7.1951
7.1794

6.4912
6.4818
6.4723
5.7260
5.7235
5.7095
5.7064
5.6290
5.6259
5.6117
5.6092

3.8821
3.8724
3.8627

2.7200
2.7154
2.7106
2.6974
2.6874
2.6769
2.6663
2.6554
2.6466
2.6339
2.6233
2.6188
2.3568

¹H NMR of 3-93cP in CDCl₃



—161.69
—157.73

138.37
138.36
135.90
135.75
129.20
125.63
125.58

—109.89

v

93.38
92.00

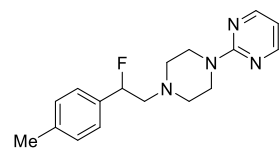
64.75
64.57

—53.54

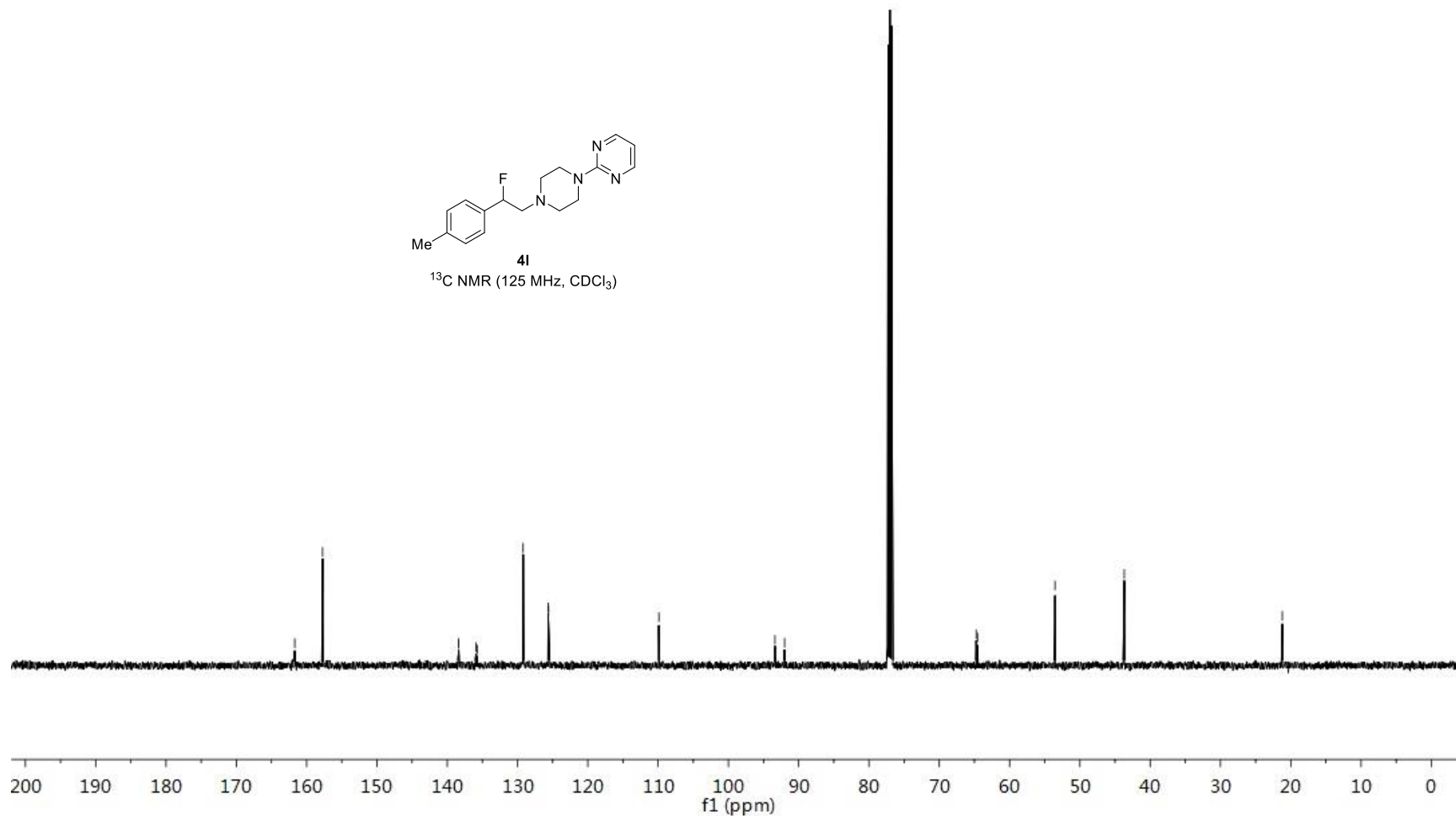
—43.68

—21.20

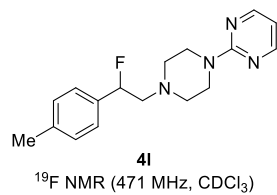
¹³CNMR gf-3-93cP in CDCl₃



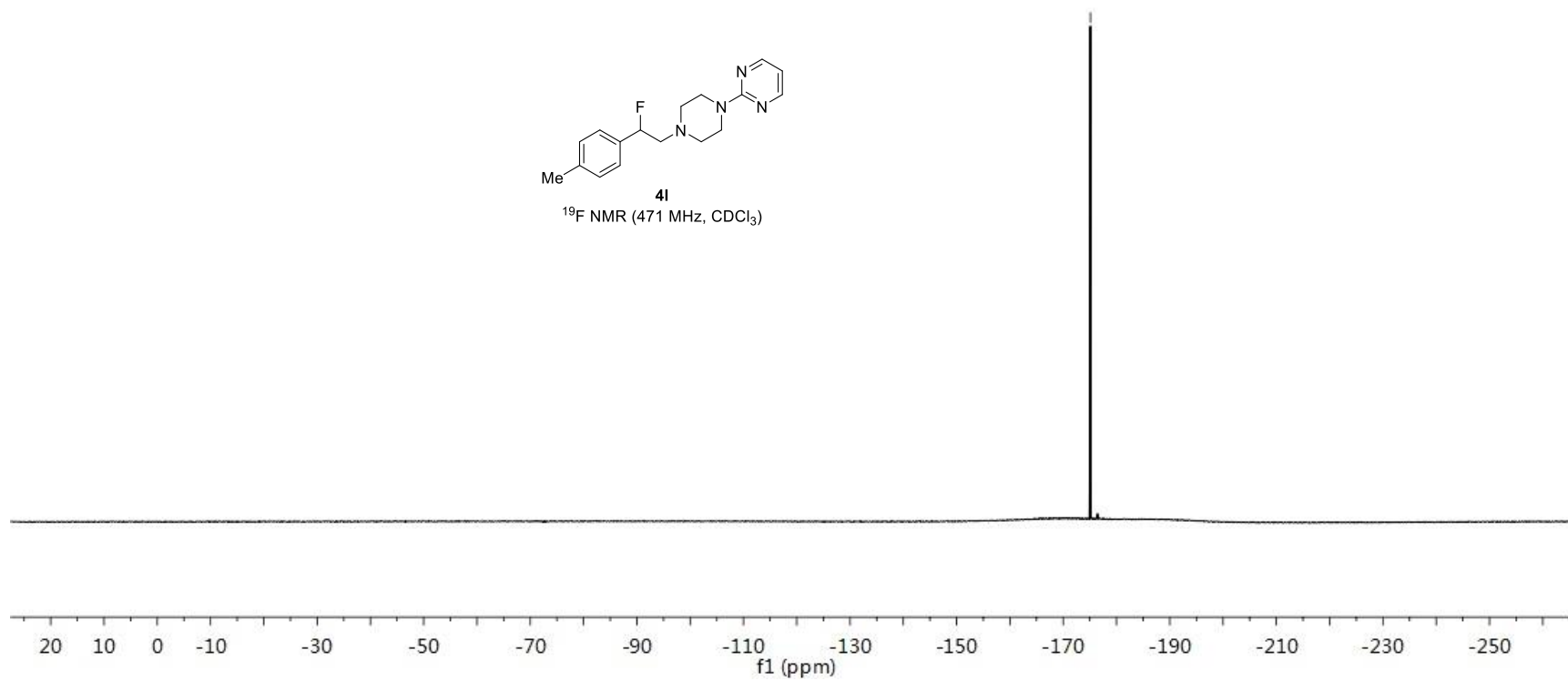
¹³C NMR (125 MHz, CDCl₃)



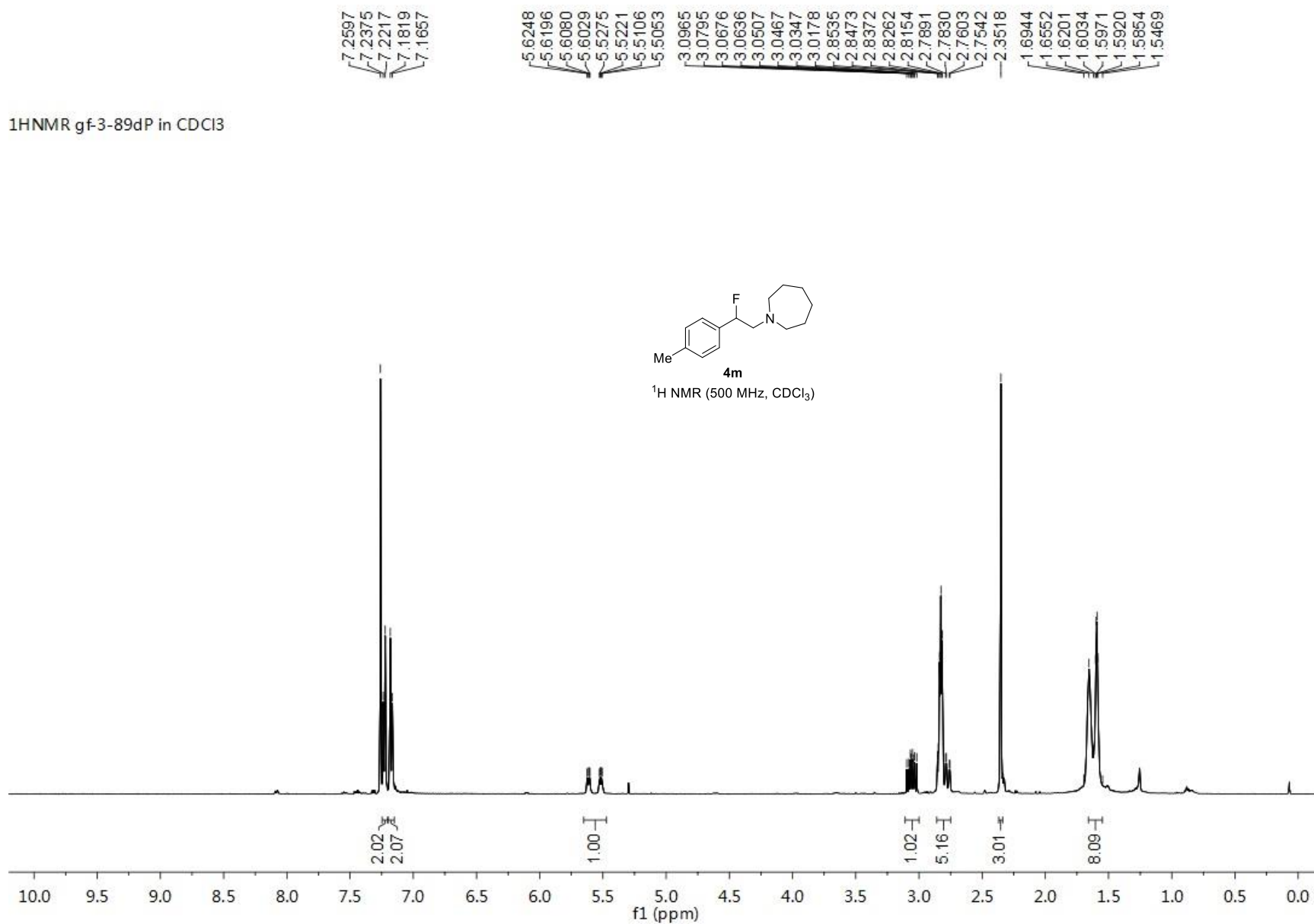
¹⁹F NMR gf-3-93cP in CDCl₃

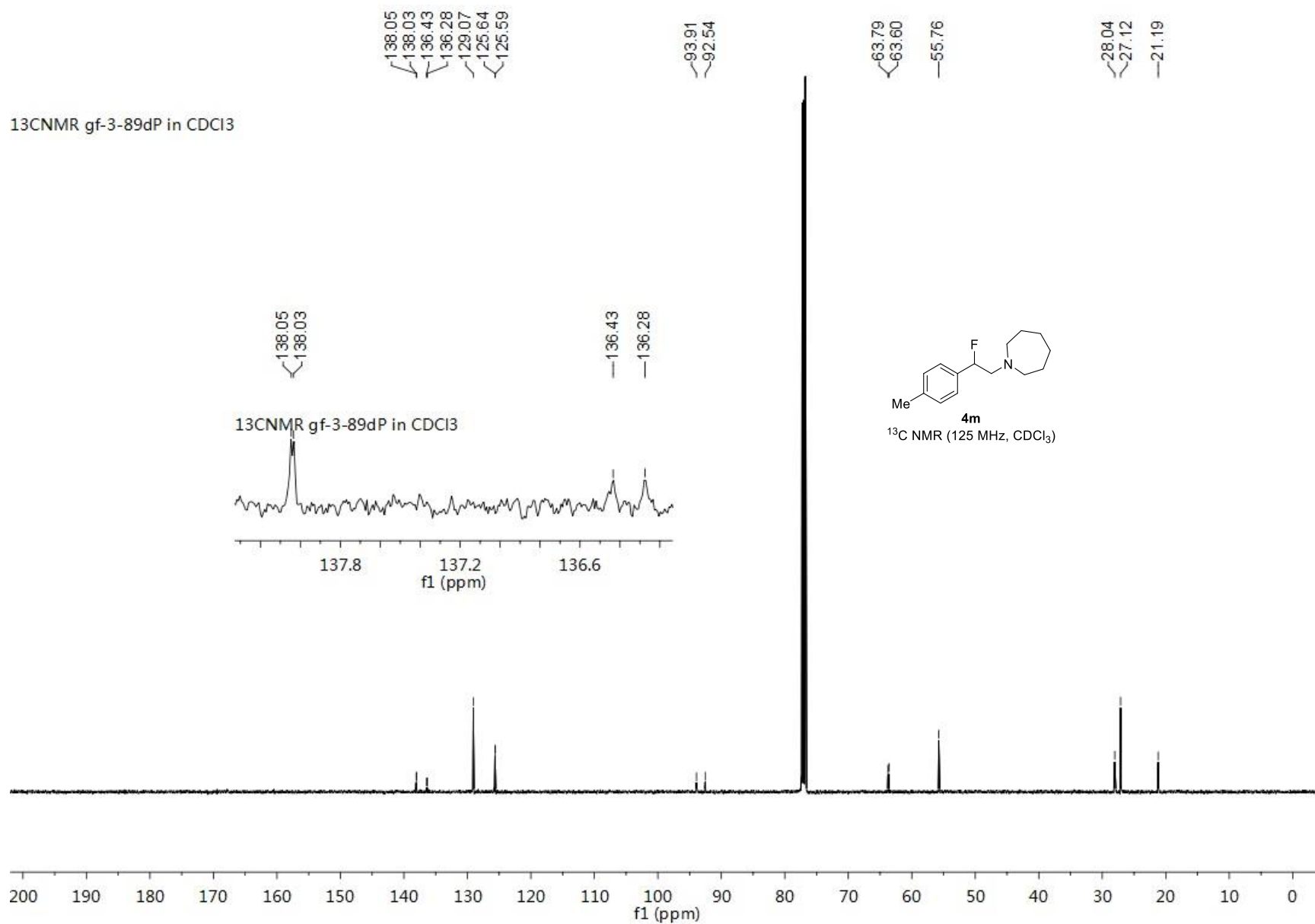


-175.1137



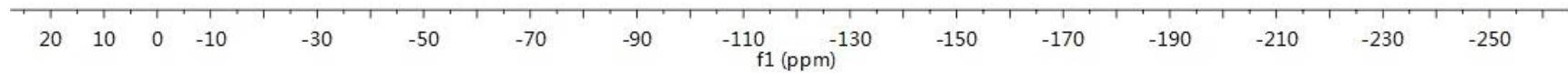
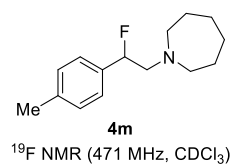
¹H NMR of compound 4m in CDCl₃

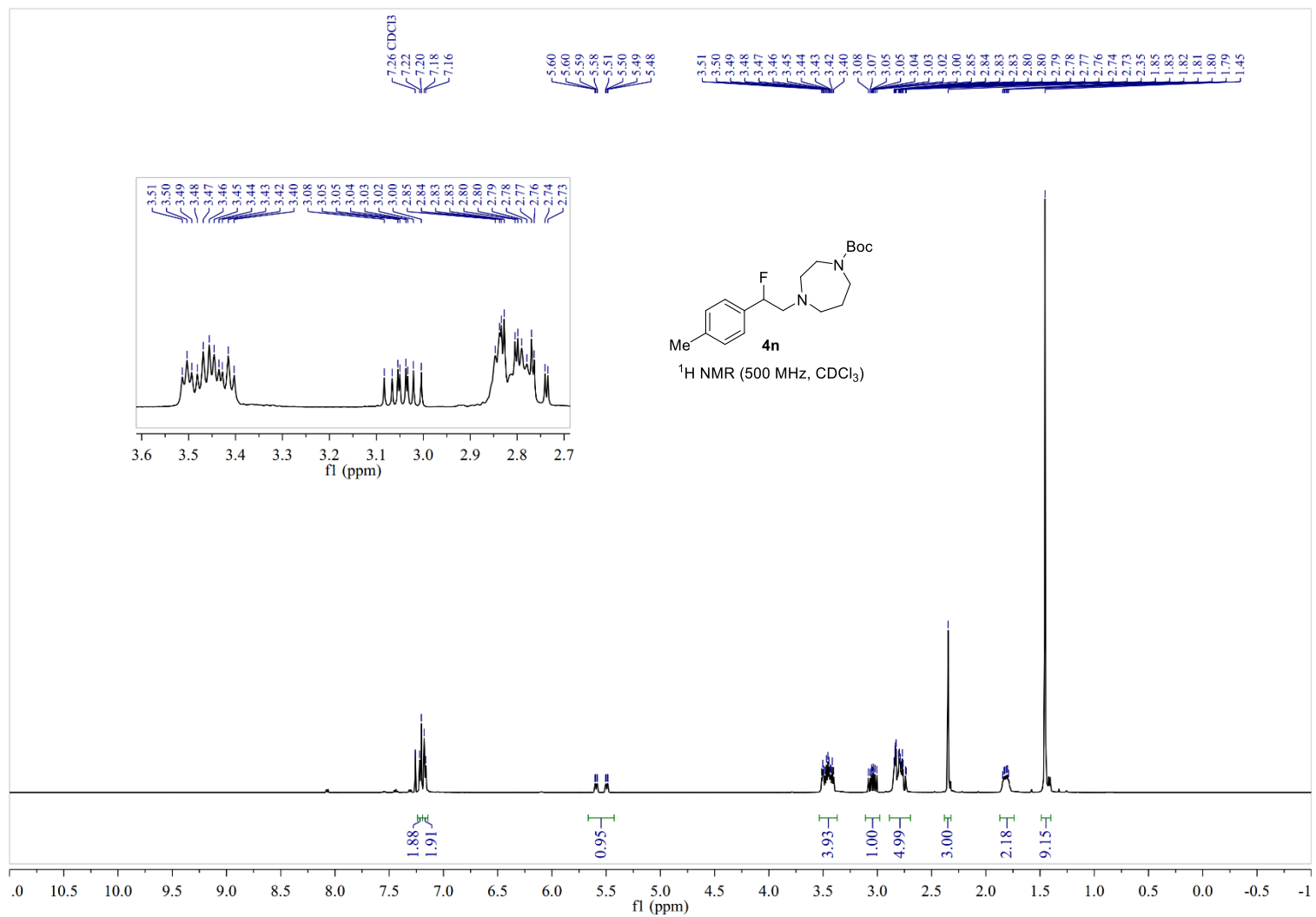


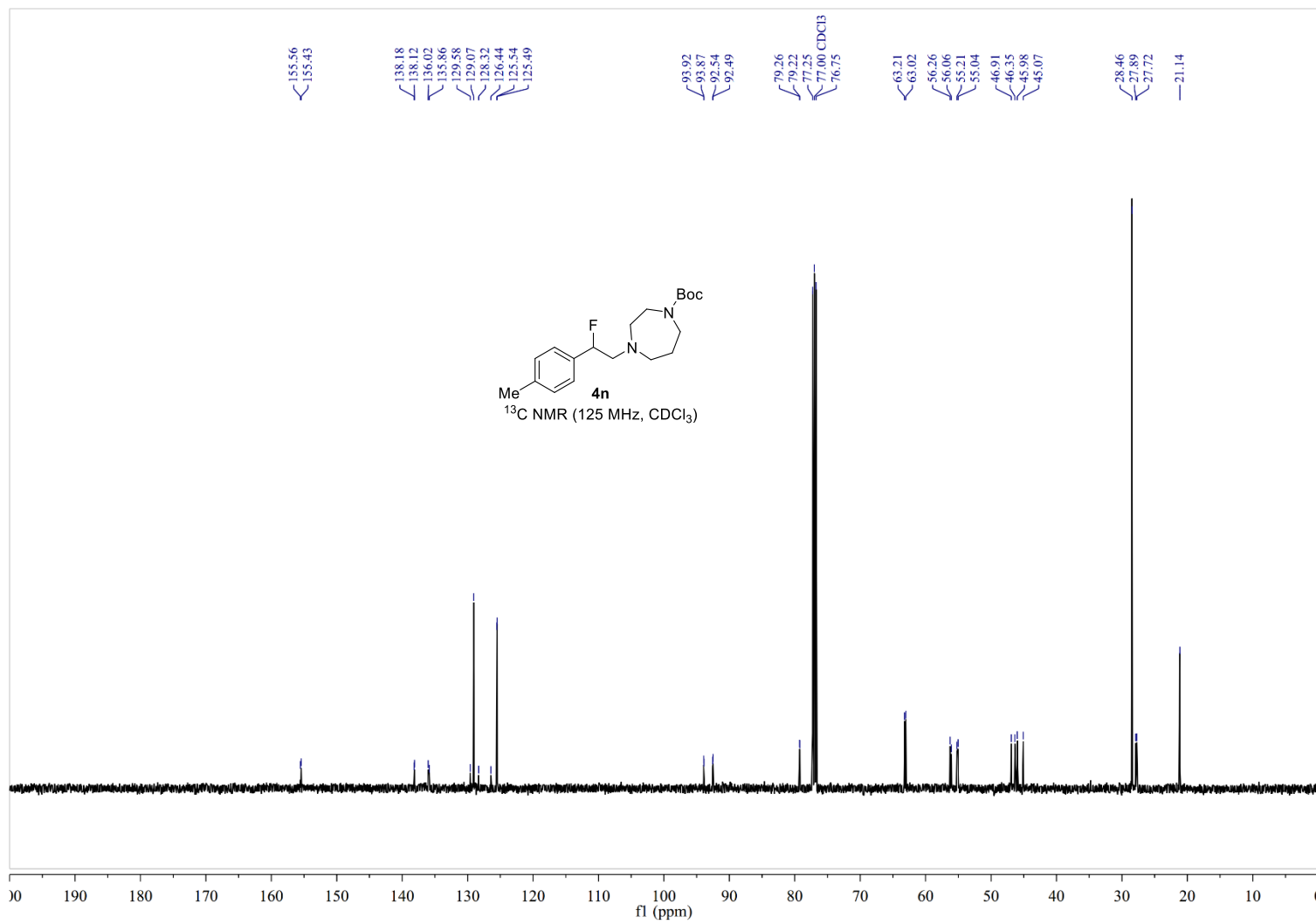


¹⁹F NMR gf-3-89dP in CDCl₃

— -176.9541

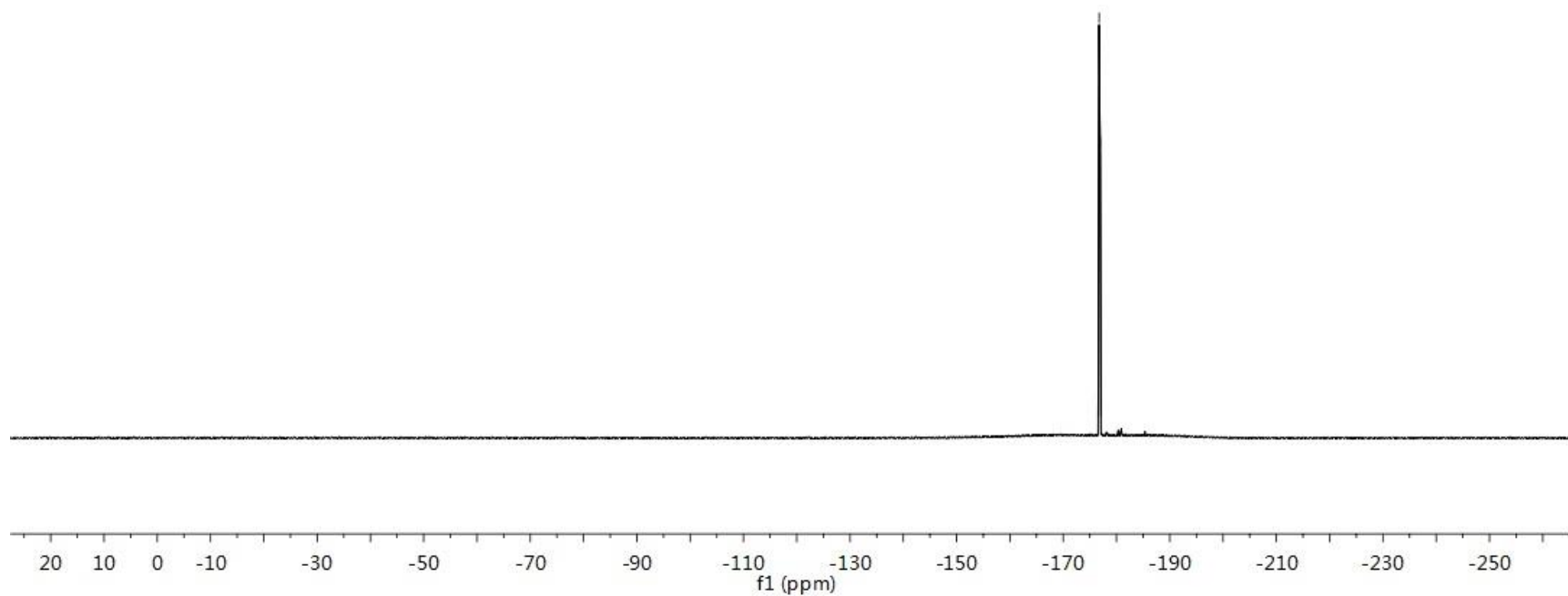
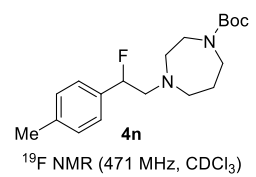






¹⁹F NMR of gf-ck-II-96dP in CDCl₃

176.7485
176.9494

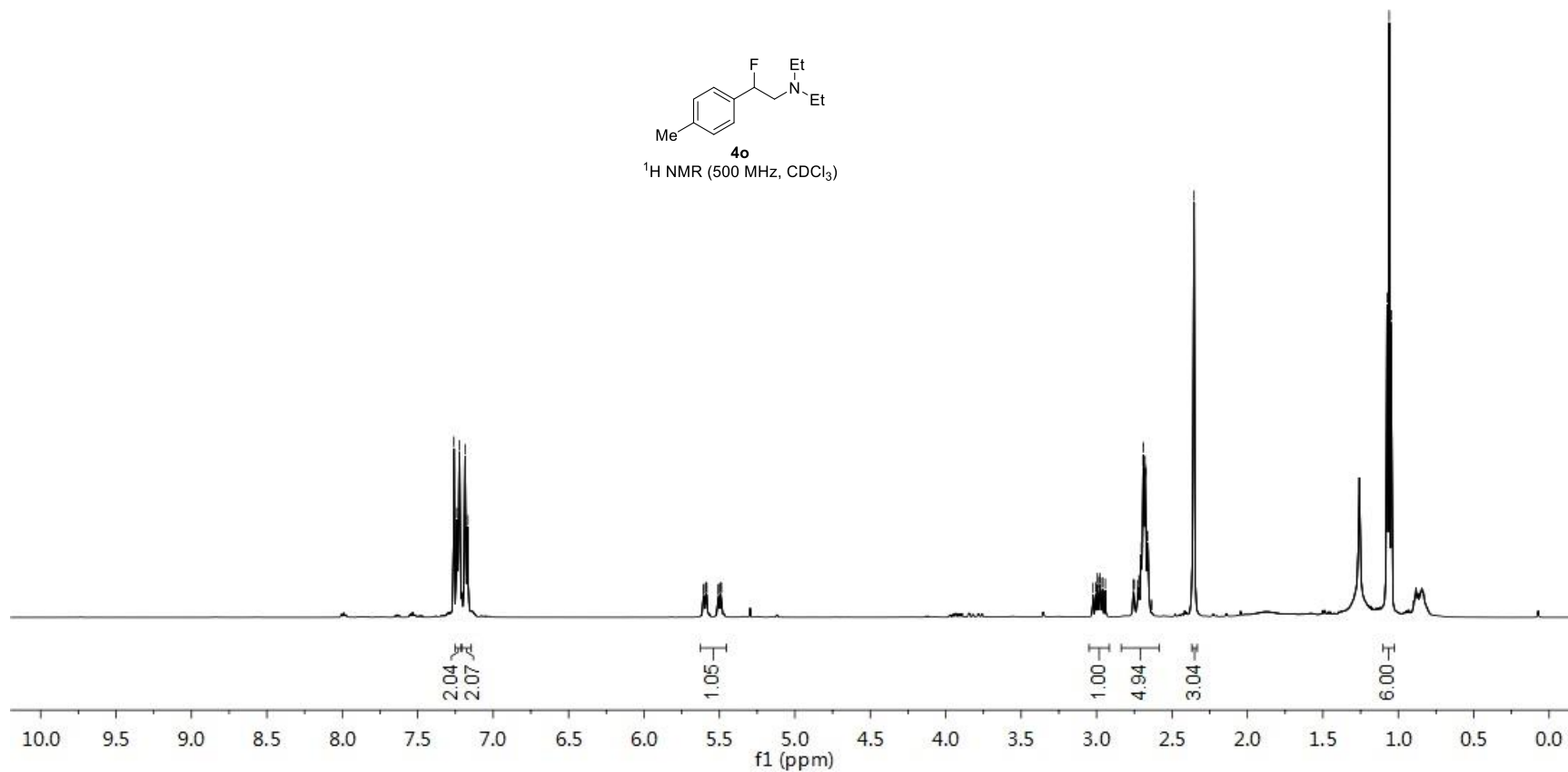
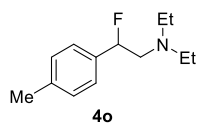


7.2602
7.2393
7.2236
7.1862
7.1704

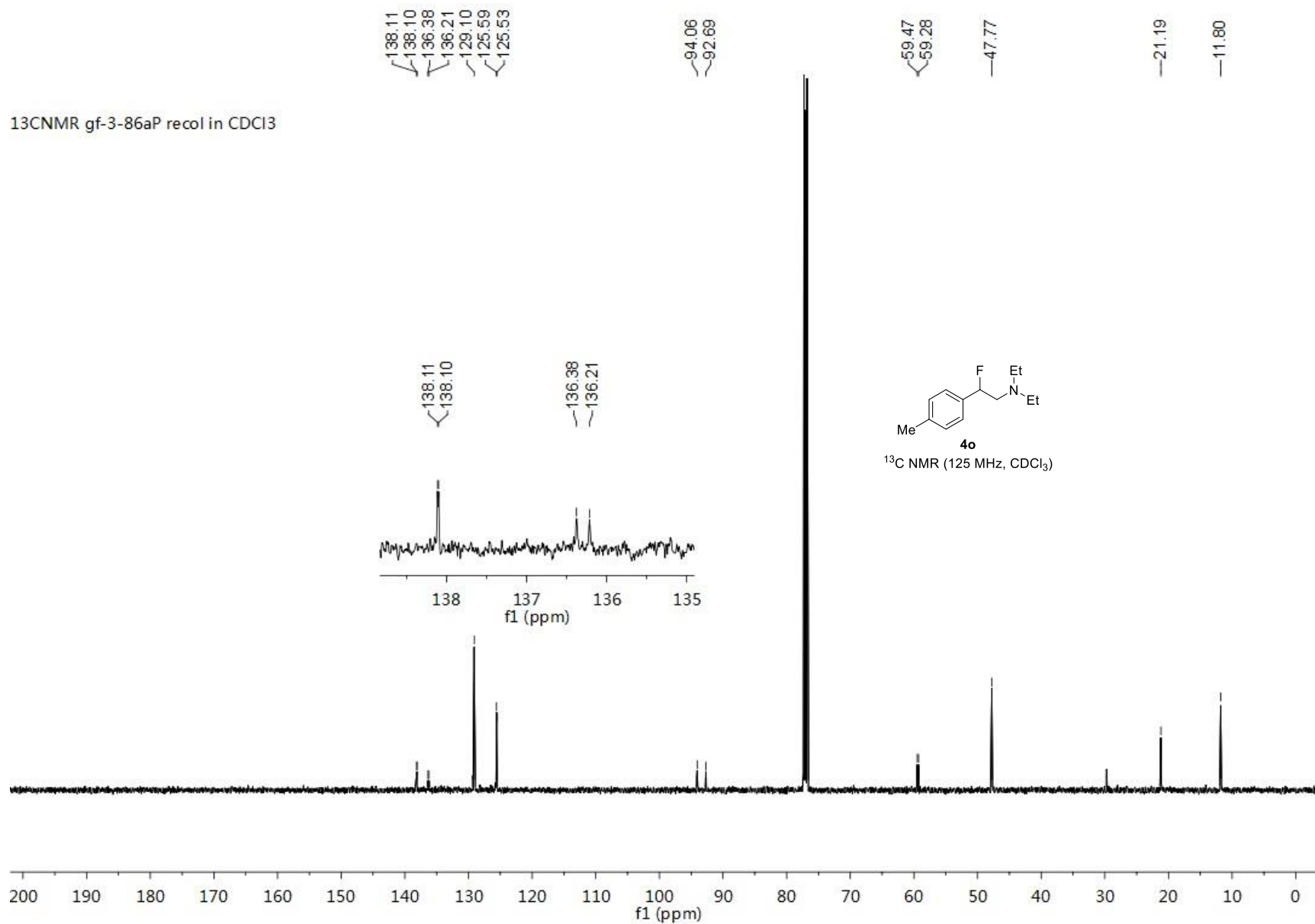
5.6065
5.6014
5.5890
5.5842
5.5091
5.5041
5.4917
5.4869

3.0232
3.0059
2.9938
2.9897
2.9765
2.9724
2.9602
2.9429
2.7558
2.7503
2.7264
2.7204
2.7031
2.6922
2.6890
2.6783
2.6750
2.6606
2.6344
2.3540
1.0745
1.0602
1.0460

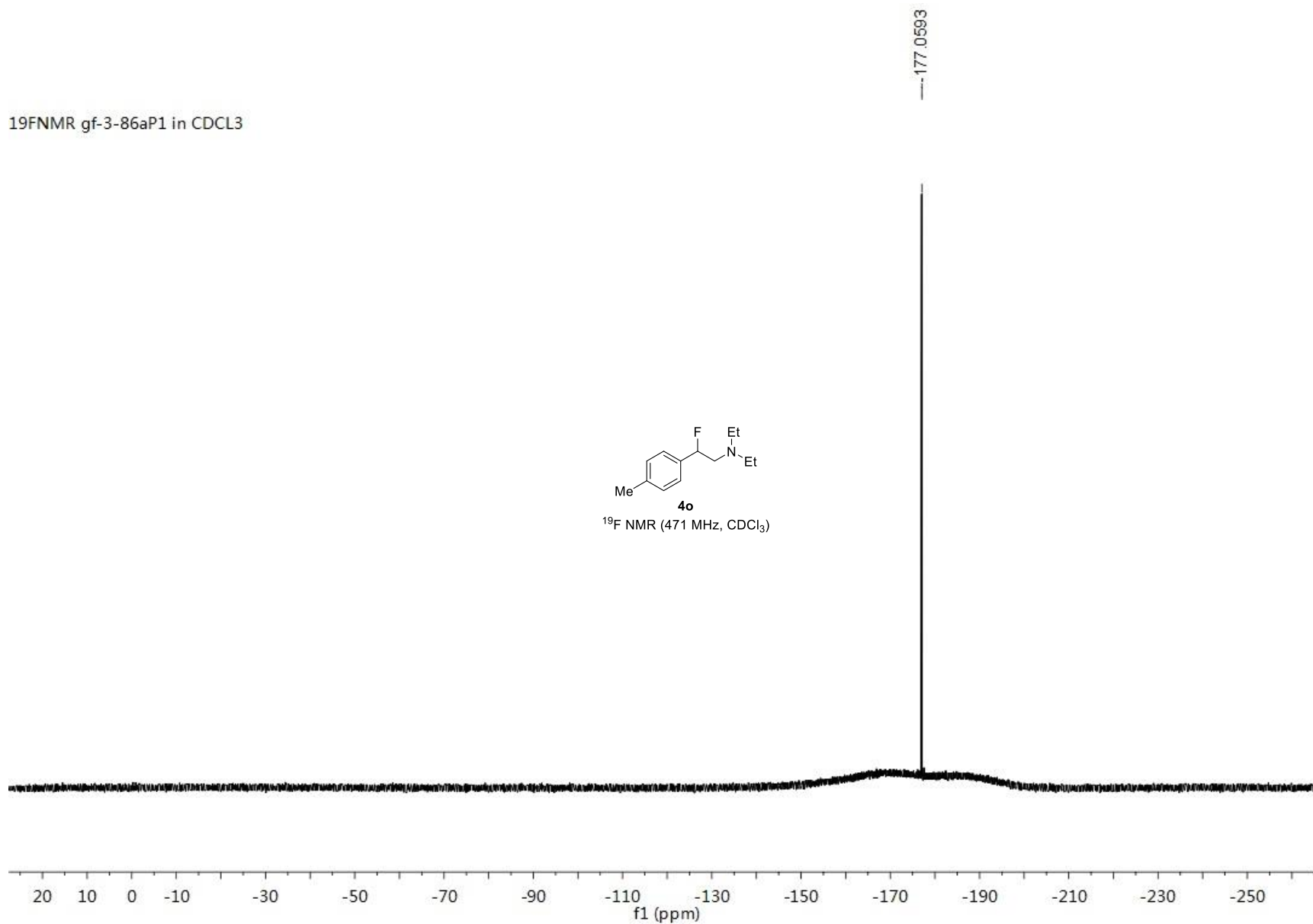
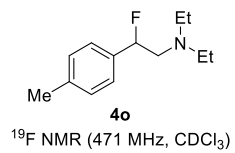
¹H NMR gf-3-86aP recol in CDCl₃



¹³CNMR gf-3-86aP recol in CDCl₃



¹⁹F NMR gf-3-86aP1 in CDCl₃

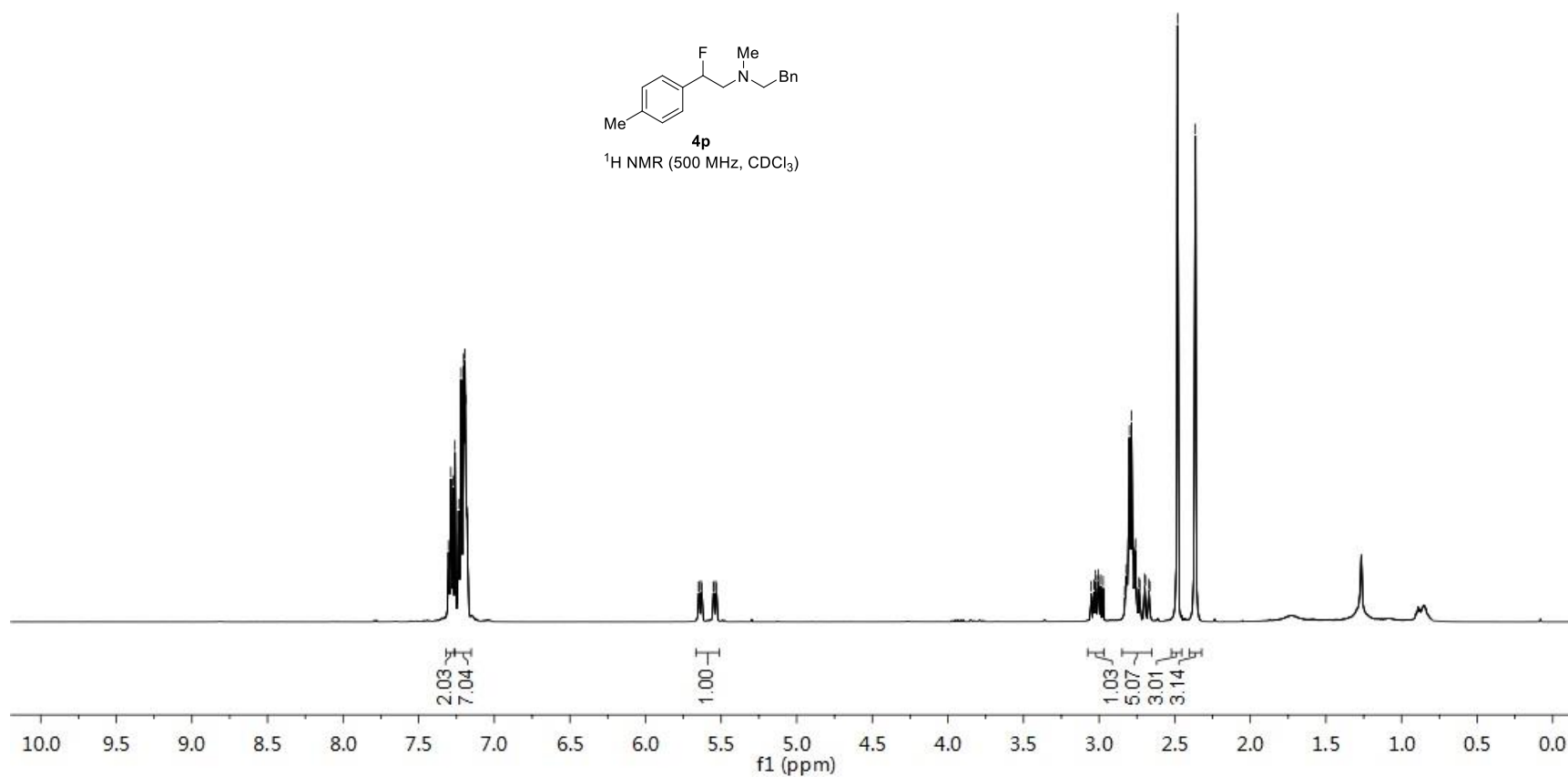
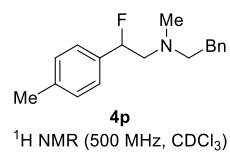


7.3029
7.2871
7.2780
7.2725
7.2605
7.2359
7.2195
7.2032
7.2000
7.1942
7.1883
7.1787

5.6491
5.6436
5.6319
5.6265
5.5518
5.5462
5.5346
5.5290

3.0525
3.0352
3.0239
3.0194
3.0066
3.0021
2.9908
2.9735
2.8293
2.8199
2.8139
2.8064
2.7990
2.7934
2.7856
2.7804
2.7750
2.7649
2.7596
2.7502
2.7362
2.7305
2.6991
2.6934
2.6706
2.6649
2.4812
2.3635

¹H NMR of **4p** in CDCl₃



¹³CNMR gf-3-85dP in CDCl₃

140.30
138.24
138.22
136.11
135.96
129.16
128.74
128.41
126.03
125.63
125.58

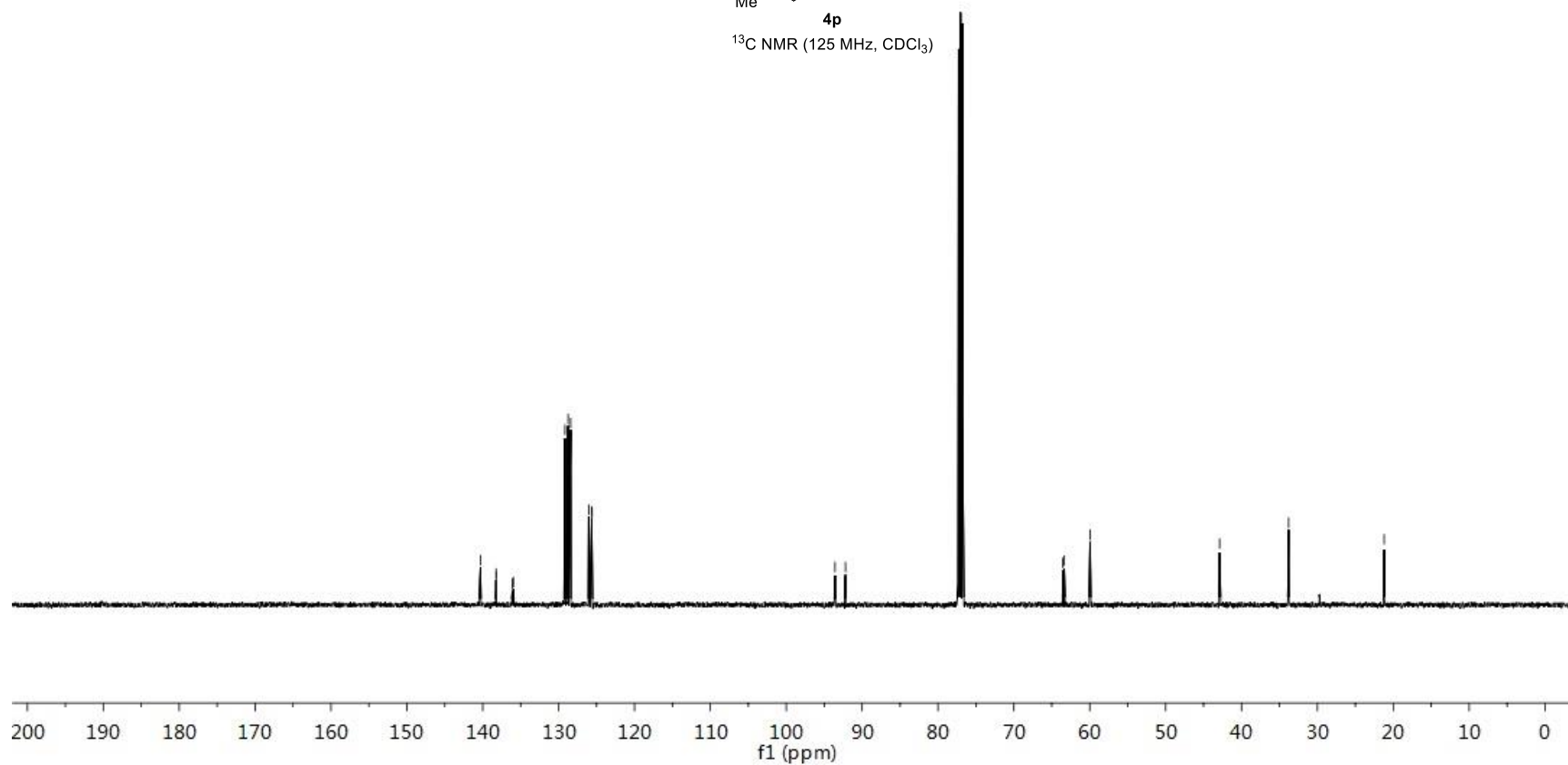
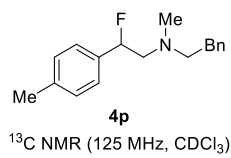
93.57
92.20

63.54
63.35
59.96

42.89

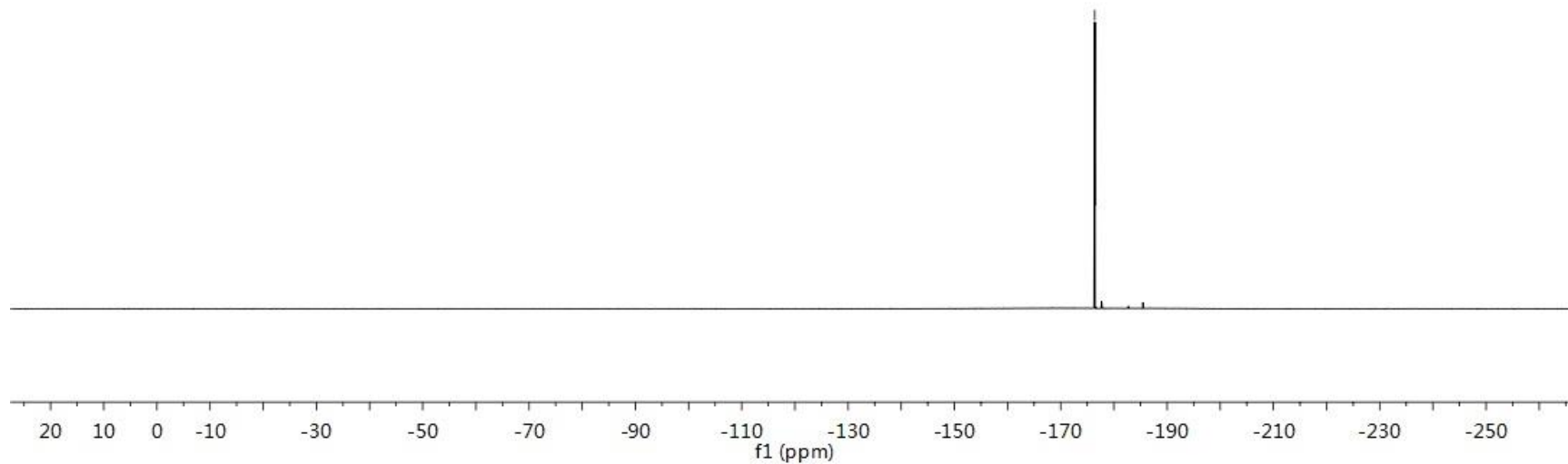
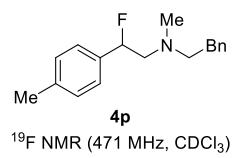
33.79

