

Identification of visual cortex cell types and species differences using single-cell RNA sequencing

Authors

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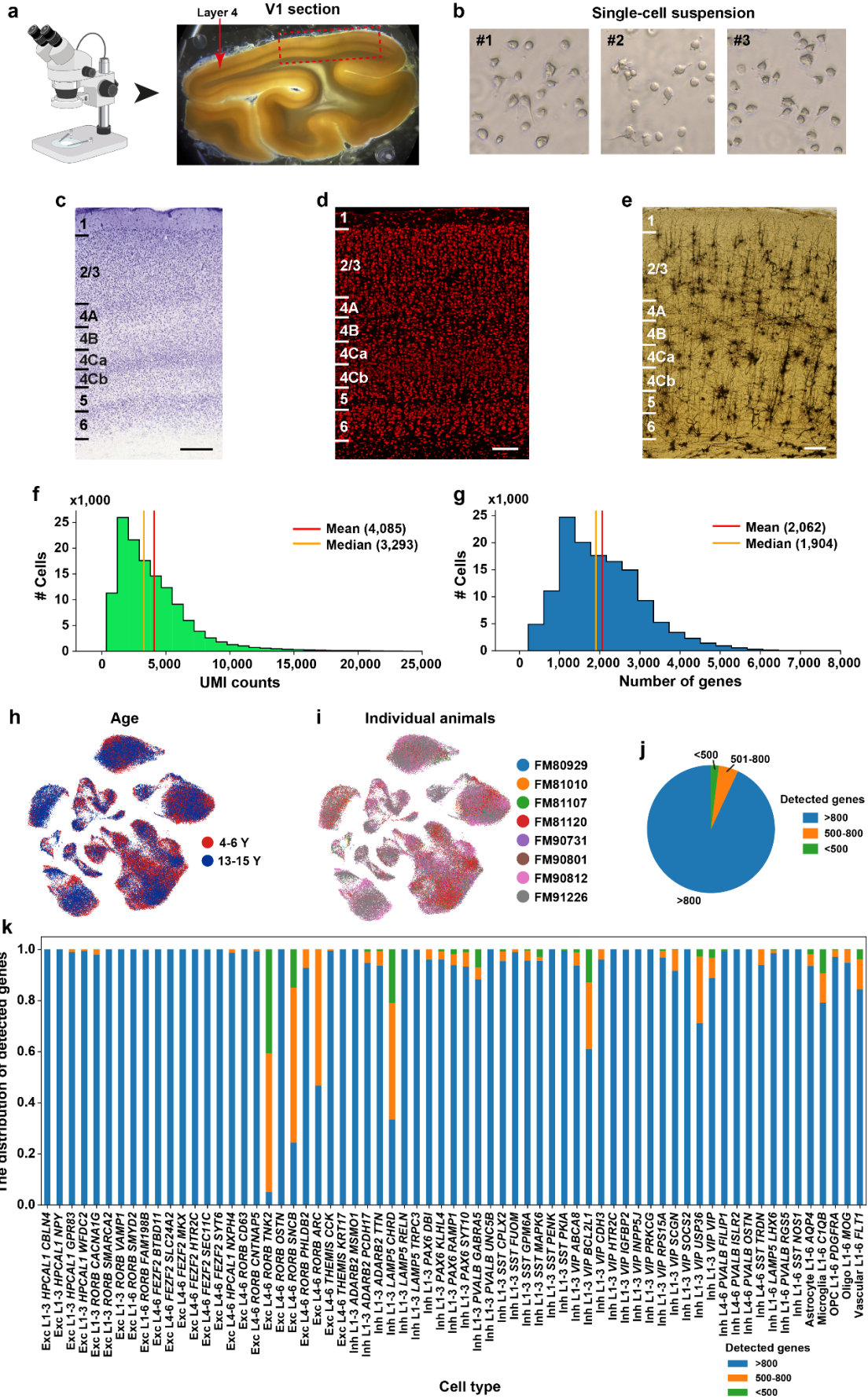
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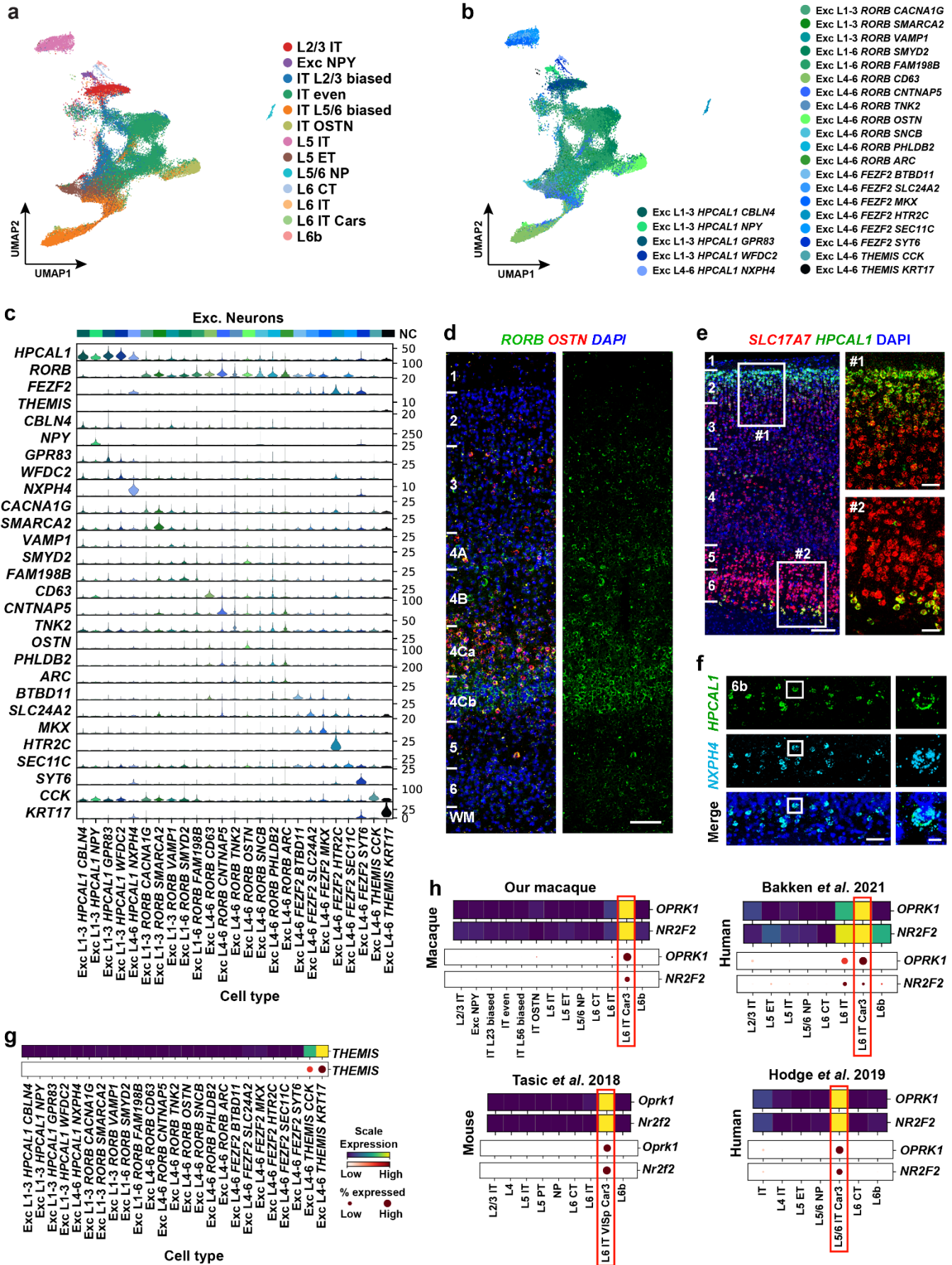
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Supplementary figures 1-11



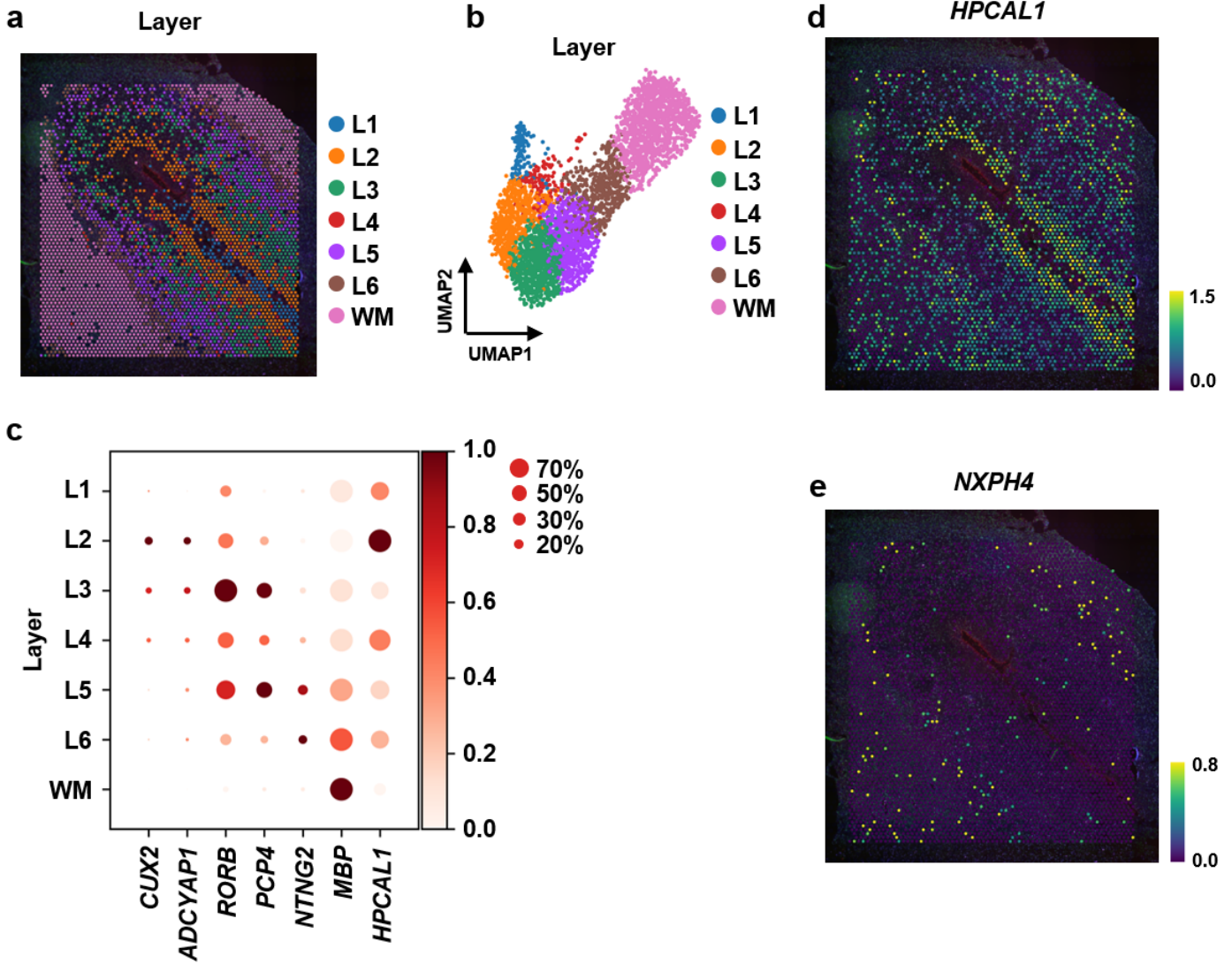
Supplementary Figure 1. Experimental design and quality control of scRNA-seq data.

- a) Stereo microscope photographs illustrating the tissue dissection strategy. The coronal section of primary visual cortex (V1) was dissected from the adult macaque brain. The drawing of the stereo microscope was created with BioRender.com.
- b) Single whole-cell isolated from adult macaque V1 regions labeled by red dashline in a. Example single-cell suspension generated from macaque V1 that exhibited diverse cellular morphologies.
- c) Nissl staining section reveals the detailed laminar architecture of macaque V1. Scale bar, 150 μm . $n = 4$ macaques.
- d) Fluorescent Nissl staining image shows detailed laminar architecture of macaque V1. Scale bar, 150 μm . $n = 4$ macaques.
- e) Golgi staining showed morphological characterization of diversity cell type in macaque V1. Scale bars, 200 μm . $n = 3$ macaques.
- f) Histogram showing the distribution of UMI counts detected across the entire scRNA-seq dataset.
- g) Histogram showing the distribution of number of genes detected across the entire scRNA-seq dataset.
- h) UMAP visualization of the animal age.
- i) UMAP plot showing 133,454 cells by the individual animal.
- j) The distribution of detected genes in each cell for the whole dataset.
- k) The distribution of detected genes in each cell for each cell type. Source data are provided as a Source Data file.



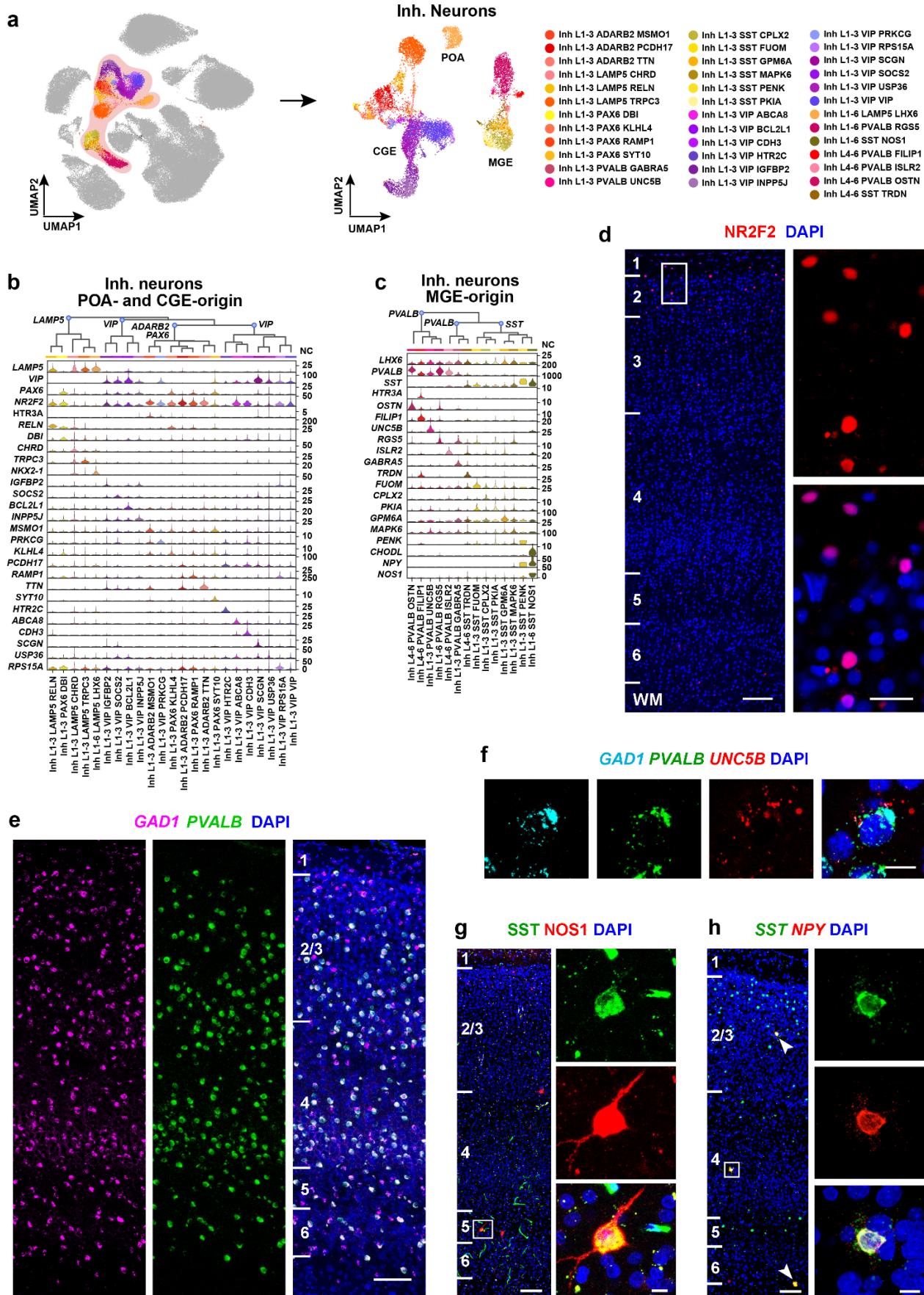
Supplementary Figure 2. Excitatory cell taxonomy and layer-specific markers in macaque V1.