21.21

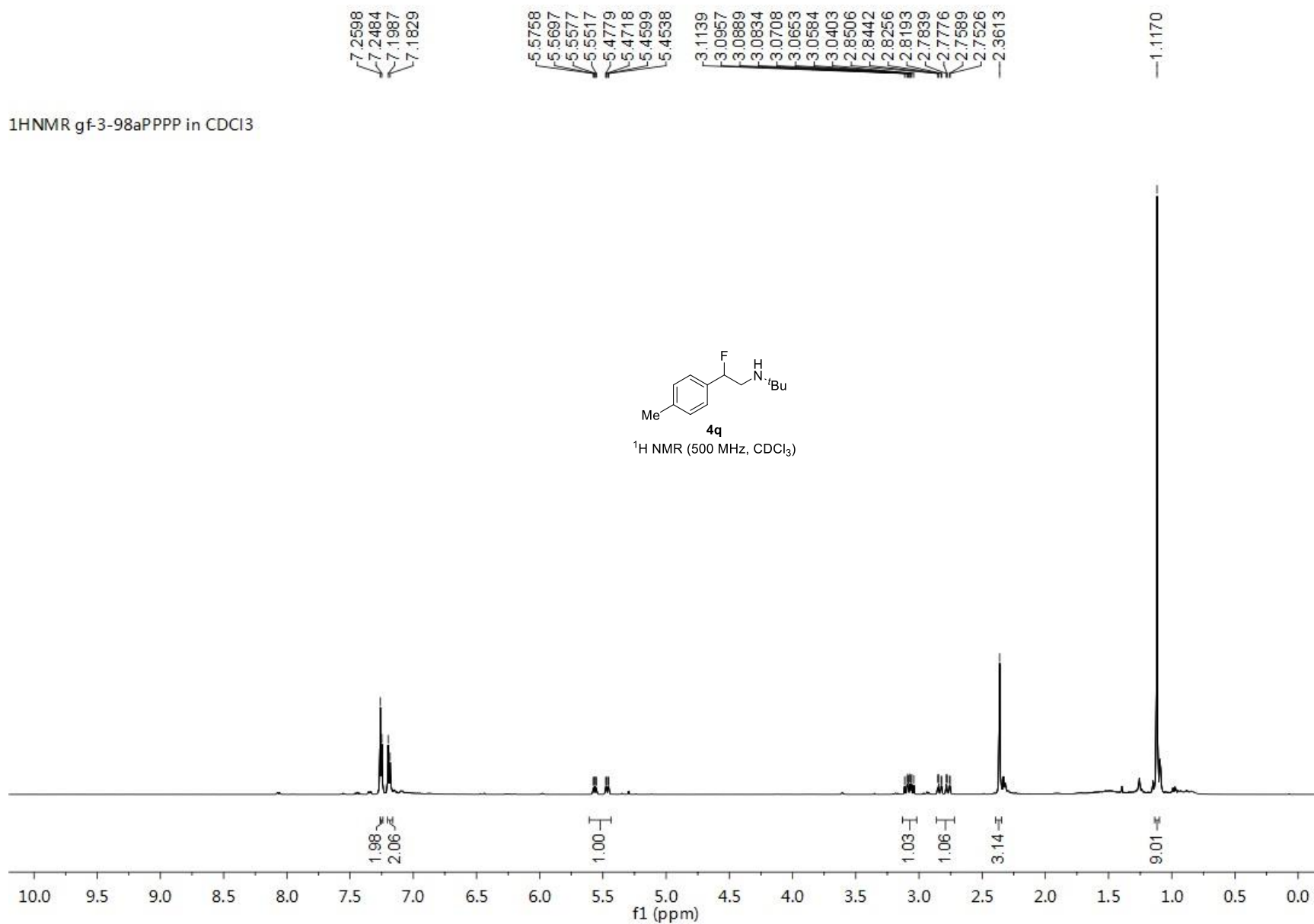


¹⁹F NMR gf-3-85dP in CDCl₃

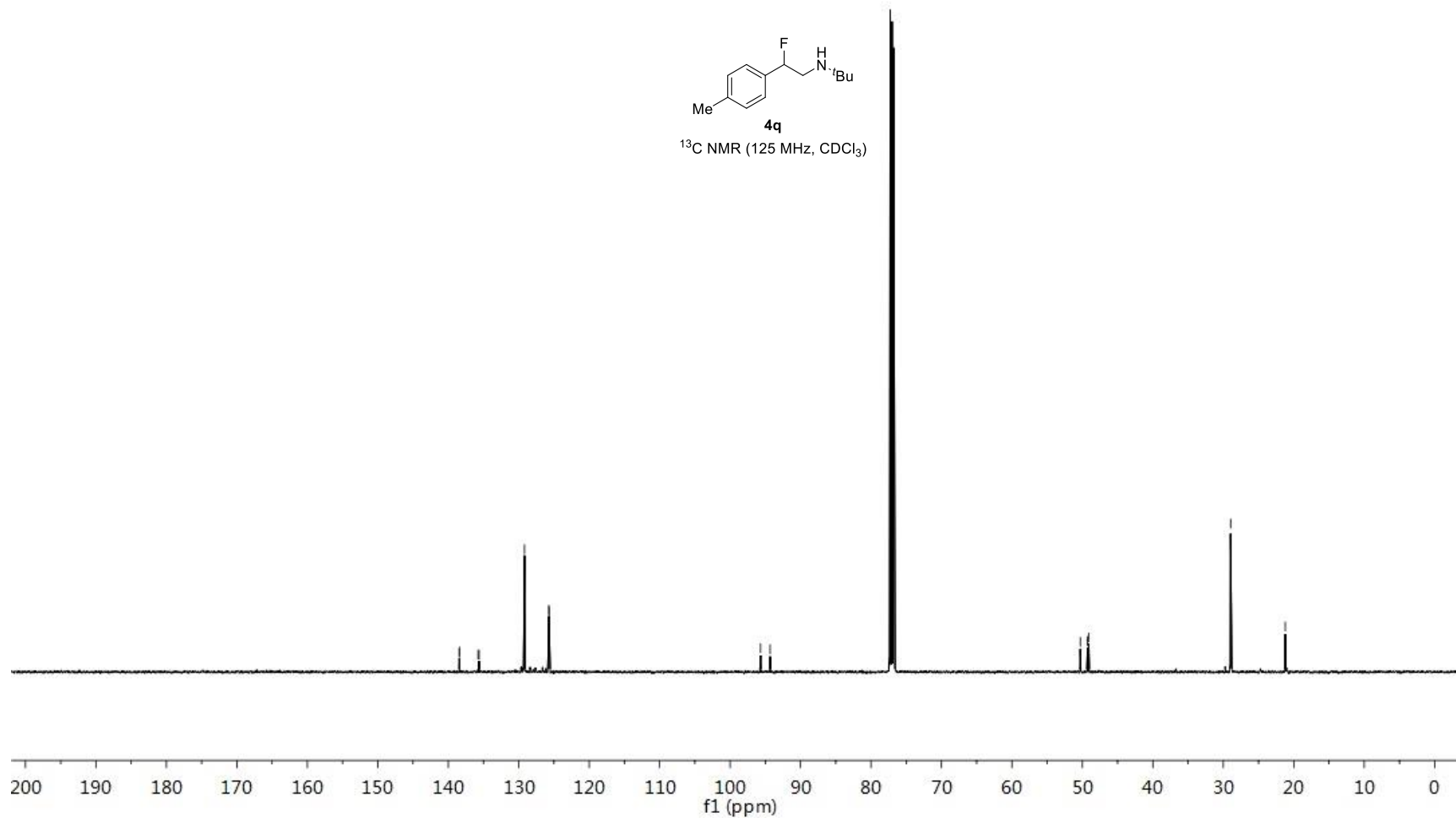
---176.4063



¹H NMR of gf-3-98aPPPP in CDCl₃

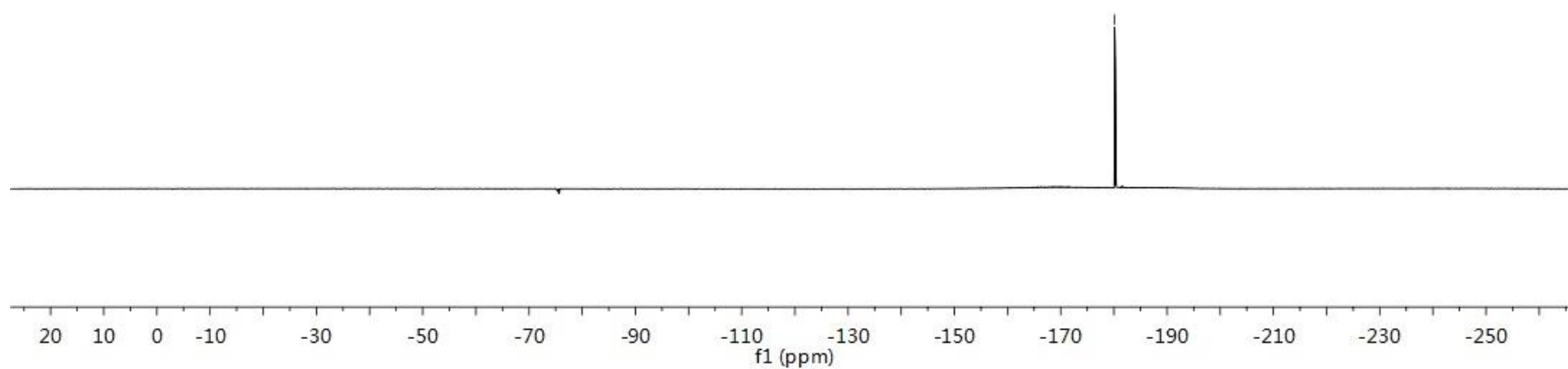
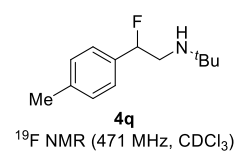


¹³CNMR gf-3-98aPPPP in CDCl₃

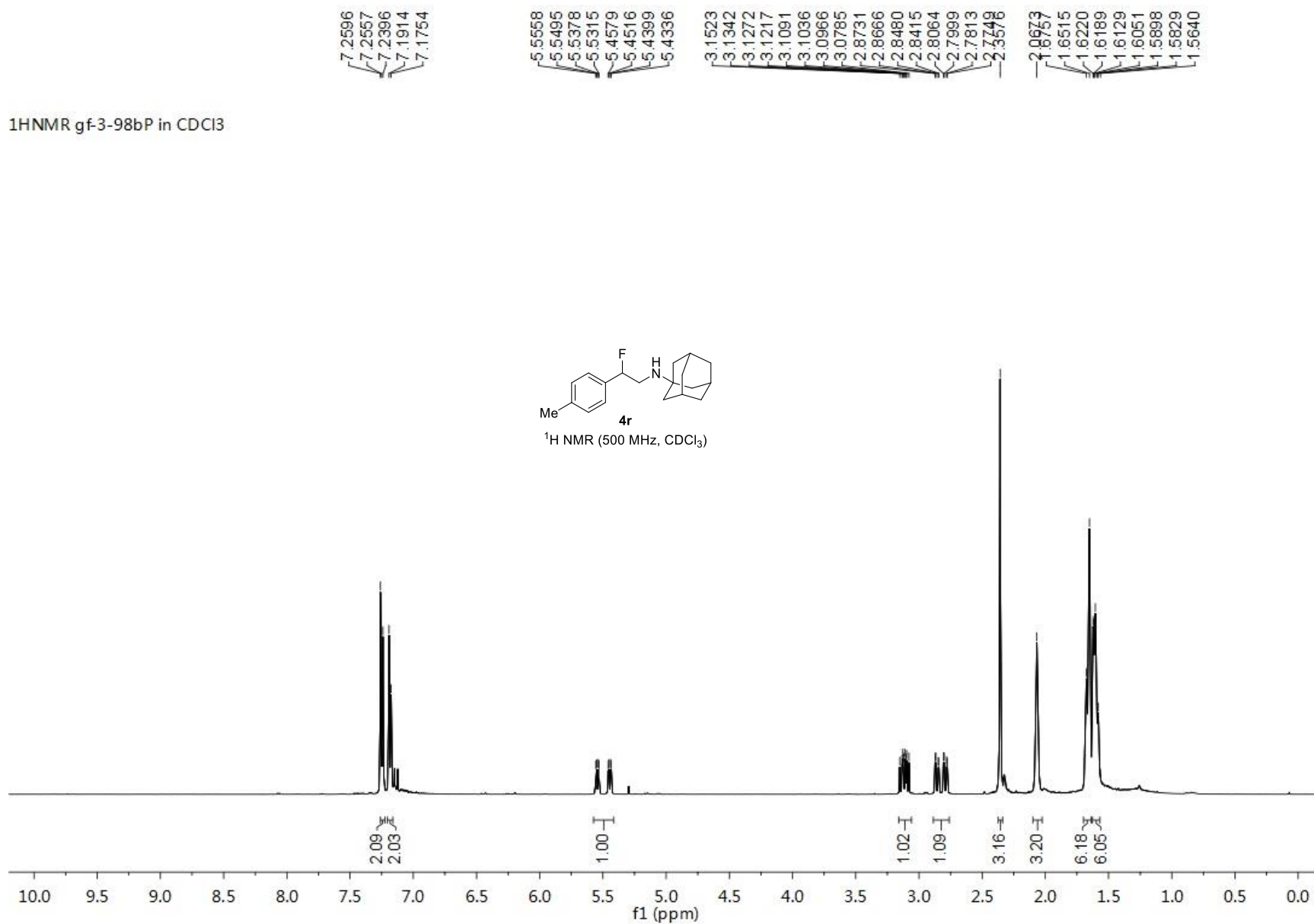


¹⁹F NMR gf-3-98aPPPP in CDCl₃

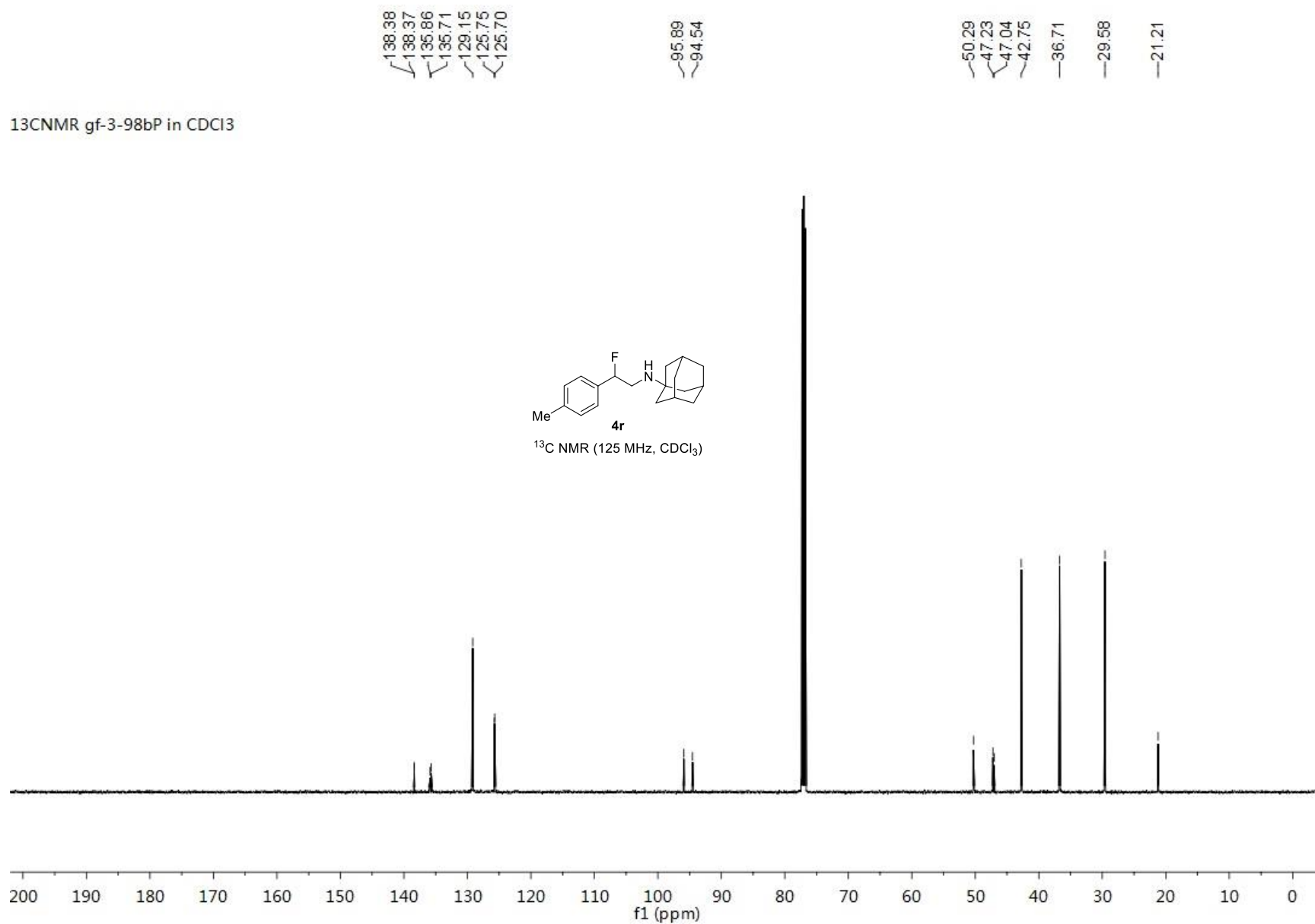
— 180.1432



¹H NMR of compound 4r in CDCl₃

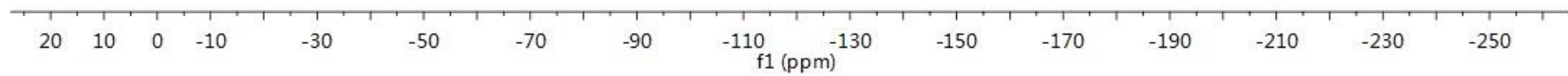
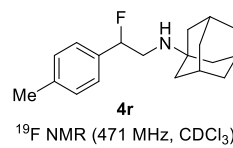


¹³C NMR gf-3-98bP in CDCl₃



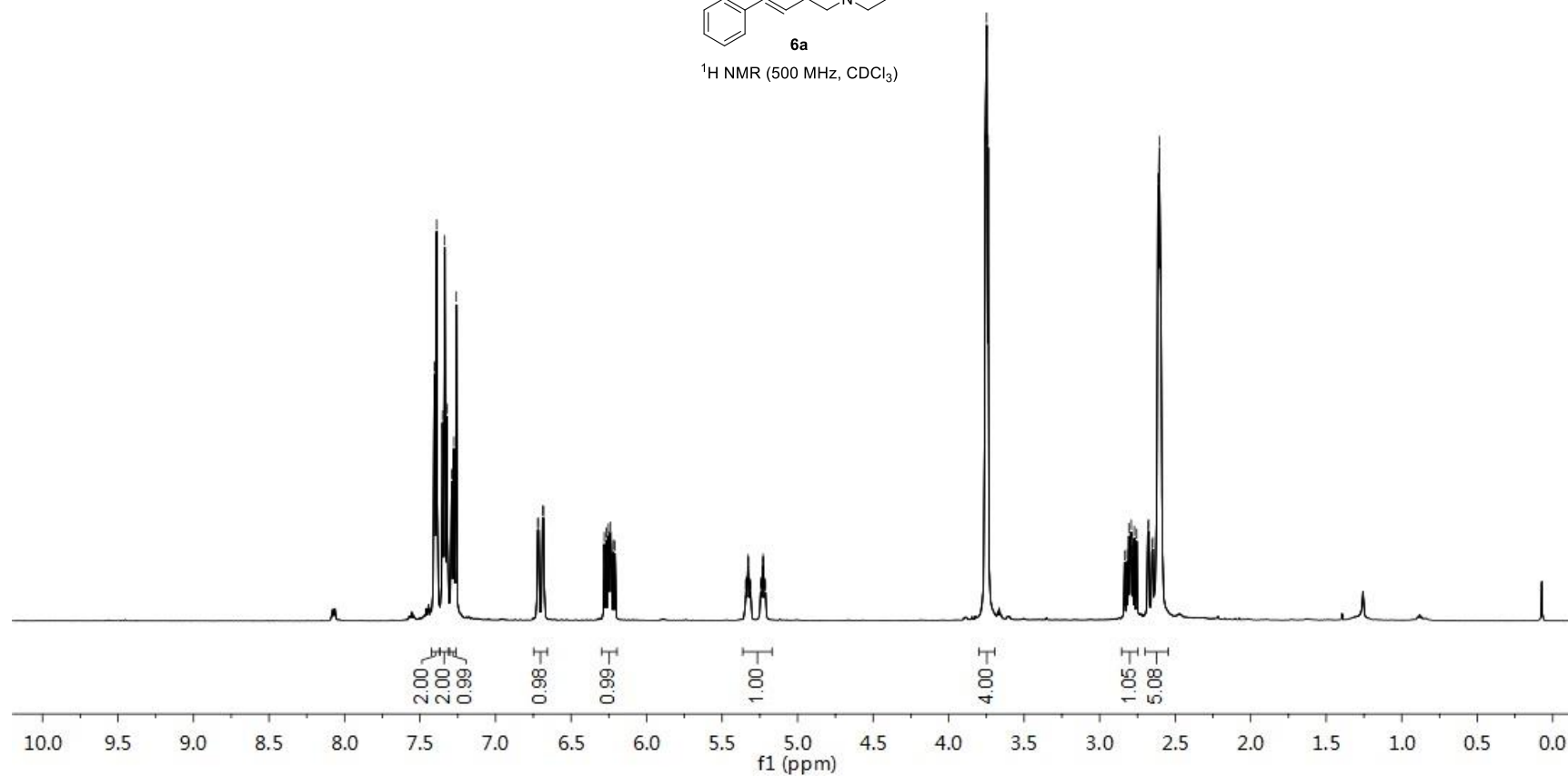
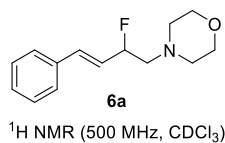
¹⁹F NMR gf-3-98bP in CDCl₃

---180.0226



7.4048
7.3901
7.3511
7.3367
7.3213
7.2904
7.2760
7.2599
6.7142
6.6888
6.6821
6.2673
6.2537
6.2483
6.2407
5.3403
5.3381
5.3306
5.3277
5.3249
5.3174
5.2412
5.2392
5.2315
5.2287
5.2259
3.7388
3.7380
3.7488
3.7394
2.8351
2.8192
2.8071
2.7985
2.7912
2.7826
2.7705
2.7546
2.6817
2.6758
2.6537
2.6478
2.6142
2.6050
2.5961
2.5867

¹H NMR gf-3-135aP in CDCl₃



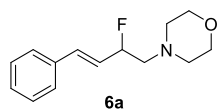
135.96
135.95
133.19
133.10
128.68
128.29
126.72
125.59
125.44

92.41
91.05

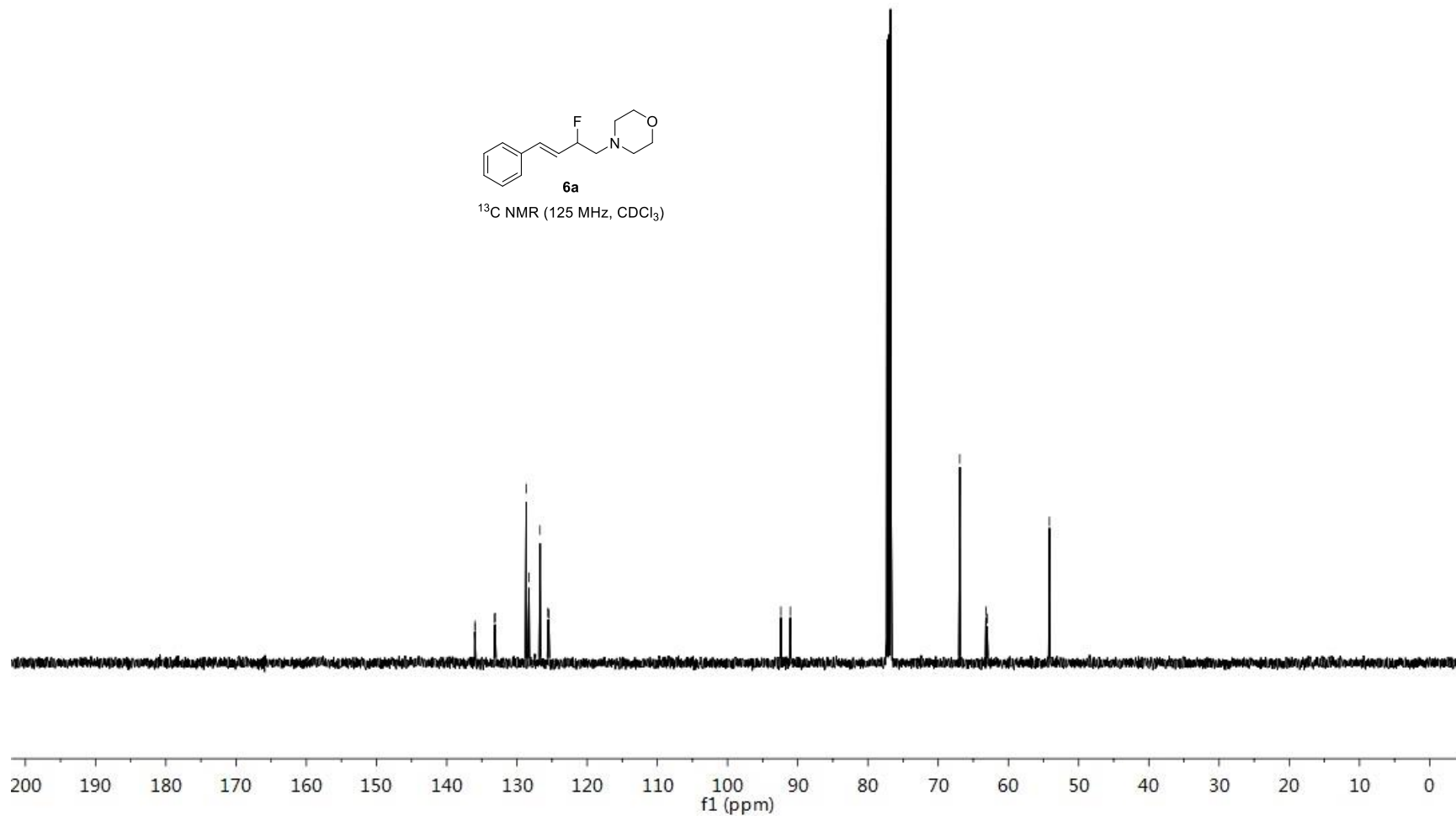
66.93
63.20
63.02

54.16

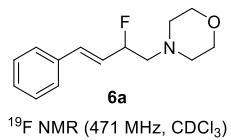
¹³CNMR gf-3-135aP in CDCl₃



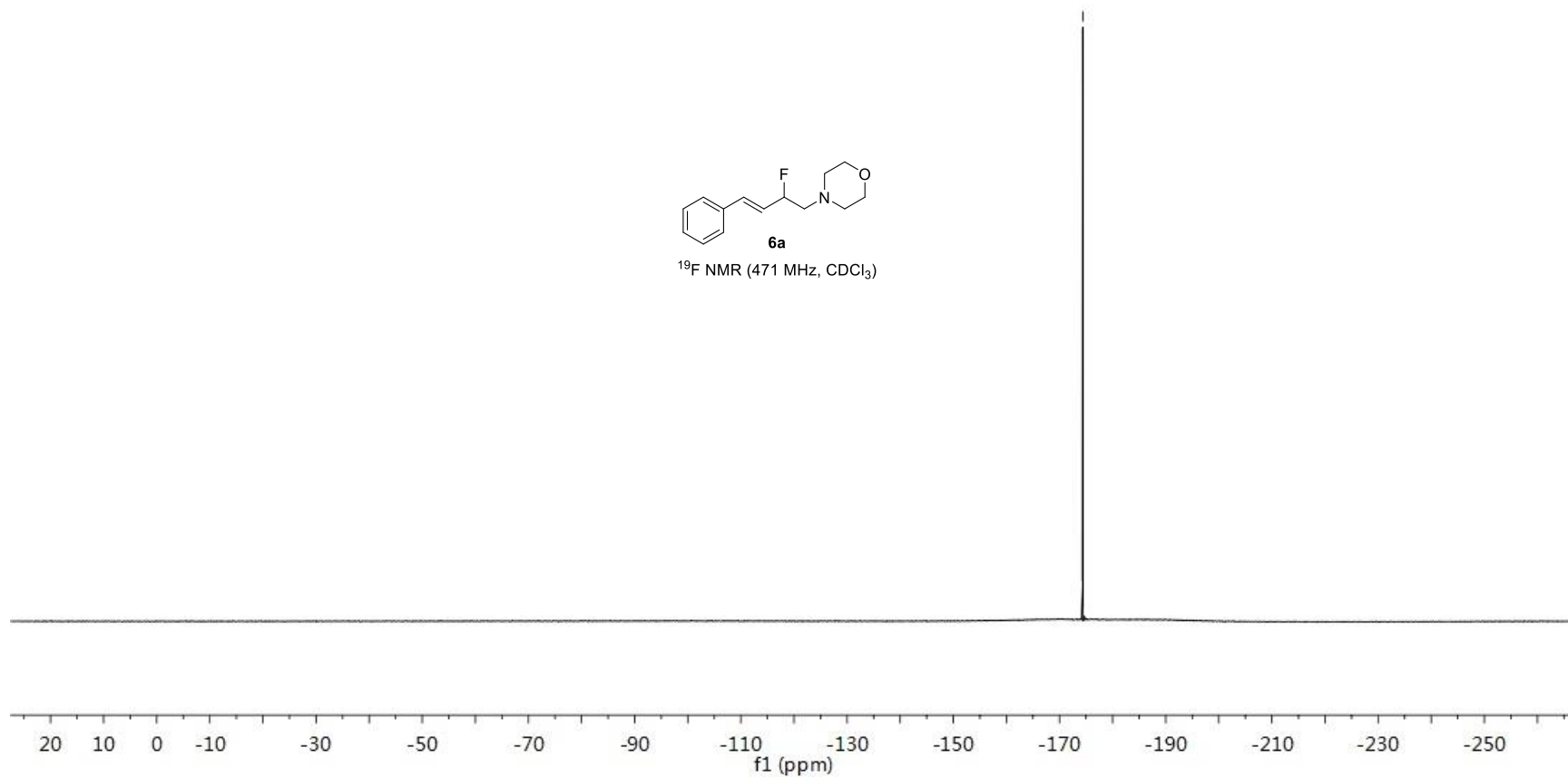
¹³C NMR (125 MHz, CDCl₃)



¹⁹F NMR gf-3-135aP in CDCl₃



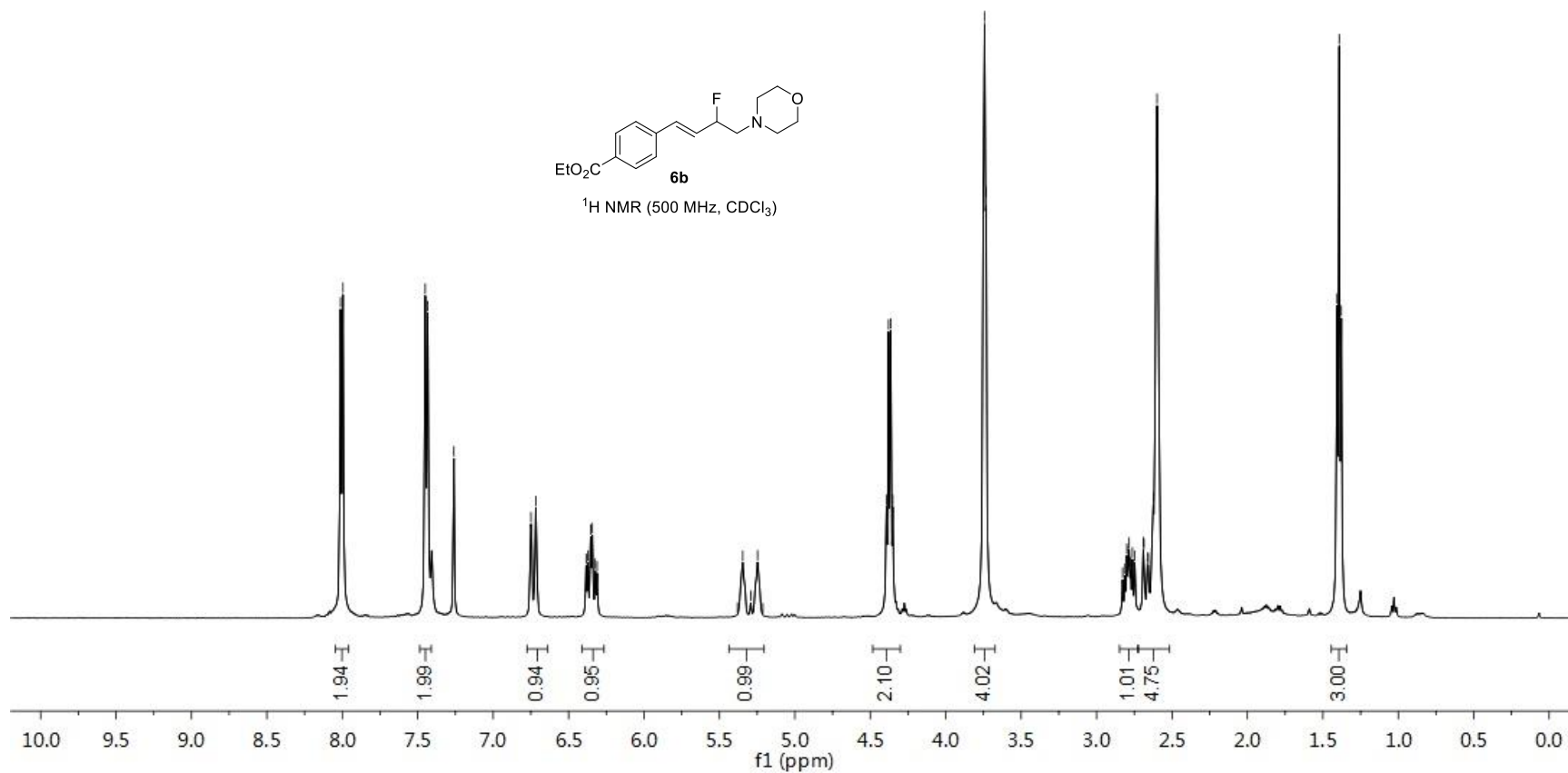
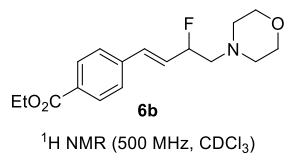
—174.3751

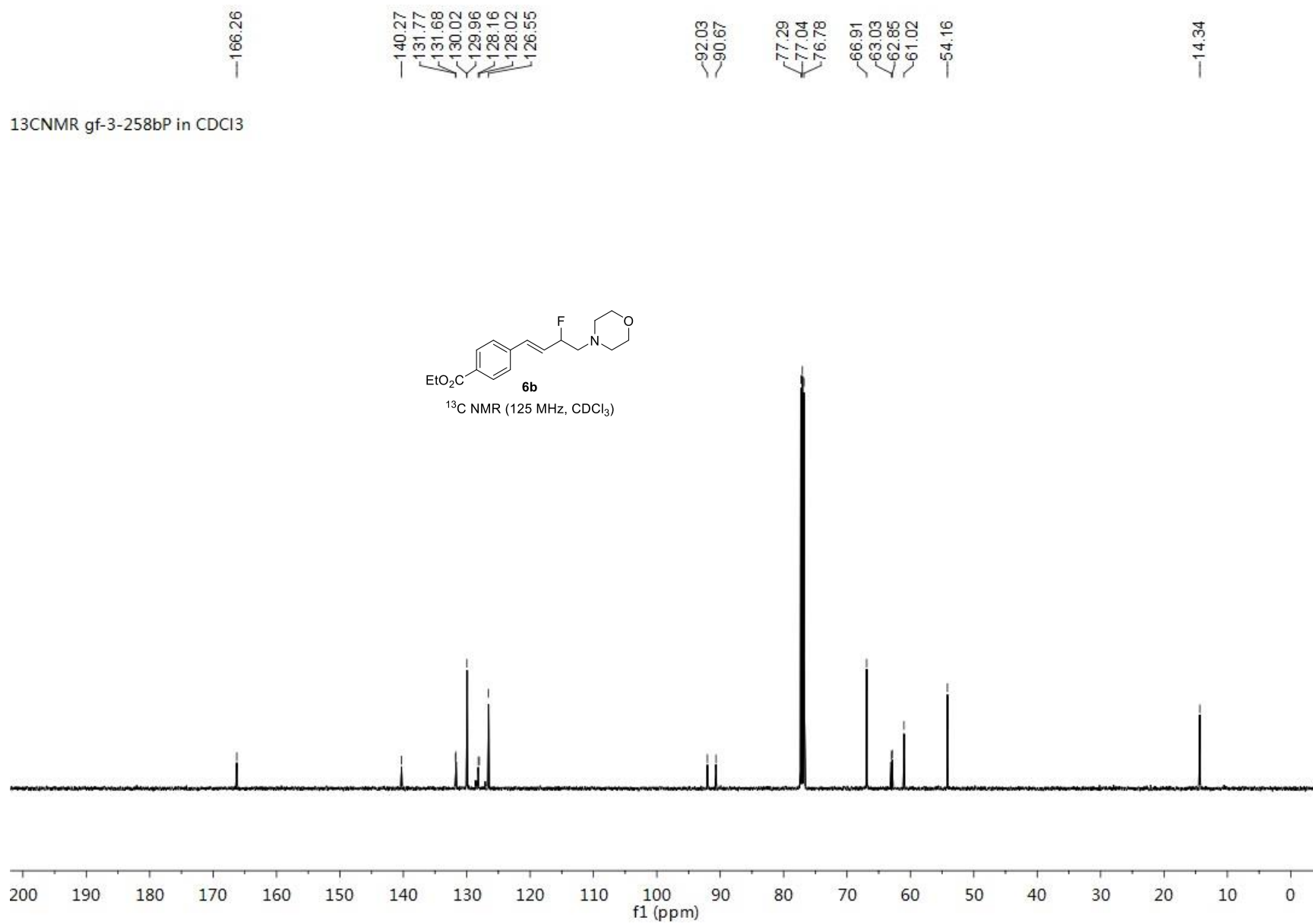


S200

8.0121
7.9959
7.4499
7.4338
7.2602
6.7496
6.7176
6.3836
6.3717
6.3540
6.3421
6.3227
6.3108
5.3809
5.3457
5.2926
5.2473
5.2115
4.3932
4.3791
4.3649
4.3508
3.7507
3.7422
3.7336
2.8118
2.7994
2.7901
2.7841
2.7749
2.7625
2.7467
2.6907
2.6857
2.6626
2.6576
2.6242
2.5991
2.4069
1.3917
1.3776

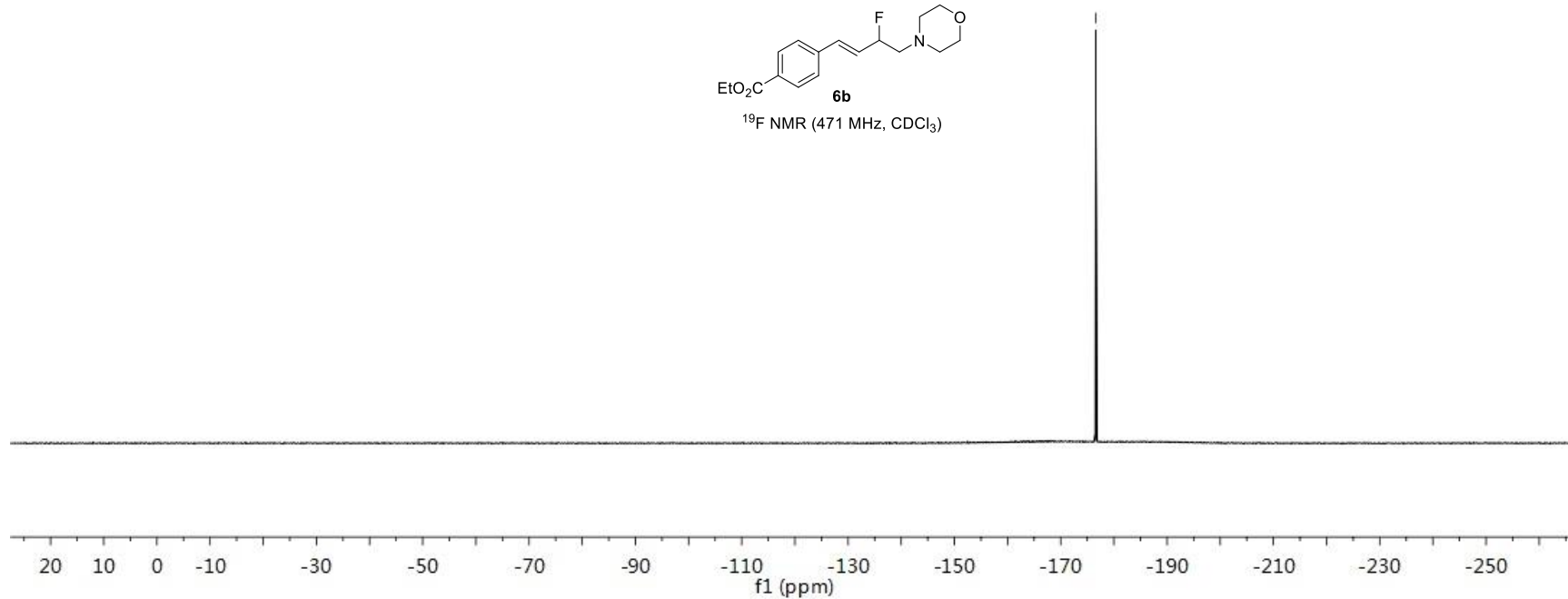
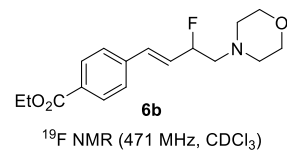
¹H NMR of 6b in CDCl₃





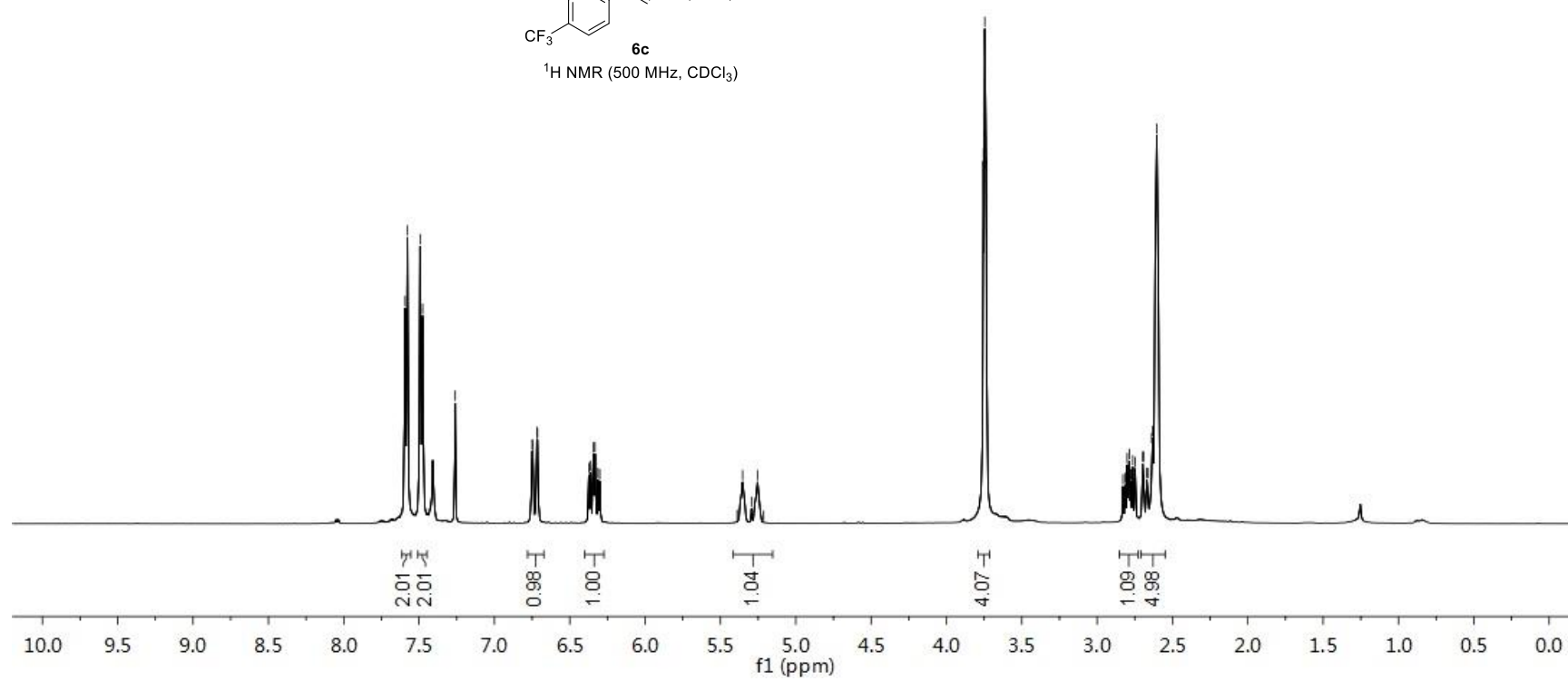
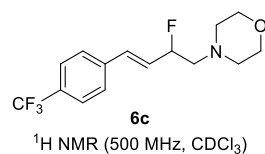
¹⁹F NMR gf-3-258bP in CDCl₃

---176.5648

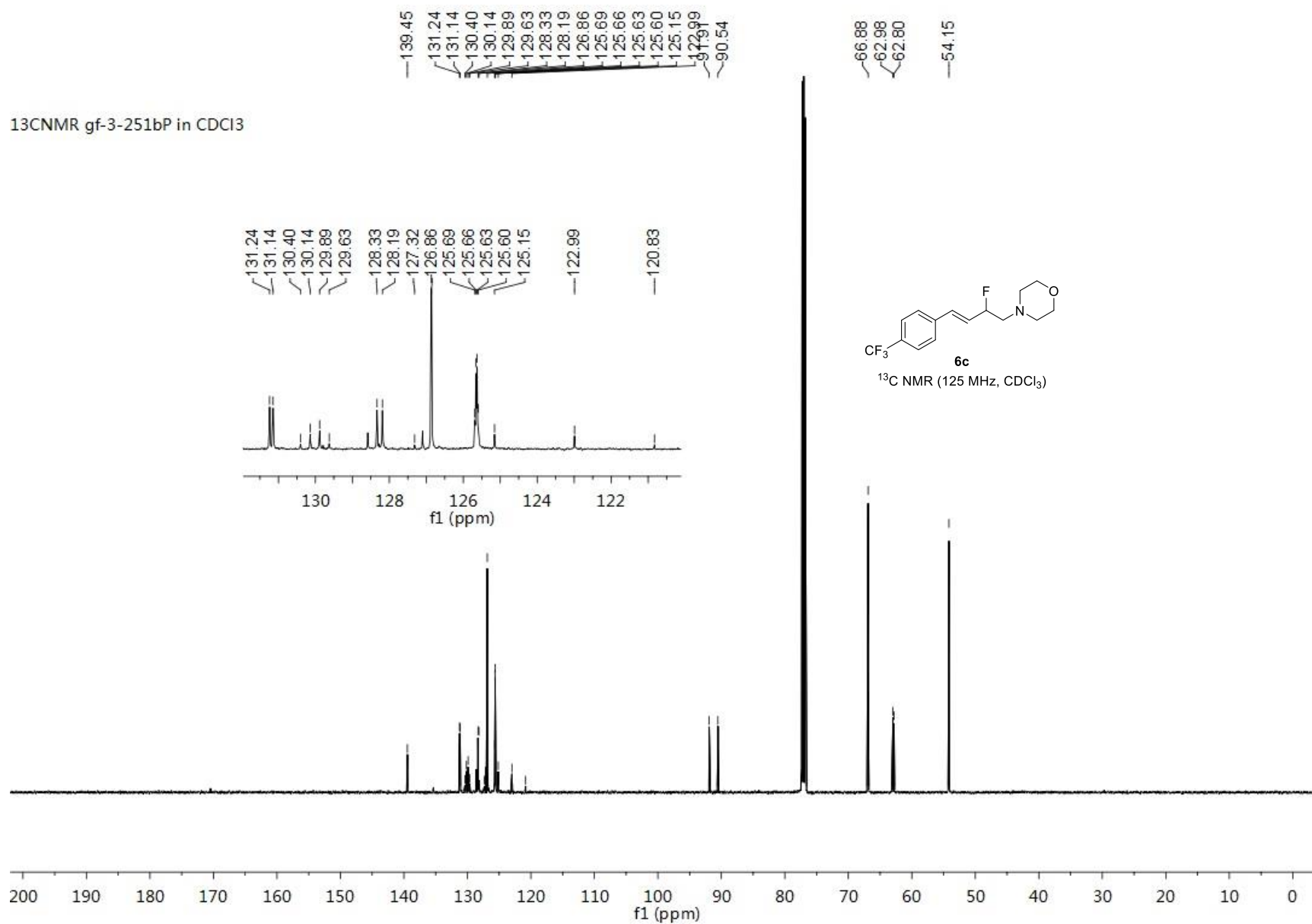


7.5930
7.5767
7.4930
7.4768
7.2602
6.7498
6.7177
6.7139
6.3733
6.3615
6.3434
6.3316
5.3600
5.3527
5.2935
5.2543
5.2132
3.7544
3.7453
3.7360
2.8287
2.8131
2.8007
2.7912
2.7852
2.7757
2.7633
2.7477
2.6980
2.6920
2.6699
2.6639
2.6377
2.6316
2.6039

¹H NMR of **6c** in CDCl₃



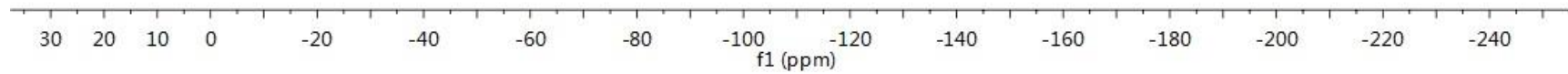
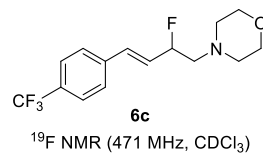
¹³CNMR gf-3-251bP in CDCl₃