- a) The UMAP plot shows the classification of excitatory neurons by their layer distributions and projections.
- b) The UMAP plot shows the excitatory neuron cell types and their layer distribution.
- c) Violin plots visualizing the distribution of marker genes expression in excitatory neurons using the same color scheme as in Fig. 1b, e. The expression levels are normalized to the maximal normalized counts (NC, 10,000 counts per cell) of each gene.
- d) RNAscope mFISH image of *RORB* (green) and *OSTN* (red) from macaque V1. Cortical layers 1-6 and white matter (WM) are labelled. The experiment was repeated three times independently with similar results. Scale bar, 100 μm . $n = 3$ macaques.
- e) RNAscope mFISH for *SLC17A7* (glutamatergic cell marker) and *HPCAL1* showing co-expression in L2/3 and L6b. Scale bars: low magnification, 100 μm ; high magnification, 25 μm . $n = 3$ macaques.
- f) RNAscope mFISH showing the co-localization of *HPCAL1* and *NXP4* in L6b. Scale bars, low magnification, 50 μm ; high magnification, 10 μm . $n = 3$ macaques.
- g) Dotplot shows the expression of *THEMIS* genes in glutamatergic neurons.
- h) The expression pattern of IT *Car3* cells in primates and mice.



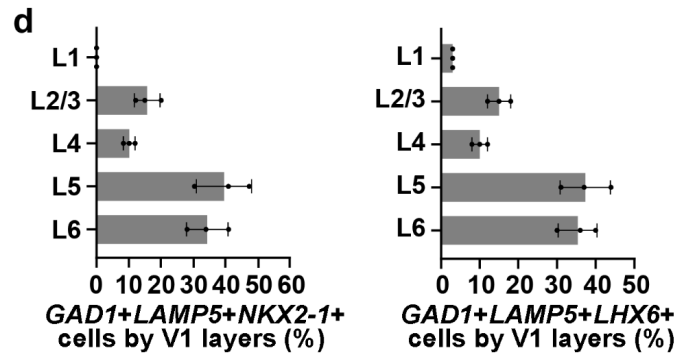
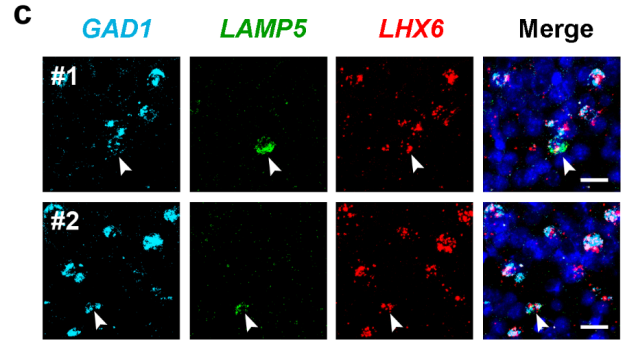
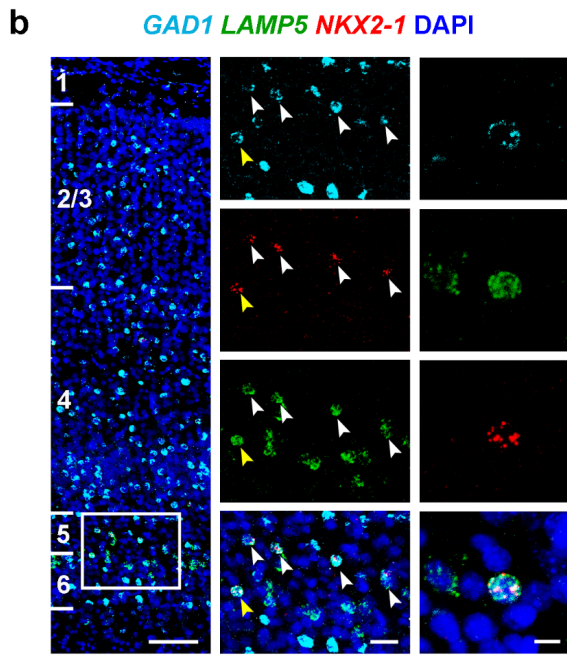
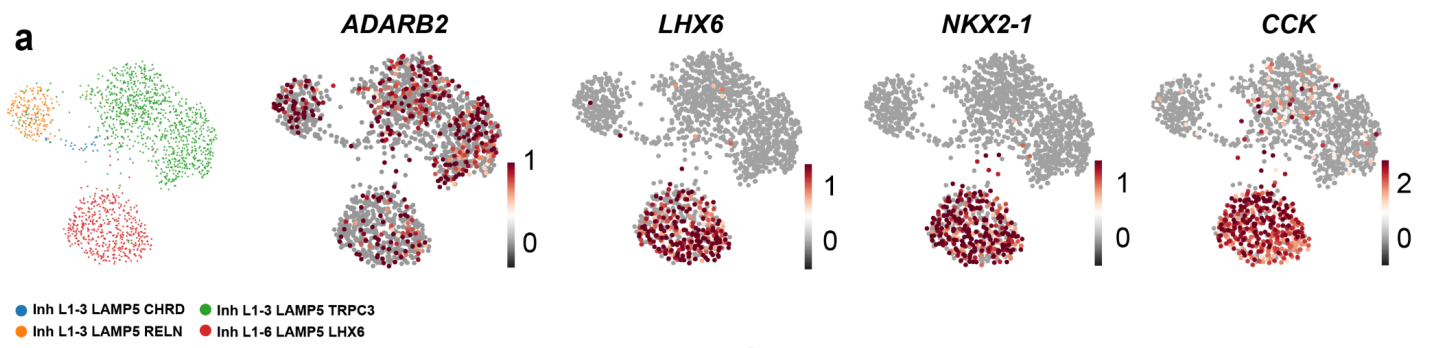
Supplementary Figure 3. *HPCAL1* and *NXPH4* labels L2 and L6b of human cerebral cortex.

- a) Leiden cluster results from the dataset, annotated by layers.
- b) The layer distribution of the Leiden results matching the expression pattern in a.
- c) The dotplot shows the expression of genes in a.
- d) *HPCAL1* gene expression is elevated in L2 and L6.
- e) *NXPH4* gene expression is elevated in L6 but not in L2.



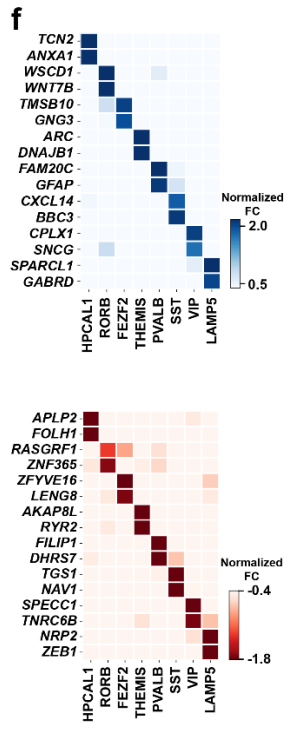
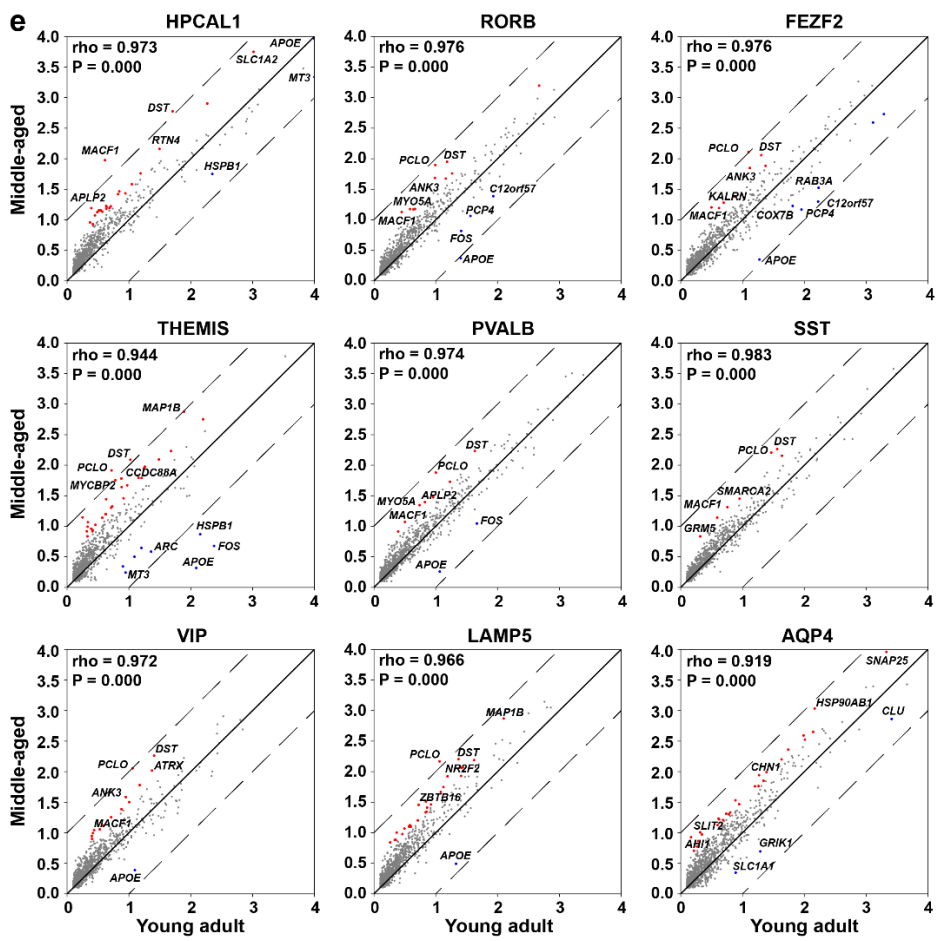
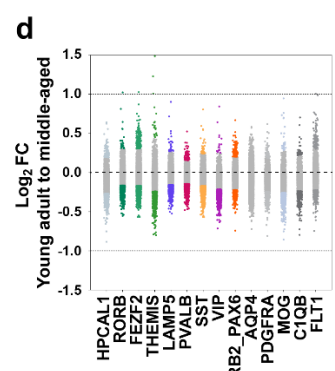
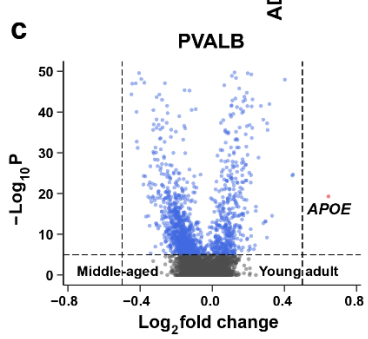
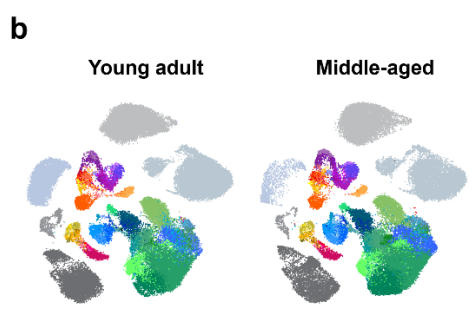
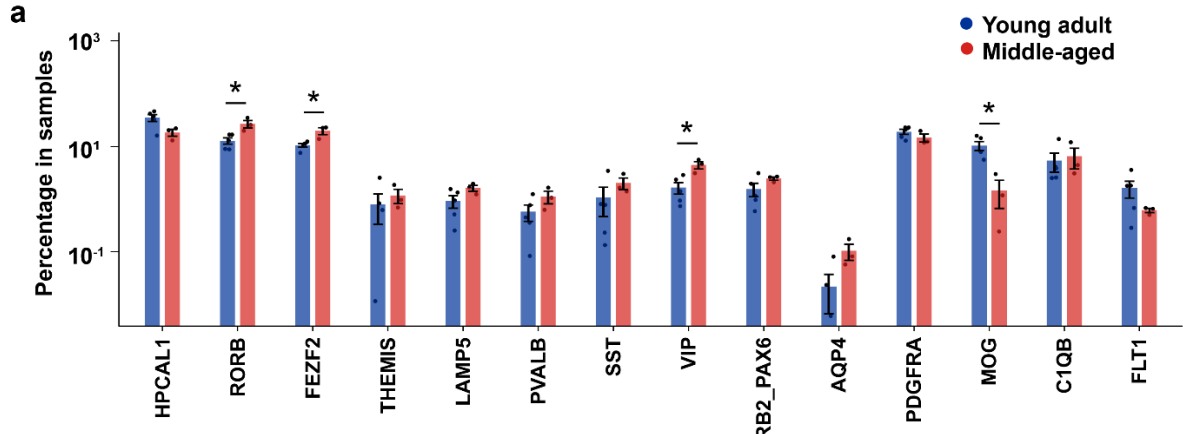
Supplementary Figure 4. Inhibitory cell taxonomy in the macaque V1.