¹⁹F NMR gf-3-251bP in CDCl₃

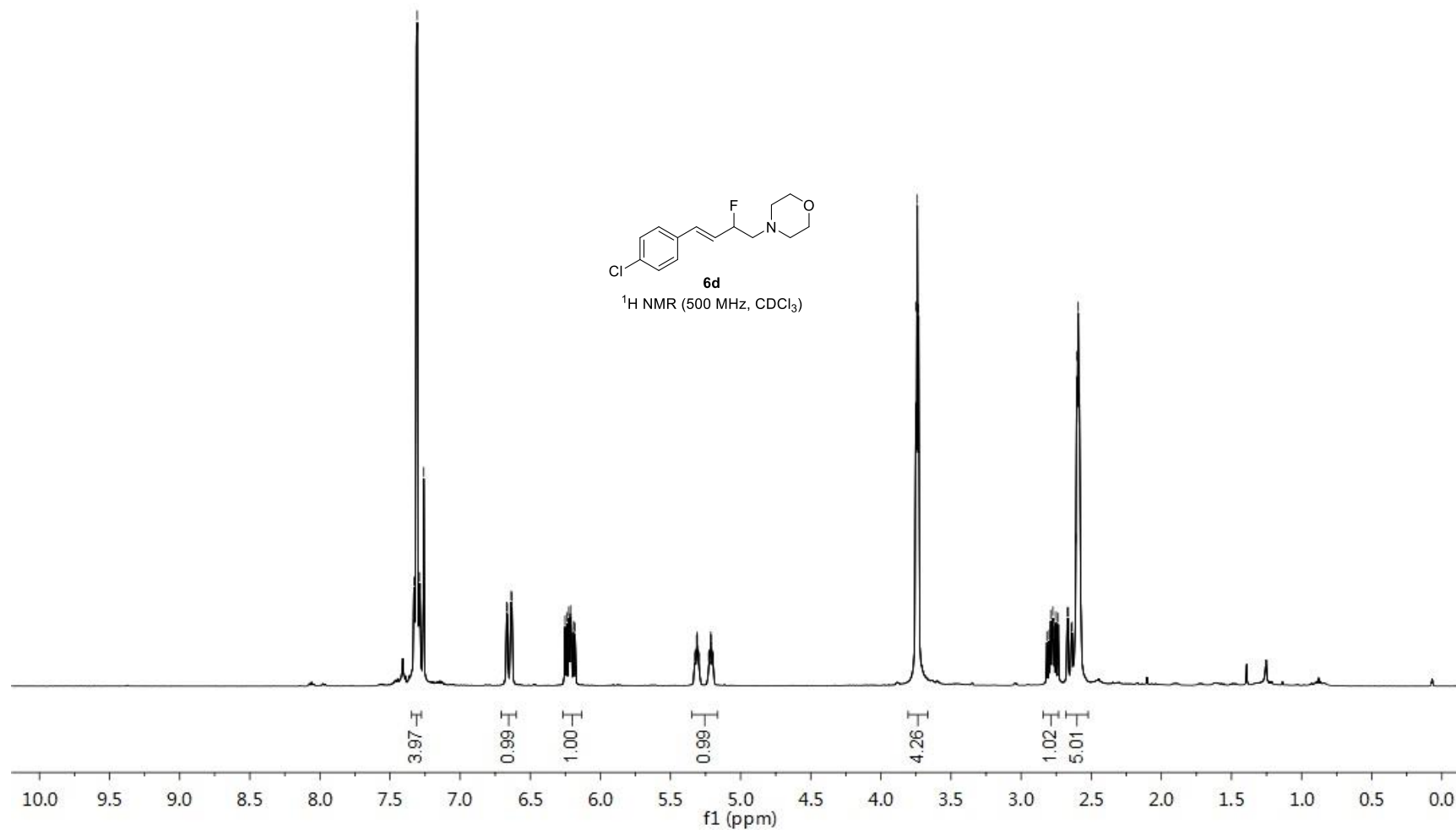
62.6069

176.9829



7.3296
7.3246
7.3119
7.3059
7.2933
7.2882
7.2598
6.6688
6.6368
6.6315
6.2401
6.2252
6.2126
6.2082
5.3233
5.3210
5.3139
5.3111
5.3084
5.3012
5.2246
5.2223
5.2150
5.2123
5.2095
3.7468
3.7407
3.7313
2.8166
2.8009
2.7886
2.7797
2.7729
2.7640
2.7517
2.7359
2.6704
2.6644
2.6424
2.6364
2.6090
2.6024
2.5928
2.5837

¹H NMR of gf-3-147aP in CDCl₃



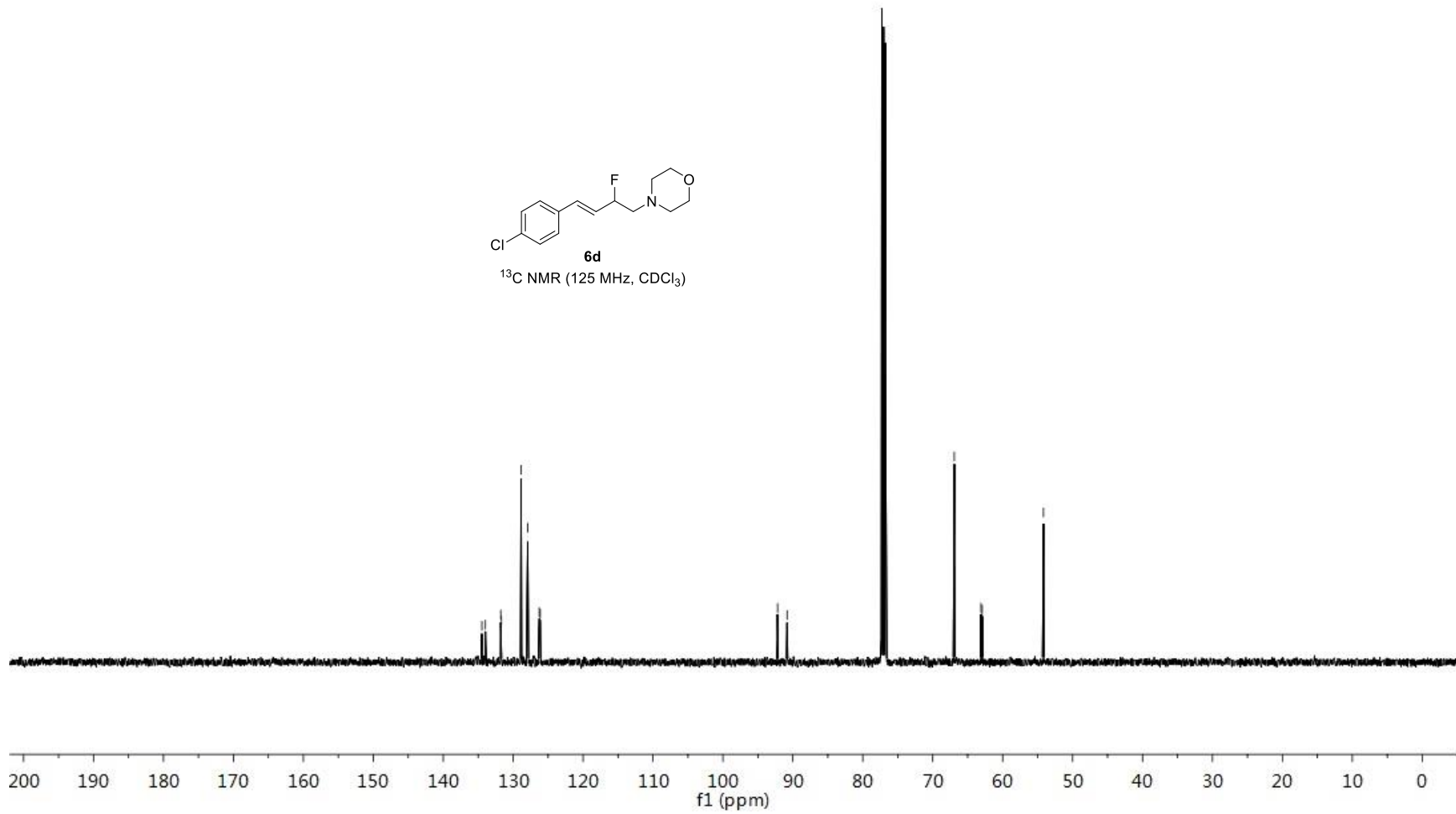
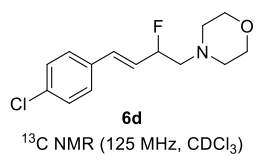
134.47
133.97
131.77
131.68
128.86
127.91
126.27
126.13

92.19
90.83

66.92
63.10
62.92

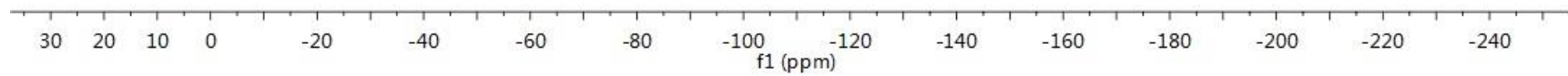
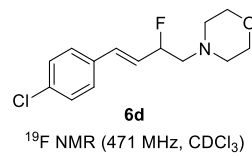
54.17

¹³CNMR gf-3-147aP in CDCl₃



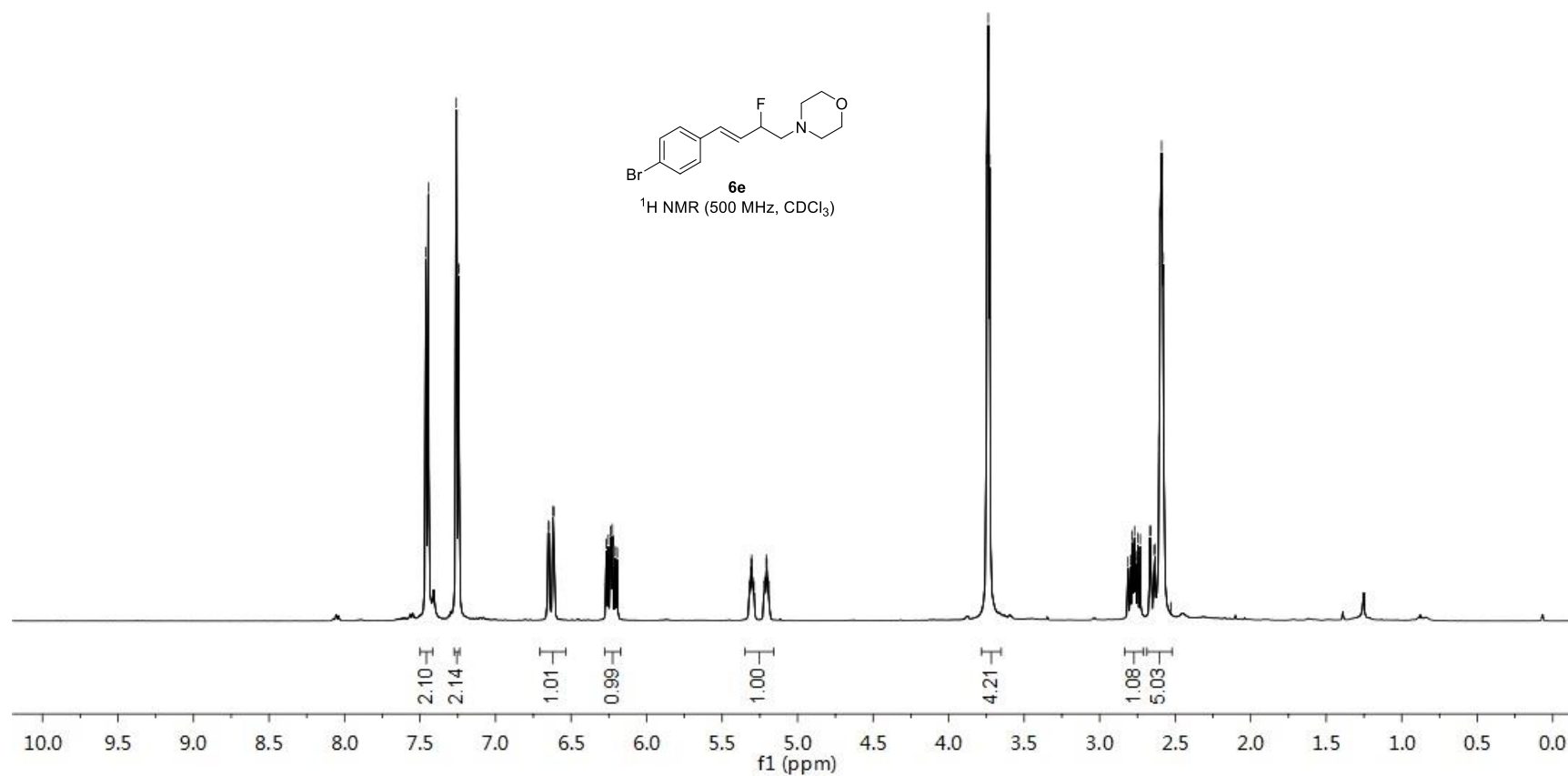
¹⁹F NMR gf-3-147aP in CDCl₃

—175.2228



7.4617
7.4449
7.2600
7.2438
6.6516
6.6458
6.6196
6.6139
6.2539
6.2387
6.2345
6.2262
5.3203
5.3144
5.3072
5.3045
5.3018
5.2945
5.2179
5.2156
5.2084
5.2057
5.2030
5.1971
3.7380
3.7286
2.8127
2.7969
2.7847
2.7757
2.7689
2.7600
2.7477
2.7319
2.6674
2.6614
2.6394
2.6334
2.6061
2.5995
2.5899
2.5807
2.5286

¹H NMR of 6e in CDCl₃



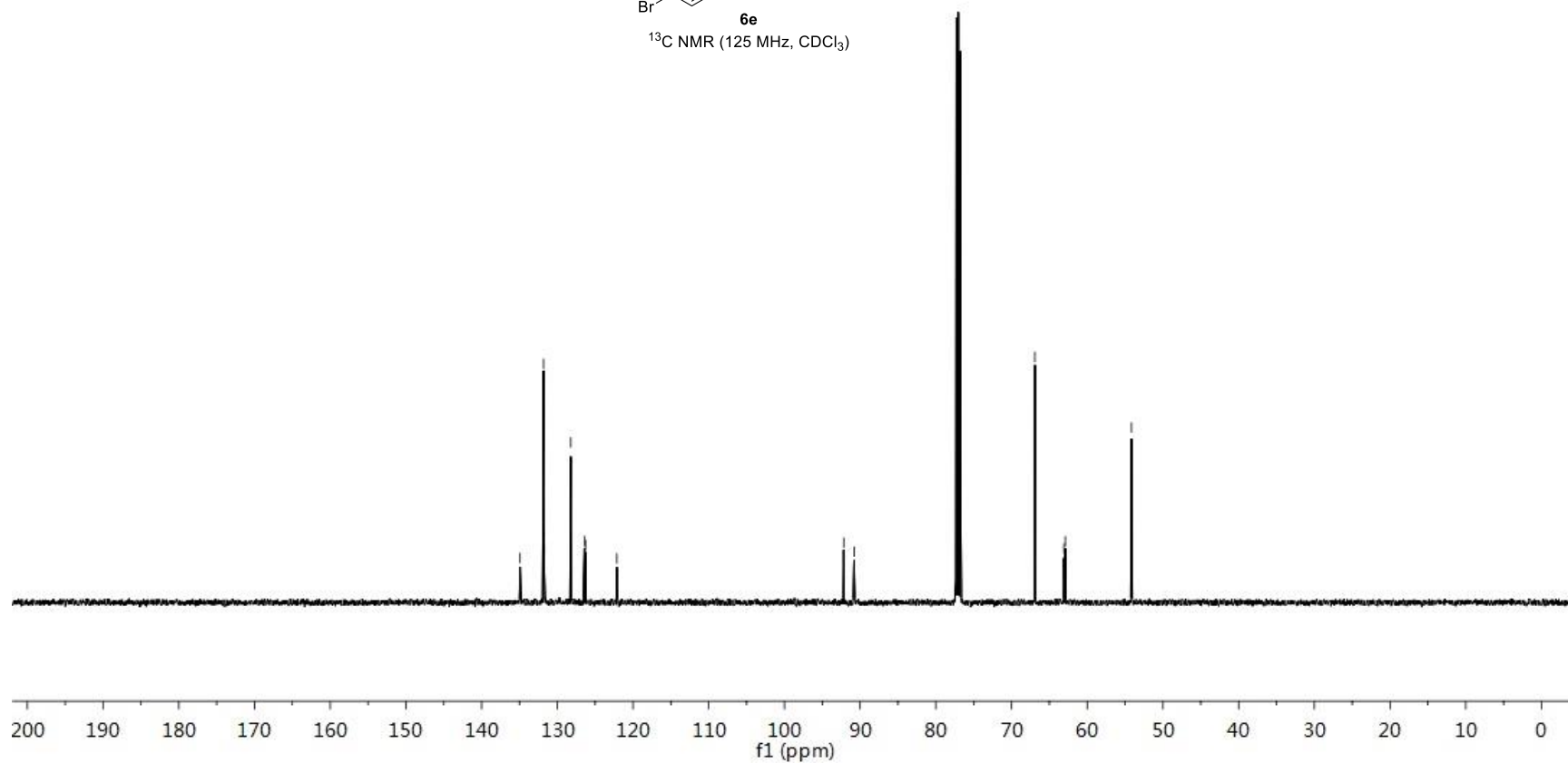
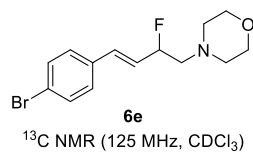
134.92
131.81
131.79
131.69
128.22
126.41
126.27
122.13

92.17
90.81

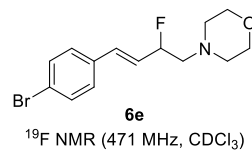
66.91
63.07
62.89

54.16

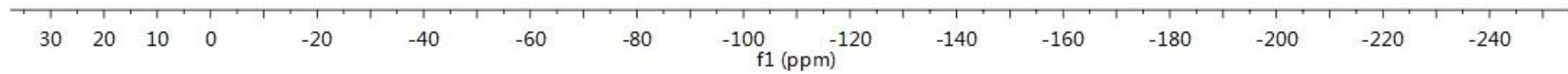
¹³CNMR gf-3-147bP in CDCl₃



¹⁹F NMR gf-3-147bP in CDCl₃



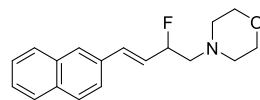
---175.4165



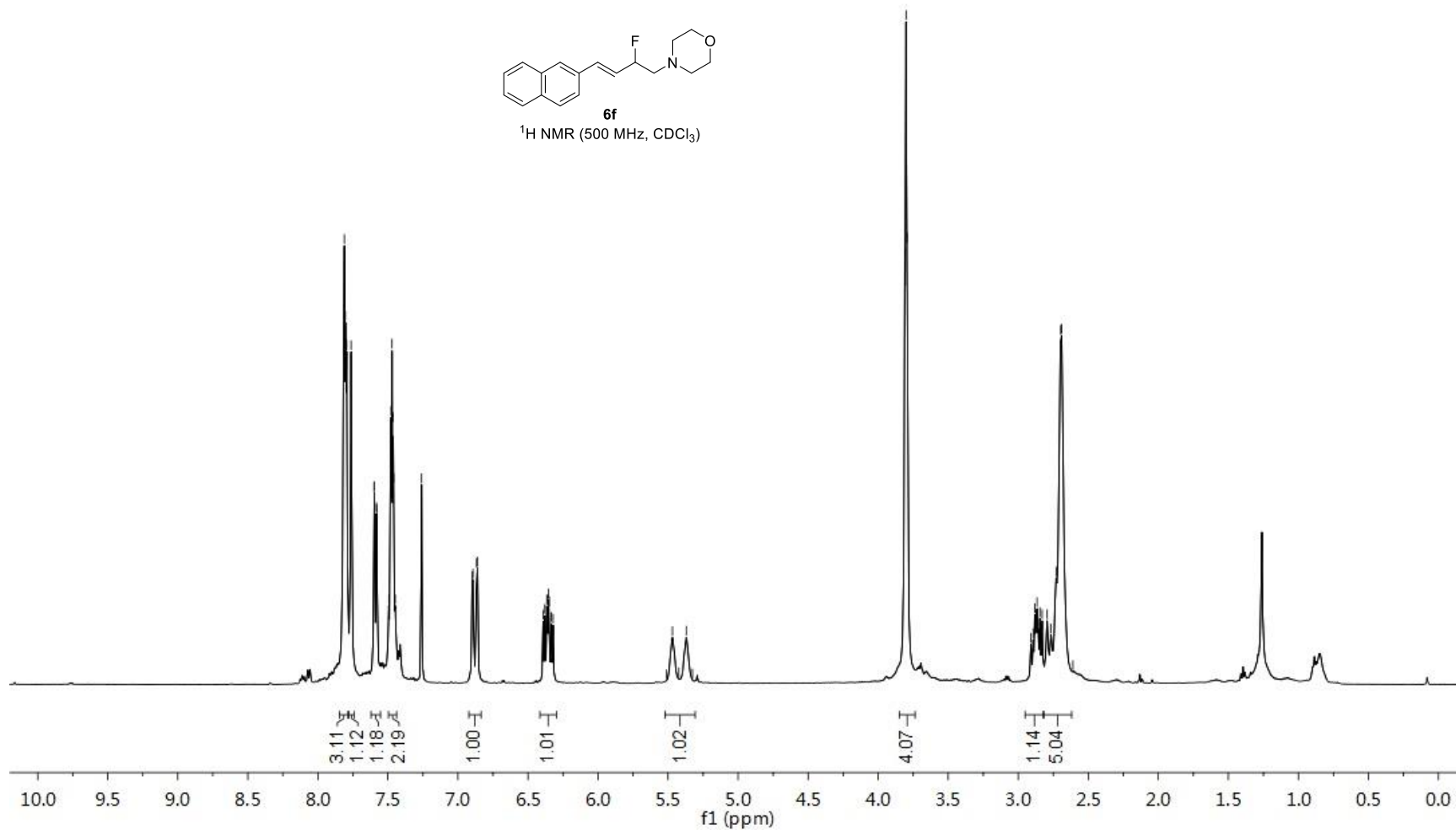
7.8168
7.8106
7.8035
7.7942
7.7627
7.5992
7.5969
7.5820
7.5799
7.4932
7.4825
7.4775
7.4705
7.4632
7.4586
7.4479
7.2599
6.8925
6.8667
6.8606
6.3795
6.3653
6.3606
6.3525
6.3478
6.3408
5.4685
5.4231
5.3699
5.3253

3.8101
3.8014
3.7926
2.9097
2.8936
2.8816
2.8735
2.8658
2.8579
2.8456
2.8295
2.7937
2.7654
2.7299
2.6975
2.6908
2.6096

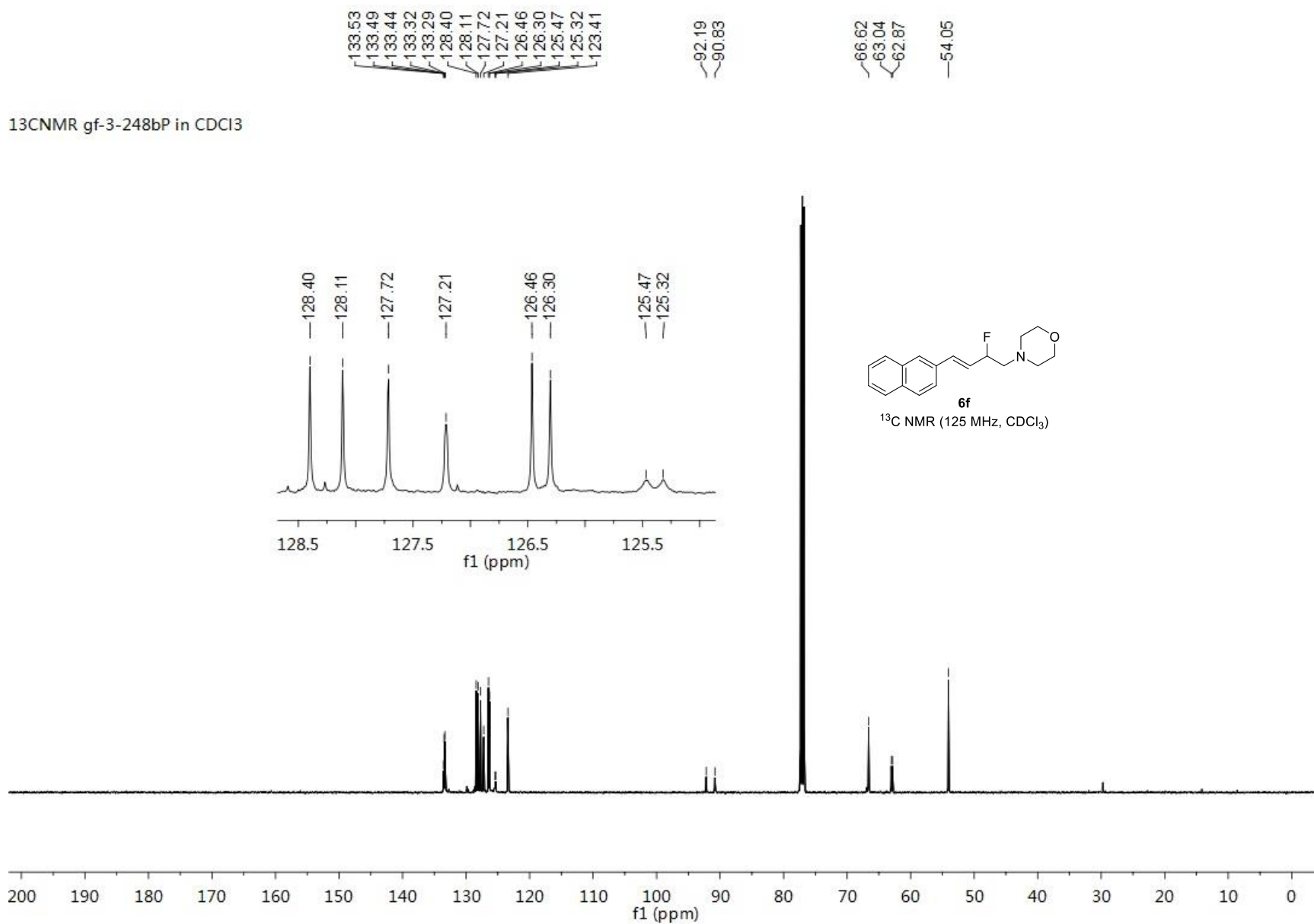
¹H NMR of gf-3-248bP in CDCl₃



6f
¹H NMR (500 MHz, CDCl₃)

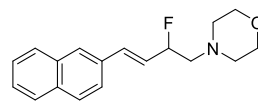


¹³CNMR gf-3-248bP in CDCl₃

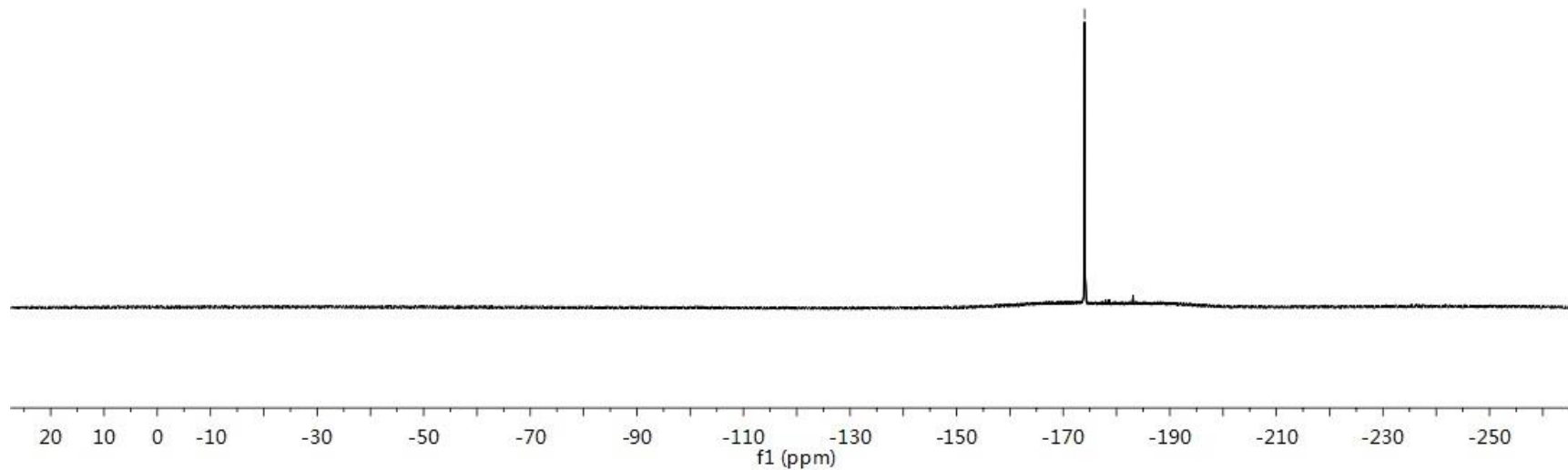


¹⁹F NMR gf-3-248bP in CDCl₃

—174.0227

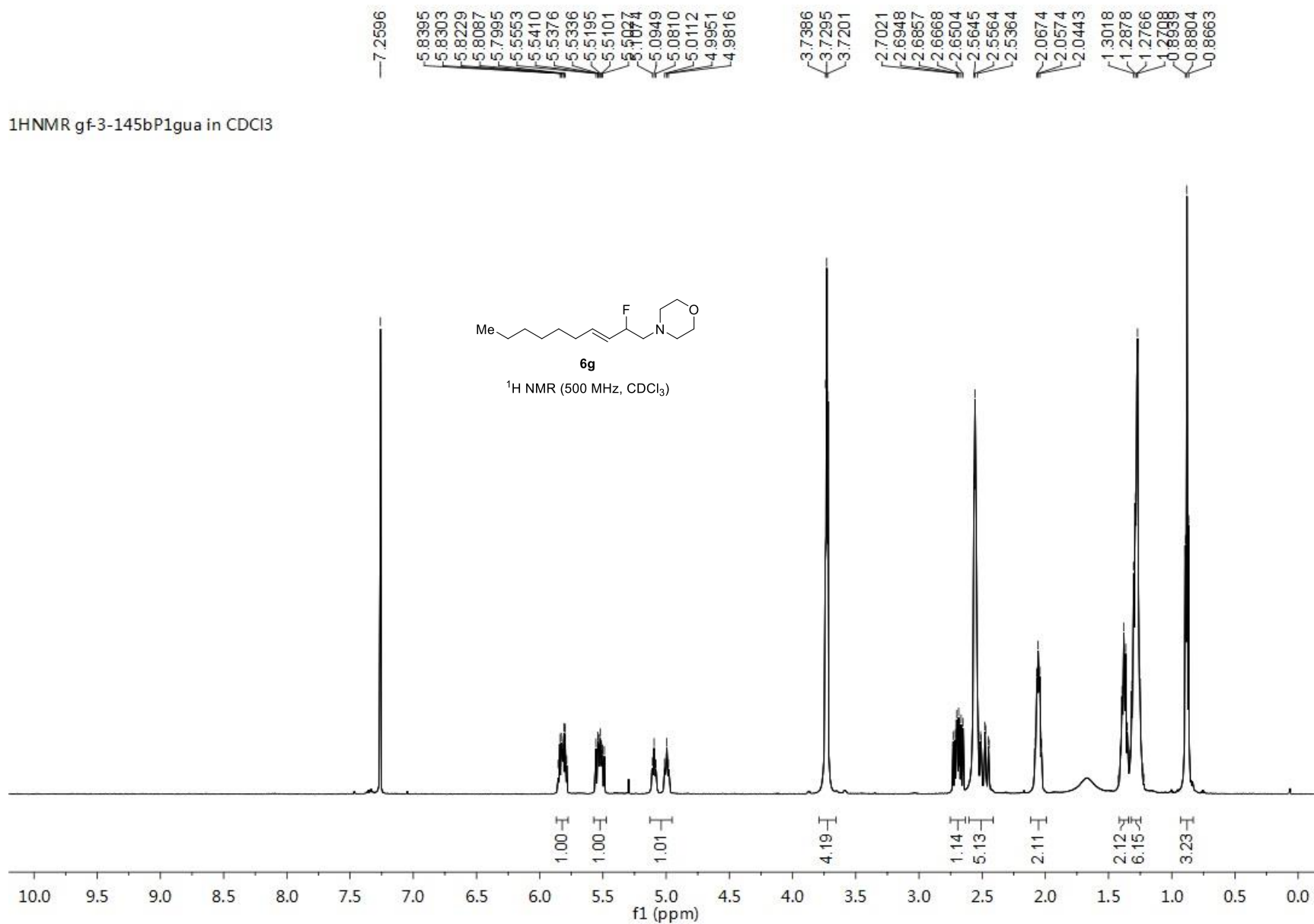


¹⁹F NMR (471 MHz, CDCl₃)

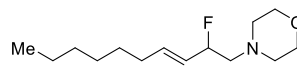


S215

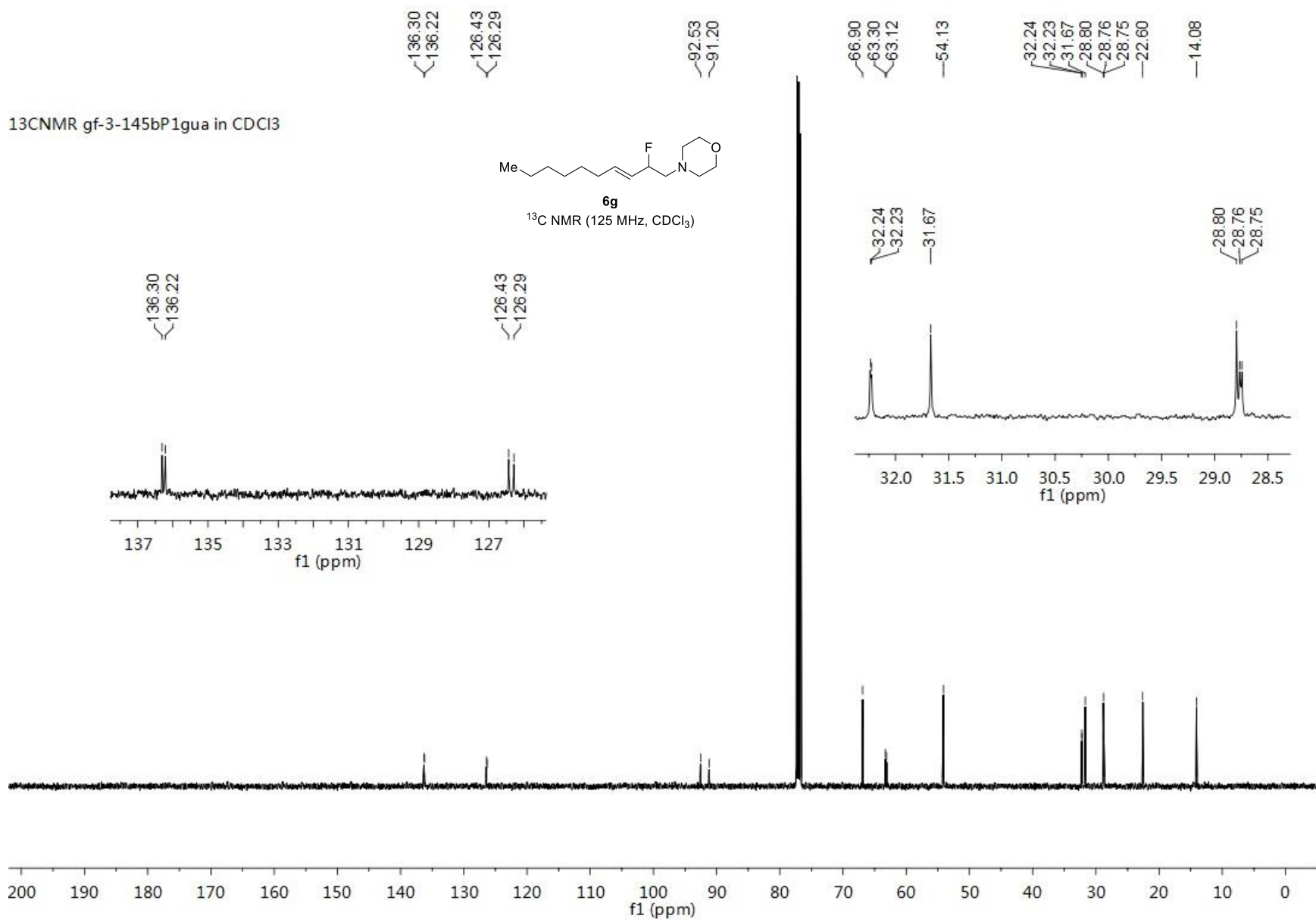
¹H NMR of 6g in CDCl₃



¹³C NMR gf-3-145bP1gua in CDCl₃

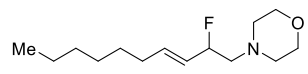


6g
¹³C NMR (125 MHz, CDCl₃)



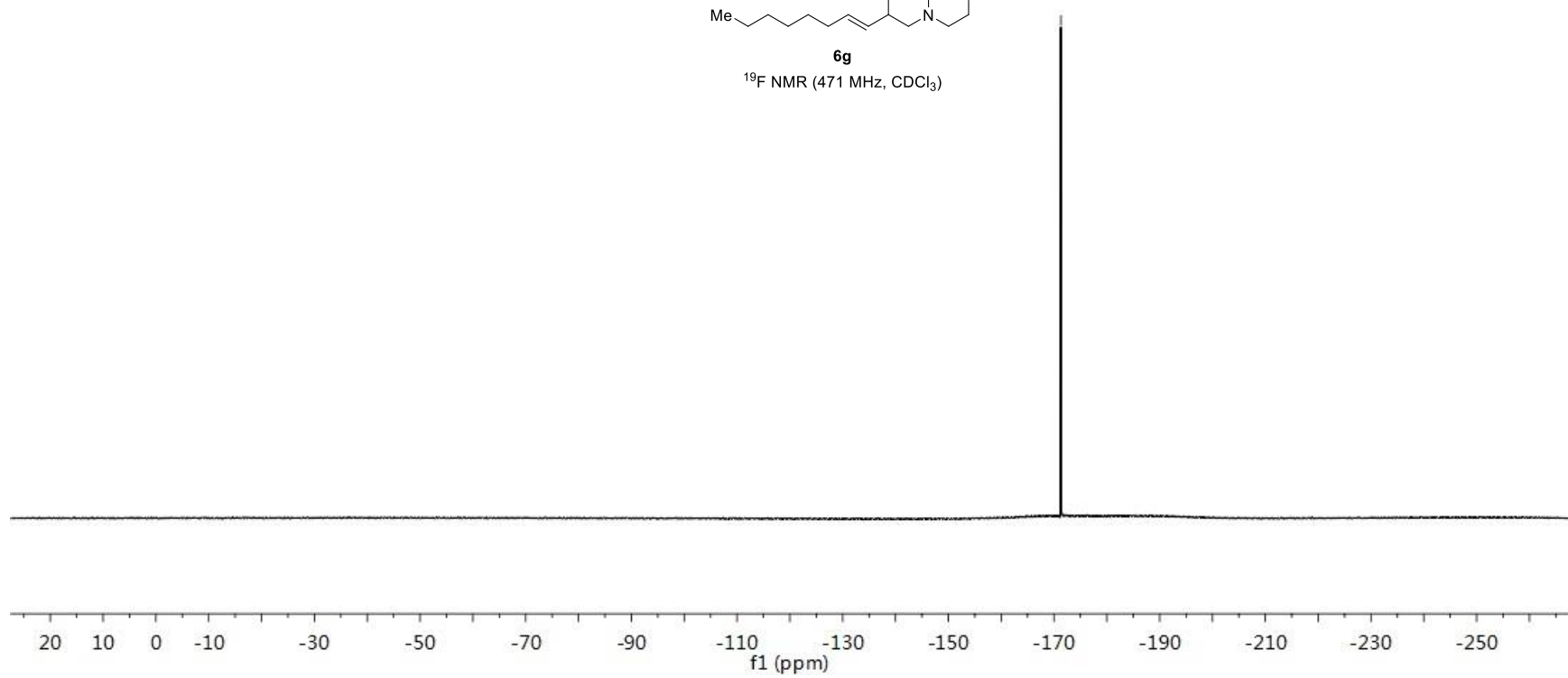
¹⁹F NMR gf-3-145bPgua in CDCl₃

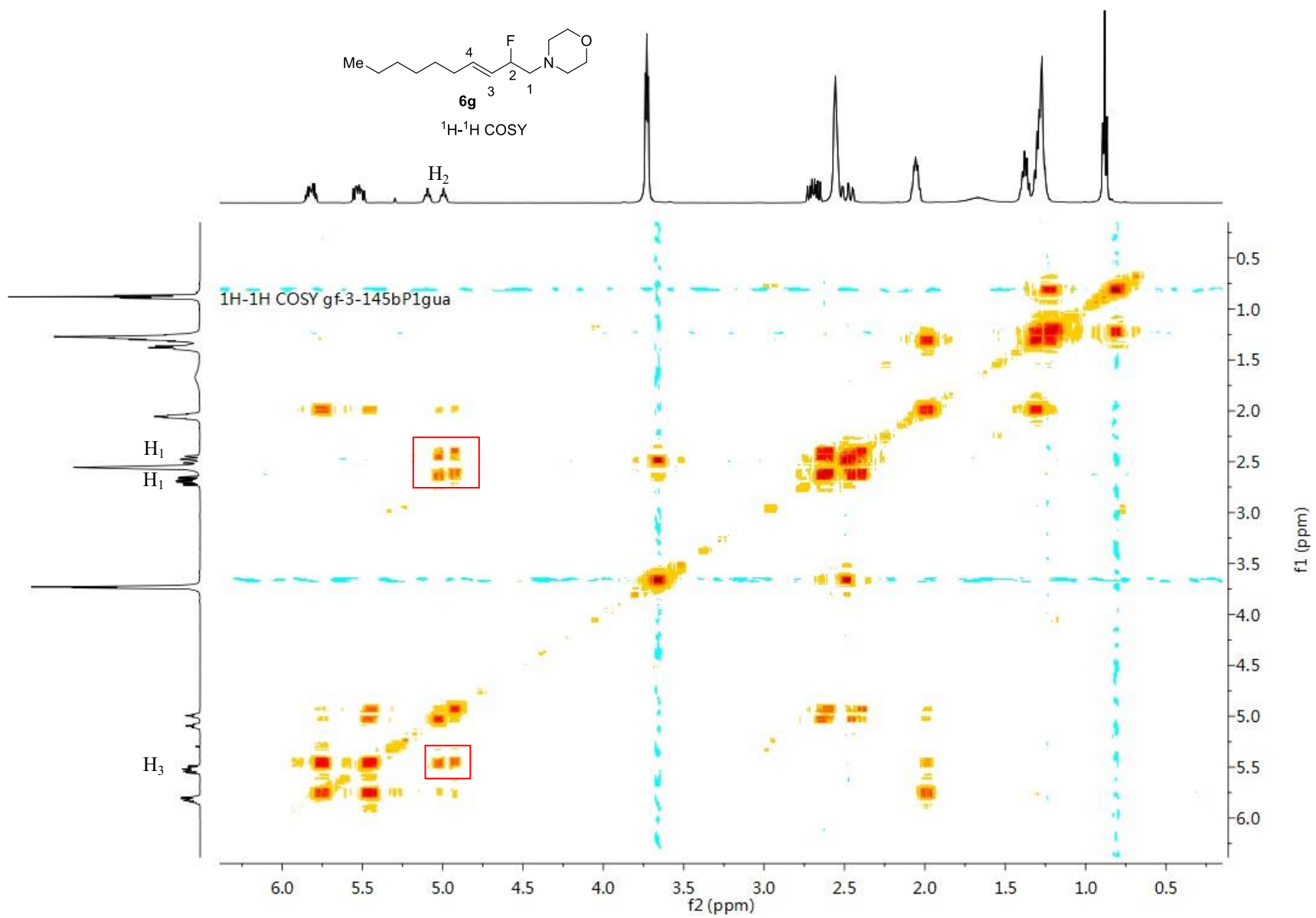
-171.3094



6g

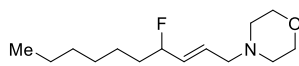
¹⁹F NMR (471 MHz, CDCl₃)



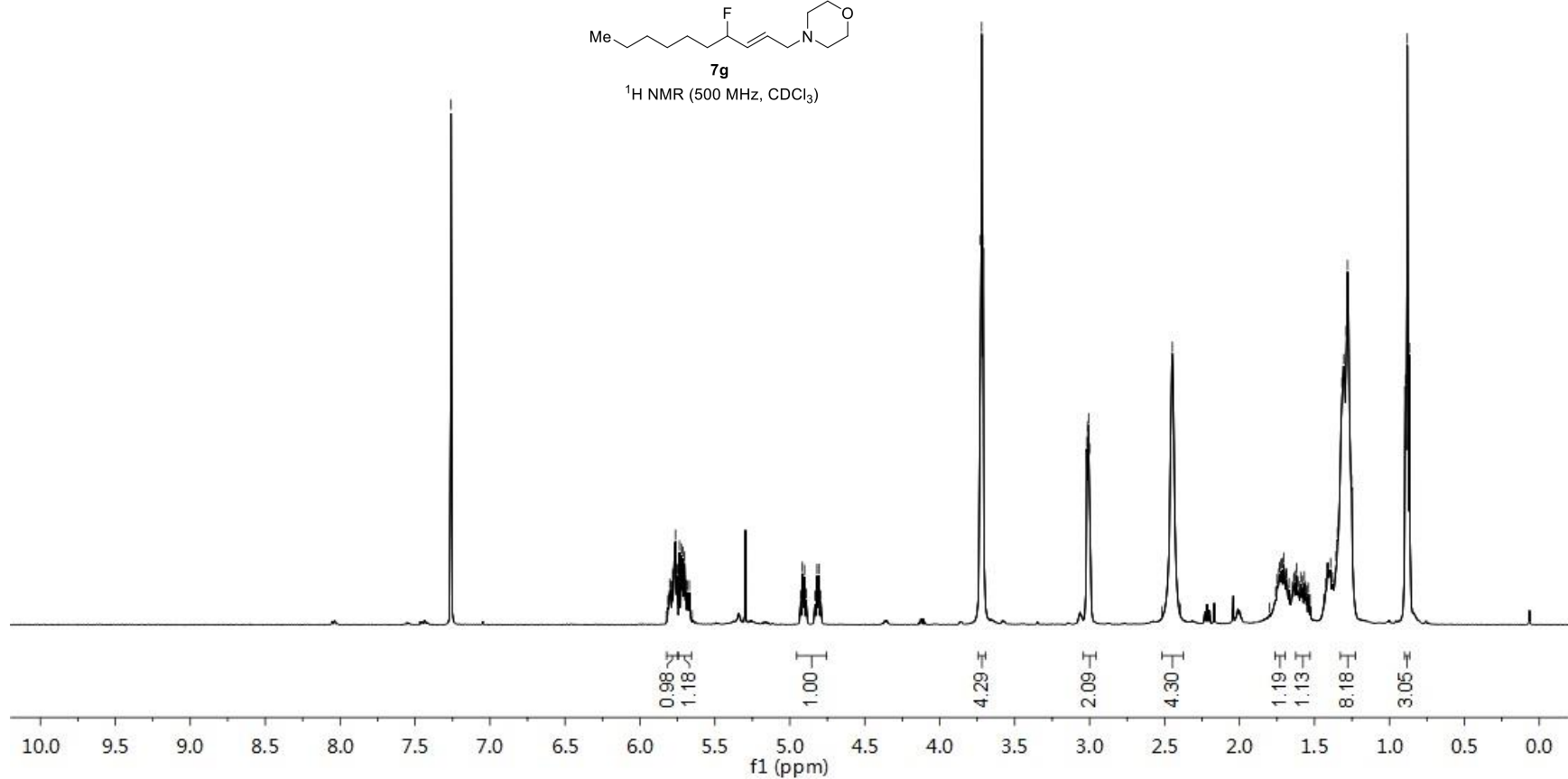


7.2602
5.8009
5.7943
5.7877
5.7824
5.7752
5.7697
5.7631
5.7573
5.7513
5.7365
5.7241
5.7142
5.7016
5.6930
5.6826
5.6699
4.9285
4.9164
4.9027
4.8908
4.8315
4.8188
4.8063
4.7933
3.7296
3.7204
3.7112
3.0187
3.0117
3.0073
2.9998
2.4478
1.7579
1.7491
1.7386
1.7328
1.7236
1.7153
1.7048
1.6895
1.6860
1.6707
1.6493
1.6395
1.6305
1.6202
1.6111
1.6027
1.5920
1.5786
1.5679
1.5588
1.5506
1.5400
1.4385
1.4251
1.3924
1.3764
1.3565
1.3458
1.3166
1.3056
1.2891
1.2799
1.2745
1.2518
0.8938
0.8807
0.8666

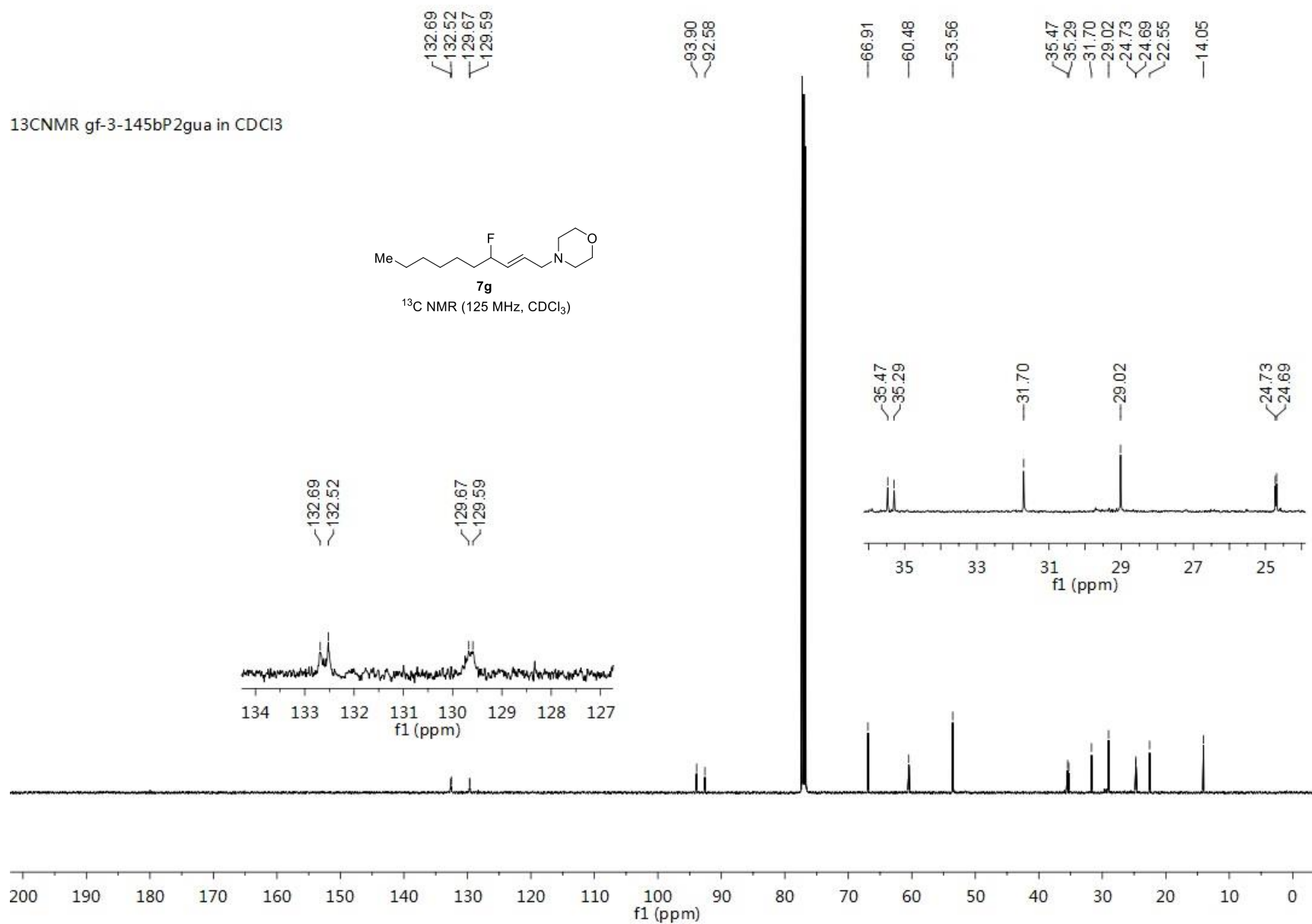
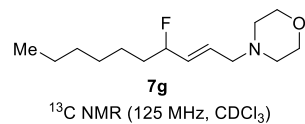
¹H NMR of 7g in CDCl₃