- a) Visualization of the inhibitory neuron cell types and their layer distribution. Left panel, t-SNE map using the same color scheme for inhibitory neurons and gray for all other cells. Right panel, UMAP visualises the inhibitory neurons clustered into POA-, CGE- and MGE-originated groups.
- b) Violin plots of the distribution of marker gene expression of POA- and CGE-originated inhibitory neurons. The expression levels are normalized to the maximal of the cell counts that normalized to 10,000 reads per cell (NC).
- c) The distribution of the marker gene expression of MGE-originated inhibitory neurons.
- d) Immunostaining for NR2F2+ cells in L1 and L2. Red, NR2F2; Blue, DAPI. Scale bars, low magnification, 100 μm ; high magnification, 25 μm . $n = 3$ macaques.
- e) RNAscope mFISH validation showing *GADI* (violet) and *PVALB* (green) is expressed across all V1 layers. Blue, DAPI. Scale bar, 100 μm . $n = 3$ macaques.
- f) mFISH shows that chandelier cells co-expressing mRNA of *GADI*, *PVALB* and *UNC5B*. Light blue, *GADI*; Green, *PVALB*; Red, *UNC5B*; Blue, DAPI. Scale bar, 10 μm . $n = 3$ macaques.
- g) Co-immunostaining shows the SST+ subclass SST+NOS1+ long-range projecting neurons, likely corresponding to the SST NPY and the SST CHODL cell types. Green, SST; Red, NOS1; Blue, DAPI. Scale bars, low magnification, 100 μm ; high magnification, 10 μm . $n = 4$ macaques.
- h) mFISH shows the *NPY* is expressed in a subset of SST+ cells, likely corresponding to SST NOS1 type and SST CHODL type. Left panel, co-labeled cells are indicated by white arrowheads and white box. Right panel, positive cell from white box in left panel. Scale bars, low magnification, 200 μm ; high magnification, 10 μm . $n = 4$ macaques.



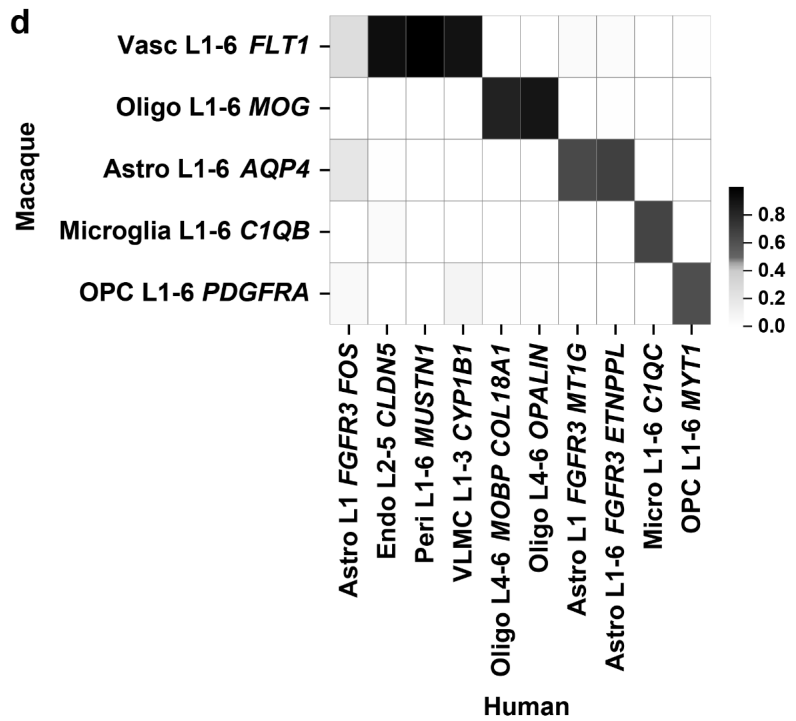
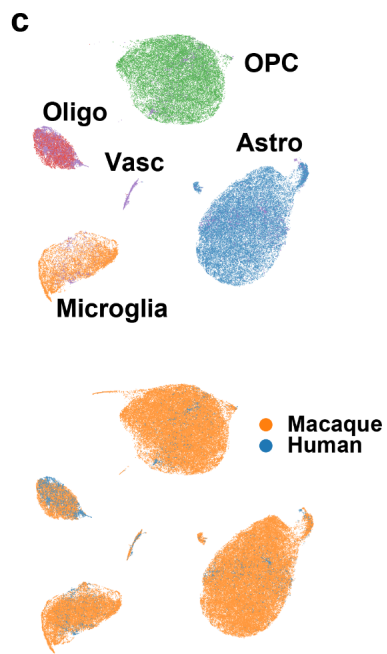
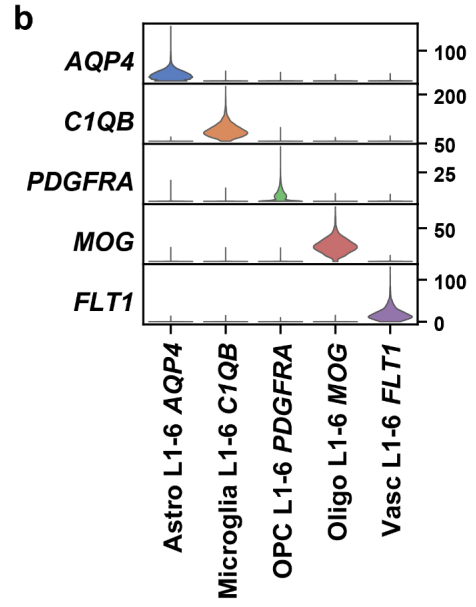
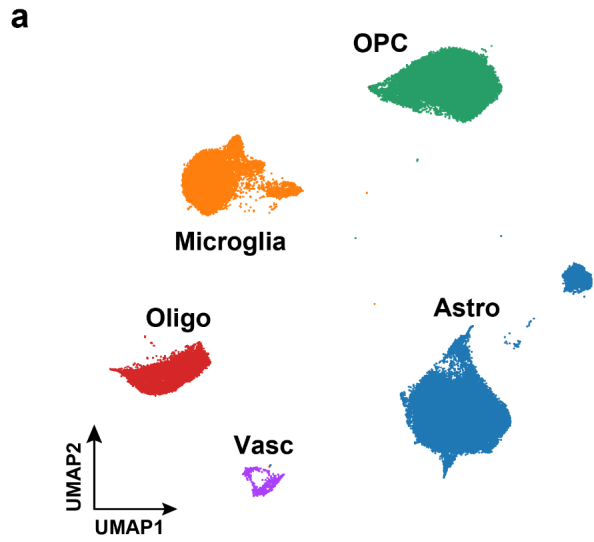
Supplementary Figure 5. *LAMP5 LHX6* cells are the equivalent to the previously reported *LAMP5 LHX6* cell type in humans and mice.

- a) UMAPs plot of the MGE (*LHX6*) and CGE (*ADARB2*) origination markers. The *LAMP5-LHX6* cell population is positive for both markers.
- b) Co-expression of *GAD1*, *LAMP5* and *NKX2-1* mRNAs in macaque V1. Light blue, *GAD1*; Green, *LAMP5*; Red, *NKX2-1*; Blue, DAPI. Middle panel, positive cells that co-express all three genes are identified by yellow and white arrowheads. Right panel, a positive cell marked by yellow arrowhead in the middle panel. Scale bars, low magnification, 100 μm ; middle magnification, 25 μm ; high magnification, 10 μm .
- c) Representative image of cells co-labeled with the *GAD1* (light blue), *LAMP5* (green) and *LHX6* (red) gene panel. Blue, DAPI. Scale bar, 10 μm . Triple-positive cells marked by white arrowheads.
- d) Left panel, count of *GAD1+LAMP5+NKX2-1+* cells across layers. Data are mean \pm s.d. and dots show the data points for individual macaques. Right panel, quantification of *GAD1+LAMP5+LHX6+* cells across layers. Data are mean \pm s.d. and individual replicates are shown as dots. $n = 3$ macaques. Source data are provided as a Source Data file.



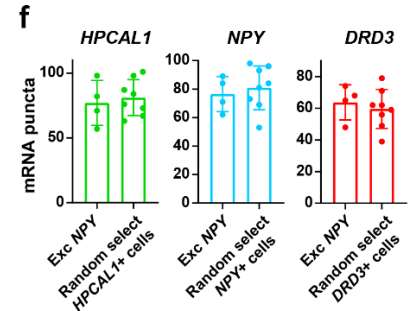
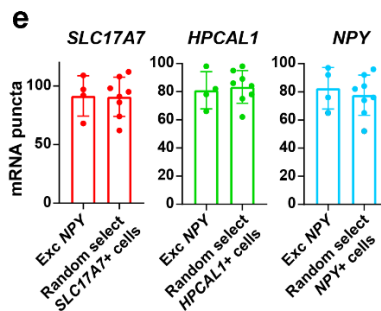
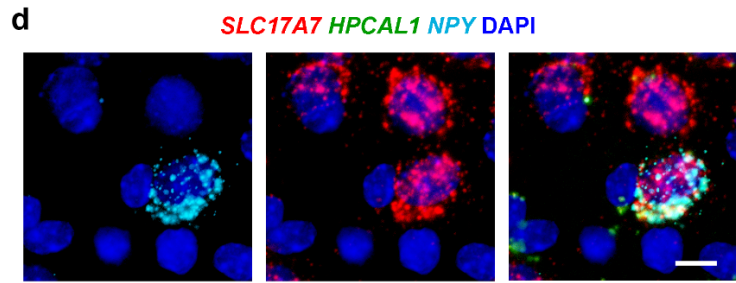
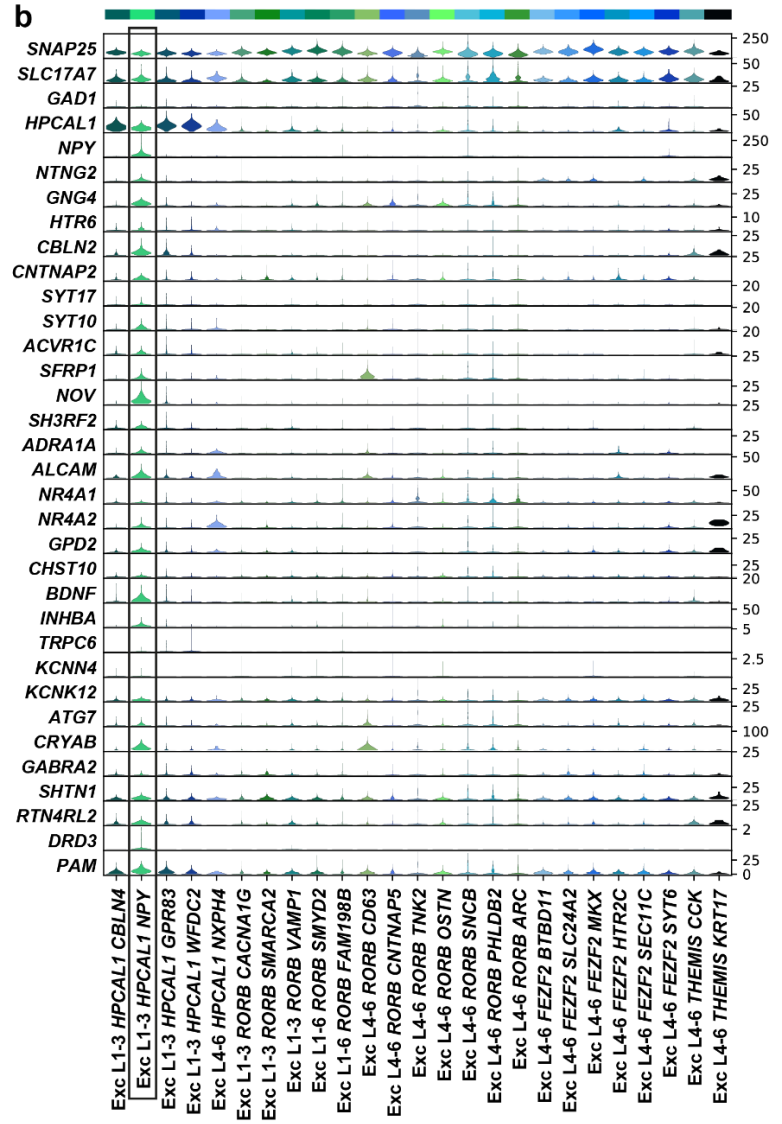
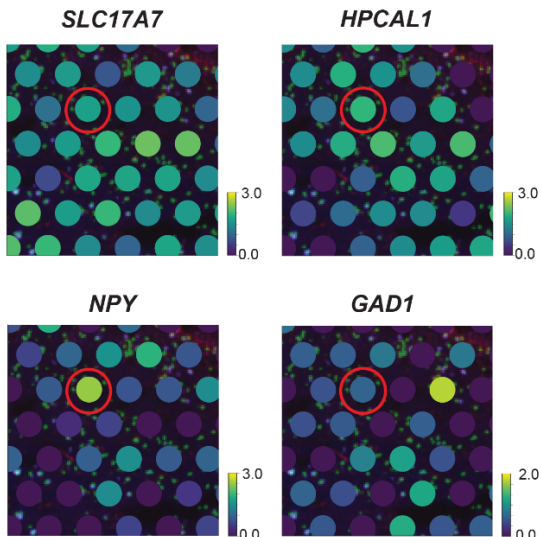
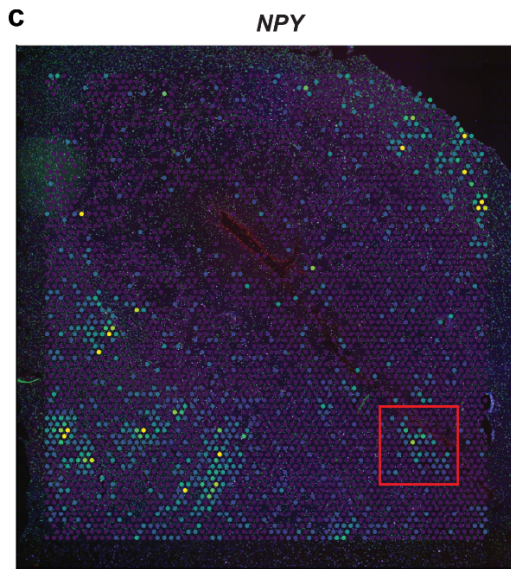
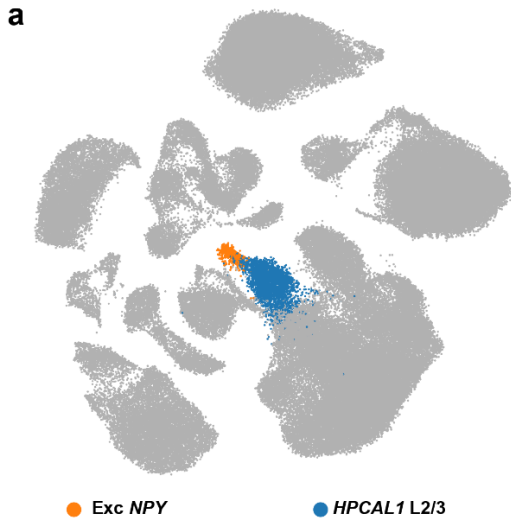
Supplementary Figure 6. The age-related changes in population distribution and gene expression.