7g
¹H NMR (500 MHz, CDCl₃)

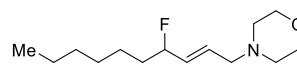


¹³CNMR gf-3-145bP2gua in CDCl₃



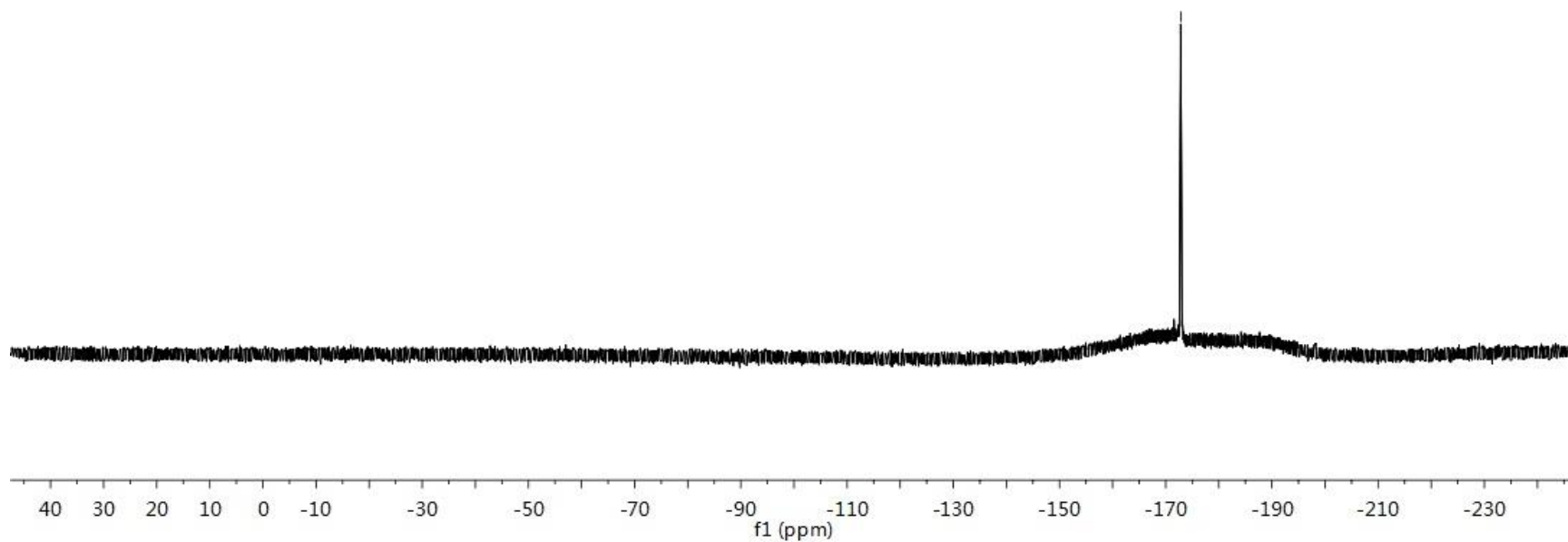
¹⁹F NMR gf-3-145bP2gua in CDCl₃

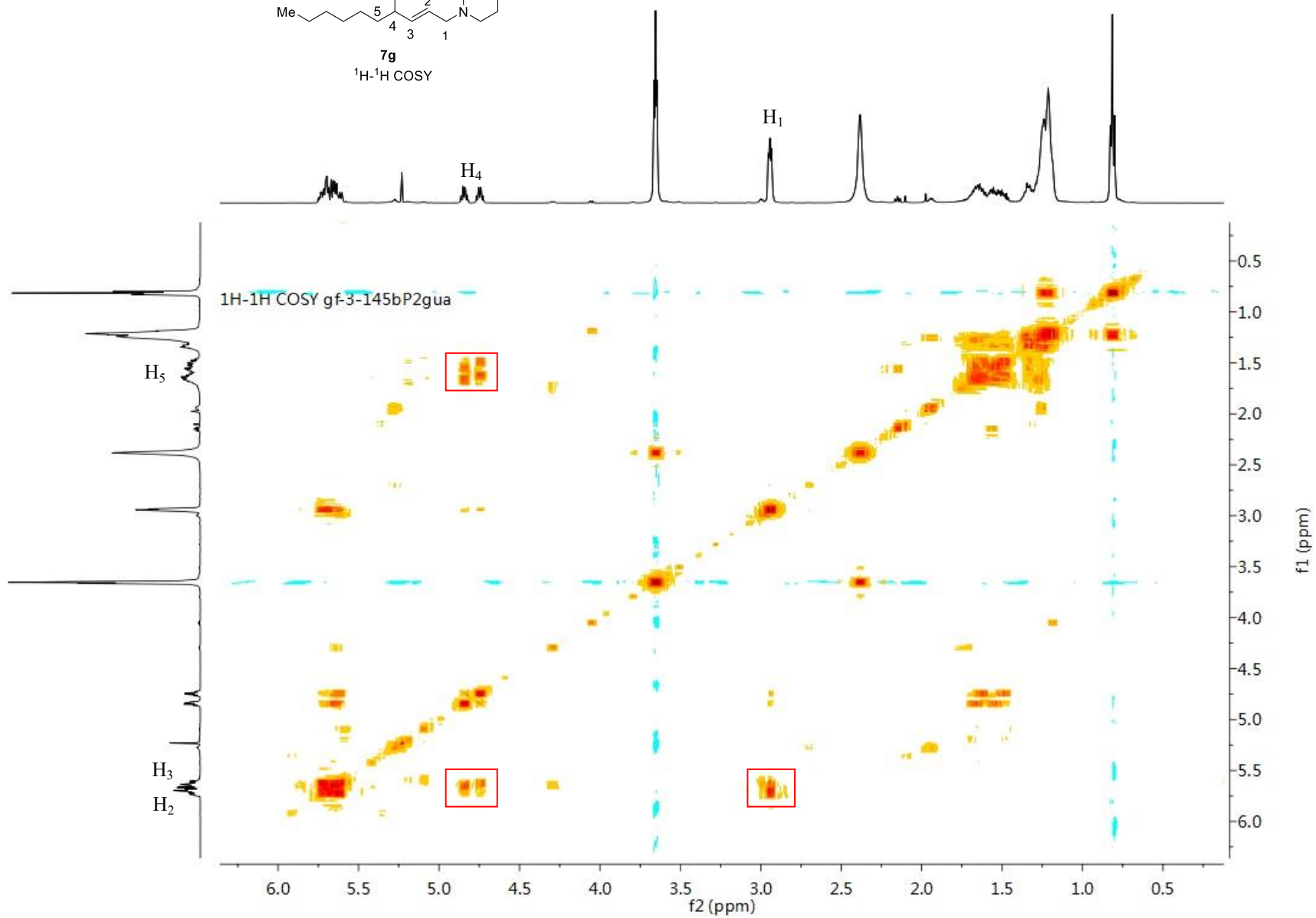
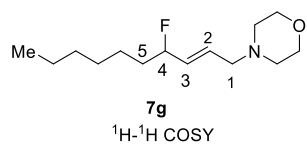
---172.8292



7g

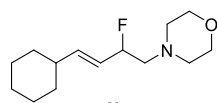
¹⁹F NMR (471 MHz, CDCl₃)



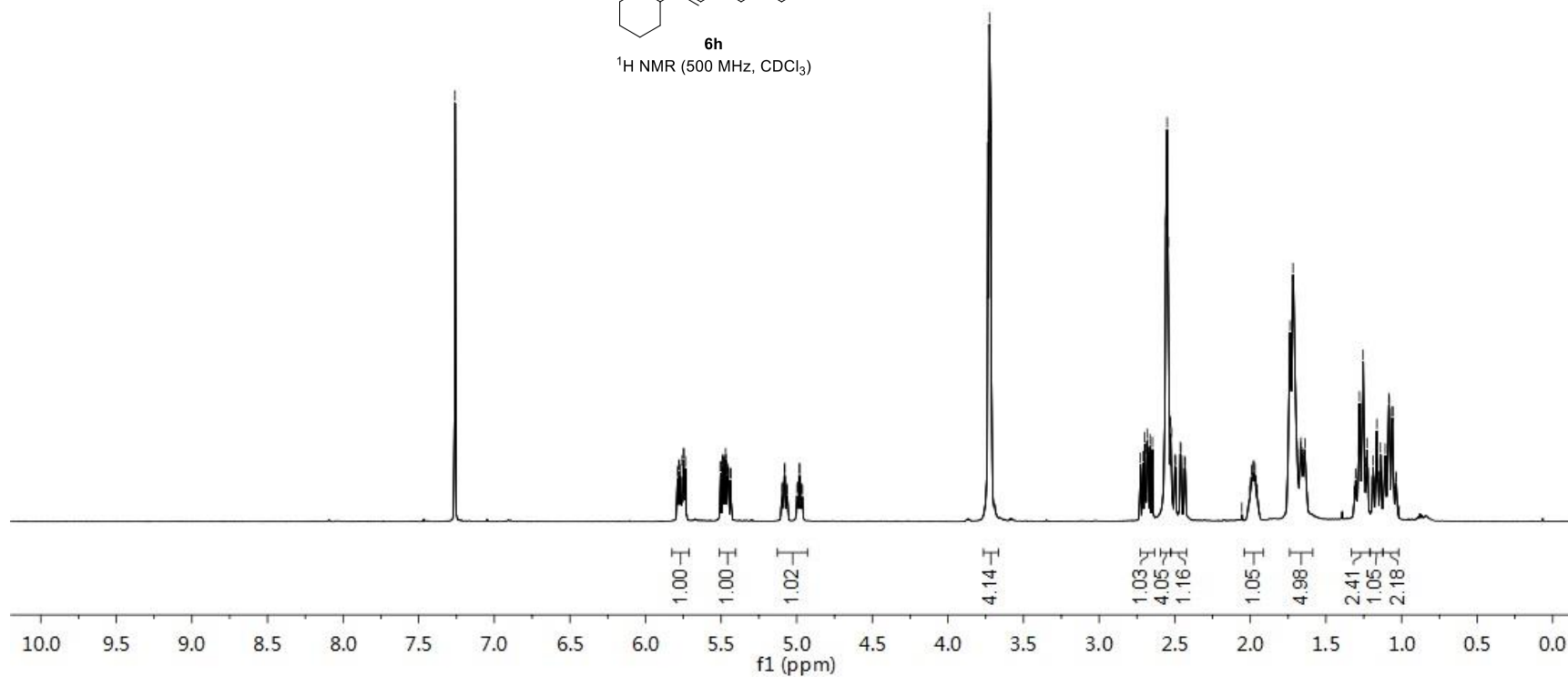


7.2599
 5.7795
 5.7764
 5.7573
 5.7483
 5.7452
 5.7362
 5.5057
 5.5032
 5.4916
 5.4890
 5.4838
 5.4812
 5.4717
 5.4698
 5.4671
 5.0778
 4.9802
 3.7345
 3.7252
 3.7159
 2.7270
 2.7105
 2.6991
 2.6919
 2.6826
 2.6753
 2.6639
 2.6474
 2.5593
 2.5506
 2.5421
 2.5273
 2.5218
 2.4993
 2.4938
 2.4635
 2.4581
 2.4355
 2.4302
 1.9842
 1.9761
 1.9691
 1.9691
 1.7374
 1.7314
 1.7180
 1.6701
 1.6669
 1.6639
 1.6607
 1.6572
 1.6453
 1.6388
 1.2859
 1.2794
 1.2729
 1.2596
 1.2538
 1.2352
 1.2286
 1.1881
 1.1696
 1.1635
 1.1571
 1.1386
 1.1074
 1.0847
 1.0808
 1.0603
 1.0579

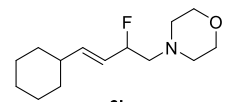
¹H NMR of 6h in CDCl₃



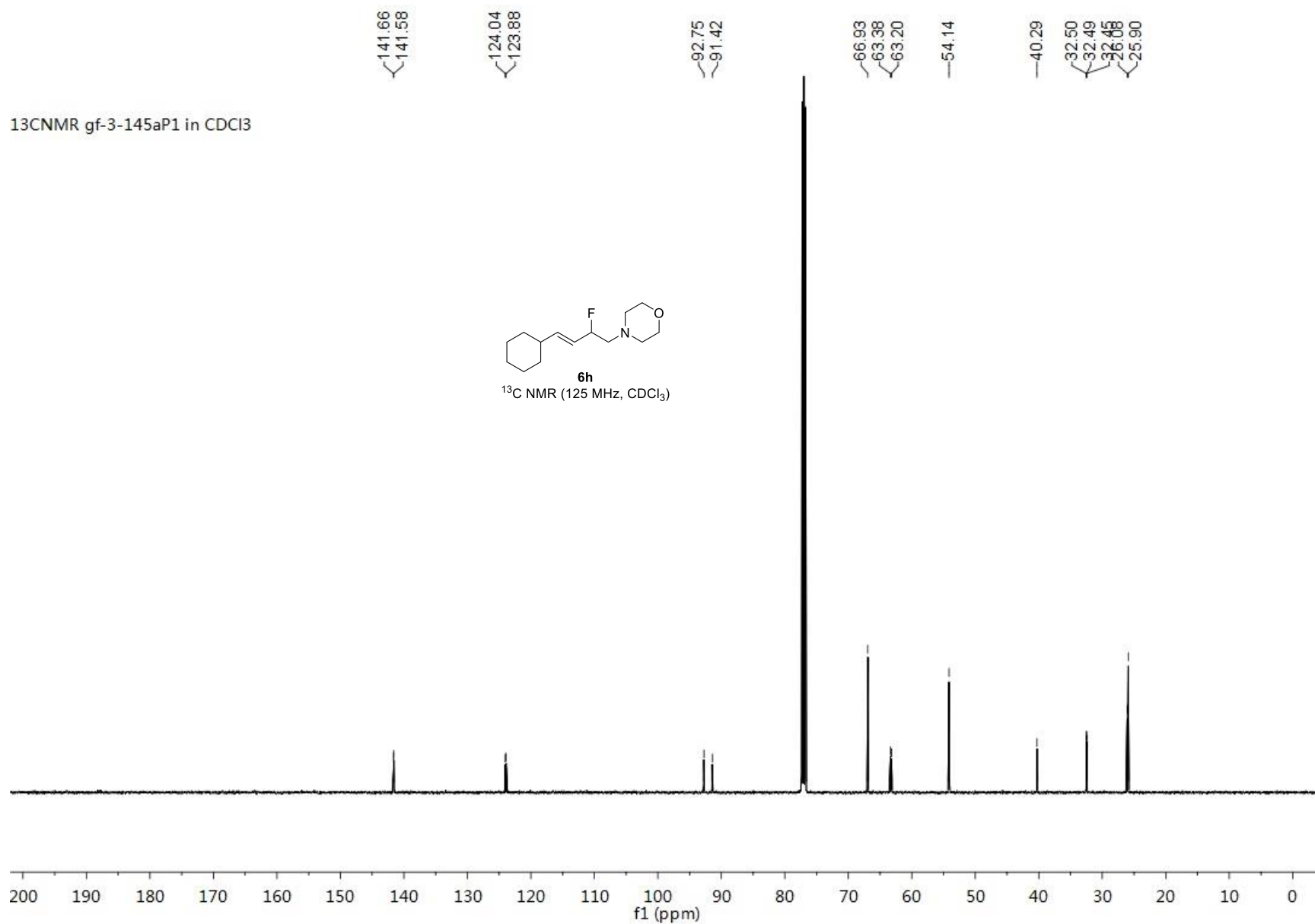
6h
¹H NMR (500 MHz, CDCl₃)



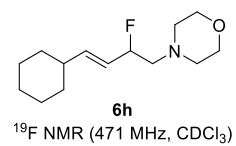
¹³CNMR gf-3-145aP1 in CDCl₃



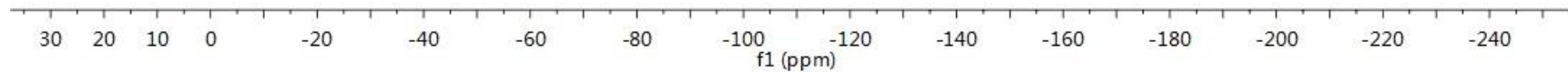
6h
¹³C NMR (125 MHz, CDCl₃)



¹⁹F NMR gf-3-145aP1 in CDCl₃

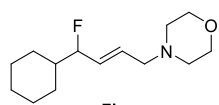


---171.4733

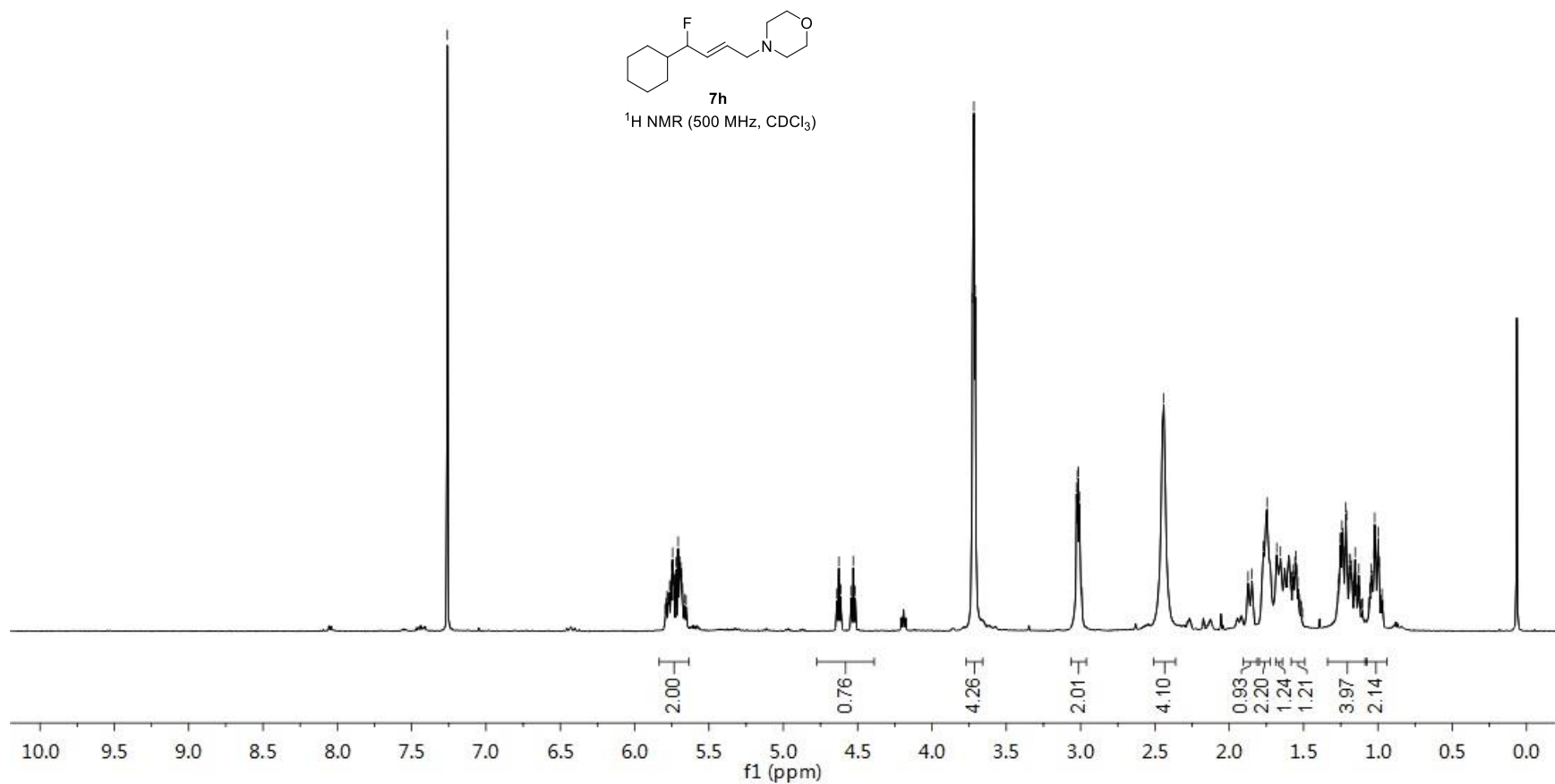


7.2596
 5.7640
 5.7565
 5.7511
 5.7445
 5.7387
 5.7329
 5.7205
 5.7074
 5.6960
 5.6924
 5.6892
 5.6840
 5.6785
 4.6259
 4.6130
 4.5428
 4.5294
 4.5161
 3.7276
 3.7184
 3.7092
 3.0274
 3.0202
 3.0163
 3.0080
 2.4407
 1.8734
 1.8478
 1.7707
 1.7456
 1.6798
 1.6554
 1.5815
 1.5749
 1.5684
 1.5576
 1.5517
 1.5451
 1.5382
 1.2671
 1.2611
 1.2532
 1.2488
 1.2422
 1.2362
 1.2299
 1.2226
 1.2167
 1.2107
 1.2048
 1.1973
 1.1915
 1.1854
 1.1773
 1.1713
 1.1593
 1.1530
 1.1464
 1.1348
 1.1281
 1.1212
 1.0530
 1.0462
 1.0400
 1.0282
 1.0217
 1.0154
 0.9977
 0.9912

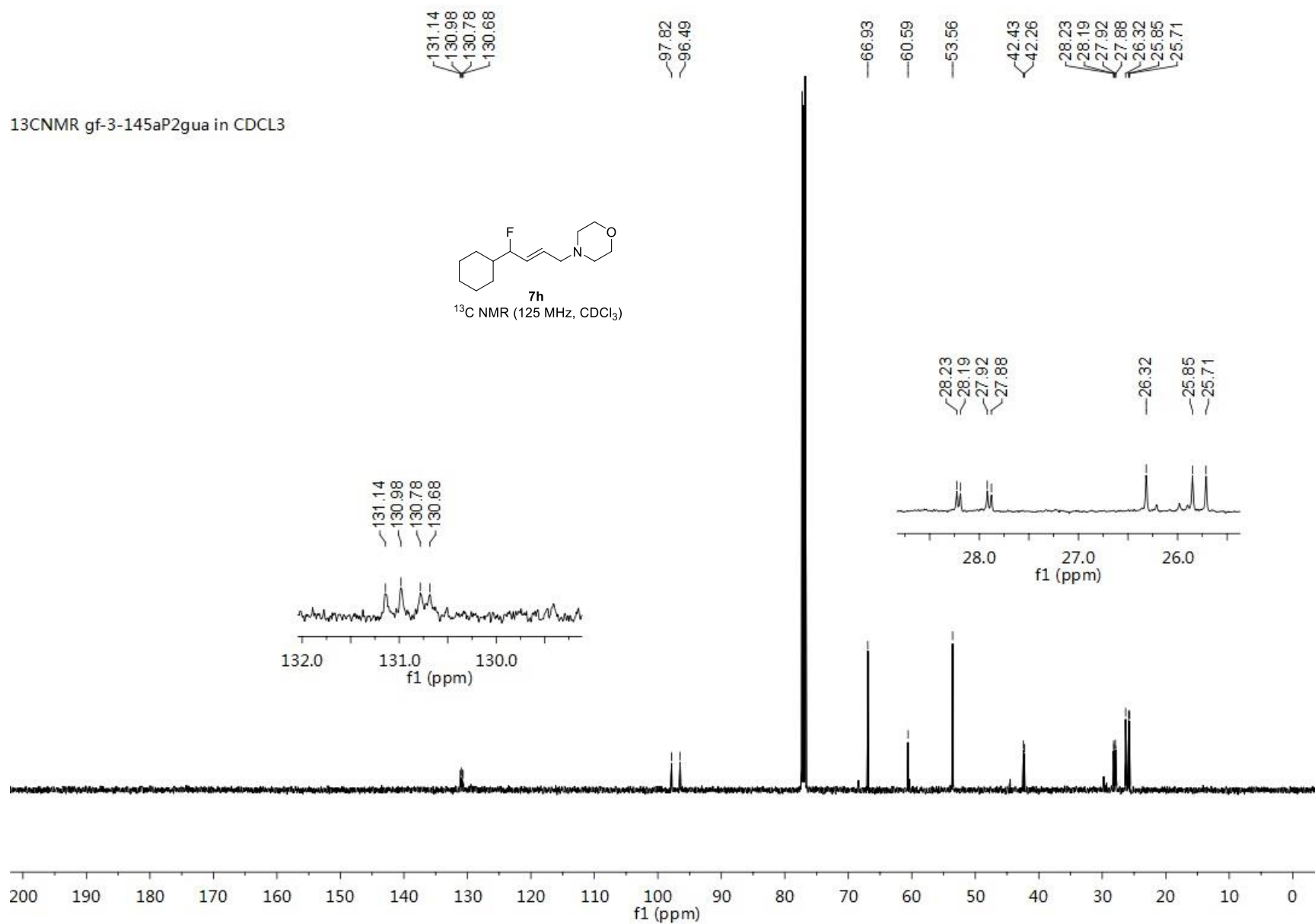
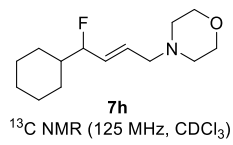
¹H NMR of 7h in CDCl₃



¹H NMR (500 MHz, CDCl₃)

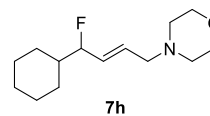


¹³C NMR gf-3-145aP2gua in CDCl₃

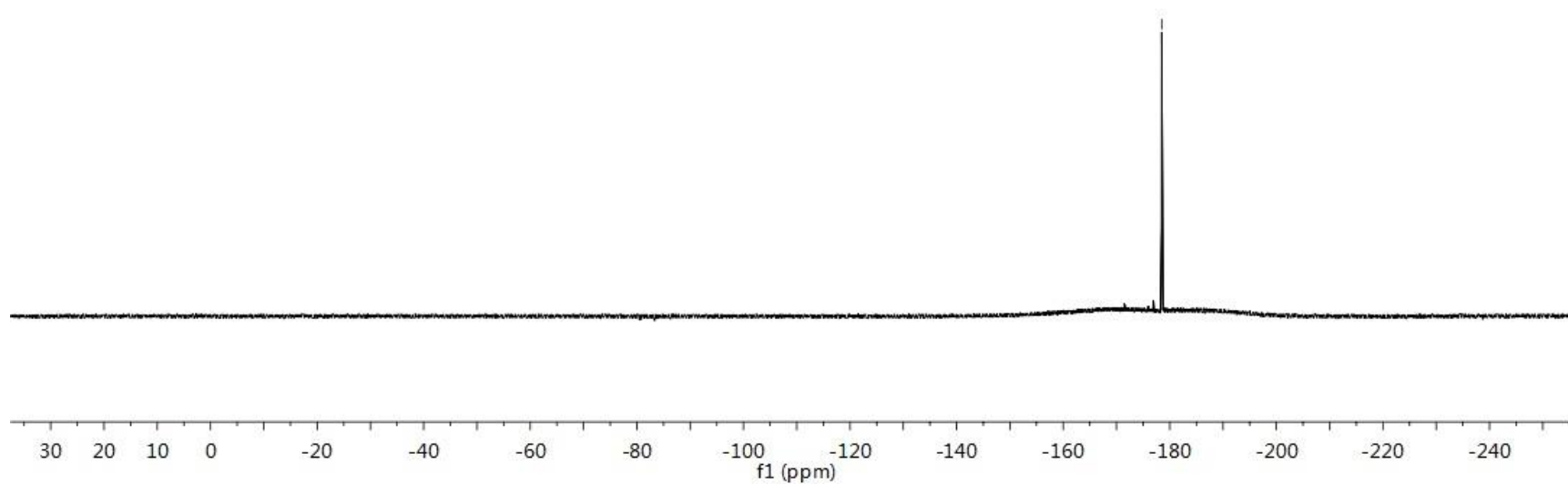


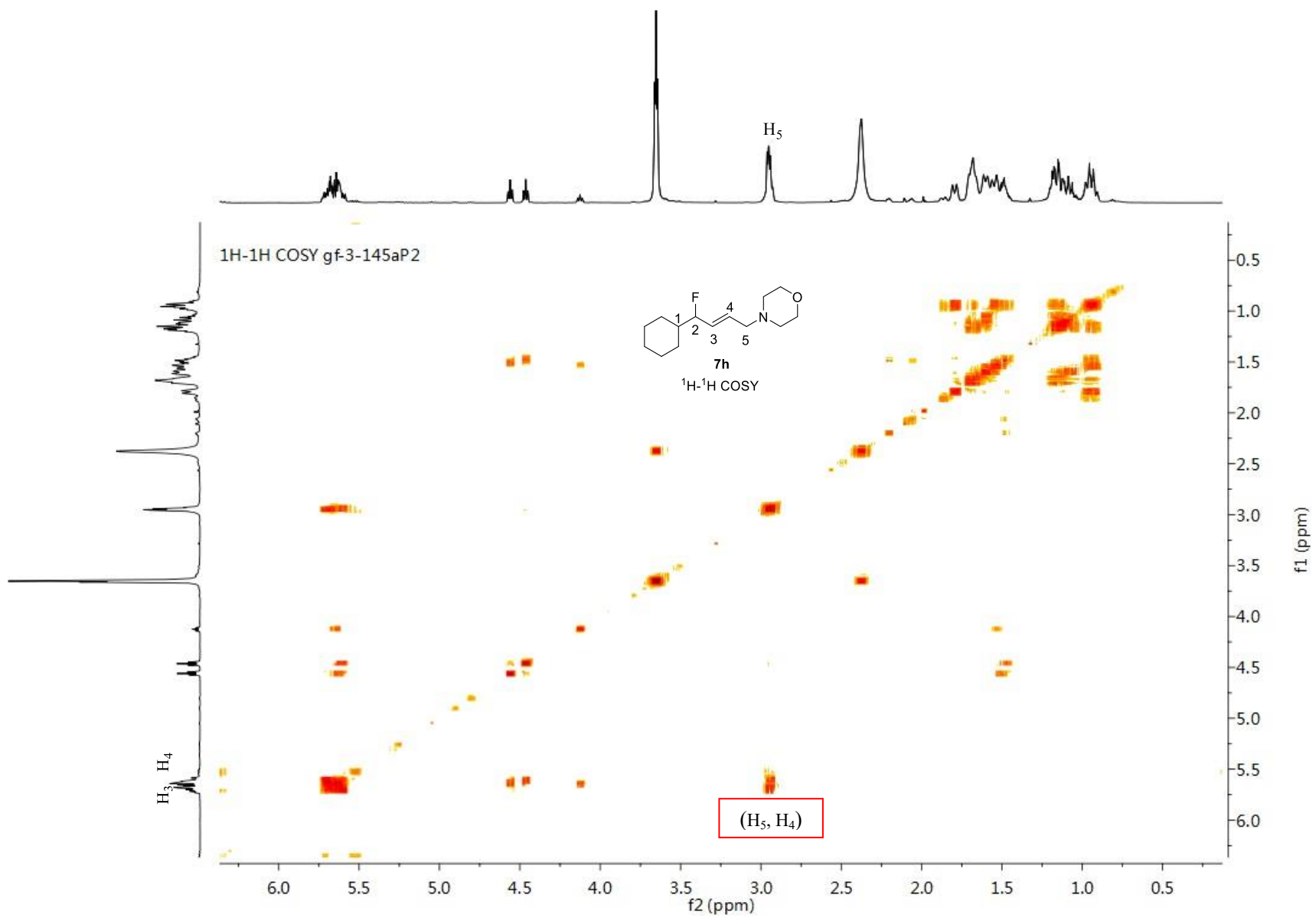
¹⁹F NMR gf-3-145aP2 in CDCl₃

---178.4836



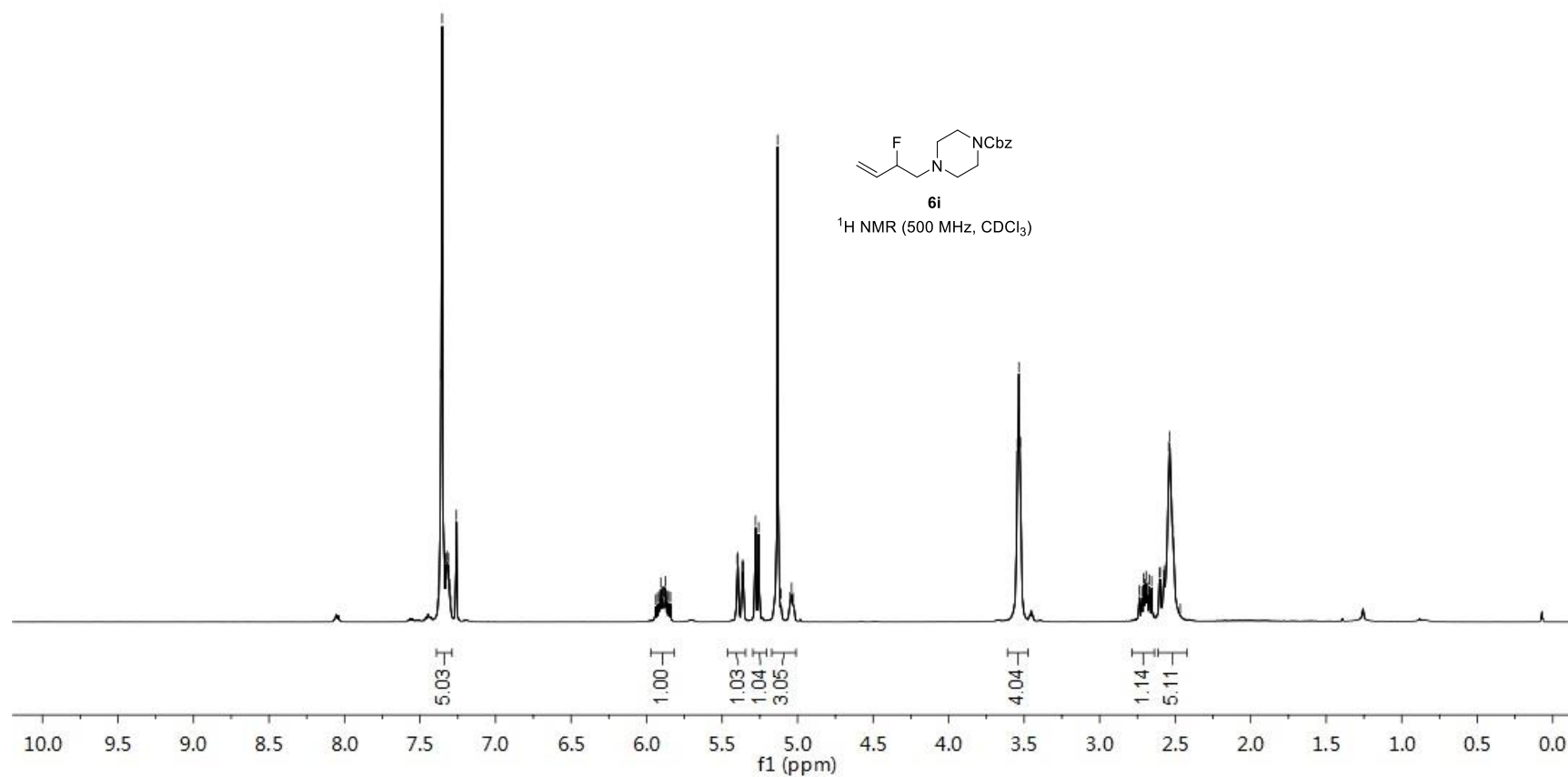
¹⁹F NMR (471 MHz, CDCl₃)

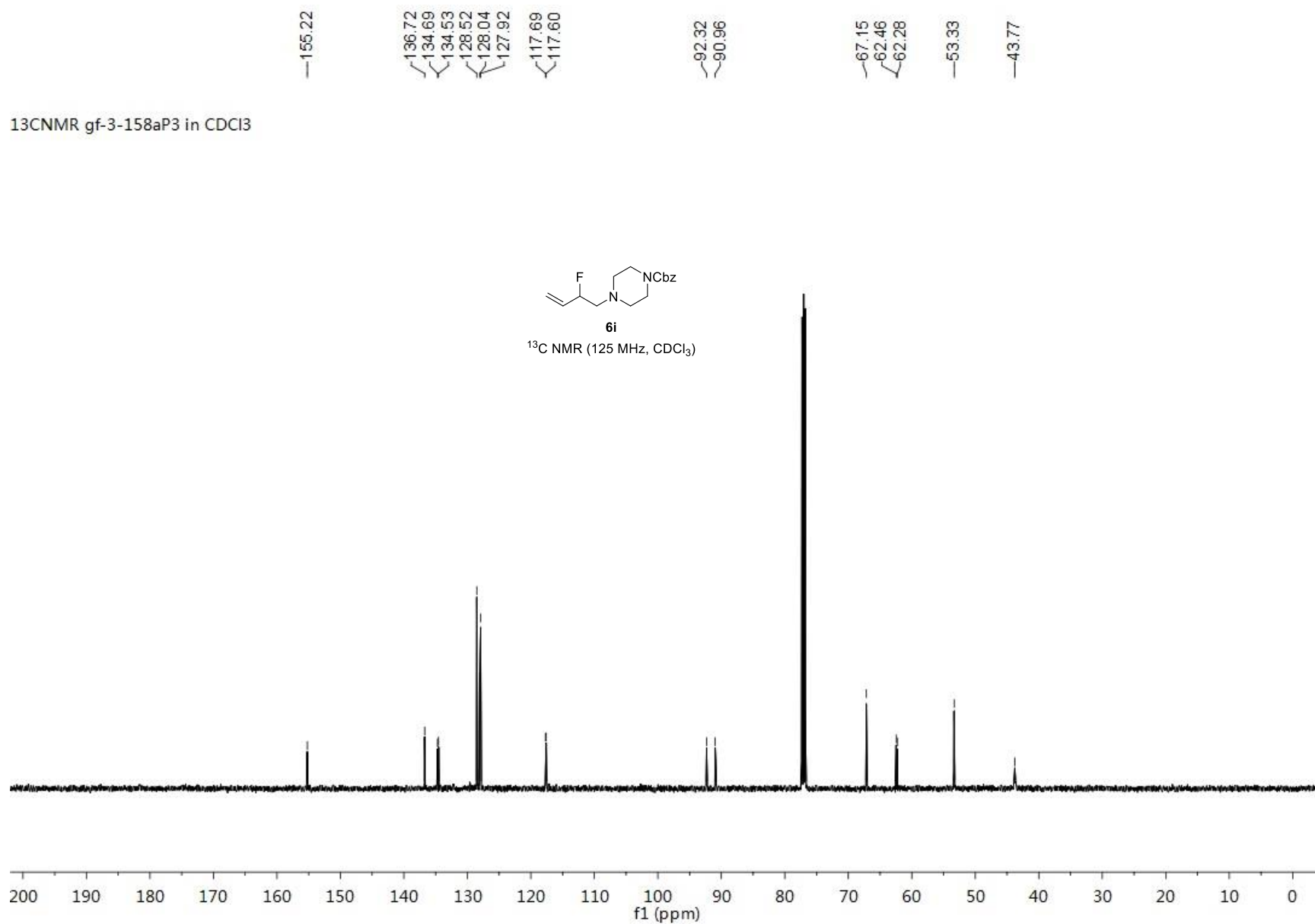




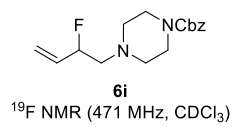
7.3789
7.3609
7.3533
7.3390
7.3300
7.3217
7.3130
7.3033
7.2960
7.2599
5.9058
5.8974
5.8932
5.8873
5.8831
5.8747
5.4003
5.3976
5.3948
5.3657
5.3630
5.3602
5.2786
5.2571
5.1324
5.1109
5.0498
5.0399
5.0344
3.5346
3.5255
2.7369
2.7209
2.7085
2.6995
2.6926
2.6836
2.6713
2.6553
2.6044
2.5989
2.5760
2.5704
2.5415
2.5365
2.5146
2.5090
2.4662

¹H NMR of gf-3-258P3 in CDCl₃

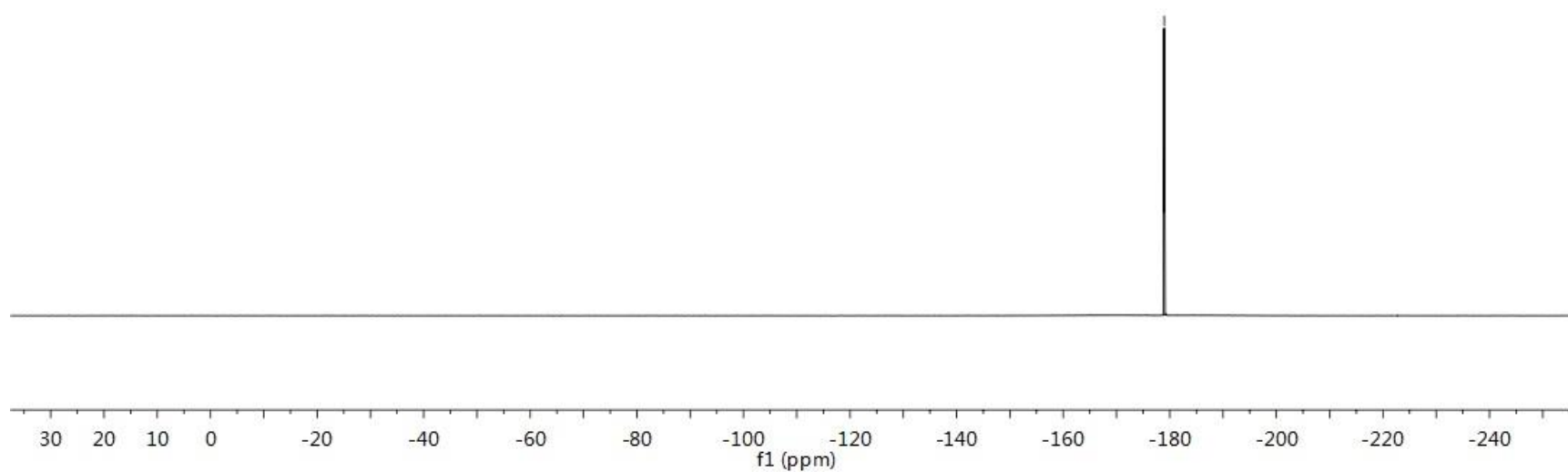




¹⁹F NMR gf-3-158aP3 in CDCl₃



---178.9649



S233

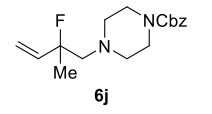
7.3744
7.3609
7.3569
7.3500
7.3401
7.3363
7.3266
7.3214
7.3187
7.3117
7.3094
7.3058
7.3004
7.2923
7.2600
5.9855
5.9634
5.9503
5.9282
5.9152
5.8931
5.3212
5.2864
5.1627
5.1406
5.1274

3.5003
3.4802
3.4804

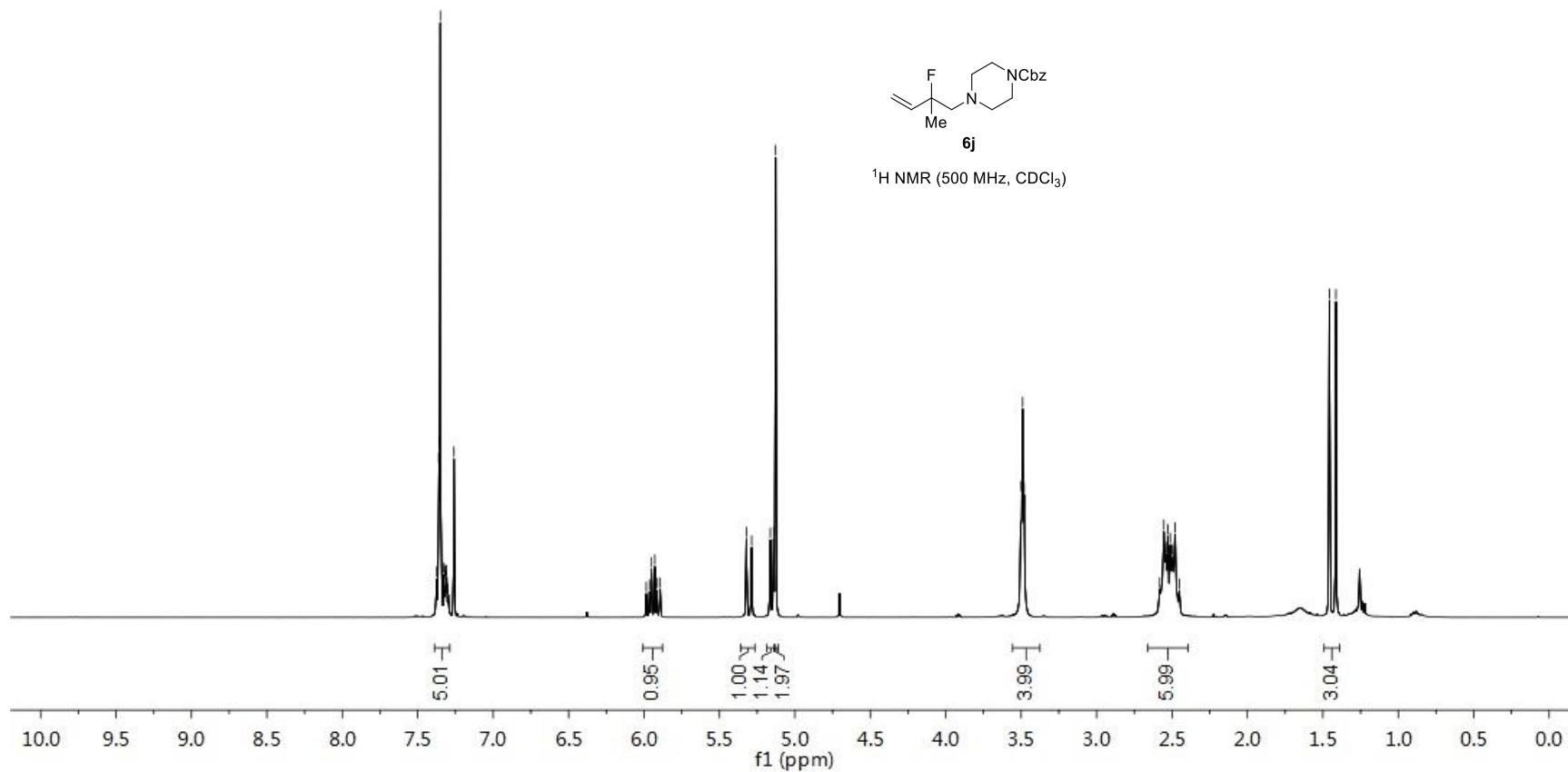
2.5822
2.5539
2.5407
2.5276
2.5118
2.4990
2.4799
2.4514

1.4567
1.4135

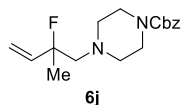
¹H NMR of **6j** in CDCl₃



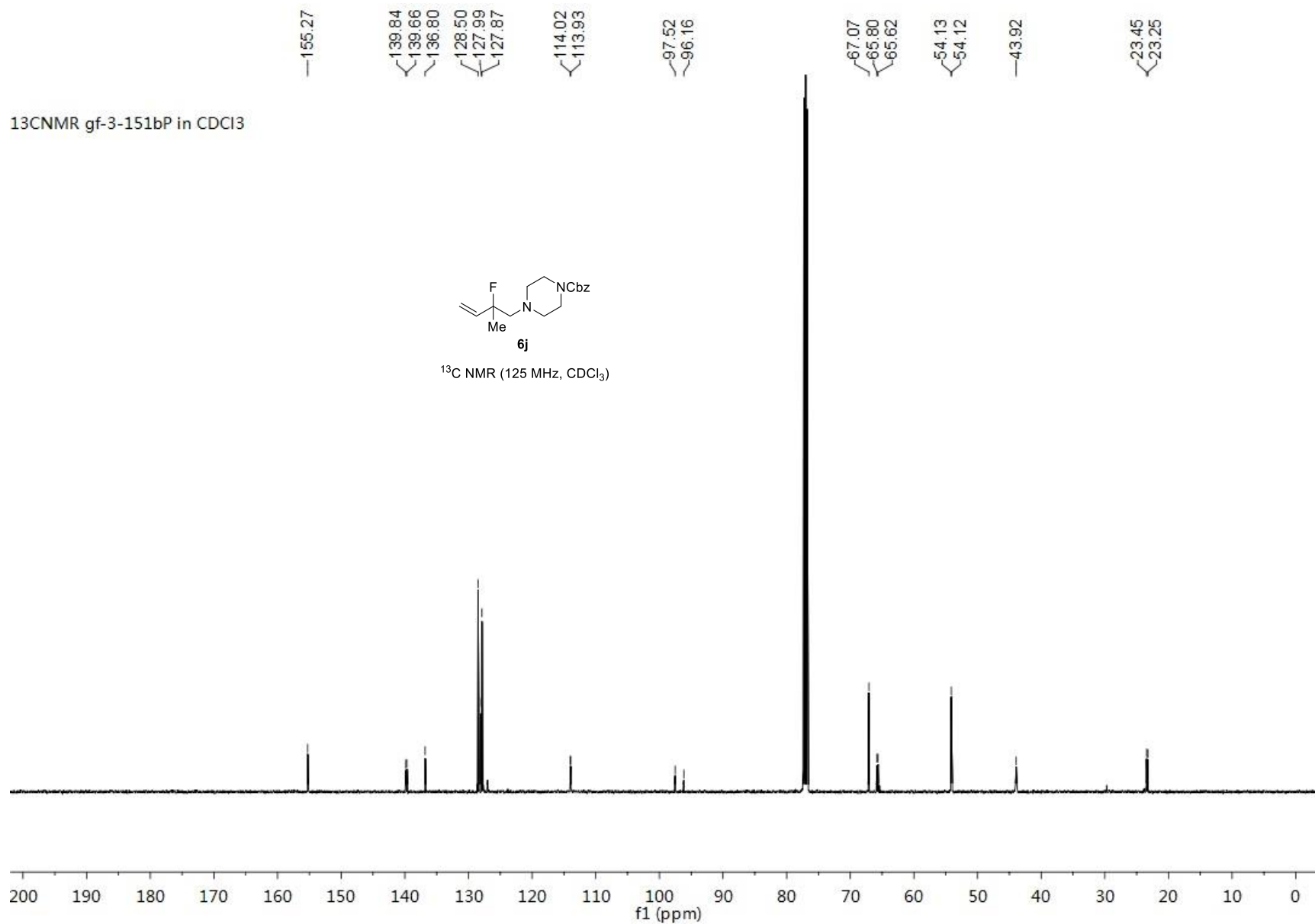
¹H NMR (500 MHz, CDCl₃)



¹³C NMR gf-3-151bP in CDCl₃

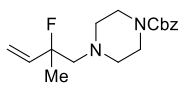


¹³C NMR (125 MHz, CDCl₃)



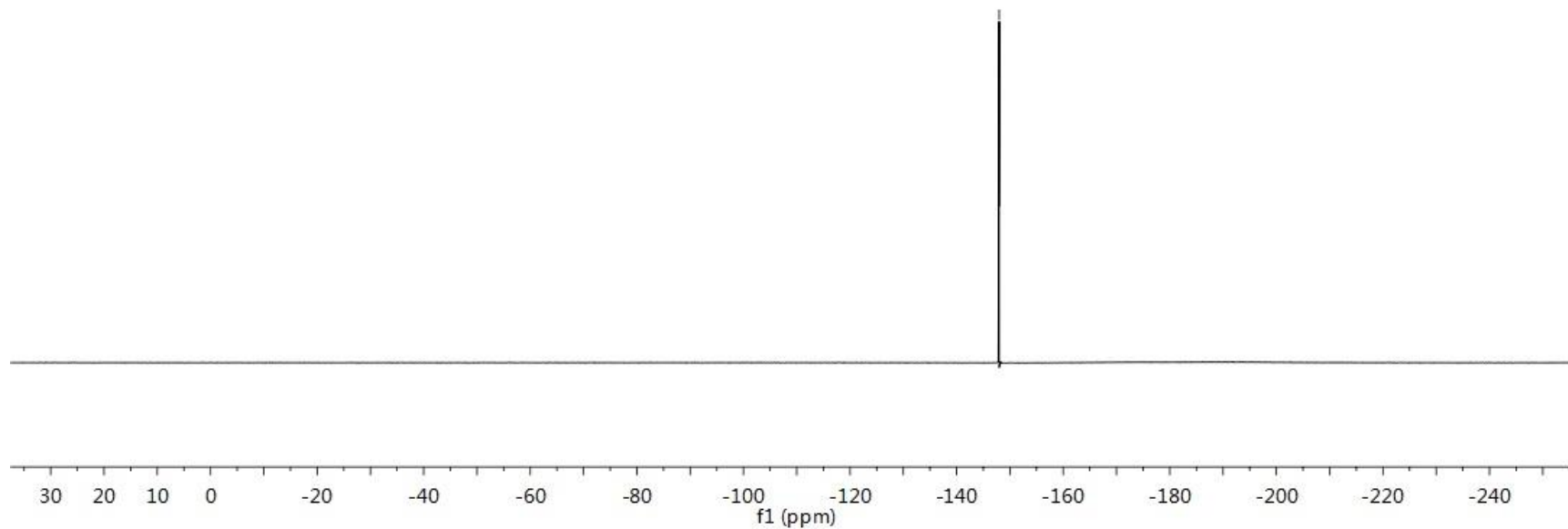
¹⁹F NMR gf-3-151bP1 in CDCl₃

— 147.9854



6j

¹⁹F NMR (471 MHz, CDCl₃)



7.3761
7.3728
7.3609
7.3598
7.3547
7.3493
7.3388
7.3281
7.3237
7.3147
7.3110
7.2997
7.2942
7.2602

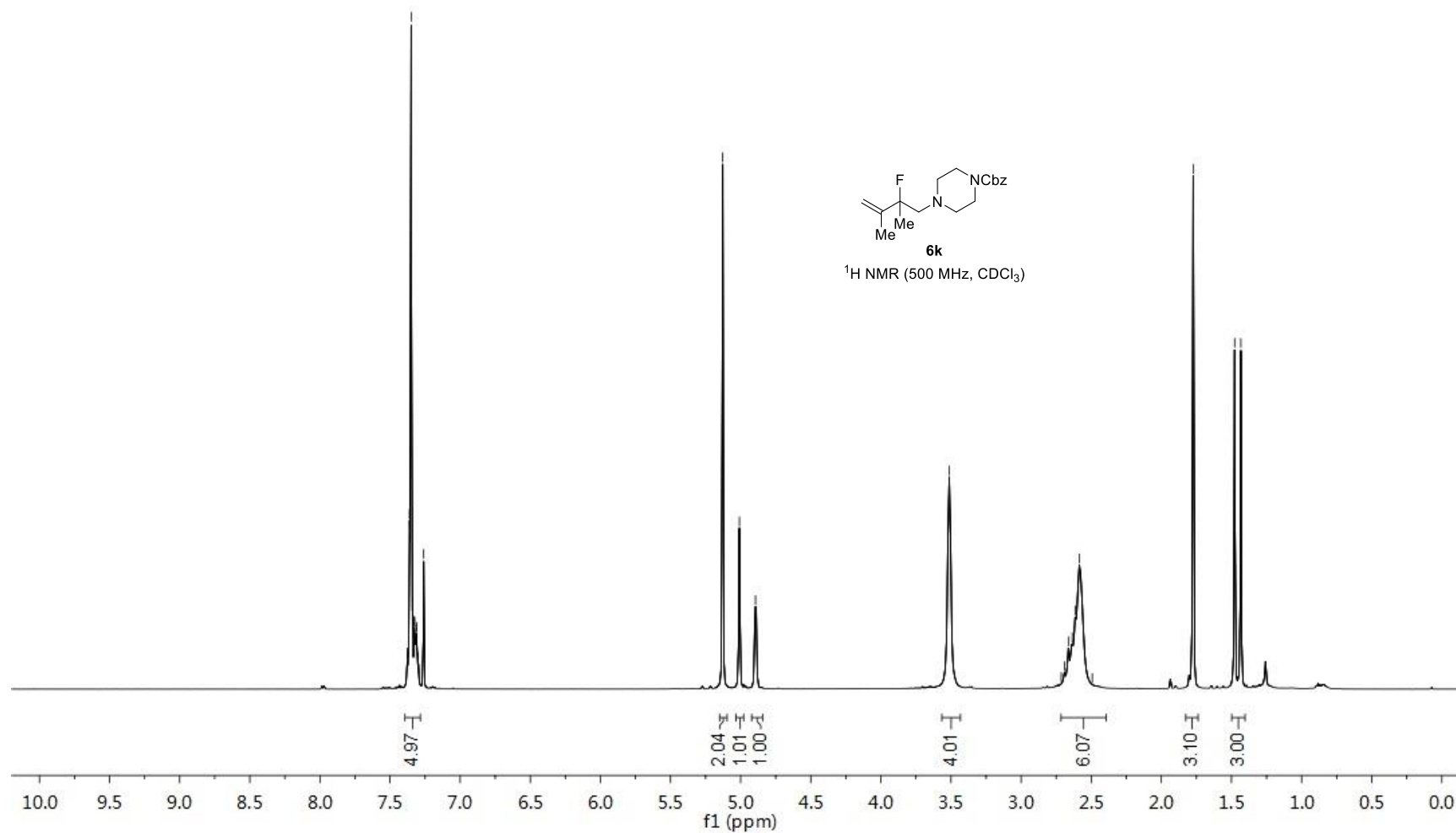
5.1277
5.0087
4.8939

3.5124

2.7141
2.6914
2.6630
2.6361
2.6151
2.5849
2.4924

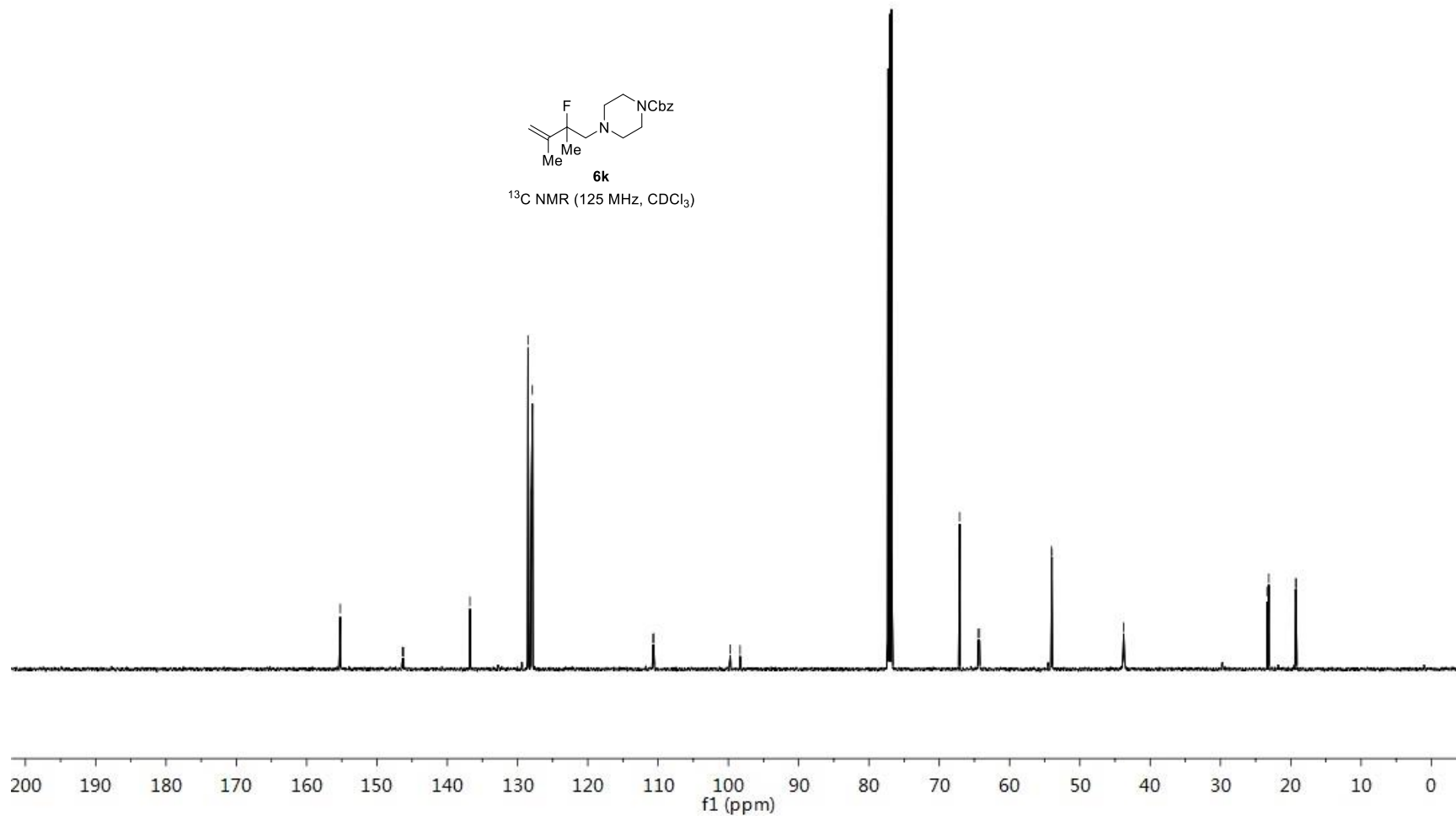
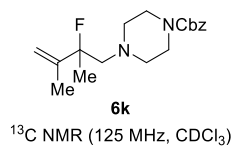
1.7729
1.4775
1.4331

¹H NMR of 6k in CDCl₃



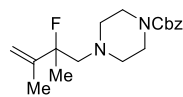
—155.23
—146.37
—146.21
—136.77
—128.50
—128.01
—127.88
—110.72
—110.63
—99.72
—98.33
—67.10
—64.46
—64.28
—54.01
—53.99
—43.76
—23.33
—23.13
—19.29
—19.25

¹³CNMR gf-3-153bP1re in CDCl₃

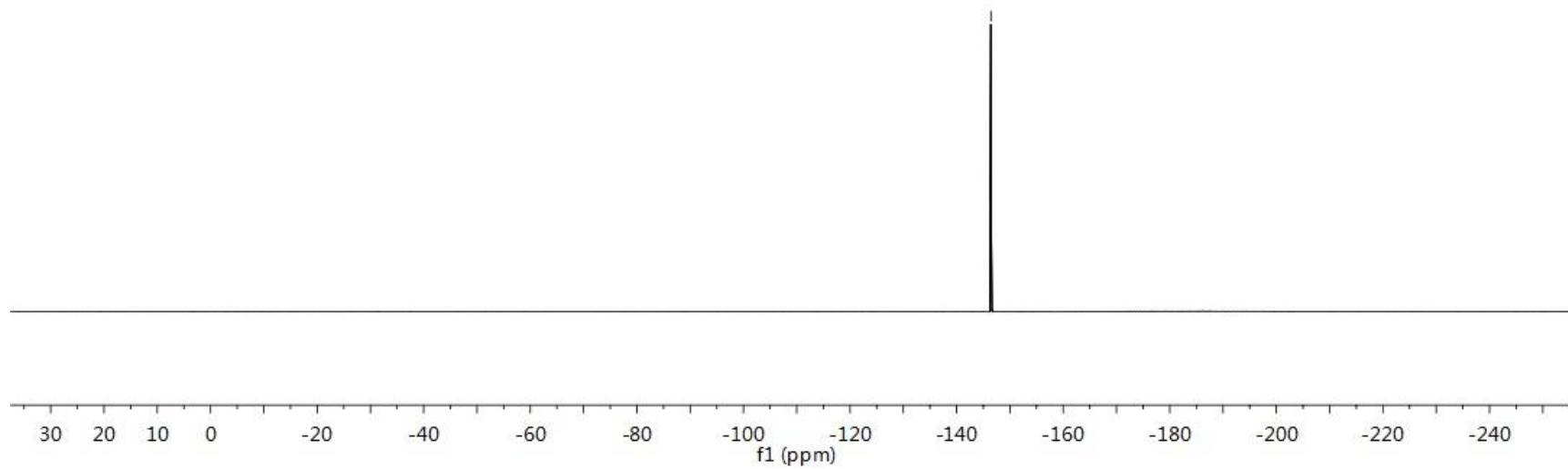


¹⁹F NMR gf-3-153bP1 in CDCl₃

146.4459

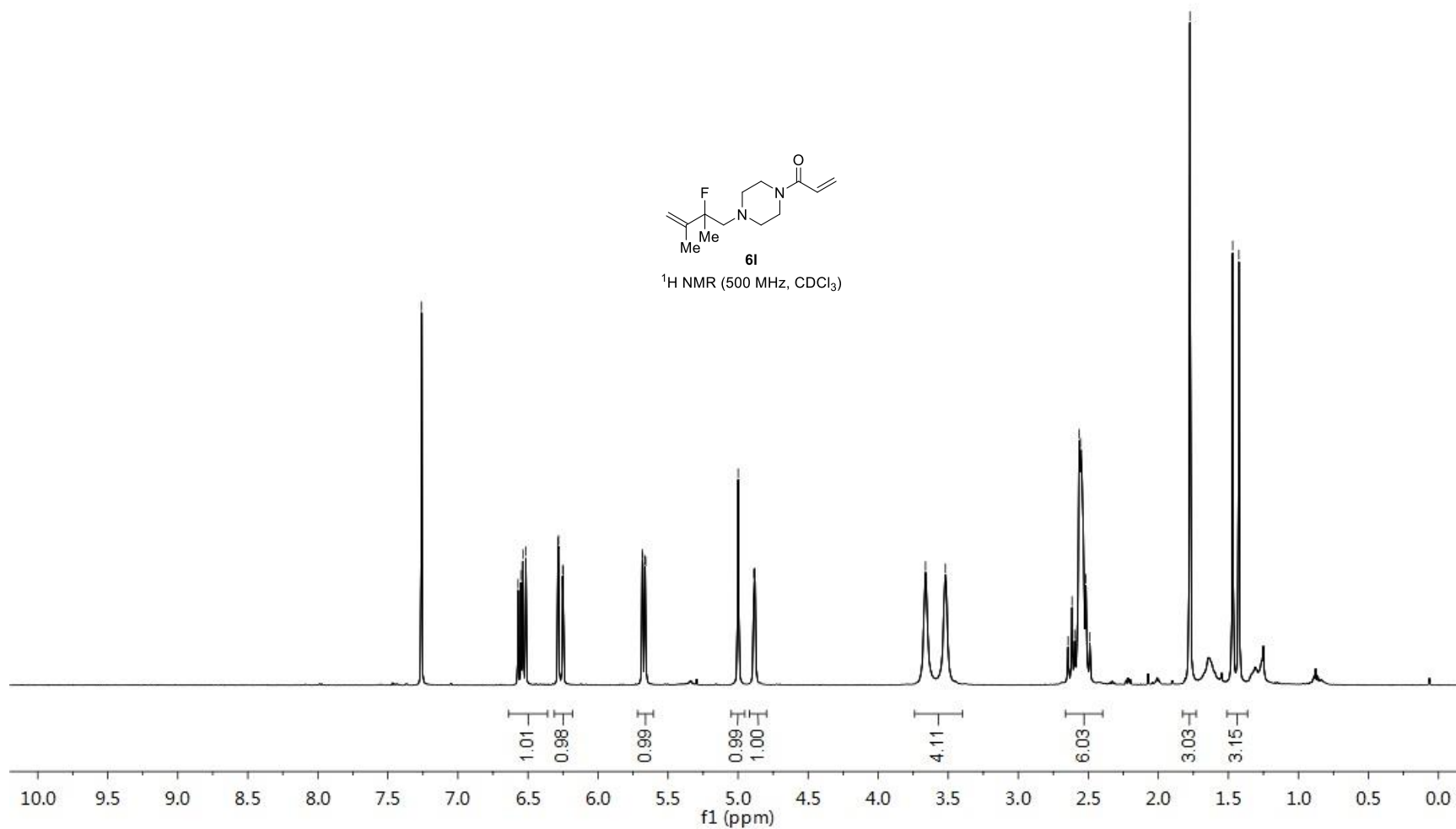
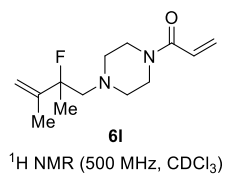


¹⁹F NMR (471 MHz, CDCl₃)



7.2597
 6.5703
 6.5491
 6.5366
 6.5155
 6.2856
 6.2818
 6.2519
 6.2481
 5.6848
 5.6810
 5.6637
 5.6599
 4.9992
 4.8867
 4.8837
 4.8821
 4.8792
 3.6618
 3.5198
 2.6449
 2.6163
 2.5973
 2.5652
 2.5538
 2.5171
 2.4885
 1.7743
 1.4693
 1.4249

¹H NMR of gf-3-229aP in CDCl₃



—165.31

146.47
146.31

127.67
127.54

110.56
110.47

99.88
98.49

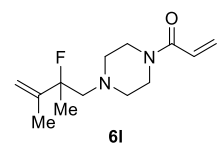
64.43
64.26

54.41
54.09

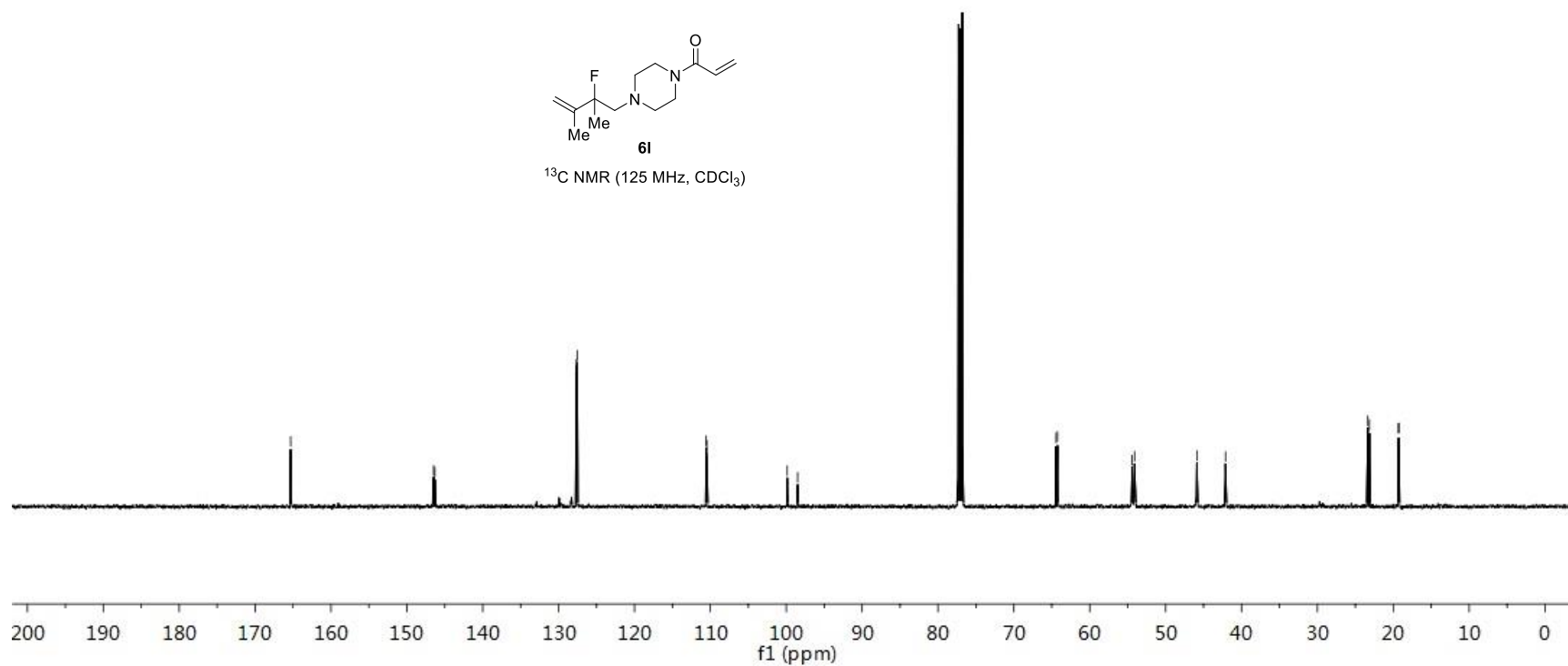
45.89
42.10

23.35
23.15
19.32
19.28

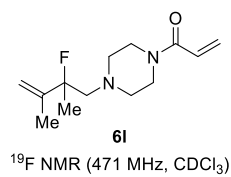
¹³CNMR gf-3-229PP in CDCl₃



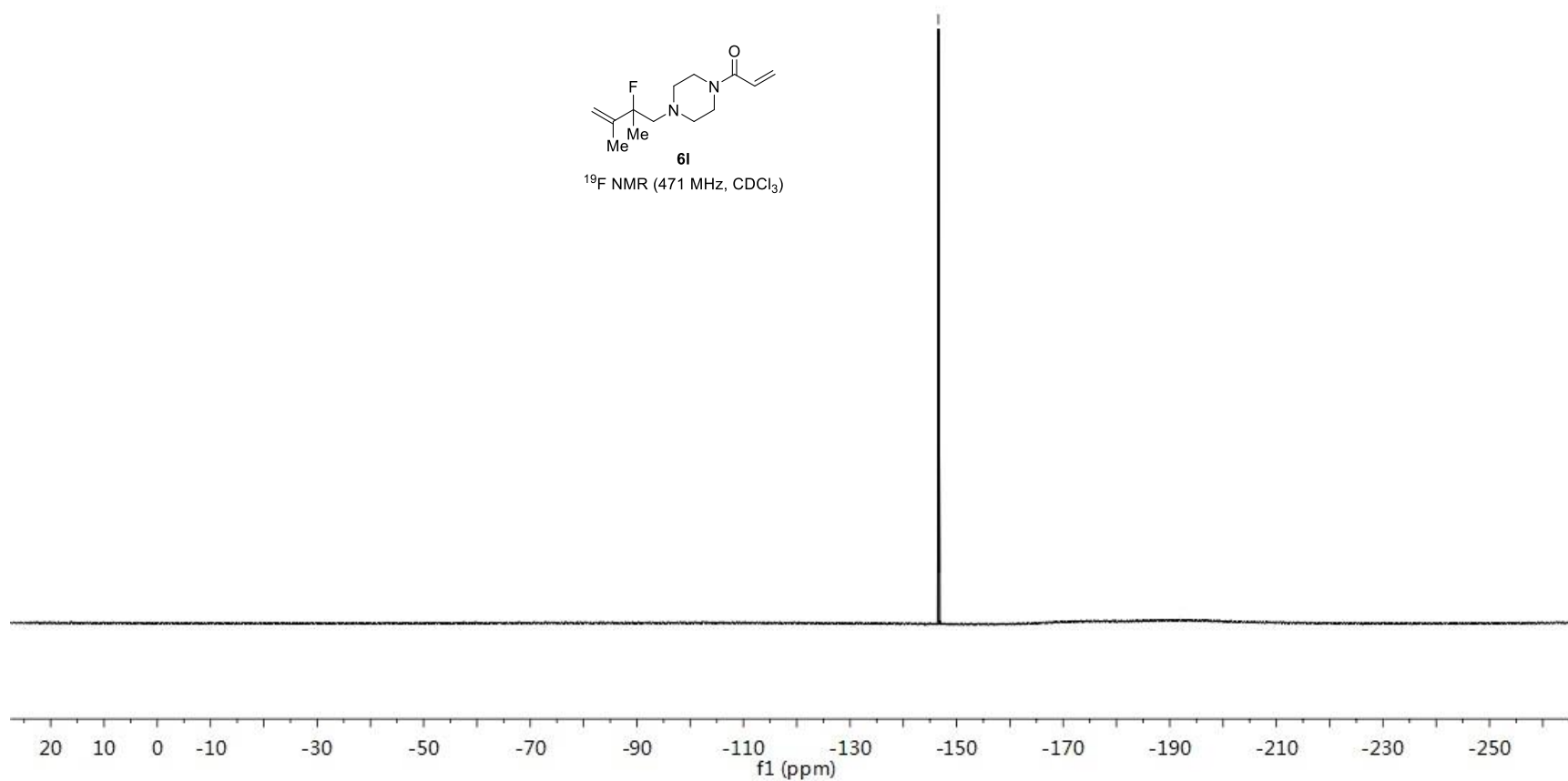
¹³C NMR (125 MHz, CDCl₃)



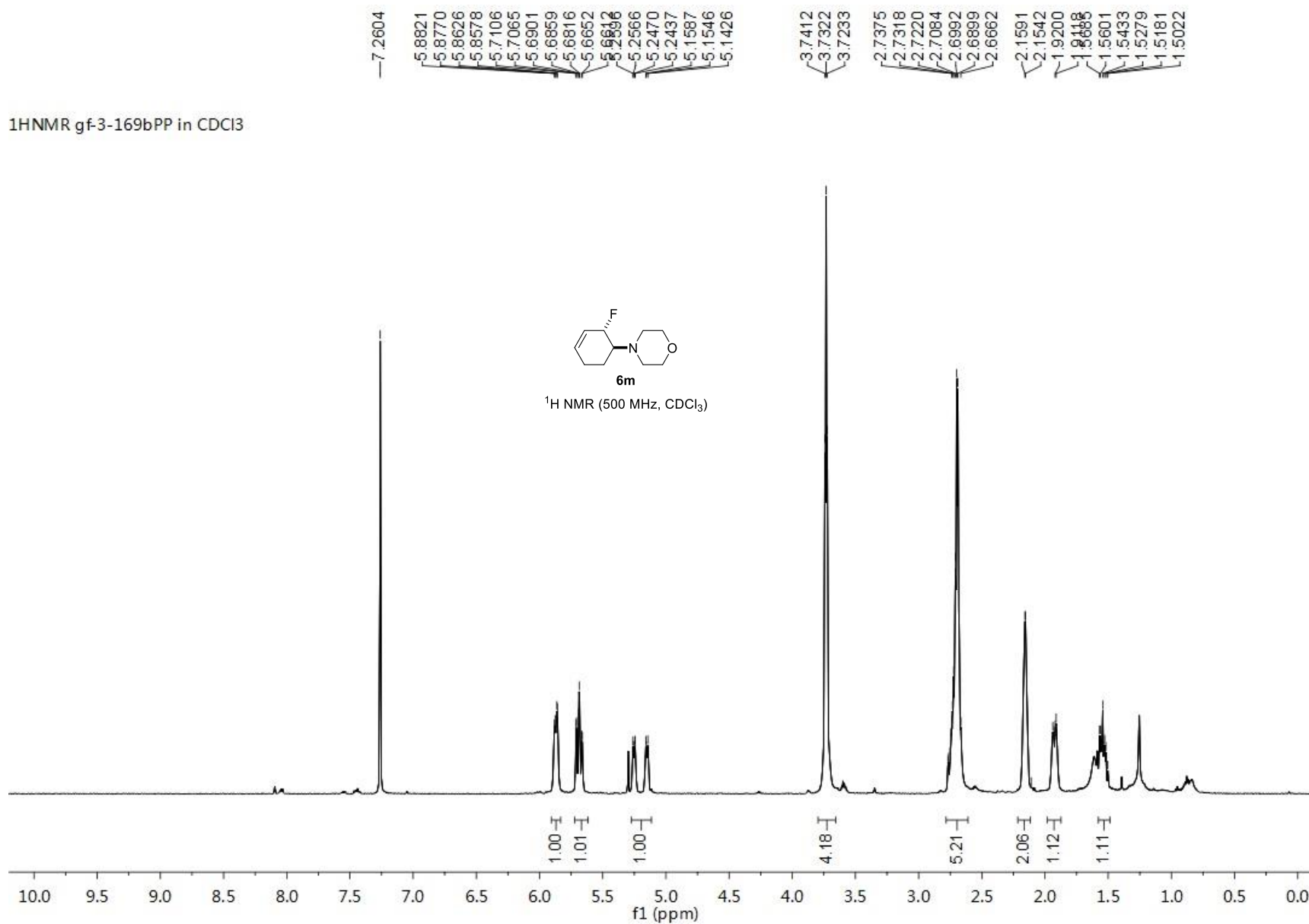
¹⁹F NMR gf-3-229aP in CDCl₃



-146.5713



¹H NMR of 6m in CDCl₃



¹³C NMR gf-3-169bPP in CDCl₃

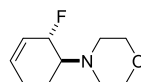
131.49
131.41
126.33
126.16

89.34
88.02

67.51
64.43
64.30

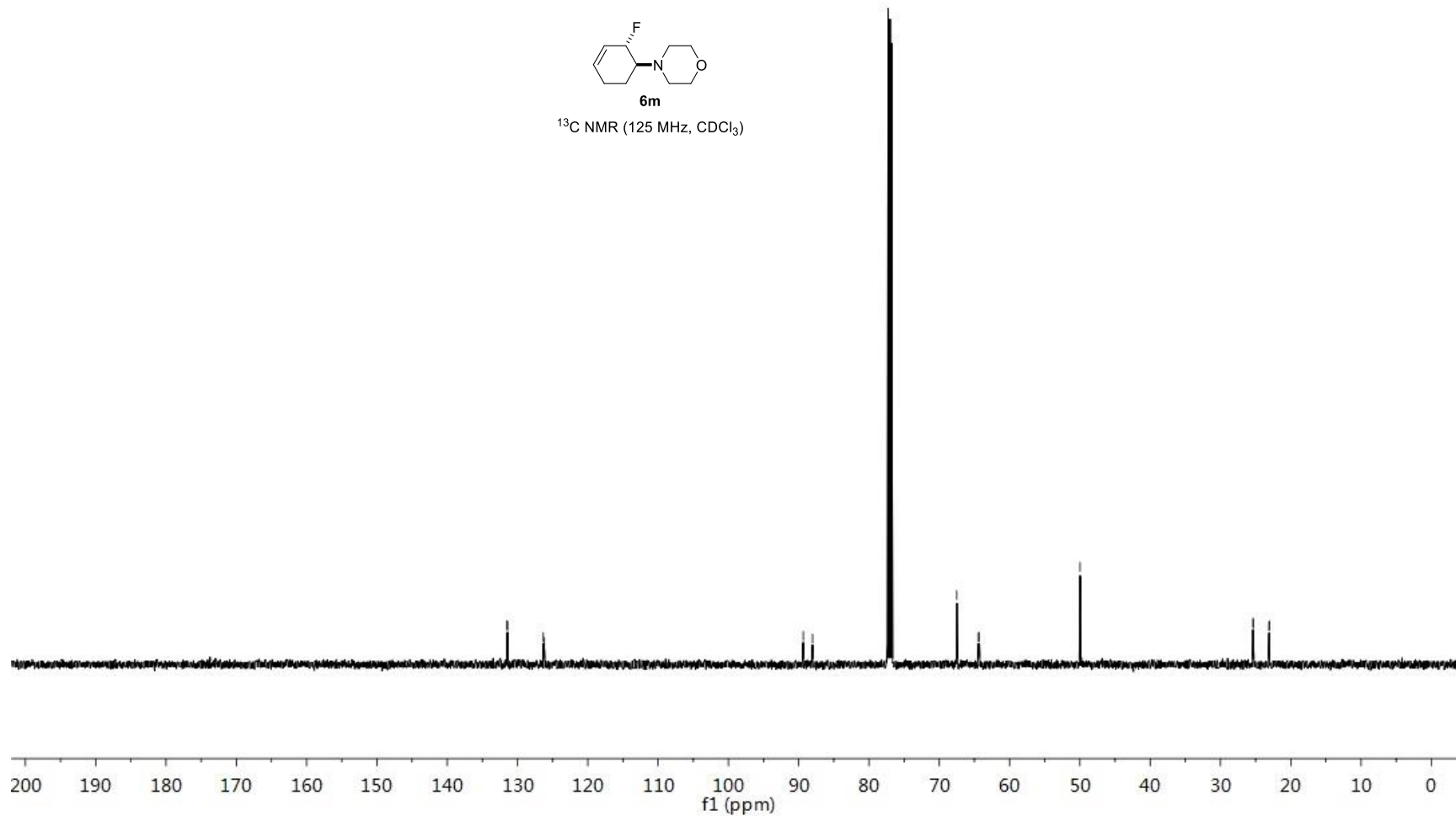
49.97

25.37
25.35
23.10
23.04

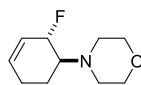


6m

¹³C NMR (125 MHz, CDCl₃)



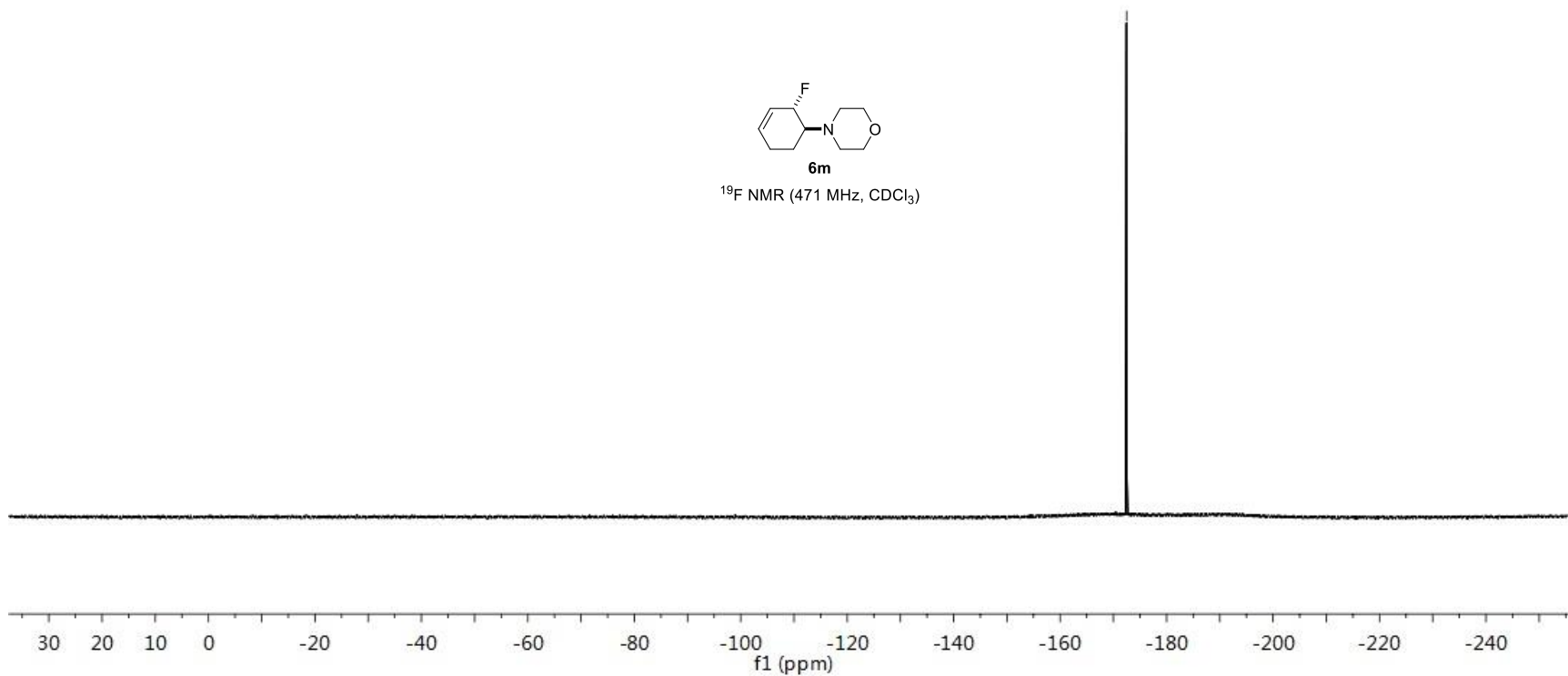
19FNMR gf-3-169bPP in CDCl3

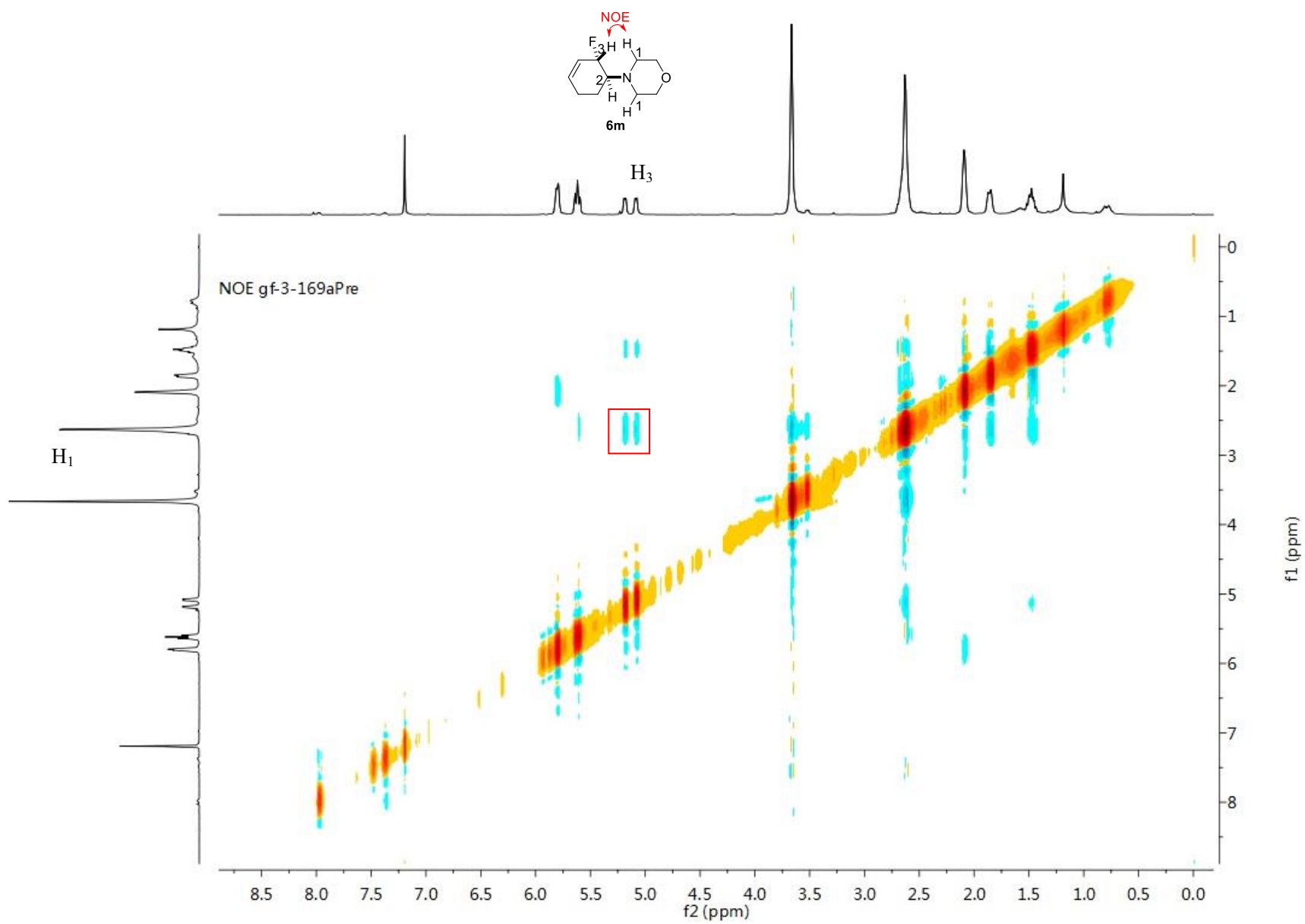


6m

¹⁹F NMR (471 MHz, CDCl₃)

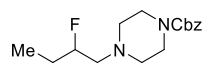
---172.4859





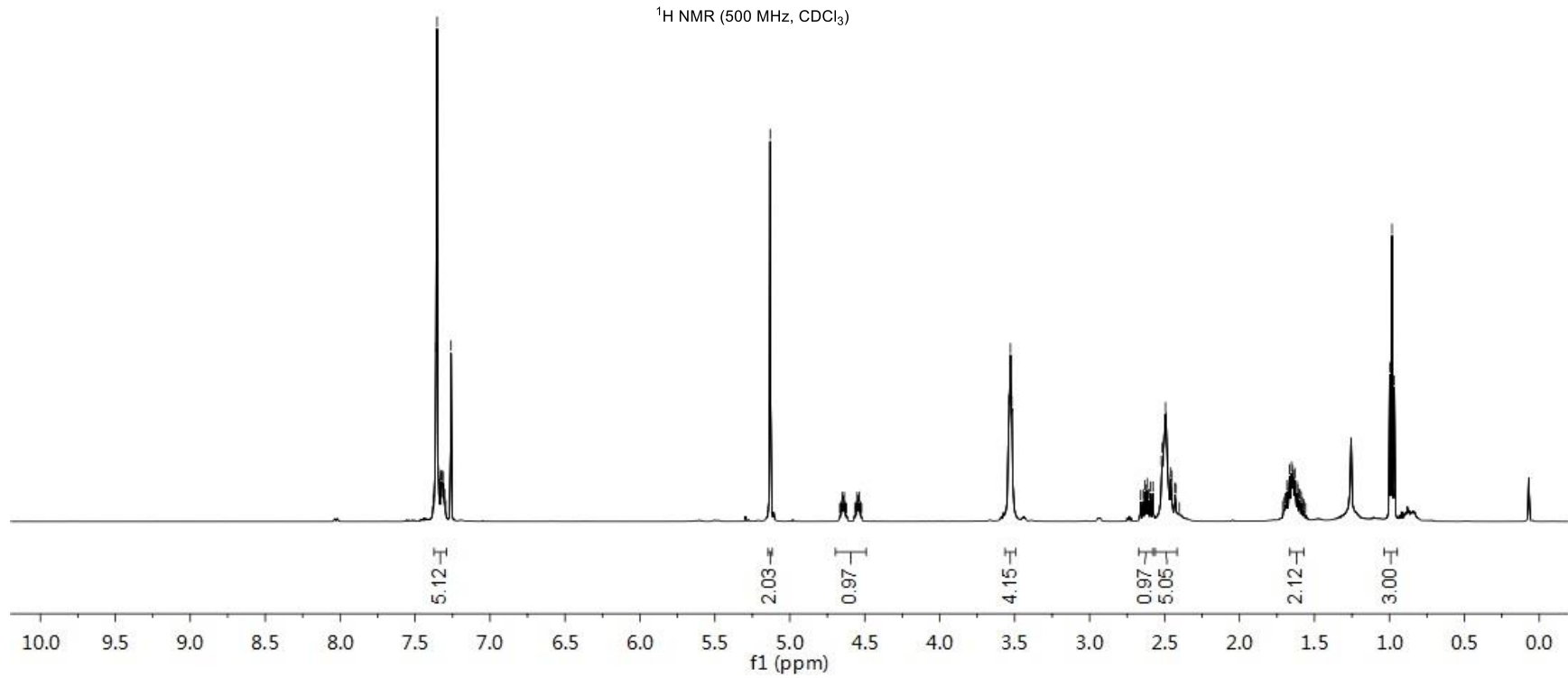
7.3789
7.3755
7.3607
7.3532
7.3385
7.3296
7.3239
7.3213
7.3125
7.3089
7.3029
7.2953
7.2603
5.1312
4.6613
4.6563
4.6515
4.6465
4.6415
4.6365
4.6318
4.6268
4.5614
4.5569
4.5516
4.5464
4.5419
4.5366
4.5314
4.5269
3.5381
3.5283
3.5190
2.6592
2.6440
2.6313
2.6209
2.6160
2.6058
2.5930
2.5778
2.5213
2.5161
2.4936
2.4886
2.4585
2.4534
2.4305
2.4254
1.7105
1.7025
1.6961
1.6895
1.6821
1.6673
1.6524
1.6447
1.6406
1.6300
1.6151
1.6123
1.6021
1.5972
1.5873
1.5833
1.5731
1.5687
0.9976
0.9827
0.9677

¹H NMR of gf-3-201aP in CDCl₃

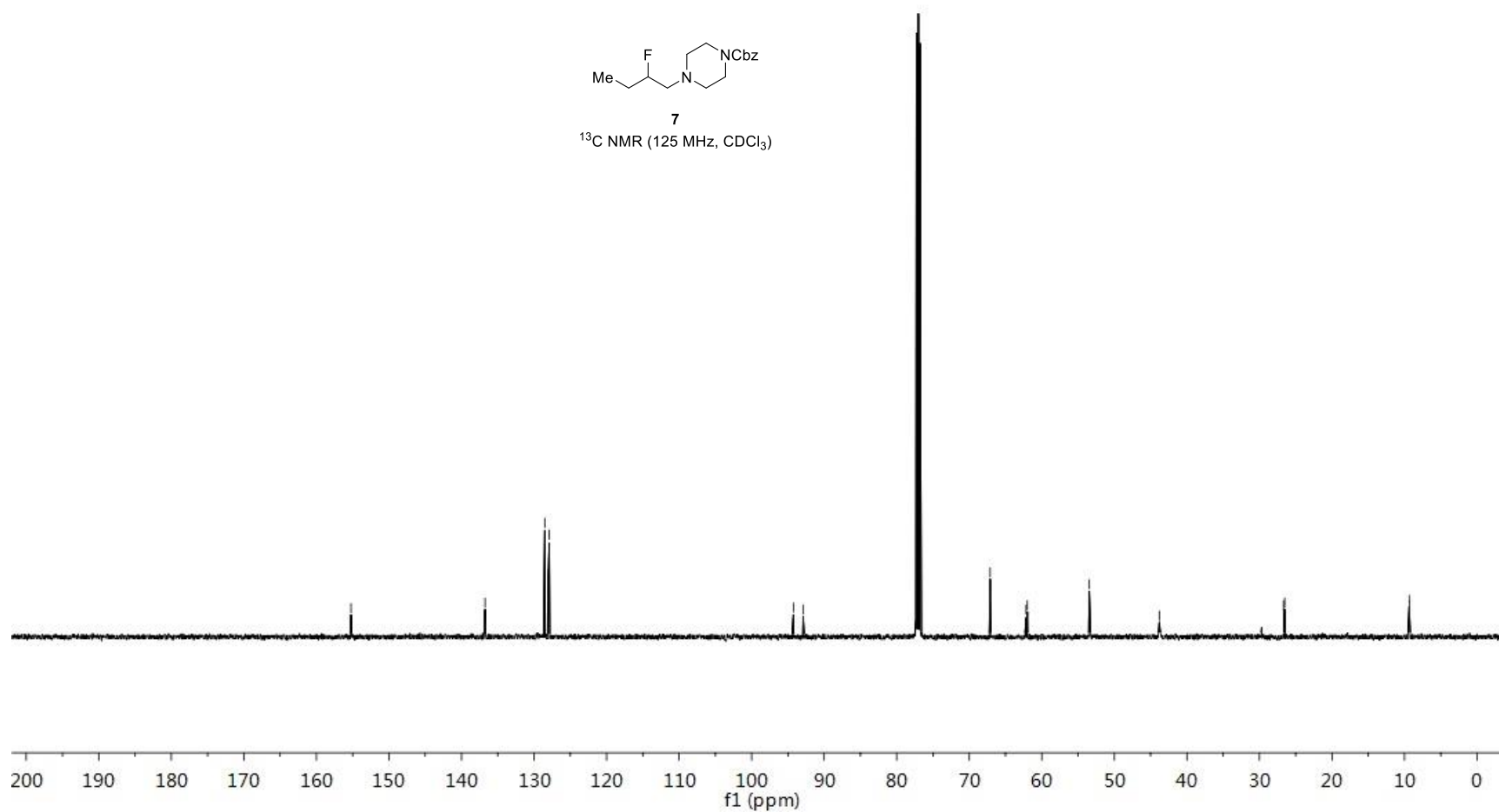


7

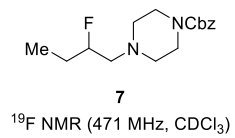
¹H NMR (500 MHz, CDCl₃)