- a) Samples distribution grouped by age (young adult macaques, 4-6 year-old, $n = 5$ macaques; middle-aged macaques, 13-15 year-old, $n = 3$ macaques). $*P < 0.05$ by two-tailed Mann-Whitney U test. Error bars, standard error of mean. The P -value for HPCAL1 is 0.136; for RORB, 0.037; for FEZF2, 0.037; for THEMIS, 0.371; for LAMP5, 0.136; for PVALB, 0.233; for SST, 0.233; for VIP, 0.037; for ADARB2_PAX6, 0.233; for AQP4, 0.072; for PDGFRA, 0.136; for MOG, 0.037; for C1QB, 0.551; for FLT1, 0.371.
- b) UMAP visualization of the 133,454 cells derived from young adult ($n = 80,495$ cells) and middle-aged ($n = 52,959$ cells) macaques V1 using the same color scheme as in Fig. 1b.
- c) Volcano plot showing the differentially expressed gene of PVALB between the young and middle-aged macaques. Genes that are significantly enriched (blue dots, determined by a two-part generalized linear model implemented by MAST) and exhibit $> 0.5 \log_2$ fold-change are marked in red.
- d) Strip charts showing the logarithmic-fold change (\log_2 FC) of all genes between young and middle-aged macaques across all cell populations. The dashed line at the top and bottom of the plot mark the 2-fold change threshold. The dashed line in the middle indicates no differential expression. Genes that are upregulated in young adult macaques have positive FCs. Genes that are differentially expressed (Two-sided Wilcoxon rank-sum test with FDR correction for multi-comparison) and $> 30\%$ max FC are colored.
- e) Pairwise comparison of gene expression between young and middle-aged macaques for selective cell populations. Red dots are genes that show $> 0.5 \log_2$ fold-change enrichment in middle-aged macaques. Blue dots are genes that show $> 0.5 \log_2$ fold change enrichment in young macaques. The solid line marks the equal expression; dashed lines indicate 2-fold enrichment. Gray dots, the mean expression of each gene of all cells of each population. All correlation rank-sum based Spearman's correlation coefficient (r) > 0.95 with $P < 0.0001$.
- f) Heatmaps showing the selective genes that are upregulated in young adult macaques (upper panel) or middle-aged macaques (lower panel) exclusively for selective cell populations. Normalized FC, normalized fold-change. Source data are provided as a Source Data file.



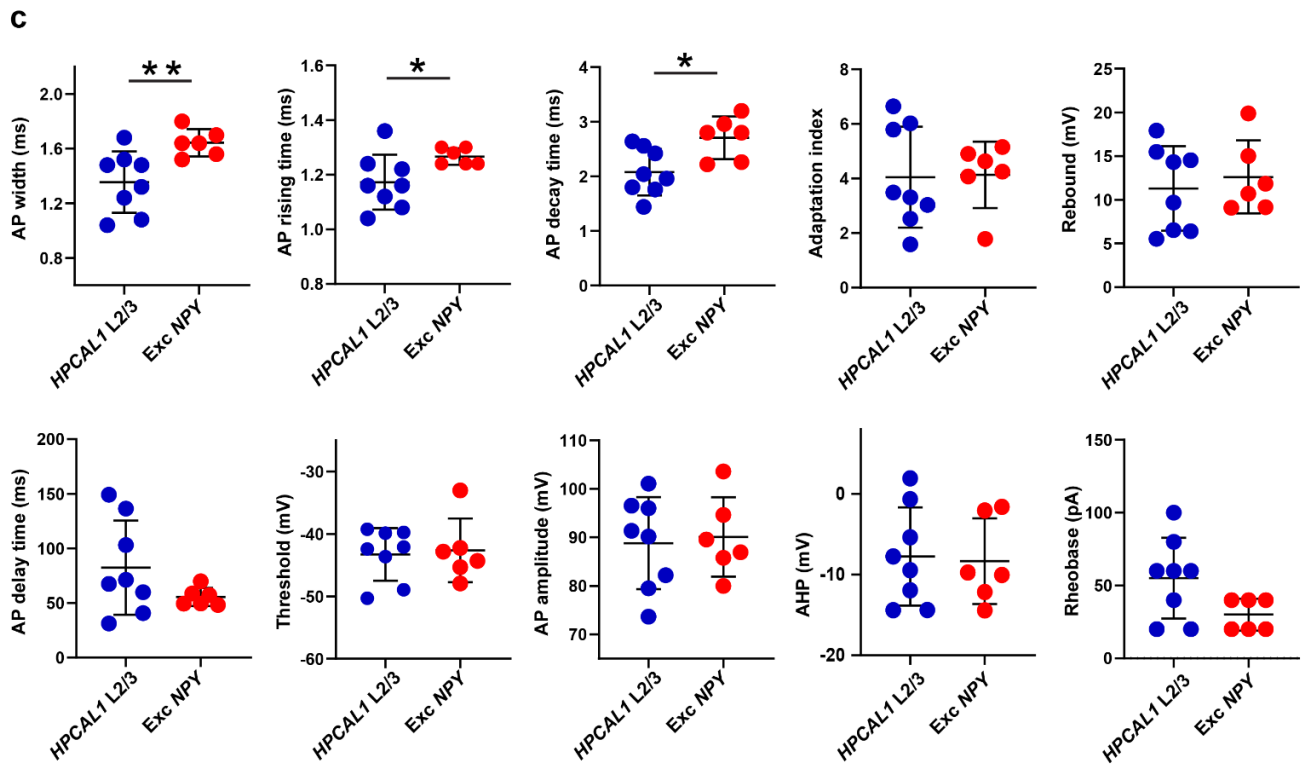
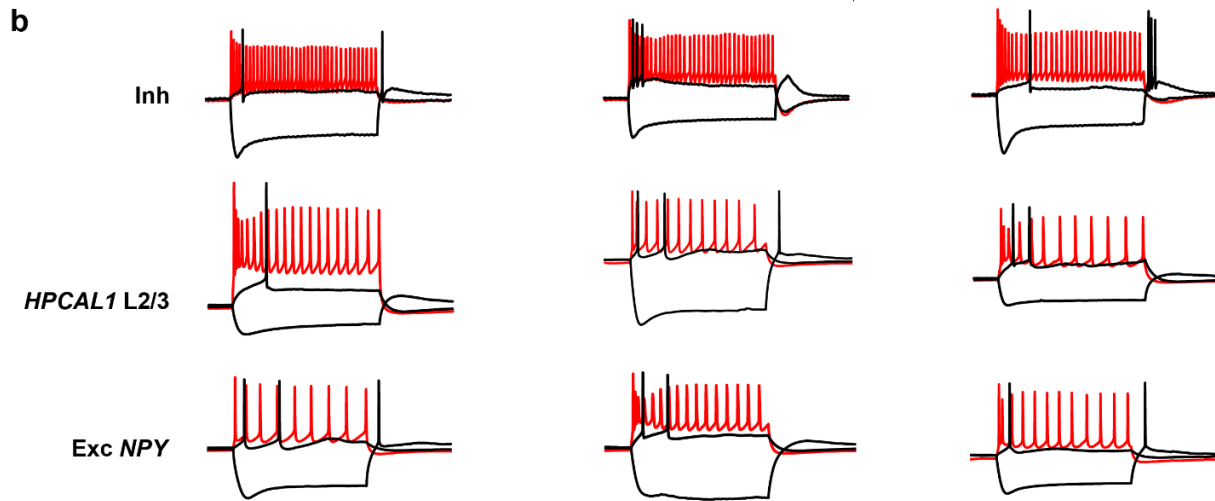
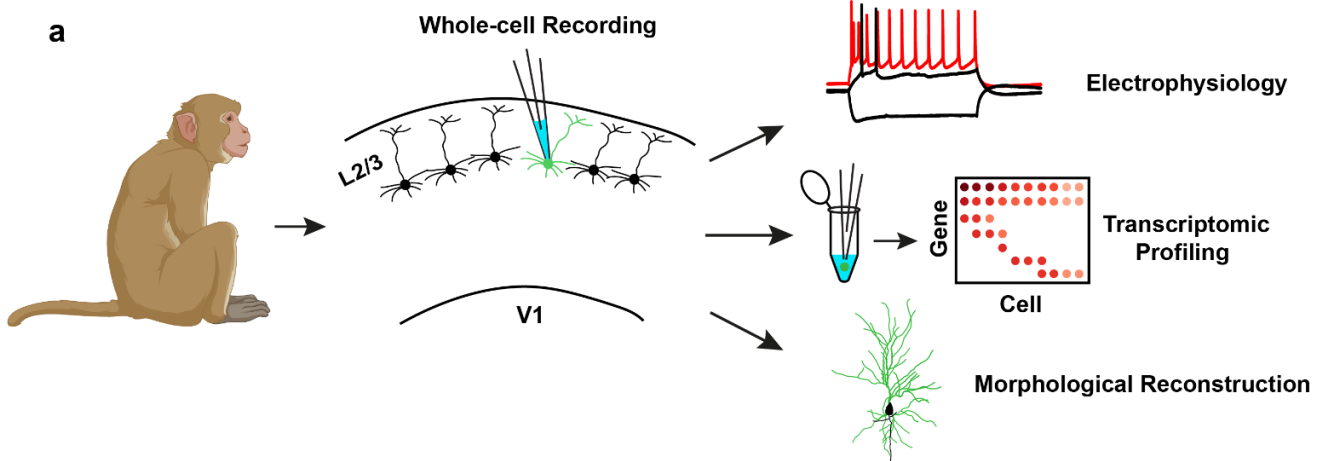
Supplementary Figure 7. Aligning non-neuronal cell types between macaque and human.