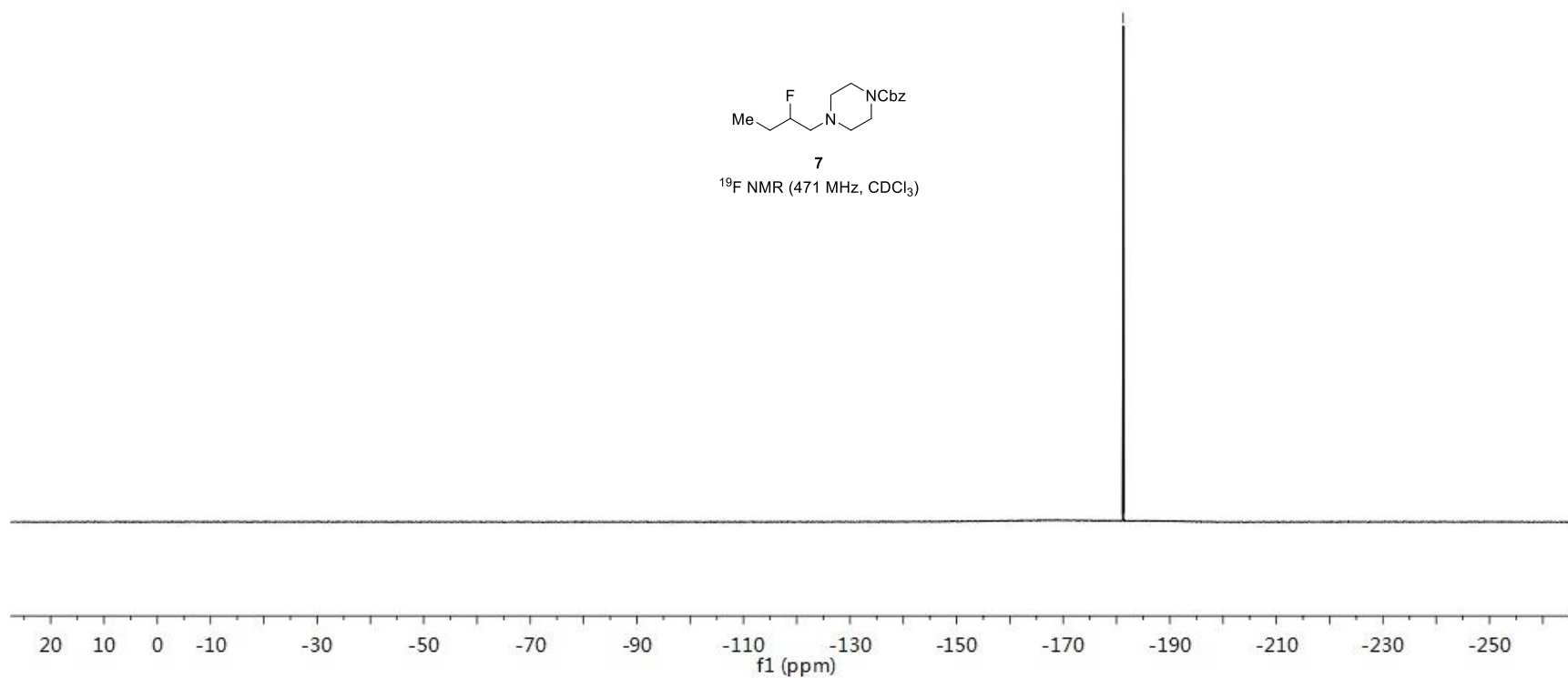
¹³C NMR gf-3-201aP in CDCl₃



¹⁹F NMR gf-3-201aP in CDCl₃



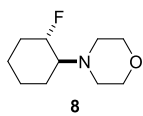
— -181.2557



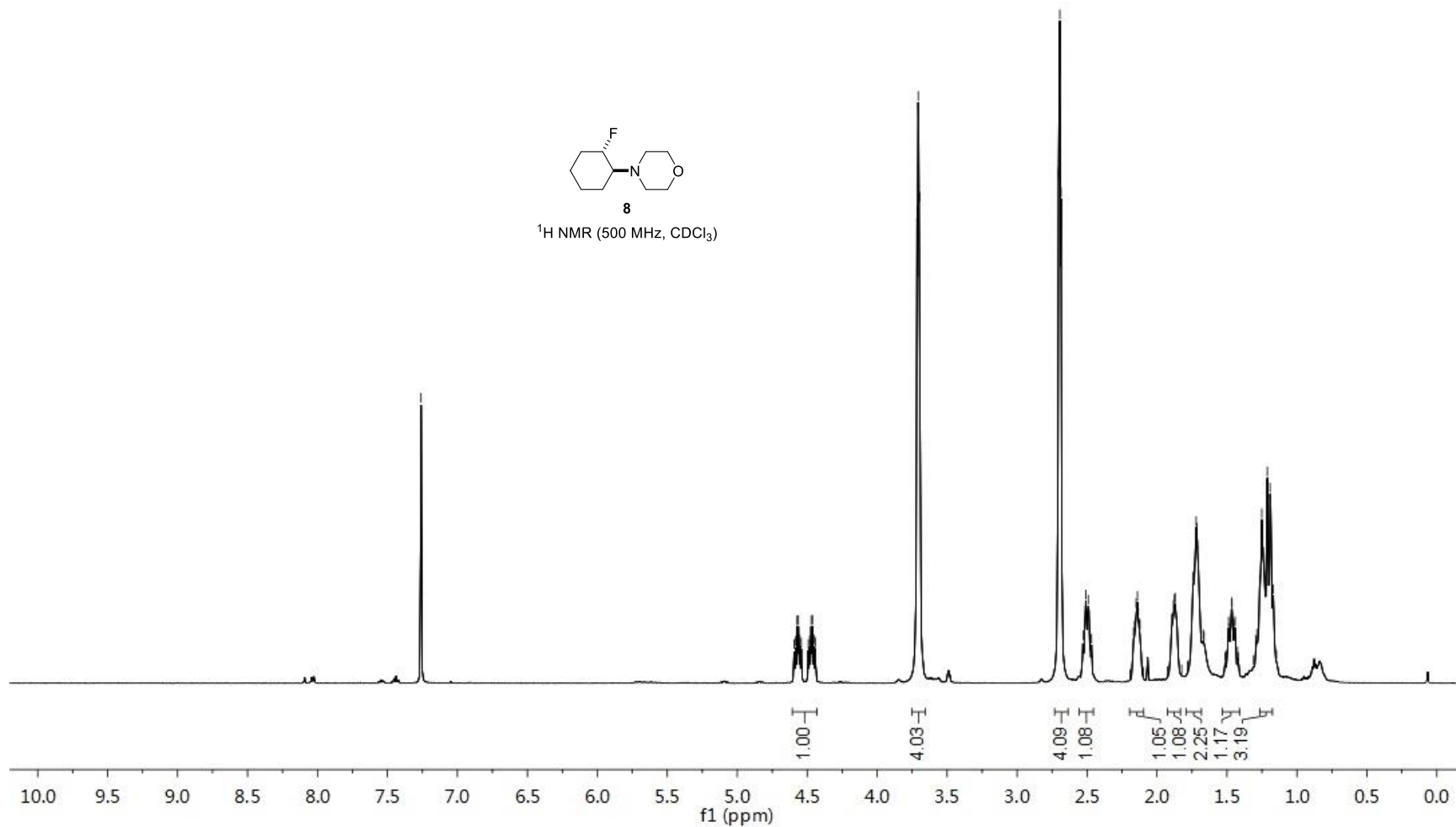
¹H NMR of compound 8 in CDCl₃

7.2599

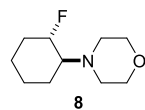
4.5924
4.5827
4.5721
4.5629
4.5522
4.5426
4.4917
4.4821
4.4715
4.4622
4.4515
4.4420
3.7147
3.7065
3.6968
2.7038
2.6948
2.6860
2.5087
2.4913
2.1494
2.1401
2.1335
1.8889
1.8820
1.8751
1.8702
1.7420
1.7201
1.7159
1.7095
1.4672
1.4635
1.2664
1.2509
1.2388
1.2115
1.1918
1.1873
1.1603



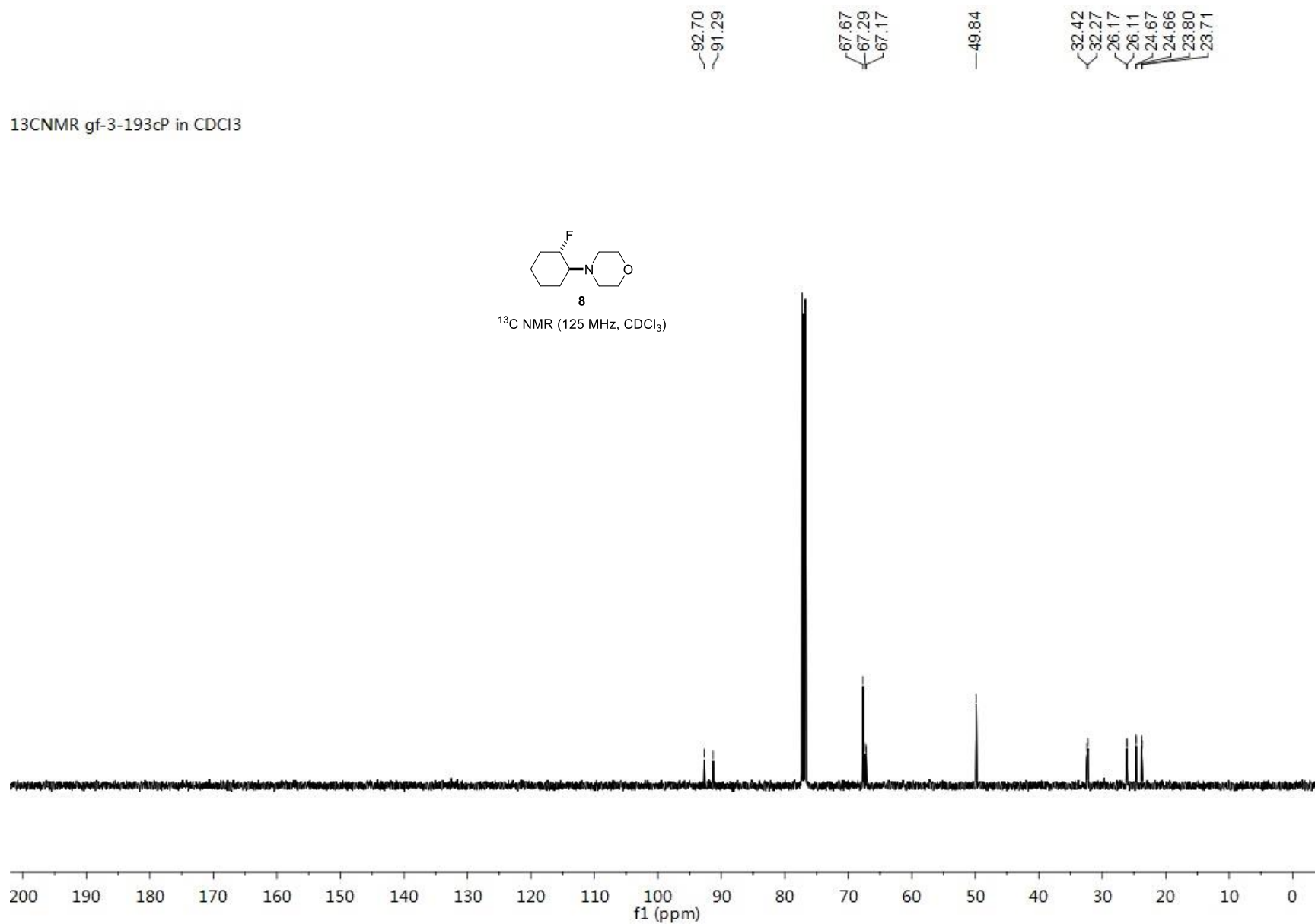
¹H NMR (500 MHz, CDCl₃)



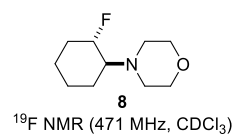
¹³CNMR gf-3-193cP in CDCl₃



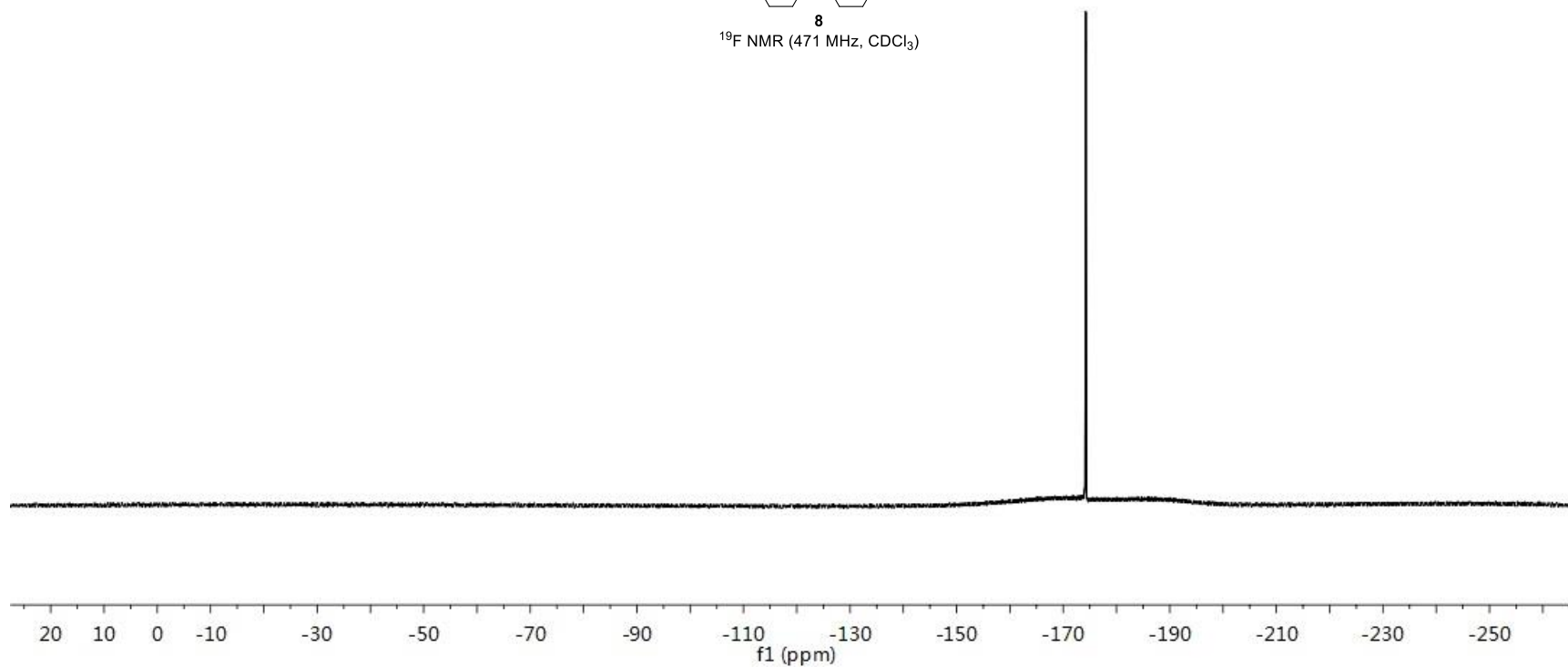
¹³C NMR (125 MHz, CDCl₃)



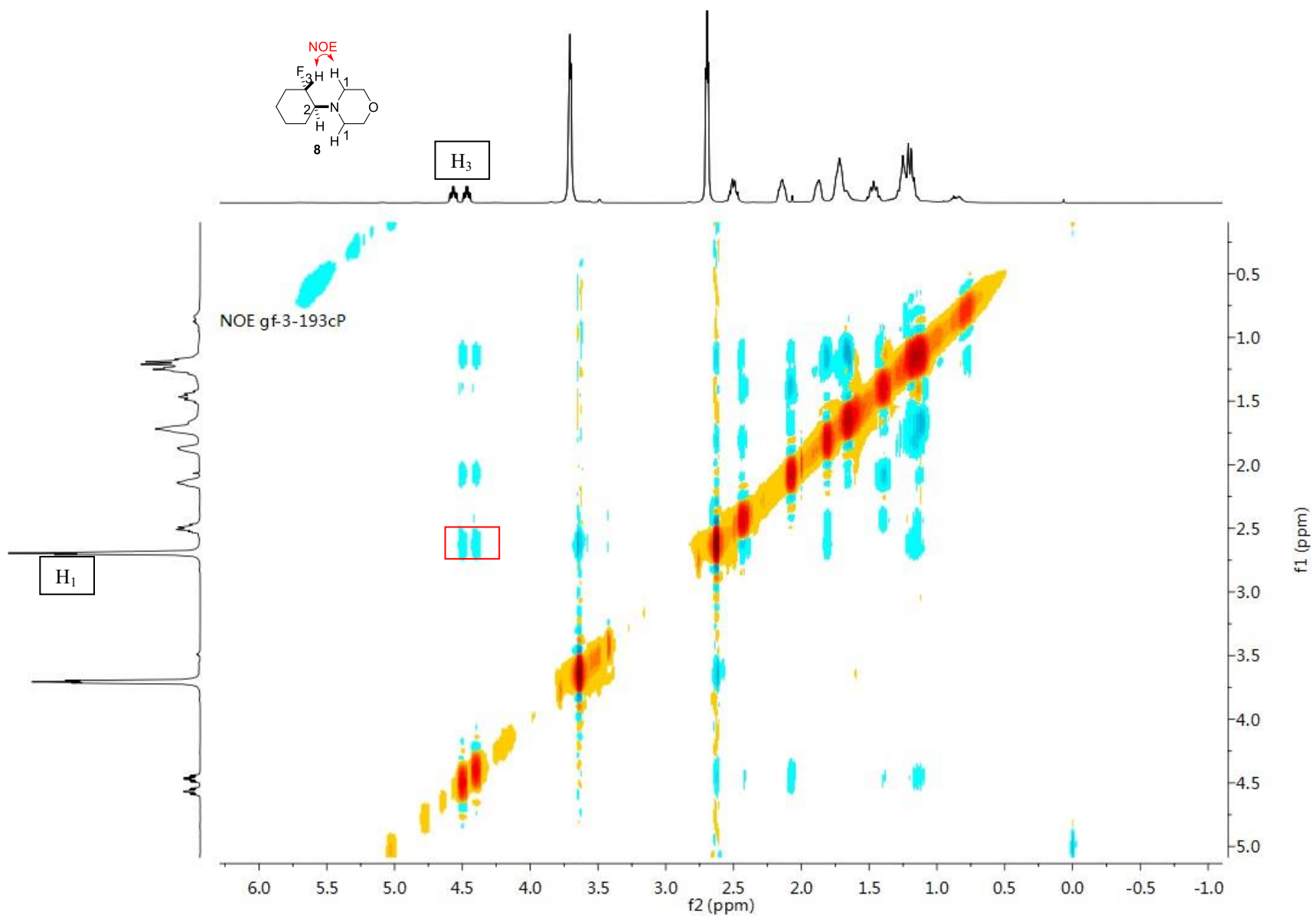
¹⁹F NMR gf-3-193cP in CDCl₃

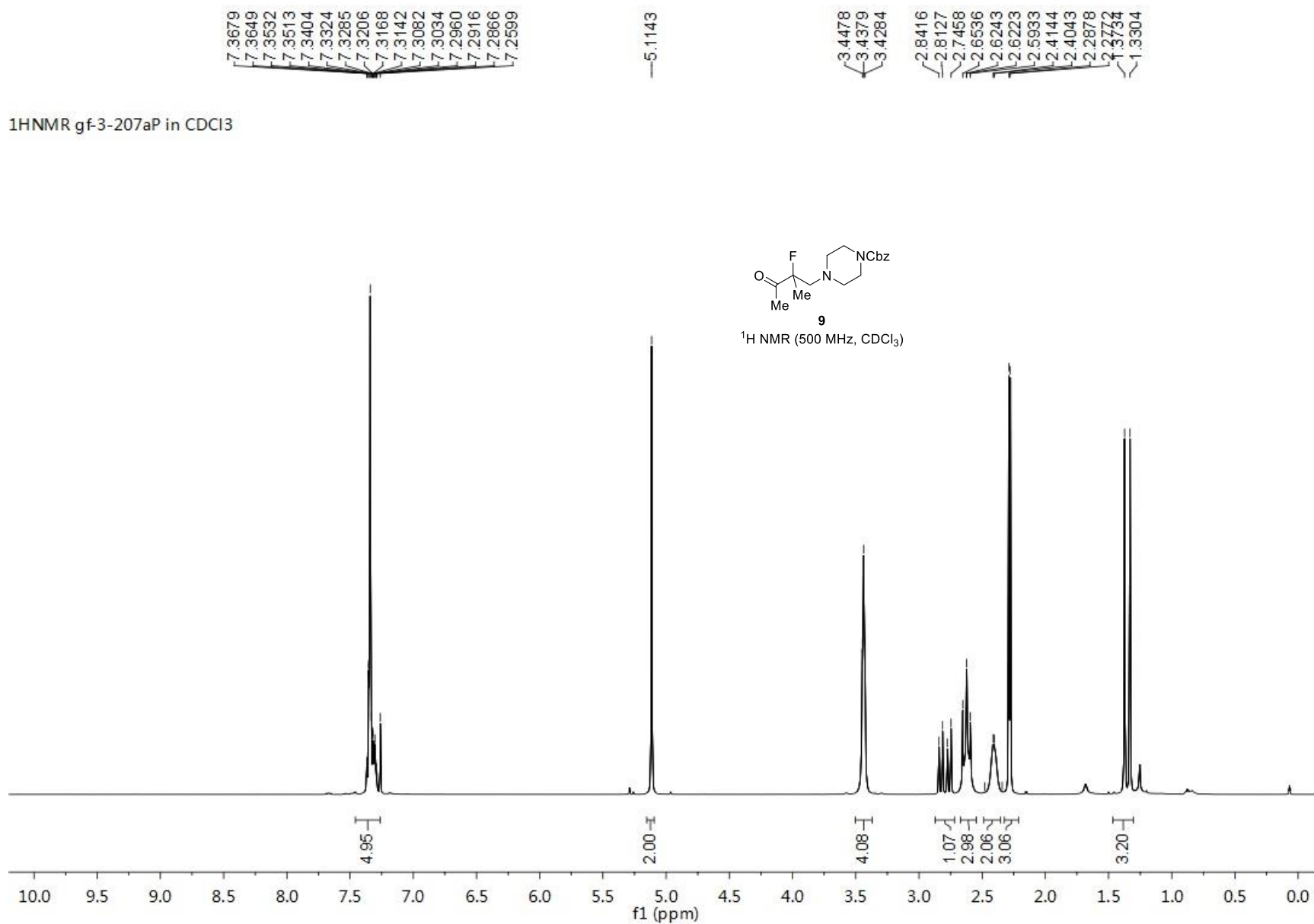


— -174.2123



S252





211.02
210.78

155.20

136.74

128.50
128.02
127.88

103.40
101.93

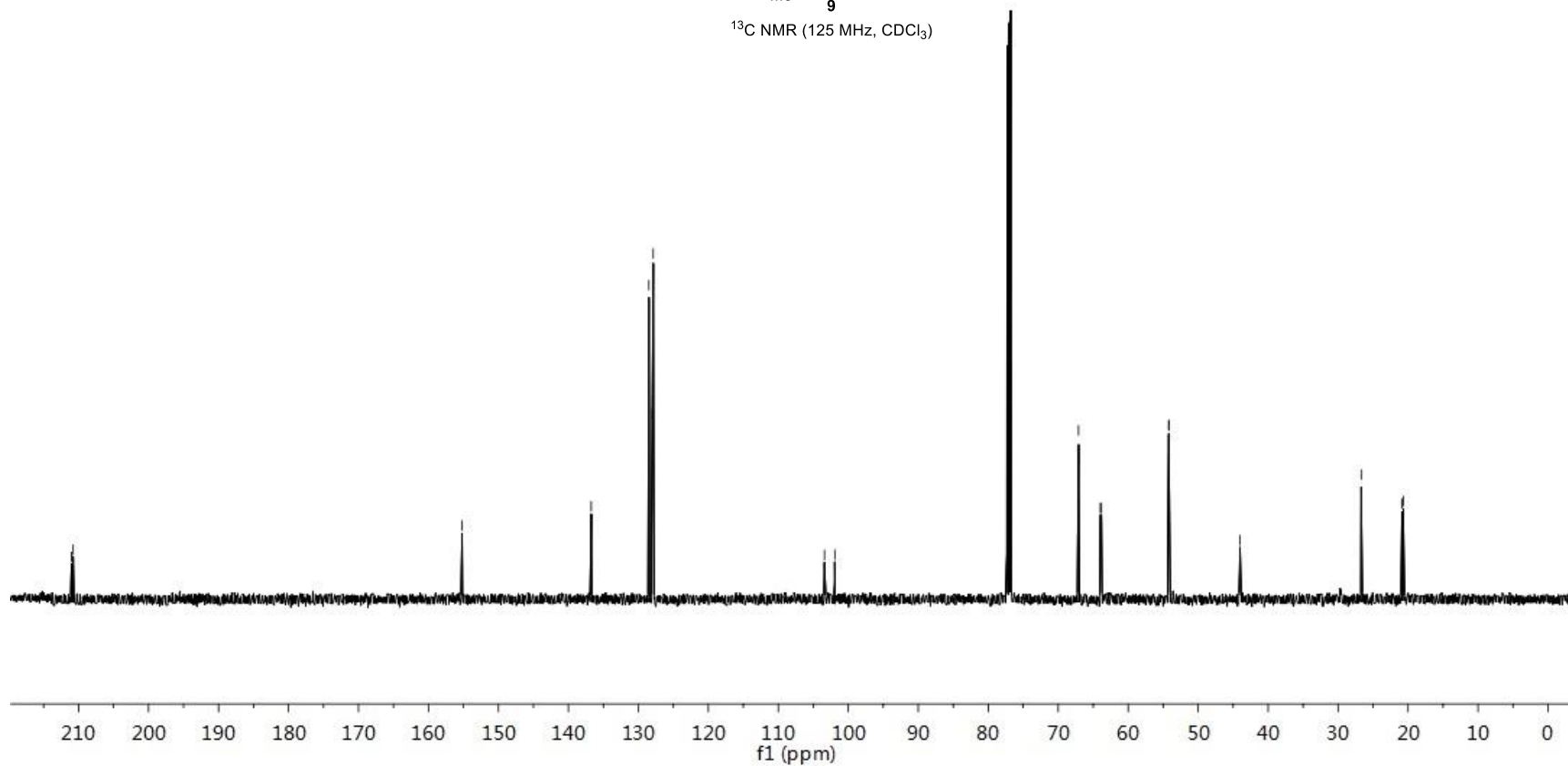
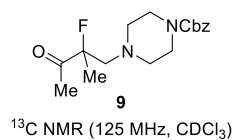
67.10
63.98
63.82

54.20
54.19

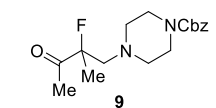
44.01

26.69
20.87
20.68

^{13}C NMR gf-3-207aP in CDCl_3

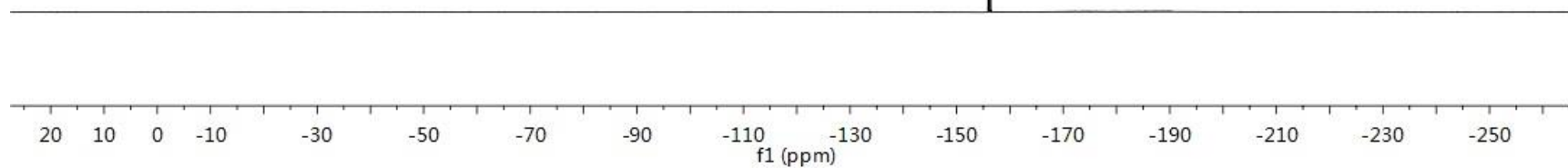


¹⁹F NMR gf-3-207aP in CDCl₃



¹⁹F NMR (471 MHz, CDCl₃)

---156.1125



7.4341
7.4175
7.2602
7.2567
7.2400
7.2223
7.1568
7.1408

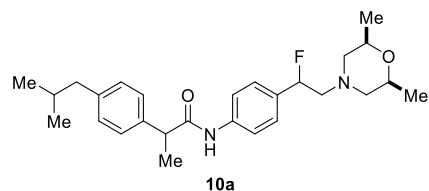
5.6490
5.6353
5.5551
5.5517
5.5380

3.7488
3.7447
3.7361
3.7315
3.7237
3.7170
3.7108
3.7062
3.7032
3.6958
3.6814
3.6673

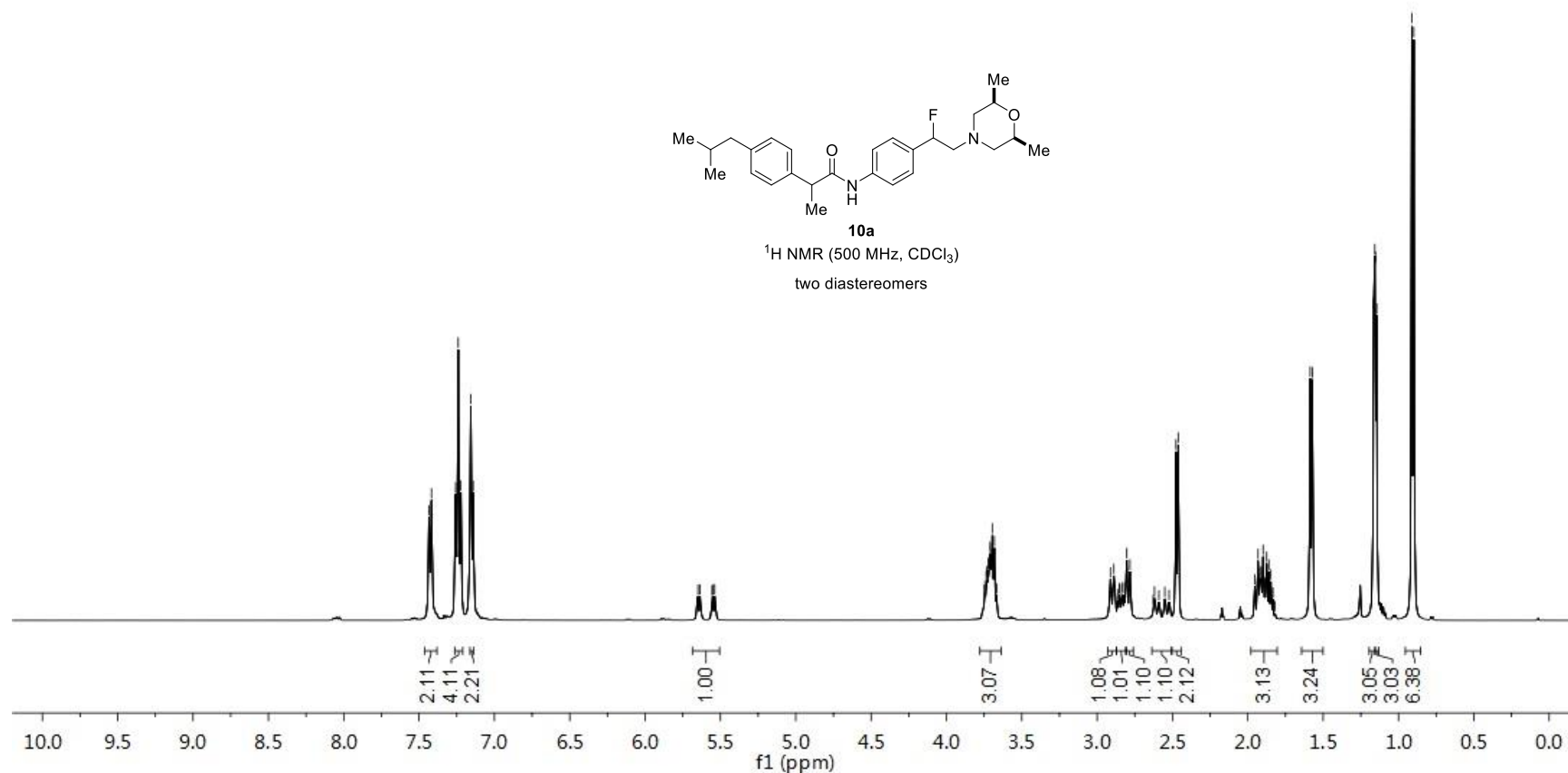
2.9118
2.8892
2.8350
2.8029
2.7821
2.4763
2.4620

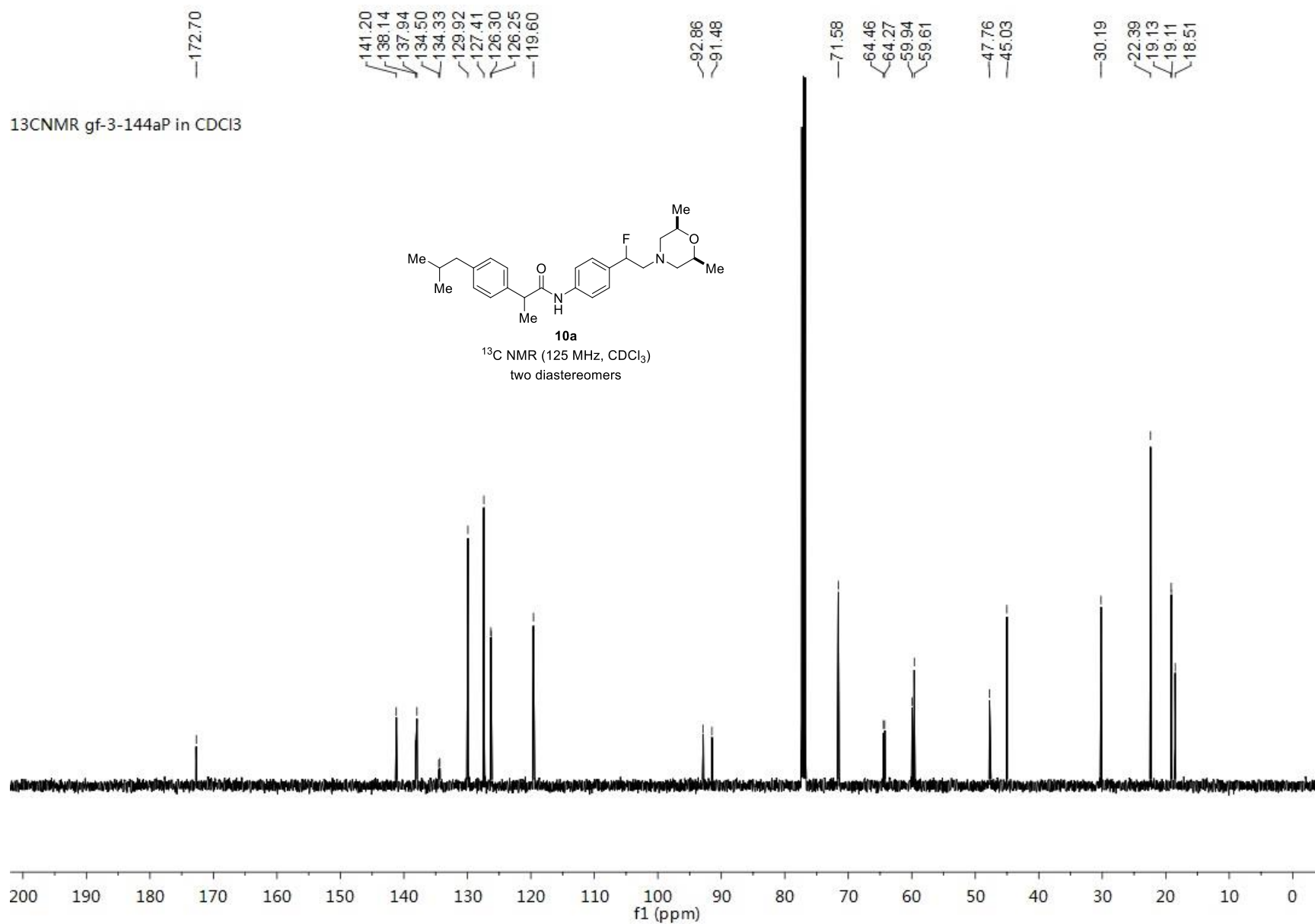
1.9322
1.8987
1.8747
1.5864
1.5721
1.1571
1.1529
1.1447
0.9115
0.8982

¹H NMR of gf-3-144aP in CDCl₃



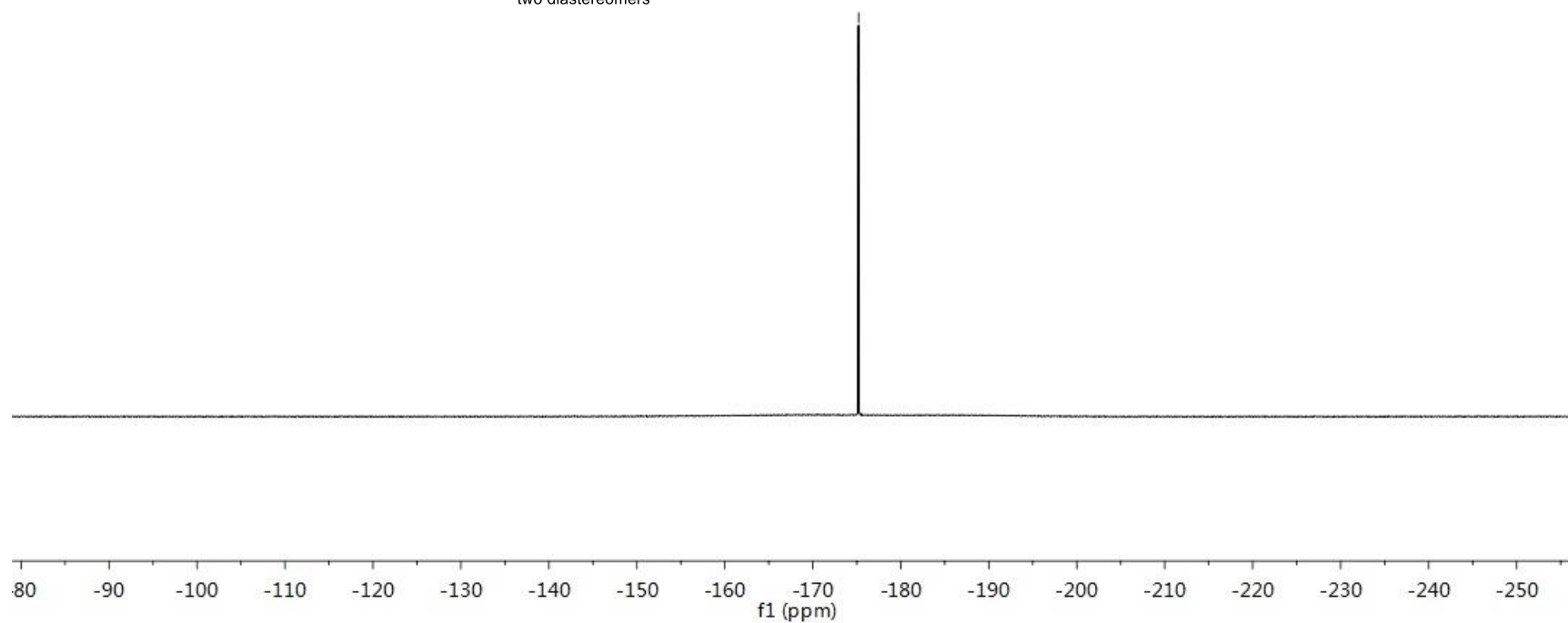
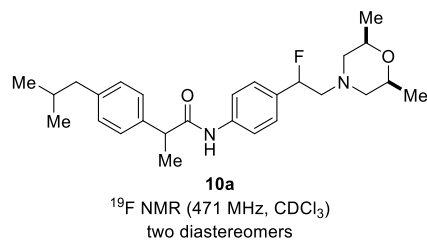
¹H NMR (500 MHz, CDCl₃)
two diastereomers





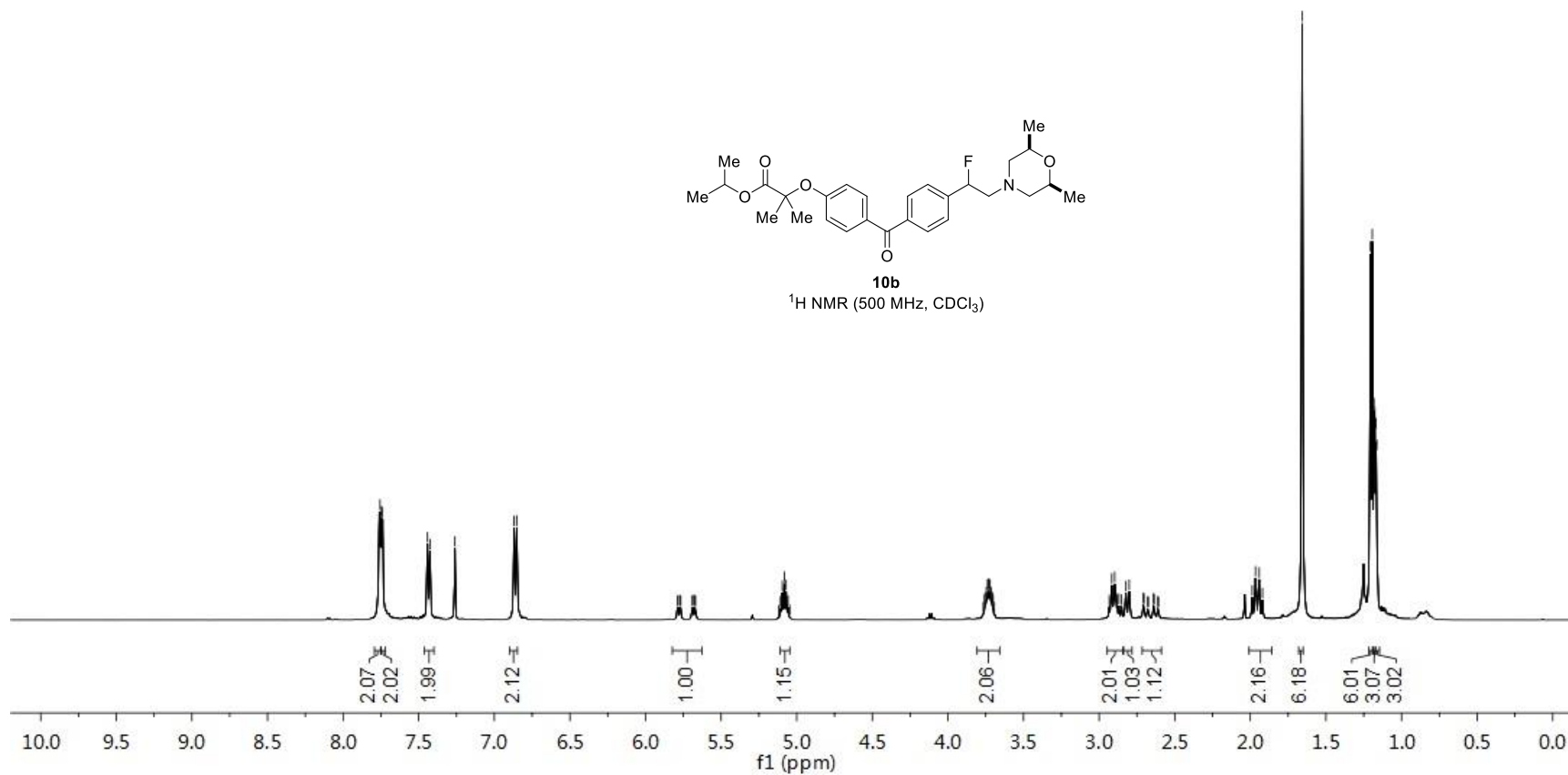
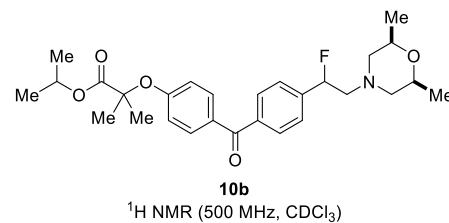
^{19}F NMR gf-3-144aP in CDCl_3

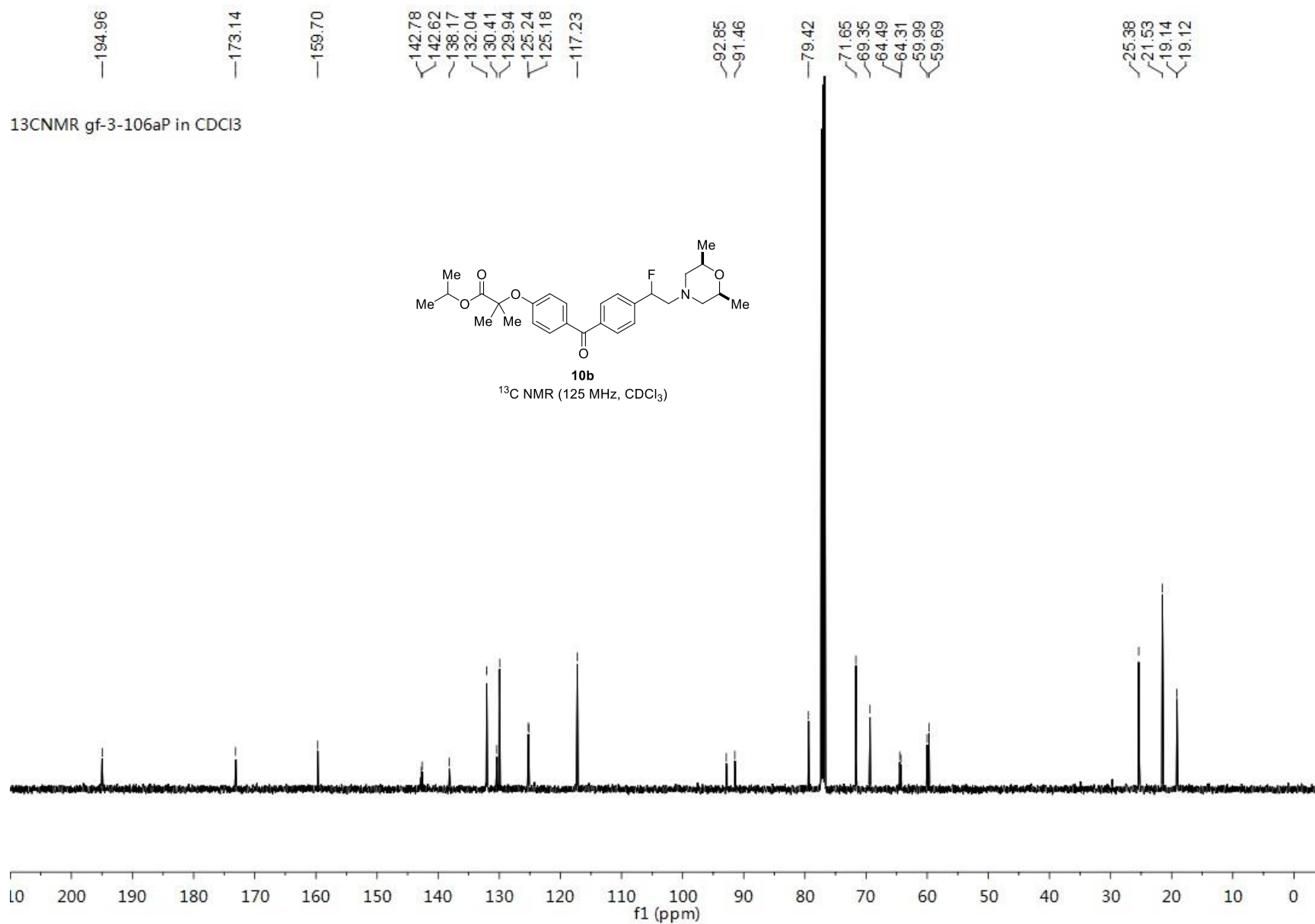
175.1791
175.2007



7.7624
7.7561
7.7466
7.7385
7.4421
7.4259
7.2604
6.8690
6.8514
5.7881
5.7839
5.7712
5.7672
5.6905
5.6864
5.6736
5.6694
5.1074
5.0949
5.0824
5.0699
5.0575
5.0454
3.7540
3.7459
3.7381
3.7338
3.7255
3.7212
3.7133
2.9162
2.9042
2.8967
2.8227
2.8009
2.7076
2.6694
2.6679
1.9663
1.9410
1.9195
1.6573
1.2059
1.1934
1.1824
1.1762
1.1698
1.1636

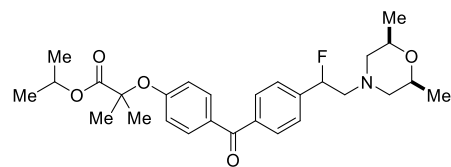
¹H NMR of gf-3-106aP in CDCl₃





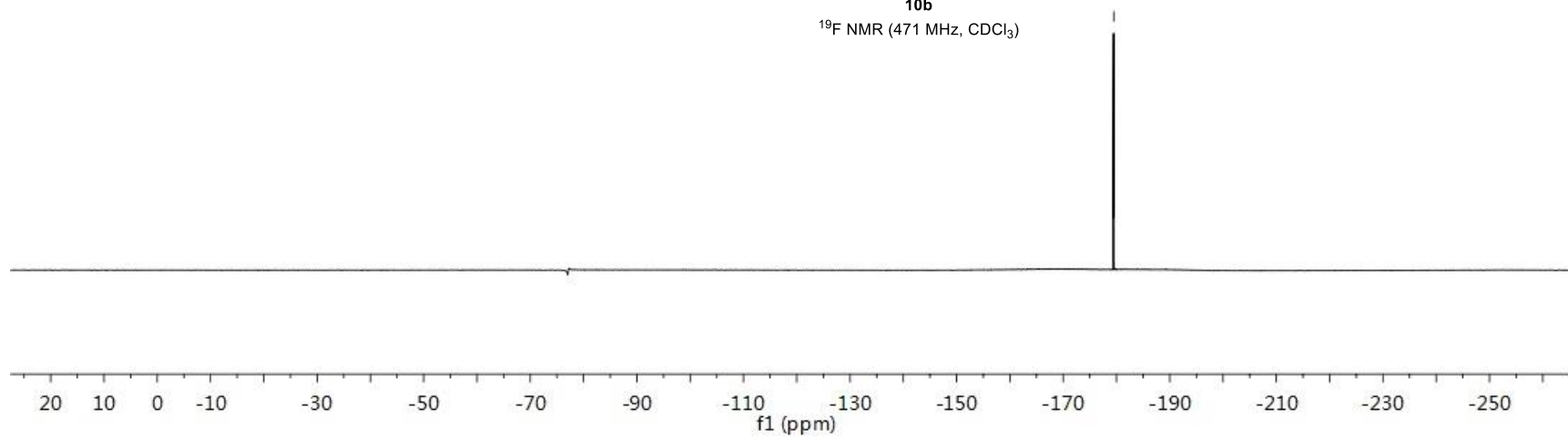
¹⁹F NMR gf-3-106aP in CDCl₃

---179.4938



10b

¹⁹F NMR (471 MHz, CDCl₃)



7.6743
7.6579
7.4929
7.4764
7.4086
7.3922
7.3751
7.2597
7.2501
7.2338
6.9366
6.8777
6.8597
6.7212
6.7067

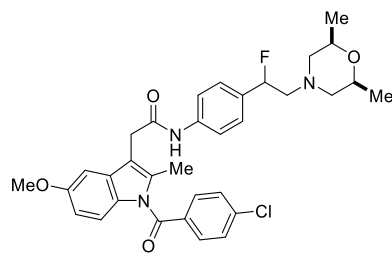
5.6443
5.6292
5.5470
5.5321

3.8000
3.7425
3.7146
3.7112
3.7022
3.6994
3.6696

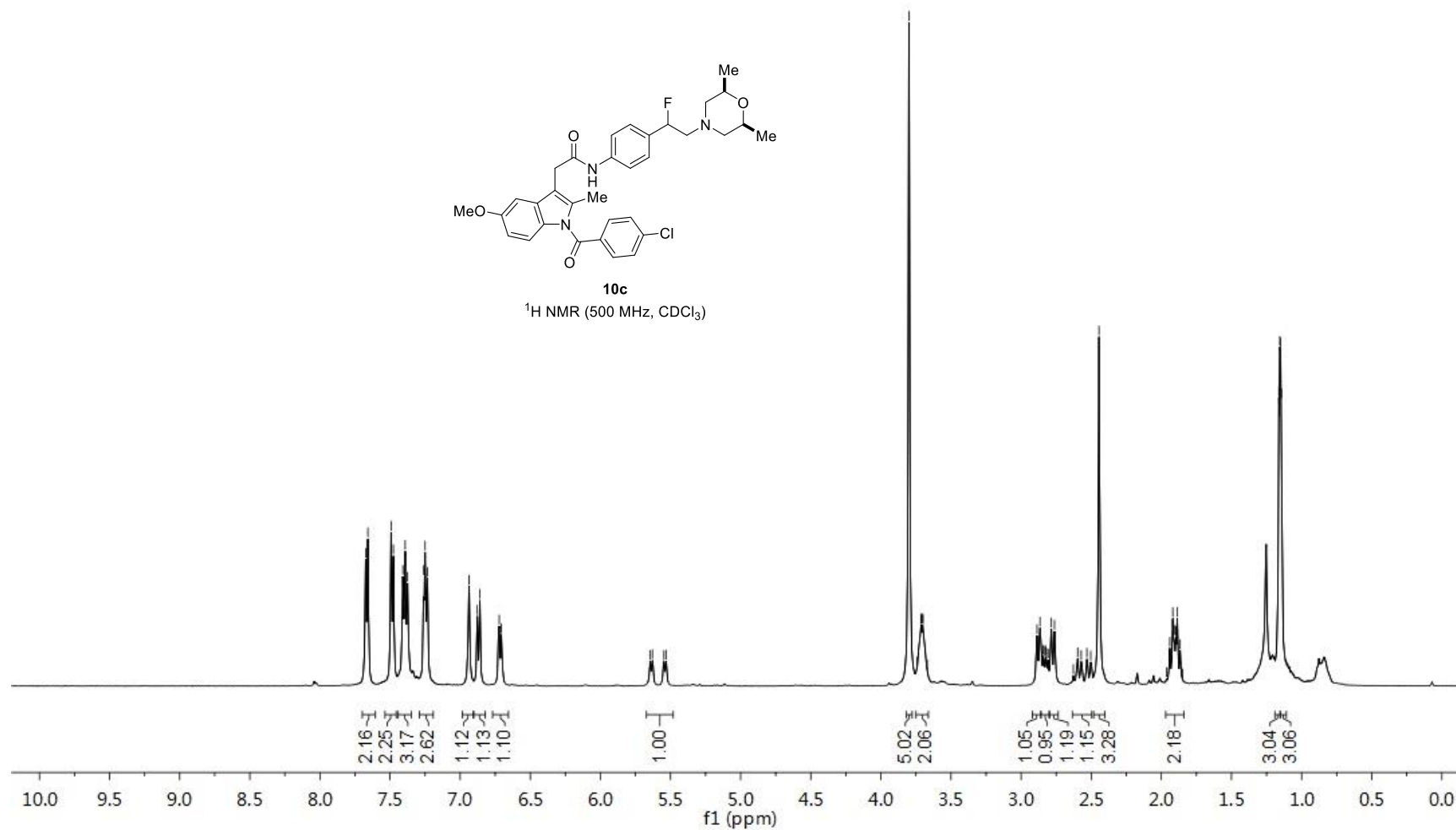
2.8866
2.8652
2.8416
2.8241
2.7843
2.7622
2.4445

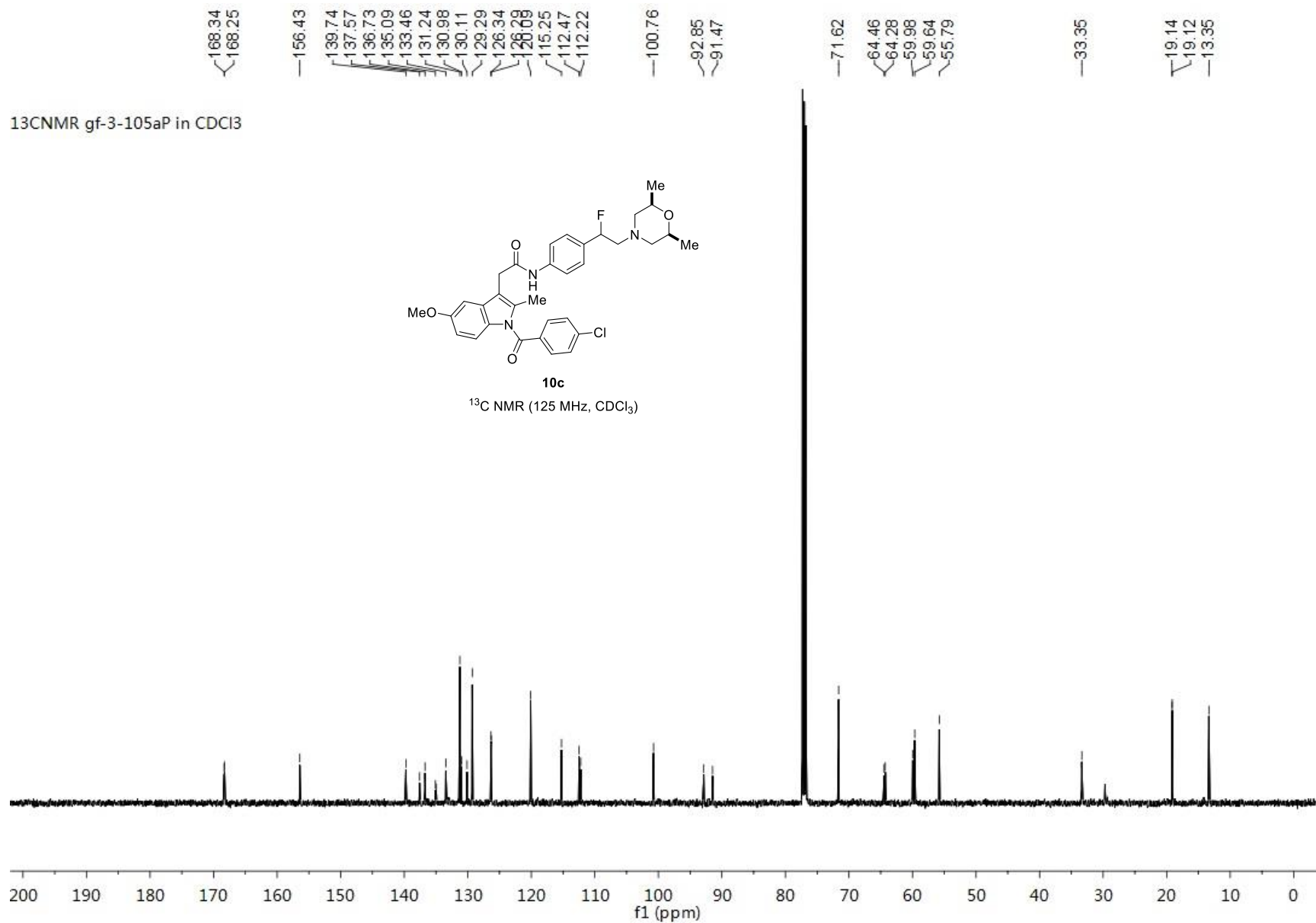
1.9399
1.9185
1.9082
1.8971
1.8820
1.1555
1.1503
1.1438

¹H NMR of gf-3-105aP in CDCl₃

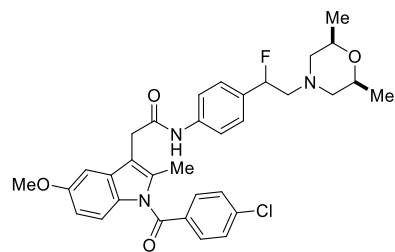


¹H NMR (500 MHz, CDCl₃)



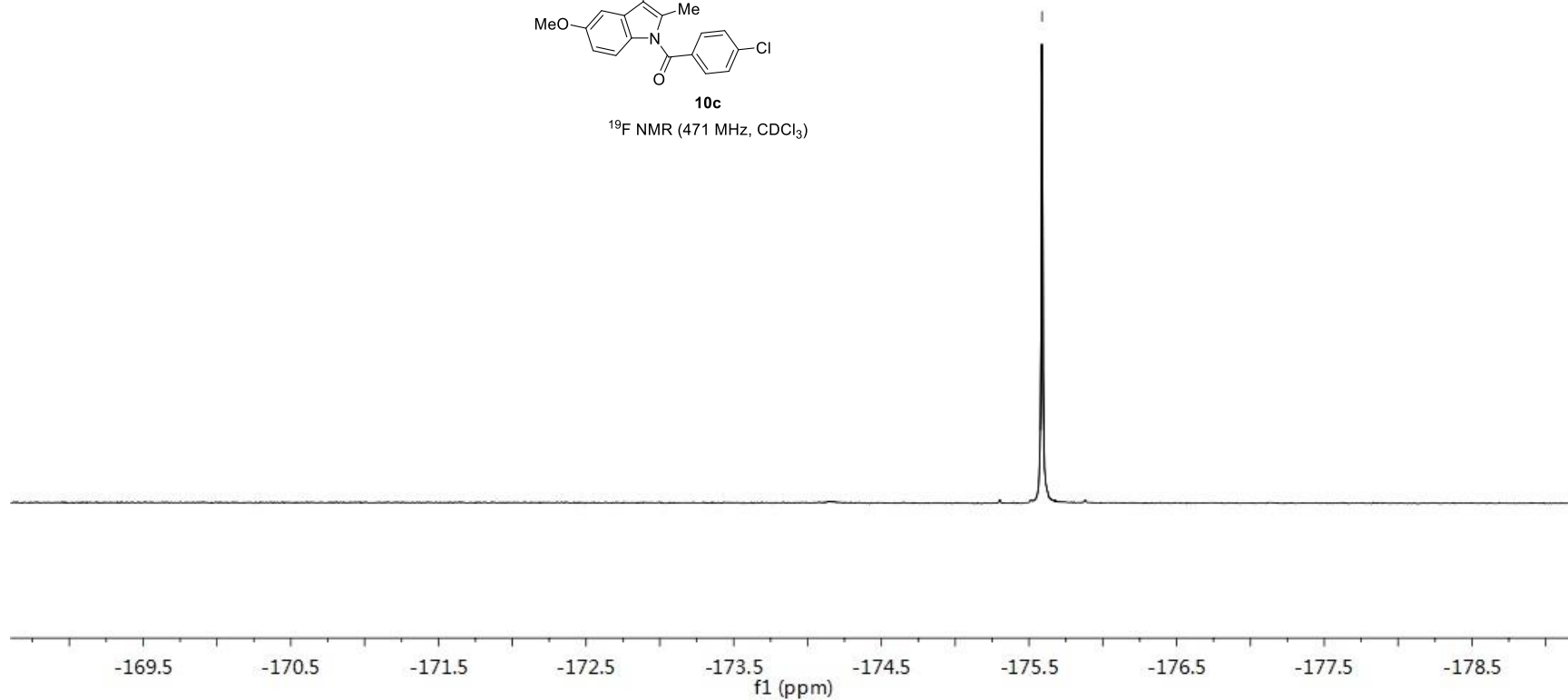


¹⁹F NMR gf-3-105aP in CDCl₃



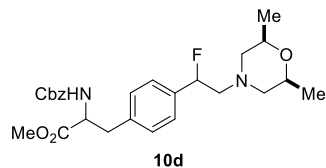
¹⁹F NMR (471 MHz, CDCl₃)

-175.5886

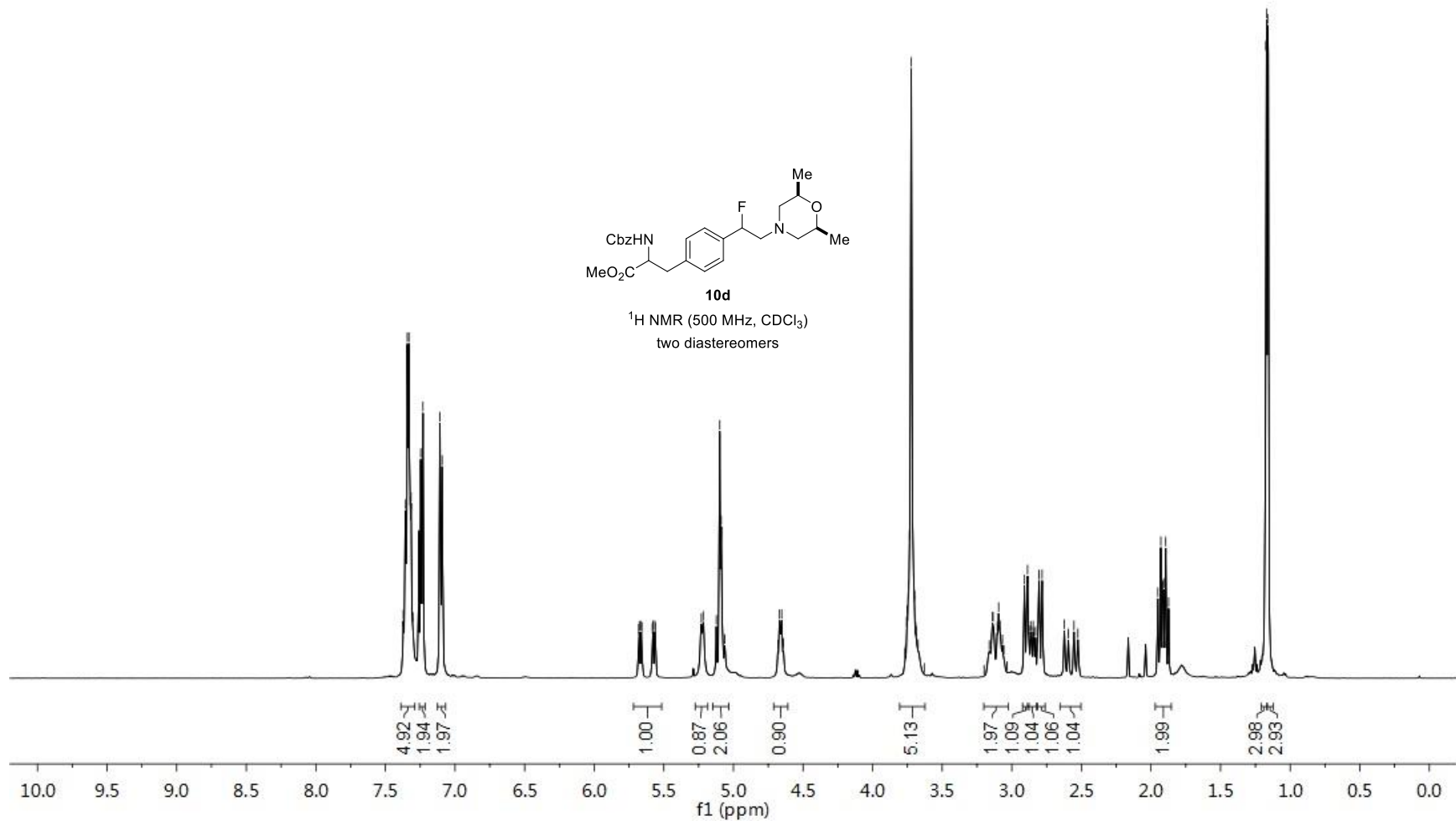


7.3714
7.3548
7.3432
7.3317
7.3220
7.3162
7.3028
7.2465
7.2307
7.1081
7.0925
5.6800
5.6762
5.6624
5.6586
5.5822
5.5784
5.5646
5.5610
5.2305
5.2170
5.1228
5.0982
5.0876
5.0625
4.6804
4.6688
4.6535
4.6418
3.7556
3.7513
3.7432
3.7352
3.7224
3.7033
3.6988
3.6904
3.6866
3.6729
3.1618
3.1381
3.1352
3.1046
3.0946
3.0844
3.0675
2.9091
2.8870
2.8757
2.8643
2.8585
2.8466
2.8412
2.8298
2.8043
2.7823
2.6229
2.5943
2.5534
2.5249
1.9510
1.9296
1.9153
1.9084
1.8944
1.8733
1.1751
1.1700
1.1625
1.1575

¹H NMR of **10d** in CDCl₃



¹H NMR (500 MHz, CDCl₃)
two diastereomers



—171.84

—155.60

137.78

137.62

136.20

136.07

129.45

128.56

128.26

128.12

125.84

125.79

93.05

91.67

71.66

71.64

67.03

64.60

64.42

60.07

59.64

54.71

52.40

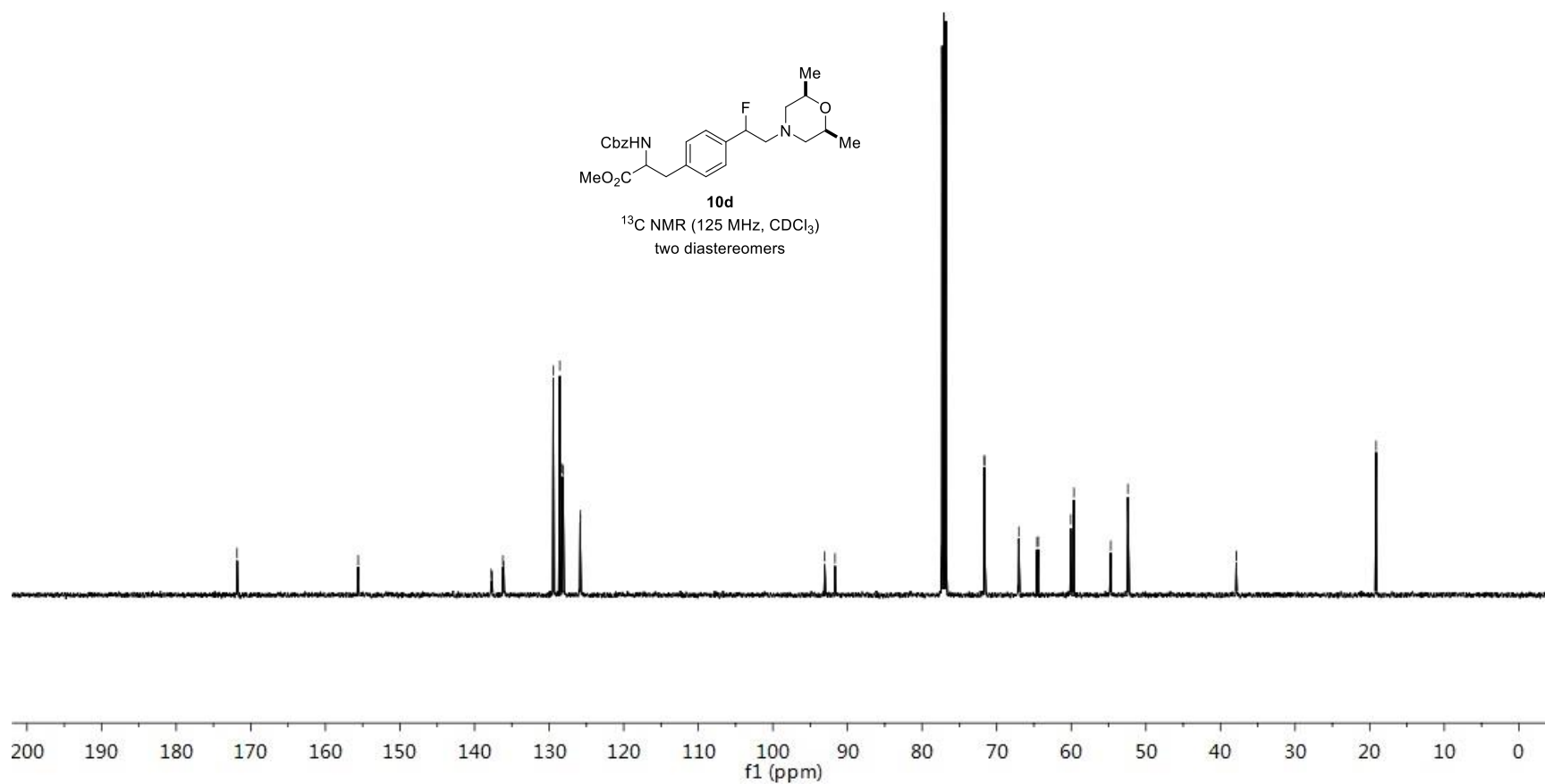
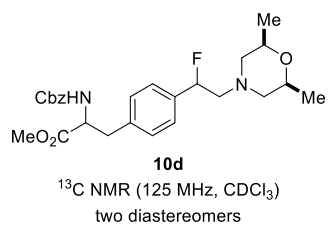
37.88

37.87

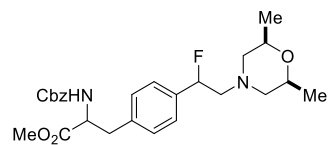
19.16

19.13

¹³CNMR gf-3-112aP in CDCl₃

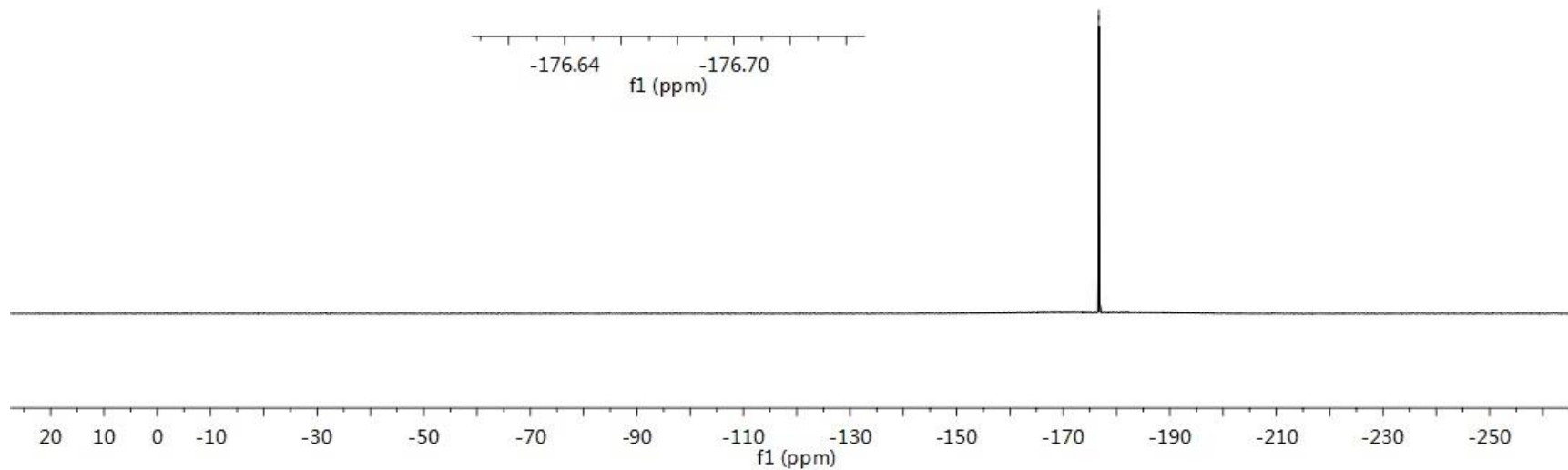
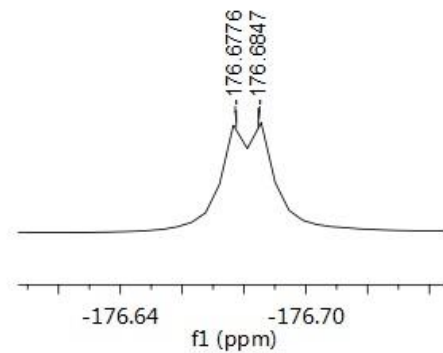


¹⁹F NMR gf-3-112aP in CDCl₃



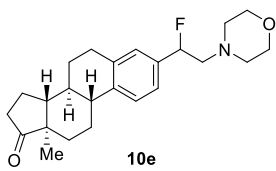
¹⁹F NMR (471 MHz, CDCl₃)
two diastereomers

176.6776
176.6847

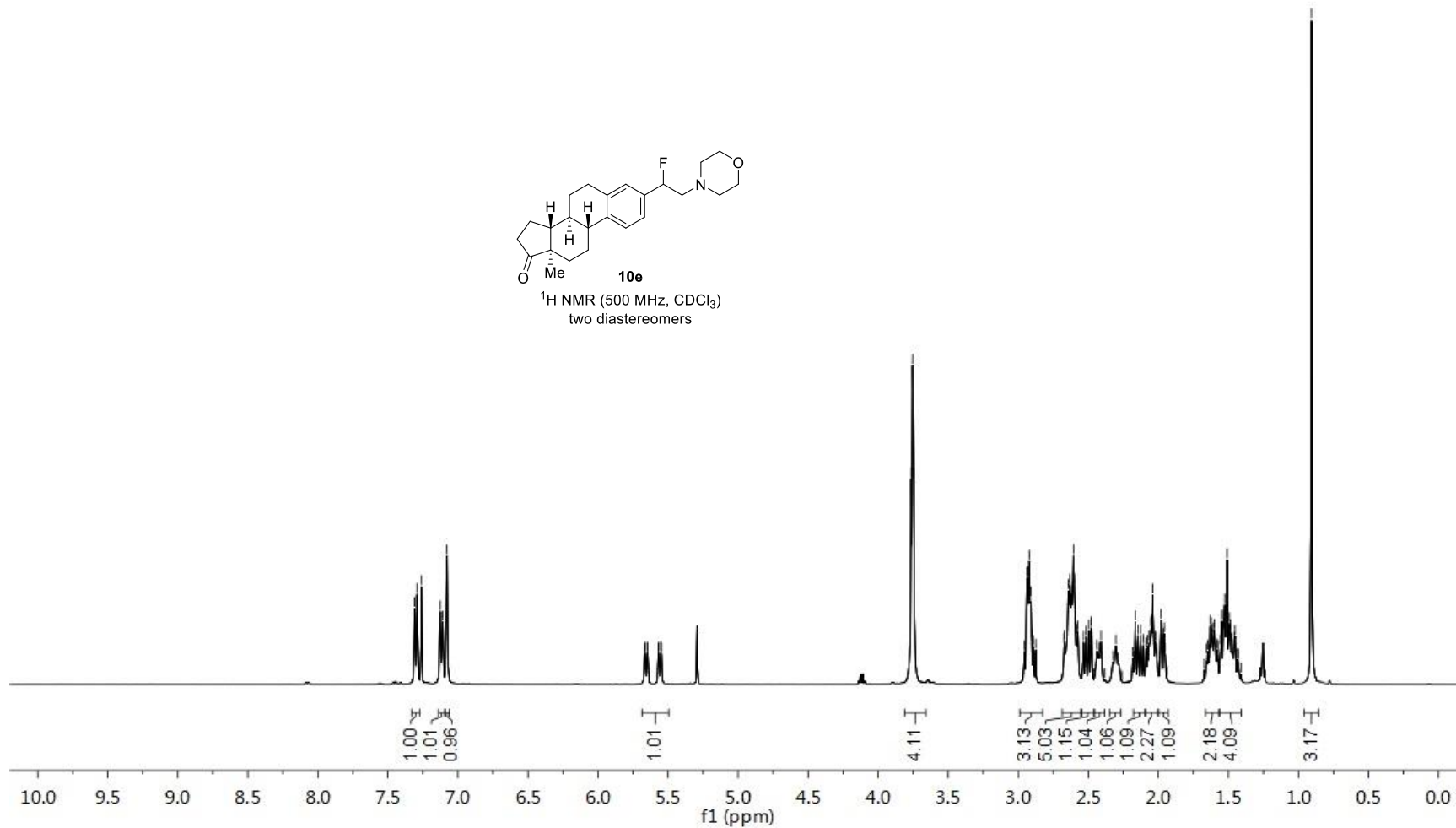


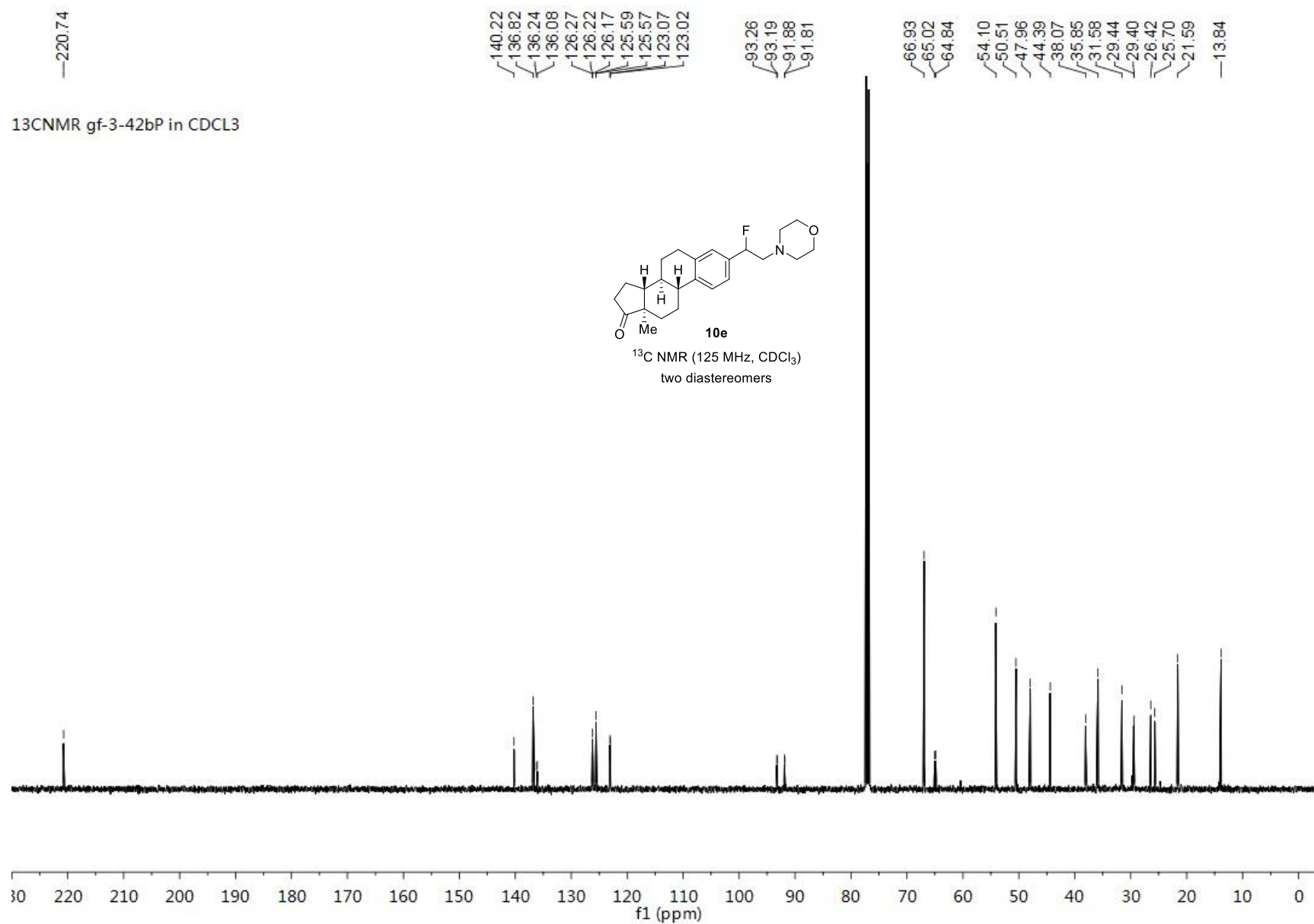
7.3088
7.2927
7.2598
7.1252
7.1091
7.0785
3.7641
3.7548
3.7455
2.9561
2.9384
2.9285
2.9217
2.9102
2.9042
2.8933
2.8753
2.6739
2.6702
2.6412
2.6308
2.6162
2.6054
2.5993
2.5837
2.5753
2.5338
2.5161
2.4955
2.4785
2.4385
2.4083
2.3028
2.1642
2.1459
2.1262
2.1086
2.0892
2.0781
2.0651
2.0538
2.0482
2.0400
2.0230
2.0175
1.9812
1.9760
1.9625
1.9574
1.6479
1.6304
1.6233
1.6176
1.6125
1.6064
1.5999
1.5786
1.5567
1.5478
1.5362
1.5261
1.5095
1.5008
1.4917
1.4855
1.4762
1.4711
1.4585
1.4527
0.9069

¹H NMR of **10e** in CDCl₃



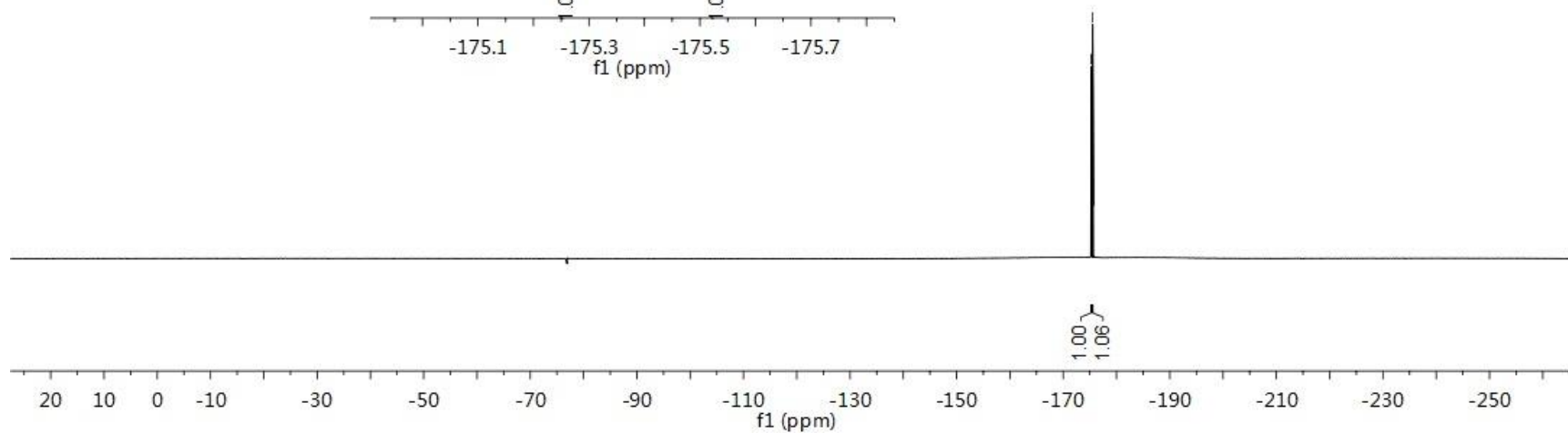
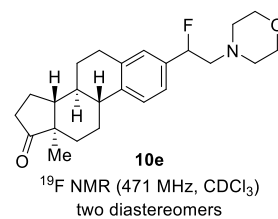
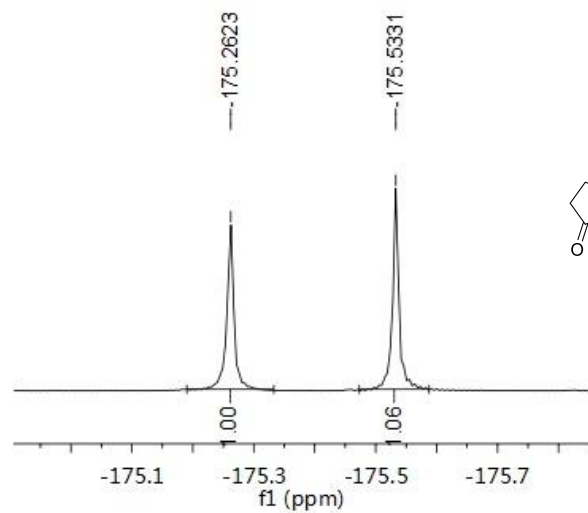
¹H NMR (500 MHz, CDCl₃)
two diastereomers





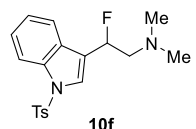
19FNMR gf-3-42bP in CDCl3

175.2623
175.5331

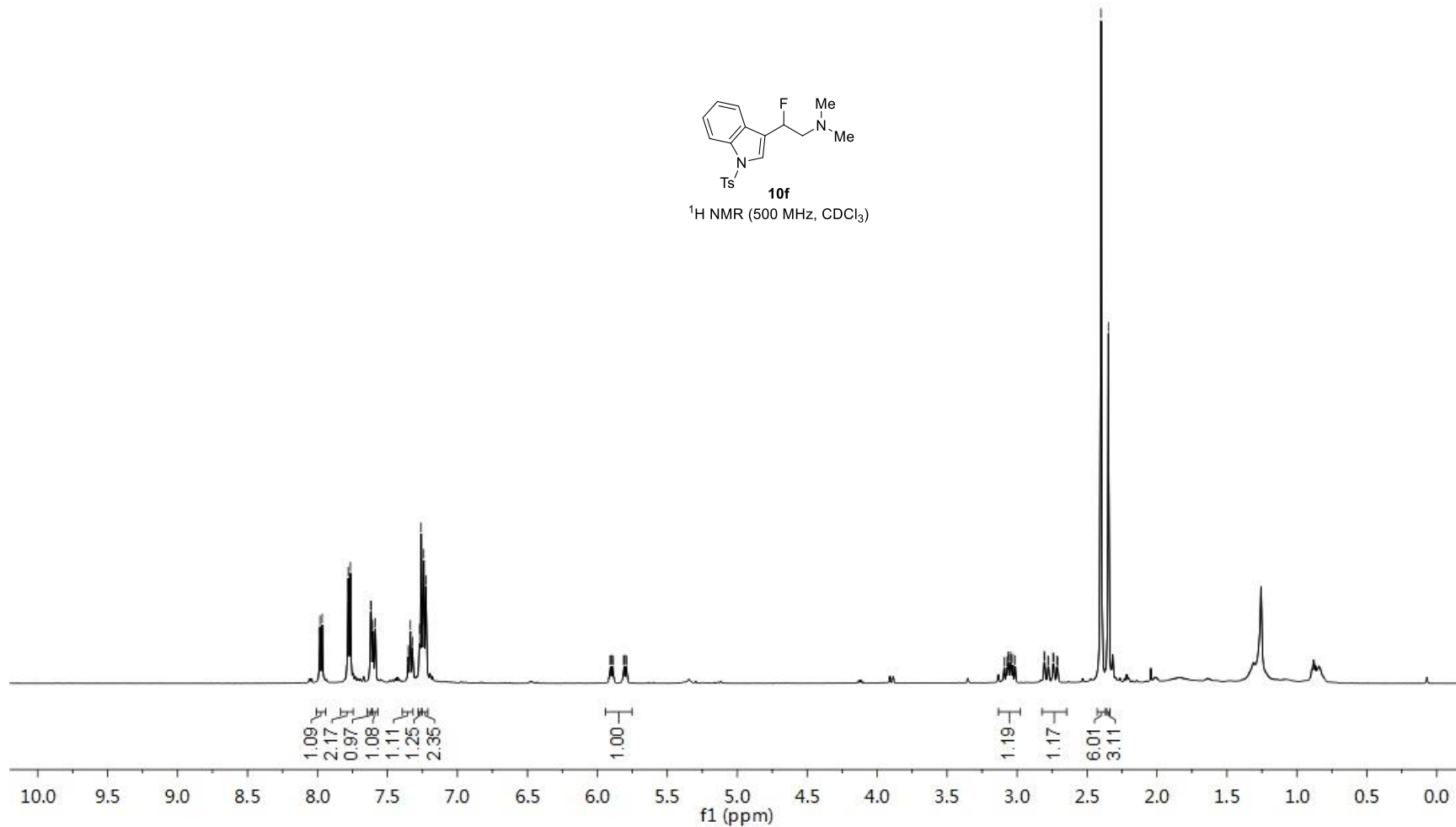


7.9827
7.9661
7.7812
7.7647
7.6198
7.6153
7.6026
7.5869
7.3515
7.3362
7.3209
7.2690
7.2598
7.2549
7.2407
7.2249
5.9090
5.9047
5.8921
5.8876
5.8121
5.8077
5.7952
5.7907
3.0928
3.0755
3.0647
3.0589
3.0476
3.0417
3.0308
3.0136
2.8073
2.8022
2.7793
2.7743
2.7431
2.7380
2.7151
2.7101
2.3998
2.3475

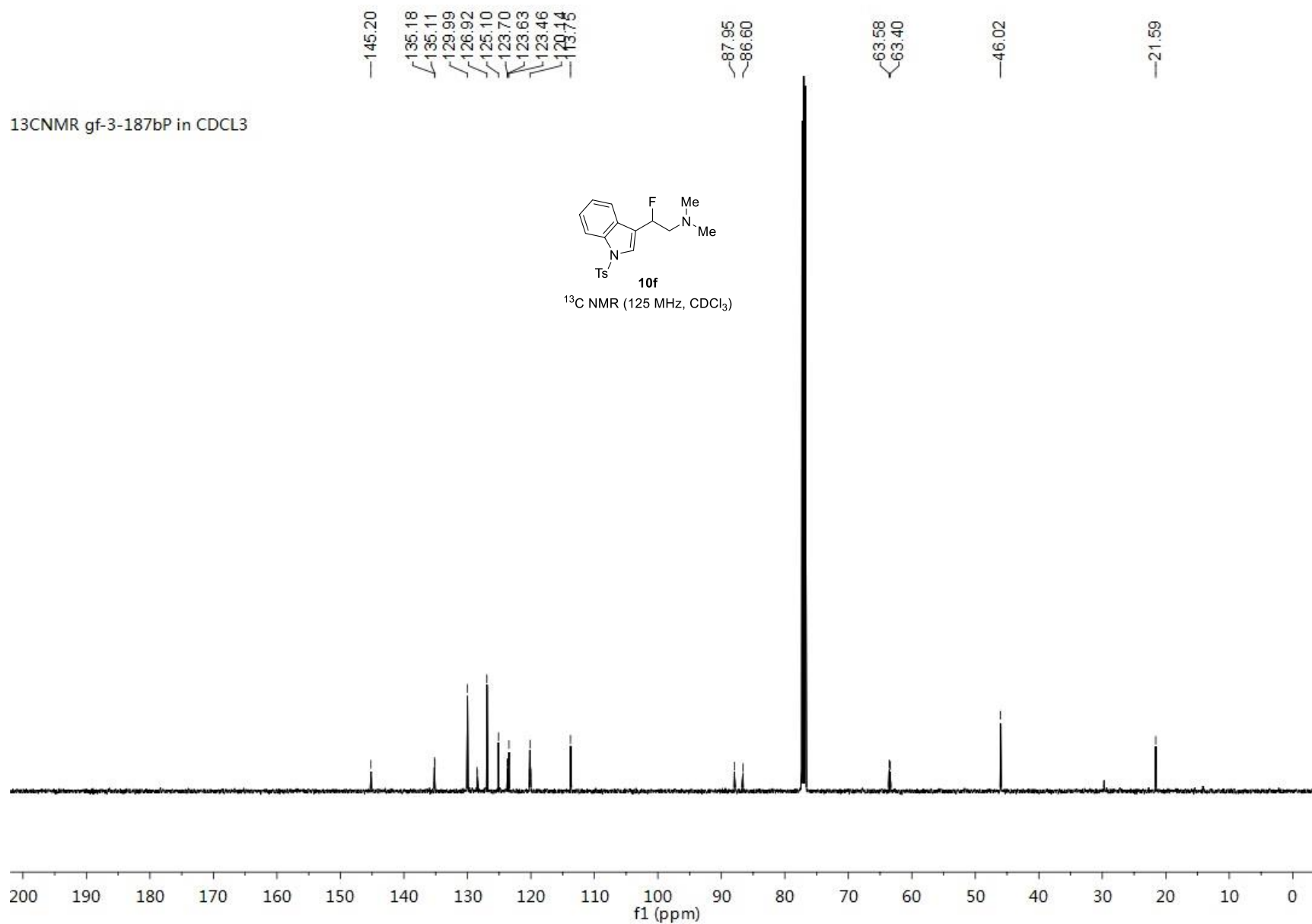
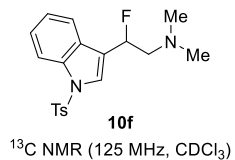
¹H NMR of gf-3-187bP in CDCl₃



¹H NMR (500 MHz, CDCl₃)

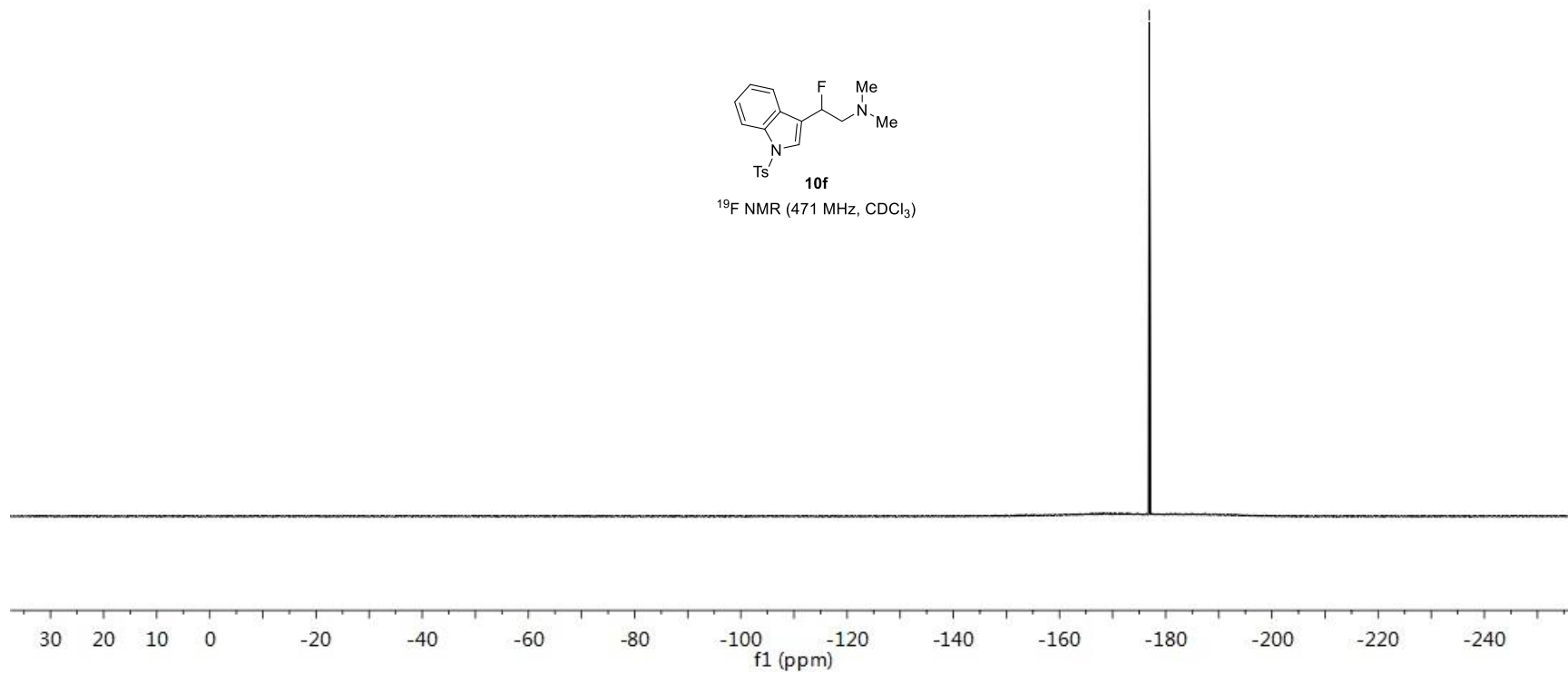
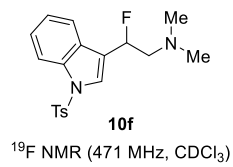


¹³CNMR gf-3-187bP in CDCl₃



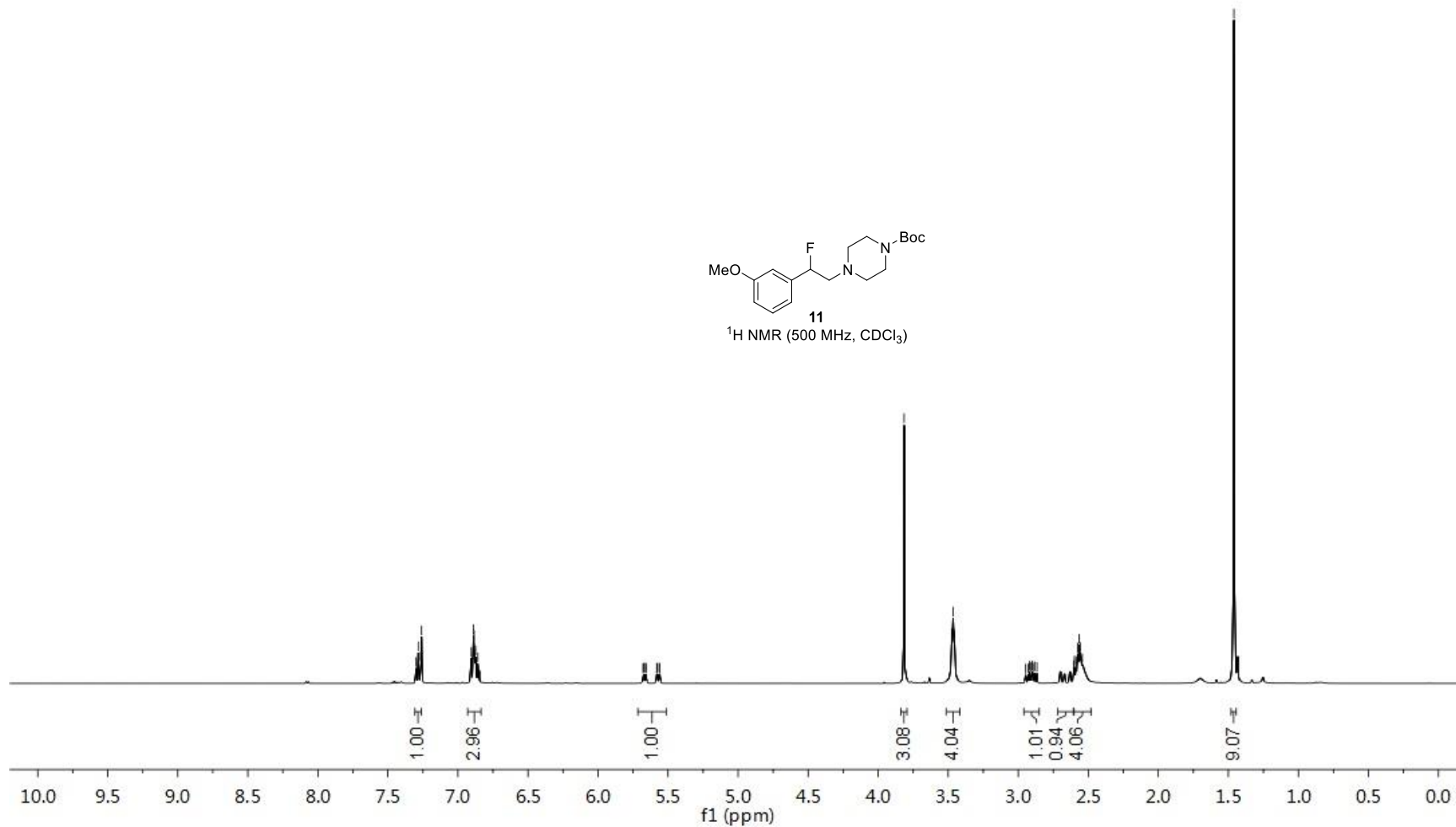
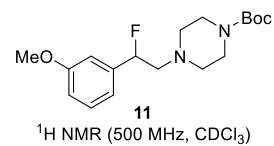
¹⁹F NMR gf-3-187bP in CDCl₃

---176.8845



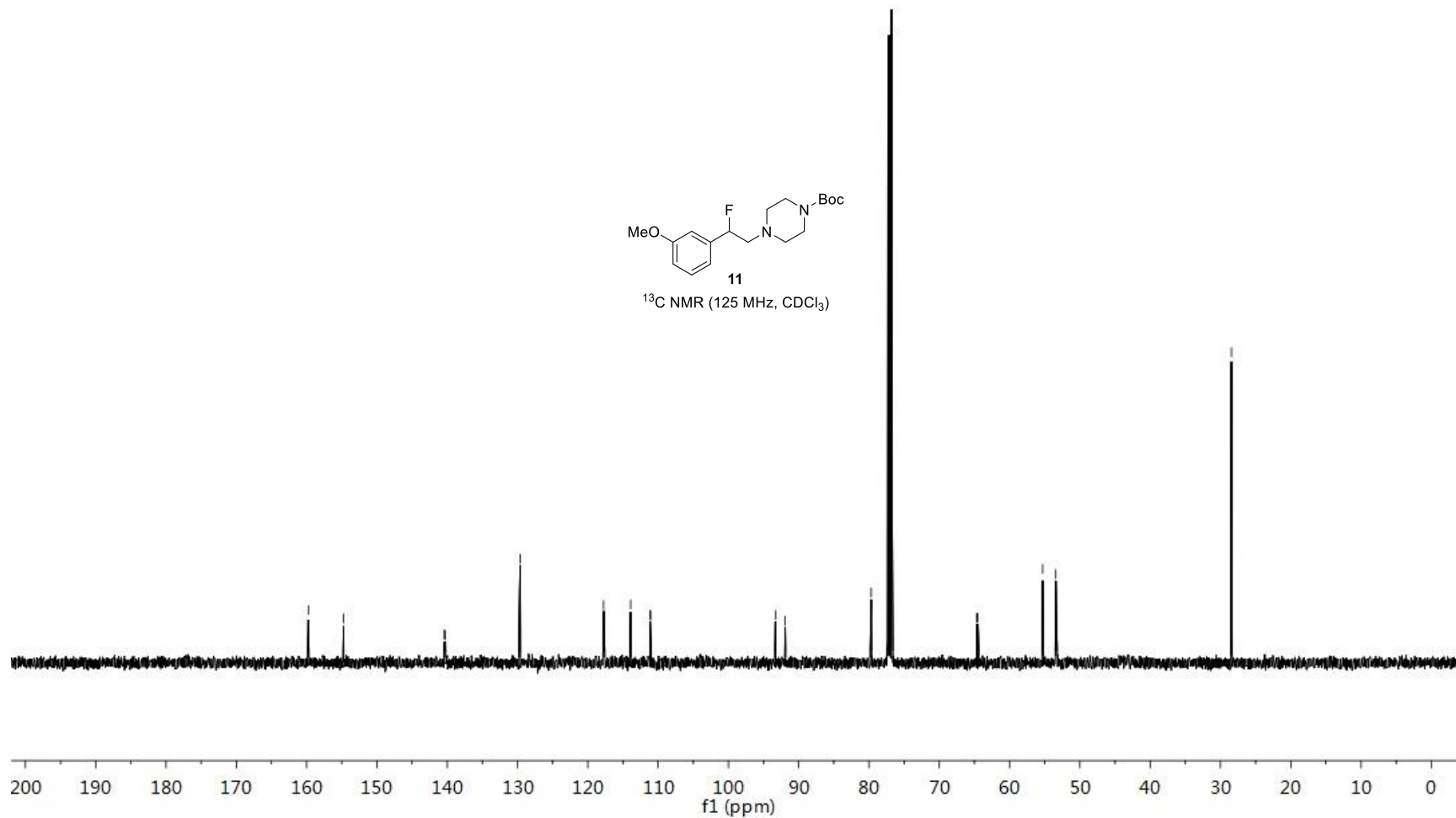
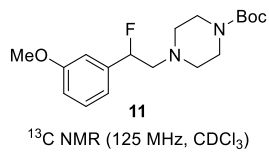
7.2983
7.2828
7.2671
7.2601
6.9047
6.8885
6.8842
6.8745
6.8589
6.8587
5.6743
5.6614
5.6570
5.5809
5.5765
5.5636
5.5592
3.8149
3.4746
3.4650
3.4560
2.9189
2.9015
2.8838
2.8664
2.6042
2.5991
2.5878
2.5760
2.5657
2.5555
2.5482
1.4604

¹H NMR of gf-3-224aP2 in CDCl₃



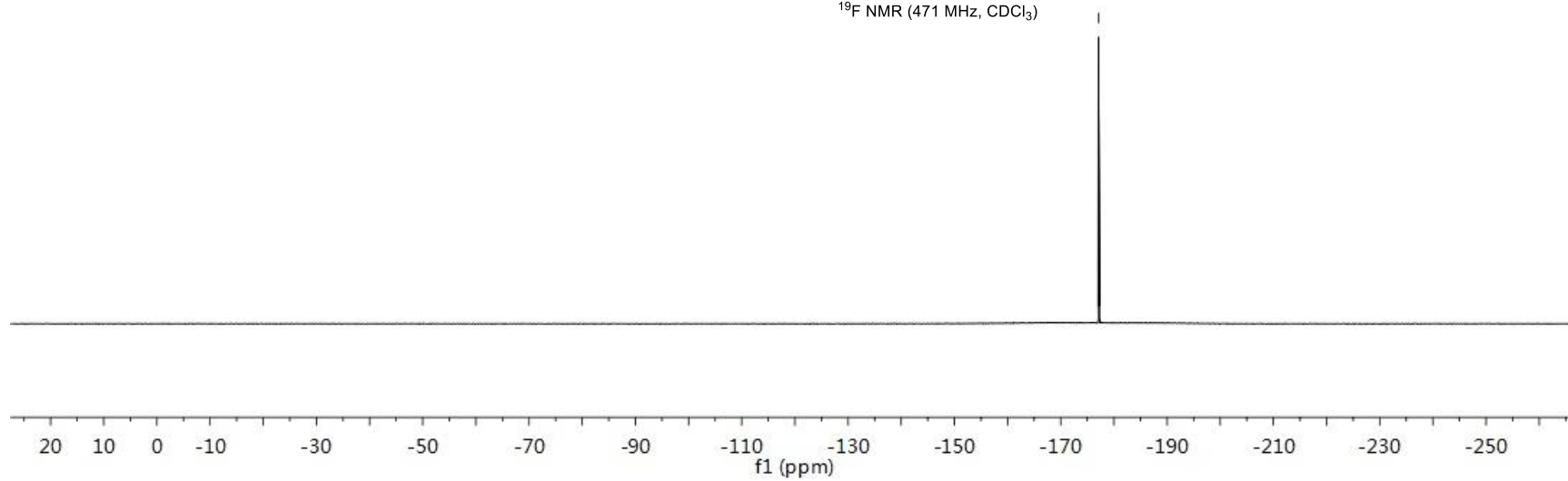
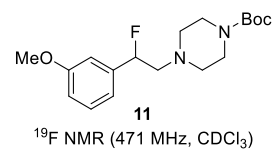
—159.74
—154.74
—140.42
—140.26
—129.62
—117.75
—117.70
—113.90
—111.12
—111.06
—93.30
—91.91
—79.68
—64.68
—64.50
—55.29
—53.43
—28.44

^{13}C NMR gf-3-224aP2 in CDCl_3

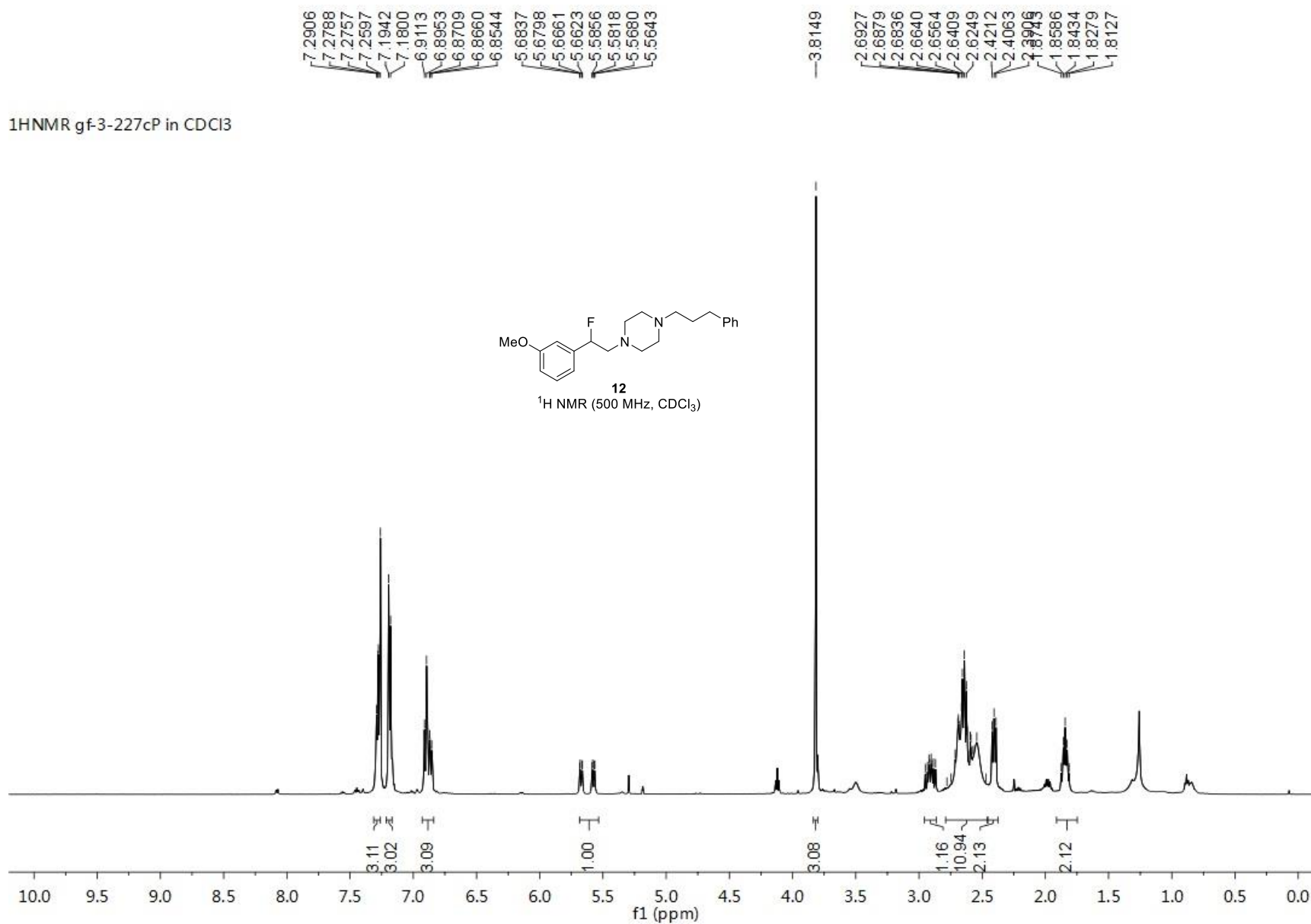


¹⁹F NMR gf-3-224aP2 in CDCl₃

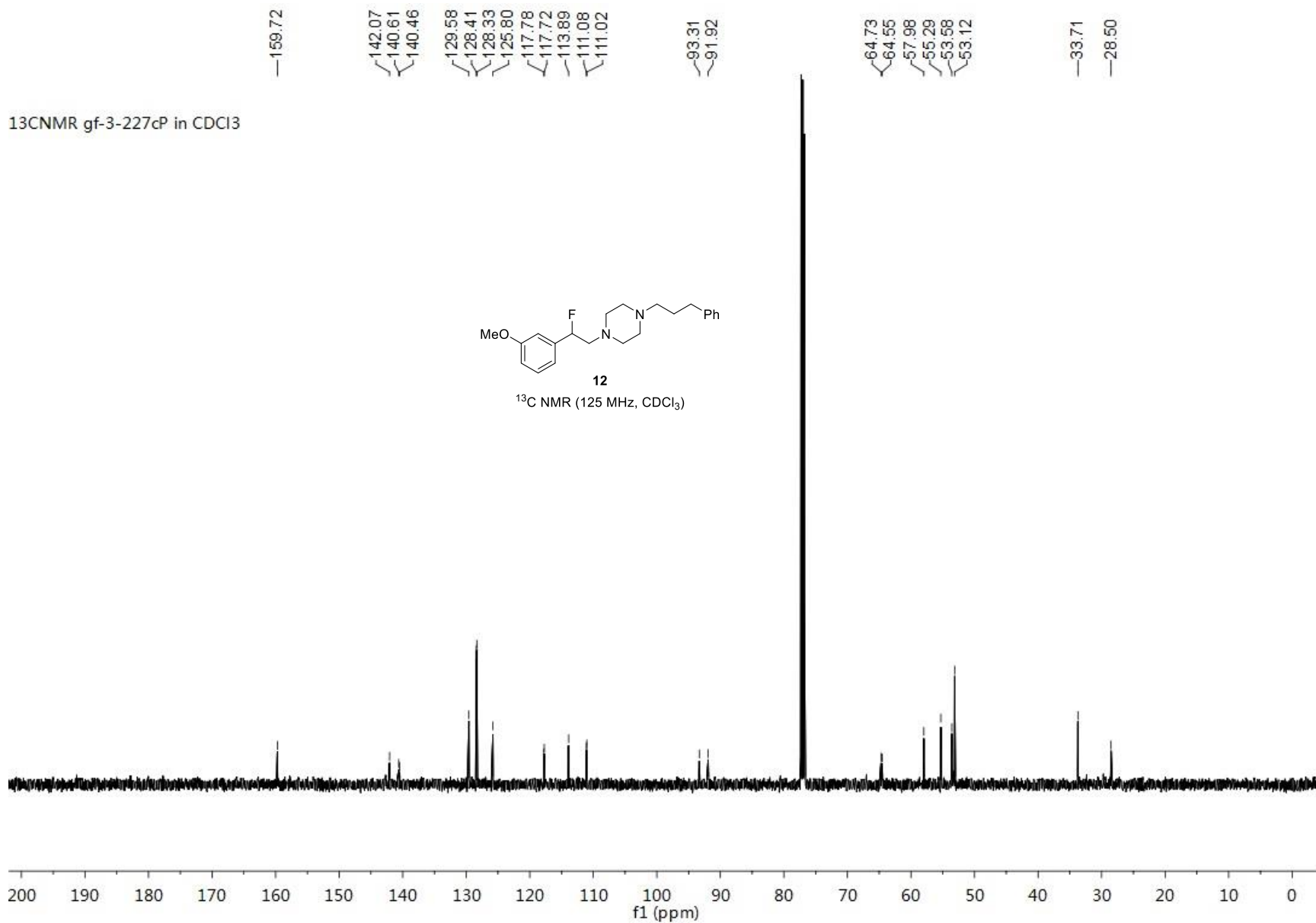
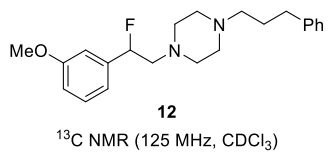
—177.0941



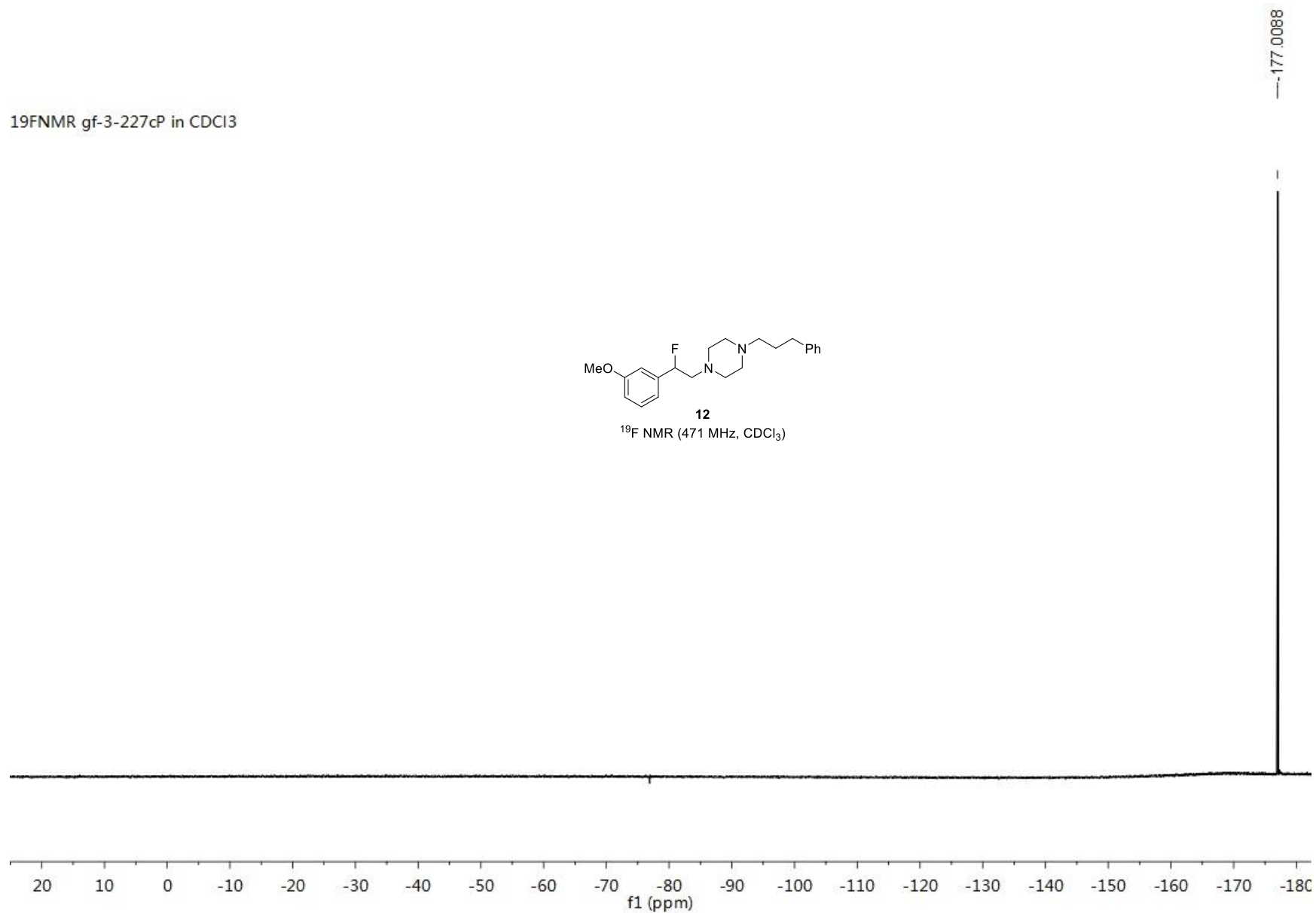
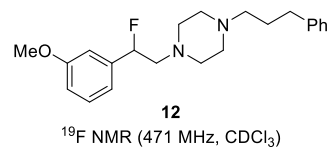
¹H NMR gf-3-227cP in CDCl₃

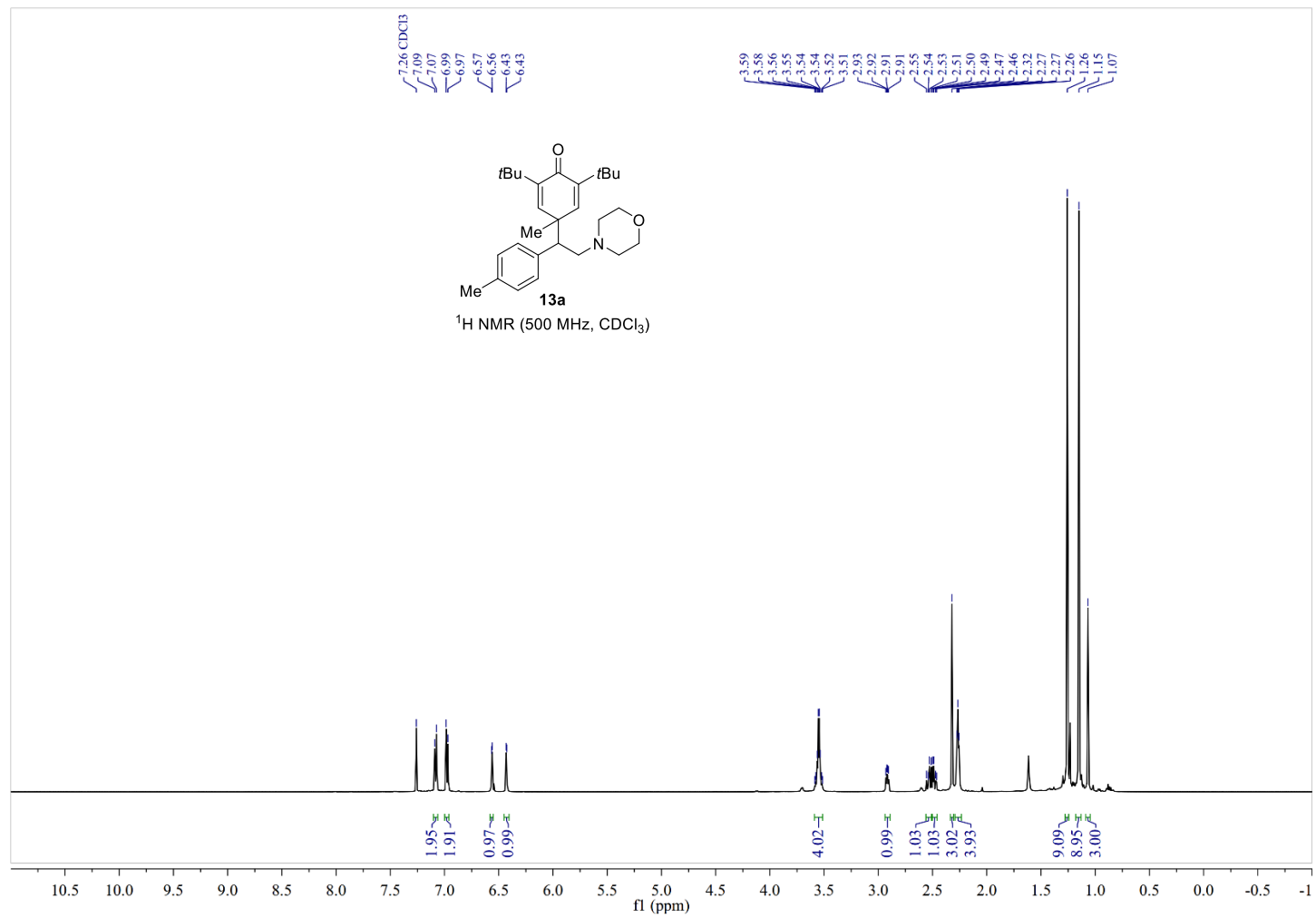


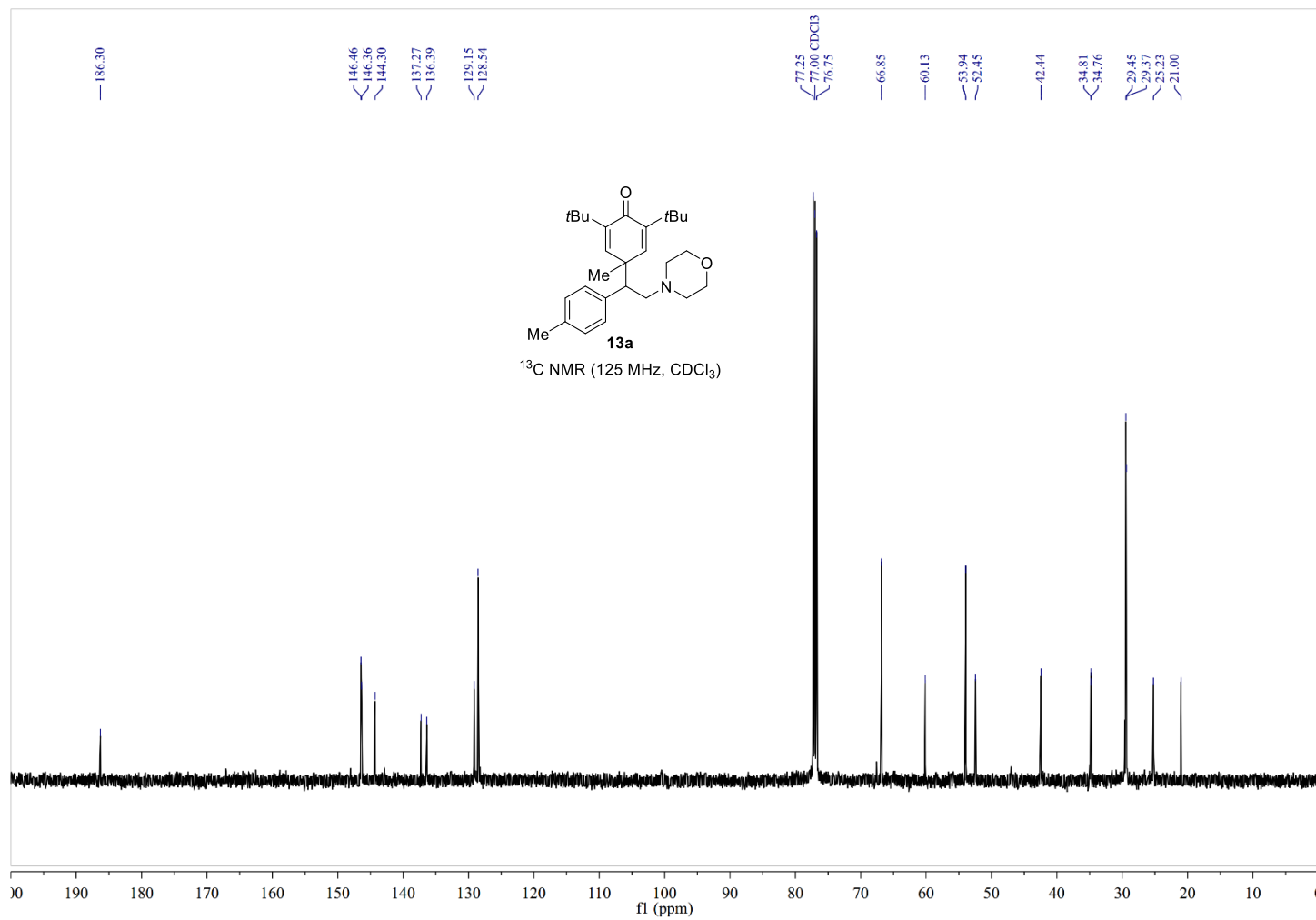
¹³CNMR gf-3-227cP in CDCl₃



¹⁹F NMR gf-3-227cP in CDCl₃



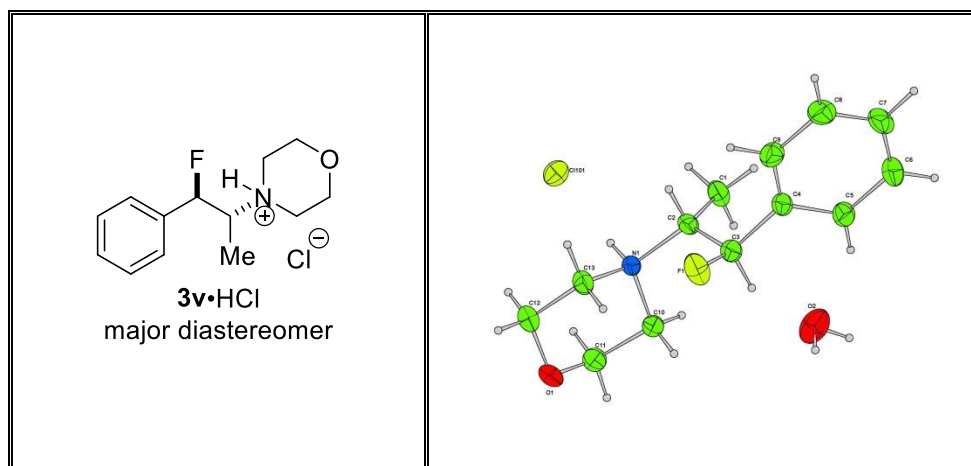




11. X-ray crystallography information

The major single diastereomer of compound **3v**•HCl was recrystallized twice in dichloromethane and hexanes. Then, a crystal of was slowly grown from dichloromethane, ethyl acetate and *n*-hexane, which is suitable for X-ray diffraction analysis. The chemical structure was showed as follows, CCDC number is 2210154. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre *via* www.ccdc.com.ac.uk/data_request/cif.

Crystal structure report for 22097 (**3v**•HCl–major single diastereomer)



The sample was submitted by Guangshou Feng (research group of Wang, Department of Chemistry, Duke University). A colorless crystal (approximate dimensions 0.230 x 0.100 x 0.040 mm³) was placed onto the tip of MiTeGen and mounted on a Bruker D8 VENTURE diffractometer and measured at 150 K.

Table 1. Crystal data and structure refinement for 22097.

Empirical formula	C13 H20.50 Cl1 F1.00 N1 O1.75	
Formula weight	273.26	
Crystal color, shape, size	colorless block fragment, 0.230 x 0.100 x 0.040 mm ³	
Temperature	150 K	
Wavelength	0.71073 Å	
Crystal system, space group	Monoclinic, C2/c	
Unit cell dimensions	a = 22.792(2) Å	a = 90°.
	b = 11.6050(13) Å	b = 111.875(3)°.
	c = 23.759(3) Å	g = 90°.
Volume	5831.8(10) Å ³	
Z	16	
Density (calculated)	1.245 Mg/m ³	
Absorption coefficient	0.266 mm ⁻¹	
F(000)	2328	

Data collection	
Diffractometer	Bruker D8 VENTURE, Bruker
Theta range for data collection	2.002 to 25.697°
Index ranges	-27<=h<=26, -14<=k<=14, -28<=l<=28
Reflections collected	84262
Independent reflections	5542 [R(int) = 0.192]
Observed Reflections	3473
Completeness to theta = 25.697°	99.7 %
Solution and Refinement	
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.99 and 0.97
Solution	Intrinsic phasing methods
Refinement method	Full-matrix least-squares on F ²
Weighting scheme	w = [s ² Fo ² + AP ² + BP] ⁻¹ , with P = (Fo ² + 2 Fc ²)/3, A = 0.115, B = 12.240
Data / restraints / parameters	5542 / 183 / 414
Goodness-of-fit on F ²	0.9871
Final R indices [I>2sigma(I)]	R1 = 0.0752, wR2 = 0.2076
R indices (all data)	R1 = 0.1109, wR2 = 0.2320
Largest diff. peak and hole	1.21 and -1.20 e.Å ⁻³

Table 2. Atomic coordinates (x 10⁴) and equivalent isotropic displacement parameters (Å²x 10³) for 22097. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
Cl1	2371(1)	4325(1)	4342(1)	57
Cl101	4124(1)	8189(1)	2817(1)	66
F1	4143(2)	4872(2)	4548(1)	86
F101	4538(4)	4343(5)	5999(5)	42
F201	4444(2)	4427(4)	5808(3)	47
O1	3365(2)	8662(3)	4296(2)	93
O2	2054(2)	4746(5)	2935(2)	105
O101	4450(2)	796(3)	5456(2)	70
O102	5000	3638(5)	7500	88
N1	3723(1)	6803(3)	3708(1)	46
N101	3685(1)	2456(2)	5755(1)	42
C1	3526(2)	5372(3)	2882(2)	65
C2	3818(1)	5581(2)	3535(1)	54
C3	3752(1)	4666(2)	3935(1)	57
C4	3936(1)	3478(2)	3791(2)	56

C5	3415(2)	2767(3)	3456(2)	64
C6	3534(2)	1683(3)	3270(2)	76
C7	4140(2)	1340(3)	3400(2)	83
C8	4632(2)	2034(4)	3720(3)	88
C9	4520(2)	3120(3)	3910(2)	71
C10	3044(2)	7002(4)	3639(2)	62
C11	2956(3)	8271(4)	3722(3)	82
C12	4006(3)	8519(5)	4355(3)	79
C13	4153(2)	7268(4)	4305(2)	63
C101	2871(4)	3805(4)	5812(5)	73
C102	3530(1)	3640(2)	5925(2)	60
C103	3877(2)	4598(4)	5814(2)	42
C104	3843(2)	5707(2)	6155(1)	52
C105	3445(8)	6608(9)	5760(3)	53
C106	3443(8)	7690(8)	5995(4)	55
C107	3745(7)	7871(8)	6589(4)	53
C108	4118(8)	7010(10)	6961(4)	54
C109	4169(9)	5924(9)	6714(4)	49
C110	3744(2)	2416(4)	5151(2)	52
C111	3885(3)	1218(4)	5006(2)	67
C112	4362(2)	729(4)	6017(2)	64
C113	4247(2)	1899(4)	6224(2)	50
C201	3058(3)	3632(4)	6200(3)	48
C203	4047(1)	4444(3)	6157(1)	48
C205	3520(5)	6320(5)	5672(2)	60
C206	3367(5)	7469(6)	5736(3)	68
C207	3572(4)	7969(5)	6305(3)	62
C208	3923(5)	7369(6)	6799(3)	65
C209	4060(6)	6220(5)	6742(2)	58

Table 3. Bond lengths [Å] and angles [°] for 22097.

H1-N1	0.884(19)	H2-N101	0.883(19)
H3-O2	0.83(2)	H4-O2	0.83(2)
H5-O102	0.841(14)	F1-C3	1.417(4)
F101-C103	1.436(9)	F201-C203	1.438(6)
O1-C11	1.410(7)	O1-C12	1.424(6)
O101-C111	1.420(6)	O101-C112	1.423(6)
N1-C2	1.515(4)	N1-C10	1.512(5)
N1-C13	1.493(5)	N101-C102	1.511(4)
N101-C110	1.492(5)	N101-C113	1.496(5)
C1-C2	1.461(5)	C1-H11	0.950
C1-H12	0.950	C1-H13	0.950
C2-C3	1.470(4)	C2-H21	0.950
C3-C4	1.517(3)	C3-H31	0.950
C4-C5	1.421(4)	C4-C9	1.320(4)
C5-C6	1.393(5)	C5-H51	0.950
C6-C7	1.359(5)	C6-H61	0.950
C7-C8	1.360(5)	C7-H71	0.950
C8-C9	1.396(5)	C8-H81	0.950
C9-H91	0.950	C10-C11	1.509(7)
C10-H101	0.950	C10-H102	0.950
C11-H111	0.950	C11-H112	0.950
C12-C13	1.505(7)	C12-H121	0.950
C12-H122	0.950	C13-H131	0.950
C13-H132	0.950	C101-C102	1.437(9)
C101-H1011	0.950	C101-H1012	0.950
C101-H1013	0.950	C102-C103	1.443(5)
C102-H1021	0.950	C102-C201	1.452(5)
C102-C203	1.441(4)	C102-H1022	0.950
C103-C104	1.537(4)	C103-C104	1.537(4)
C103-H1032	0.950	C104-C105	1.471(7)
C104-C109	1.282(7)	C104-C203	1.537(4)
C104-C205	1.316(6)	C104-C209	1.427(6)
C105-C106	1.375(7)	C105-H1051	0.950
C106-C107	1.336(8)	C106-H1061	0.950
C107-C108	1.395(8)	C107-H1071	0.950
C108-C109	1.414(7)	C108-H1081	0.950
C109-H1091	0.950	C110-C111	1.496(6)
C110-H1101	0.950	C110-H1102	0.950
C111-H1111	0.950	C111-H1112	0.950
C112-C113	1.499(6)	C112-H1121	0.950
C112-H1122	0.950	C113-H1131	0.950

C113-H1132	0.950	C201-H2011	0.950
C201-H2012	0.950	C201-H2013	0.950
C203-H2031	0.950	C205-C206	1.401(6)
C205-H2051	0.950	C206-C207	1.381(6)
C206-H2061	0.950	C207-C208	1.342(6)
C207-H2071	0.950	C208-C209	1.387(6)
C208-H2081	0.950	C209-H2091	0.950

C11-O1-C12	110.0(4)	H4-O2-H3	97(8)
C111-O101-C112	108.6(3)	H5-O102-H5#1	114(11)
H1-N1-C2	105.1(15)	H1-N1-C10	105.5(15)
C2-N1-C10	111.0(3)	H1-N1-C13	105.8(15)
C2-N1-C13	119.1(3)	C10-N1-C13	109.3(3)
H2-N101-C102	106.4(15)	H2-N101-C110	106.5(15)
C102-N101-C110	113.5(3)	H2-N101-C113	105.1(15)
C102-N101-C113	114.5(3)	C110-N101-C113	110.1(3)
C2-C1-H11	109.2	C2-C1-H12	109.9
H11-C1-H12	109.5	C2-C1-H13	109.3
H11-C1-H13	109.5	H12-C1-H13	109.5
N1-C2-C1	112.83(10)	N1-C2-C3	116.15(9)
C1-C2-C3	116.76(10)	N1-C2-H21	102.9
C1-C2-H21	102.9	C3-C2-H21	102.5
C2-C3-F1	111.47(10)	C2-C3-C4	114.01(10)
F1-C3-C4	105.31(10)	C2-C3-H31	108.3
F1-C3-H31	108.8	C4-C3-H31	108.8
C3-C4-C5	114.23(8)	C3-C4-C9	125.34(8)
C5-C4-C9	120.21(8)	C4-C5-C6	118.78(9)
C4-C5-H51	119.8	C6-C5-H51	121.4
C5-C6-C7	119.60(9)	C5-C6-H61	119.9
C7-C6-H61	120.5	C6-C7-C8	120.75(9)
C6-C7-H71	119.8	C8-C7-H71	119.4
C7-C8-C9	120.17(9)	C7-C8-H81	119.9
C9-C8-H81	119.8	C8-C9-C4	120.48(9)
C8-C9-H91	119.8	C4-C9-H91	119.7
N1-C10-C11	108.4(4)	N1-C10-H101	109.6
C11-C10-H101	109.1	N1-C10-H102	109.8
C11-C10-H102	110.5	H101-C10-H102	109.5
C10-C11-O1	111.6(5)	C10-C11-H111	108.3
O1-C11-H111	107.8	C10-C11-H112	110.1
O1-C11-H112	109.5	H111-C11-H112	109.5
O1-C12-C13	110.8(4)	O1-C12-H121	108.7

C13-C12-H121	109.5	O1-C12-H122	109.1
C13-C12-H122	109.3	H121-C12-H122	109.5
C12-C13-N1	109.6(4)	C12-C13-H131	109.7
N1-C13-H131	109.5	C12-C13-H132	109.5
N1-C13-H132	108.9	H131-C13-H132	109.5
H1011-C101-H1012	109.5	H1011-C101-H1013	109.5
H1012-C101-H1013	109.5	N101-C102-C101	113.76(10)
N101-C102-C103	117.01(9)	C101-C102-C103	117.93(10)
N101-C102-H1021	103.3	C101-C102-H1021	96.9
C103-C102-H1021	103.6	N101-C102-C201	113.59(10)
N101-C102-C203	116.90(9)	C201-C102-C203	117.92(10)
N101-C102-H1022	101.4	C201-C102-H1022	102.2
C203-C102-H1022	100.8	C102-C103-C104	114.27(10)
F101-C103-C104	105.34(10)	F101-C103-H1032	107.2
C104-C103-H1032	108.2	C103-C104-C105	113.58(10)
C103-C104-C109	125.87(10)	C105-C104-C109	120.17(10)
C203-C104-C205	126.05(8)	C203-C104-C209	113.60(8)
C205-C104-C209	120.30(8)	C104-C105-C106	119.03(10)
C104-C105-H1051	119.5	C106-C105-H1051	121.4
C105-C106-C107	119.60(10)	C105-C106-H1061	119.7
C107-C106-H1061	120.7	C106-C107-C108	120.70(10)
C106-C107-H1071	119.8	C108-C107-H1071	119.5
C107-C108-C109	119.95(10)	C107-C108-H1081	120.0
C109-C108-H1081	120.0	C108-C109-C104	120.05(10)
C108-C109-H1091	119.9	C104-C109-H1091	120.0
N101-C110-C111	110.8(4)	N101-C110-H1101	109.2
C111-C110-H1101	108.8	N101-C110-H1102	109.0
C111-C110-H1102	109.7	H1101-C110-H1102	109.5
C110-C111-O101	111.0(4)	C110-C111-H1111	108.9
O101-C111-H1111	108.9	C110-C111-H1112	109.1
O101-C111-H1112	109.4	H1111-C111-H1112	109.5
O101-C112-C113	111.1(4)	O101-C112-H1121	109.6
C113-C112-H1121	110.2	O101-C112-H1122	108.6
C113-C112-H1122	107.7	H1121-C112-H1122	109.5
C112-C113-N101	110.6(3)	C112-C113-H1131	110.2
N101-C113-H1131	109.6	C112-C113-H1132	108.0
N101-C113-H1132	108.9	H1131-C113-H1132	109.5
H2011-C201-H2012	109.5	H2011-C201-H2013	109.5
H2012-C201-H2013	109.5	C104-C203-C102	114.41(10)
C104-C203-F201	105.39(10)	C104-C203-H2031	108.3
F201-C203-H2031	110.3	C104-C205-C206	120.03(9)
C104-C205-H2051	120.0	C206-C205-H2051	120.0
C205-C206-C207	119.94(9)	C205-C206-H2061	120.0

C207-C206-H2061	120.1	C206-C207-C208	120.74(9)
C206-C207-H2071	119.4	C208-C207-H2071	119.8
C207-C208-C209	119.74(9)	C207-C208-H2081	120.4
C209-C208-H2081	119.9	C104-C209-C208	119.15(9)
C104-C209-H2091	119.6	C208-C209-H2091	121.2

Symmetry transformations used to generate equivalent atoms: #1 $-x+1,y,-z+3/2$

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for 22097. The anisotropic displacement factor exponent takes the form: $-2p^2[h^2 a^*2U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U ¹¹	U ²²	U ³³	U ²³	U ¹³	U ¹²
Cl1	46(1)	54(1)	68(1)	4(1)	16(1)	-5(1)
Cl101	69(1)	67(1)	64(1)	15(1)	26(1)	-1(1)
F1	137(3)	60(2)	57(2)	1(1)	34(2)	-6(2)
F101	35(4)	44(5)	43(6)	-3(4)	8(4)	-2(3)
F201	45(3)	47(3)	52(4)	-4(2)	22(3)	-2(2)
O1	116(3)	61(2)	134(4)	-35(2)	84(3)	-9(2)
O2	85(3)	133(4)	83(3)	13(3)	15(2)	-30(3)
O101	65(2)	56(2)	97(3)	-12(2)	40(2)	12(2)
O102	99(4)	78(4)	64(3)	0	4(3)	0
N1	54(2)	36(2)	54(2)	-2(2)	28(2)	-2(2)
N101	37(2)	34(2)	58(2)	-4(2)	20(2)	-1(1)
C1	92(3)	48(3)	55(3)	-6(2)	29(3)	-5(2)
C2	69(3)	39(2)	63(3)	-4(2)	36(2)	1(2)
C3	70(3)	47(3)	63(3)	0(2)	34(2)	-4(2)
C4	71(3)	39(2)	65(3)	3(2)	35(2)	-2(2)
C5	66(3)	44(3)	87(3)	3(2)	33(3)	4(2)
C6	89(4)	47(3)	89(4)	-5(3)	30(3)	-5(3)
C7	124(5)	40(3)	107(4)	4(3)	67(4)	13(3)
C8	88(4)	56(3)	140(5)	16(3)	65(4)	25(3)
C9	60(3)	58(3)	100(4)	6(3)	36(3)	3(2)
C10	57(2)	49(3)	90(3)	-9(2)	37(2)	-1(2)
C11	78(3)	50(3)	137(5)	-8(3)	63(4)	3(3)
C12	92(4)	60(3)	92(4)	-27(3)	43(3)	-17(3)
C13	76(3)	54(3)	62(3)	-14(2)	29(2)	-12(2)
C101	62(6)	64(5)	89(6)	-10(6)	23(5)	-8(5)
C102	59(2)	42(2)	93(3)	-8(2)	43(2)	0(2)
C103	46(4)	37(4)	50(5)	0(4)	27(4)	1(4)
C104	58(2)	36(2)	73(3)	-6(2)	36(2)	-6(2)
C105	64(6)	43(5)	58(5)	1(5)	28(5)	13(6)
C106	68(5)	53(6)	48(7)	7(6)	27(7)	2(6)
C107	67(7)	41(5)	50(6)	4(5)	21(5)	13(4)
C108	73(10)	36(6)	48(6)	9(5)	16(5)	17(6)
C109	58(7)	32(6)	58(5)	-1(5)	24(5)	18(6)
C110	56(2)	45(2)	47(2)	-4(2)	11(2)	3(2)
C111	88(3)	51(3)	65(3)	-15(2)	32(3)	-2(3)
C112	53(2)	42(3)	83(3)	6(2)	11(2)	7(2)
C113	51(2)	46(2)	47(2)	3(2)	13(2)	0(2)
C201	51(3)	42(3)	65(4)	-14(3)	38(3)	-4(3)

C203	56(4)	42(3)	50(4)	-6(3)	24(3)	-4(3)
C205	74(5)	52(5)	64(4)	-8(4)	37(4)	1(4)
C206	74(5)	56(5)	71(6)	19(5)	24(5)	4(4)
C207	73(6)	45(4)	71(5)	-1(4)	30(5)	7(4)
C208	79(7)	51(5)	64(5)	-2(4)	24(4)	6(5)
C209	74(6)	35(5)	69(4)	-4(3)	33(4)	6(4)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for 22097.

	x	y	z	U(eq)
H1	3782(11)	7243(19)	3430(10)	68(2)
H2	3362(10)	2009(19)	5728(10)	63(2)
H3	2100(40)	4610(80)	3290(16)	158(2)
H4	1720(20)	4370(70)	2770(40)	158(2)
H5	5190(30)	3240(60)	7320(30)	131(2)
H11	3601	4597	2800	80
H12	3702	5877	2672	80
H13	3084	5503	2751	80
H21	4258	5558	3609	65
H31	3324	4647	3902	69
H51	2996	3032	3368	79
H61	3191	1194	3049	93
H71	4223	616	3258	97
H81	5054	1766	3828	103
H91	4865	3611	4122	86
H101	2956	6581	3942	75
H102	2768	6755	3249	75
H111	3054	8683	3423	100
H112	2530	8421	3675	100
H121	4274	8807	4739	100
H122	4076	8942	4043	100
H131	4580	7190	4339	74
H132	4091	6844	4621	74
H1011	2798	4564	5922	94
H1012	2707	3262	6015	94
H1013	2667	3709	5386	94
H1021	3673	3608	6355	72
H1022	3303	3964	5537	72
H1032	3723	4765	5393	56
H1051	3185	6426	5353	70
H1061	3223	8300	5734	73
H1071	3706	8594	6759	68
H1081	4350	7163	7378	68
H1091	4441	5352	6963	63
H1101	4079	2909	5157	61
H1102	3359	2674	4850	61
H1111	3544	728	4988	79
H1112	3929	1219	4623	79

H1121	4722	384	6315	77
H1122	3999	271	5961	77
H1131	4182	1840	6595	59
H1132	4610	2359	6283	59
H2011	2977	4399	6290	61
H2012	3212	3194	6564	61
H2013	2679	3296	5928	61
H2031	4281	4224	6565	62
H2051	3385	5979	5281	78
H2061	3125	7904	5389	88
H2071	3468	8749	6344	79
H2081	4072	7724	7187	79
H2091	4298	5773	7087	72

Table 6. Torsion angles [°] for 22097.

C112-O101-C111-C110	63.3(5)	C111-O101-C112-C113	-63.1(5)
C110-N101-C102-C201	137.7(4)	C110-N101-C102-C203	-79.7(4)
C113-N101-C102-C201	-94.8(4)	C113-N101-C102-C203	47.9(4)
C102-N101-C110-C111	-178.9(4)	C113-N101-C110-C111	51.2(5)
C102-N101-C113-C112	179.8(3)	C110-N101-C113-C112	-50.9(4)
C12-O1-C11-C10	62.2(6)	C11-O1-C12-C13	-61.3(6)
C201-C102-C203-F201	-174.8(4)	N101-C102-C203-F201	44.2(4)
N101-C102-C203-C104	163.8(3)	C201-C102-C203-C104	-55.2(4)
C209-C104-C203-F201	-116.9(7)	C209-C104-C203-C102	119.9(6)
C205-C104-C203-C102	-62.6(7)	C209-C104-C205-C206	-2.5(14)
C203-C104-C209-C208	178.1(9)	C203-C104-C205-C206	-179.8(7)
C205-C104-C203-F201	60.5(8)	C205-C104-C209-C208	0.5(15)
N101-C110-C111-O101	-58.2(6)	O101-C112-C113-N101	57.6(5)
C104-C205-C206-C207	2.0(16)	C205-C206-C207-C208	0.7(16)
C206-C207-C208-C209	-2.7(16)	C207-C208-C209-C104	2.1(17)
C10-N1-C2-C1	71.5(4)	C10-N1-C2-C3	-67.1(4)
C13-N1-C2-C1	-160.2(4)	C13-N1-C2-C3	61.2(4)
C2-N1-C10-C11	-171.3(4)	C13-N1-C10-C11	55.4(5)
C2-N1-C13-C12	175.2(4)	C10-N1-C13-C12	-55.8(5)
N1-C2-C3-F1	-53.3(4)	N1-C2-C3-C4	-172.4(3)
C1-C2-C3-F1	169.7(3)	C1-C2-C3-C4	50.6(4)
F1-C3-C4-C5	136.7(3)	F1-C3-C4-C9	-48.8(4)
C2-C3-C4-C5	-100.8(3)	C2-C3-C4-C9	73.8(4)
C3-C4-C5-C6	176.3(3)	C9-C4-C5-C6	1.4(6)
C3-C4-C9-C8	-175.6(4)	C5-C4-C9-C8	-1.3(6)
C4-C5-C6-C7	-1.4(6)	C5-C6-C7-C8	1.3(7)
C6-C7-C8-C9	-1.2(8)	C7-C8-C9-C4	1.2(8)
N1-C10-C11-O1	-59.1(6)	O1-C12-C13-N1	58.7(6)

Symmetry transformations used to generate equivalent atoms: #1 -x+1,y,-z+3/2