- a) UMAP visualization of non-neuronal cells, coloured by cell type. Astro, astrocytes; Oligo, oligodendrocytes; OPC, oligodendrocyte progenitor cells; Vasc, vascular cells.
- b) The distribution of the marker gene expression of non-neuronal cell types. The expression levels are normalized to the maximal of the cell counts that normalized to 10,000 reads per cell (NC).
- c) UMAP visualization of unsupervised alignment and clustering of the combination of macaque ($n = 73,381$ cells) and human non-neurons ($n = 4,753$ cells) using Harmony. Non-neuronal cells are colored by cell type (upper panel) and species (lower panel).
- d) Cell-type homology of macaque and human non-neuronal cells, estimated by shared cluster membership between the two species. Endo, endothelial cells; Peri, pericytes; VLMC, vascular leptomeningeal cells; Micro, microglia.



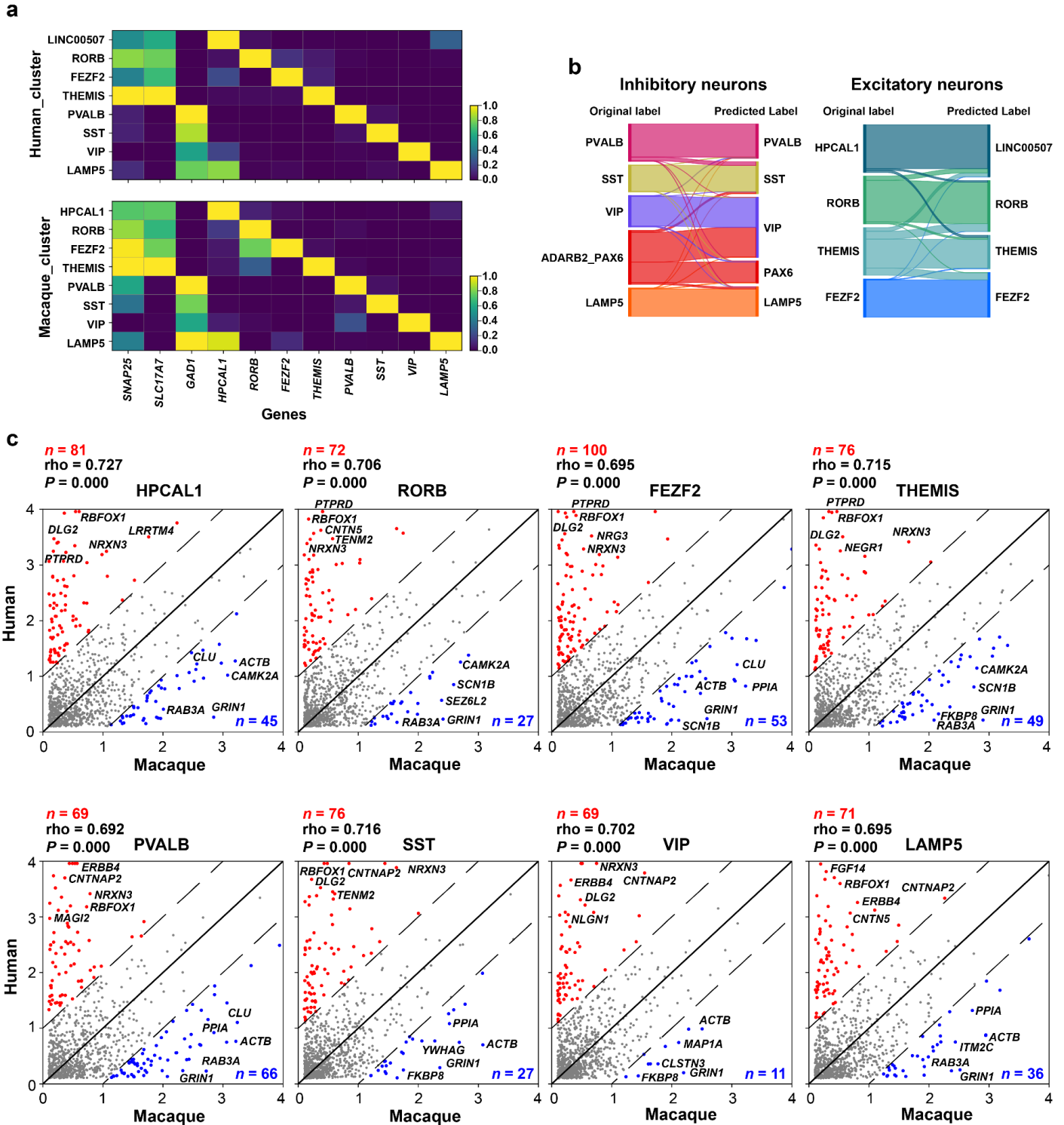
Supplementary Figure 8. Exc *NPY* cells are a distinct cell type and may also exist in the human cortex.

- a) UMAP plot visualizing the Exc *NPY* (Exc L1-3 *HPCAL1 NPY*) cell type (orange) and the *HPCAL1* L2/3 subclass (blue). Other cell subclasses are colored in gray.
- b) Violin plots show the distribution of Exc *NPY* marker genes expression across all excitatory cell types. The color label at the top matches the color scheme in Fig. 1b, e. The black square indicates the *HPCAL1* L2/3 subclass, which contains only one cell type Exc *NPY* (Exc L1-3 *HPCAL1 NPY*).
- c) Top panel, the expression of *NPY* of the human spatial transcriptomic dataset downloaded from the 10x Genomics database. Bottom panel: expression of *NPY*, *HPCAL1*, *SLC17A7* and *GAD1* genes in the wells indicated in the red box in the top panel. Notice the well in the red circle is *SLC17A7+HPCAL1+NPY+* but *GAD1-*, matching the expression pattern of Exc *NPY* cells.
- d) RNAscope mFISH of another *SLC17A7+HPCAL1+NPY+* positive cell. Red, *SLC17A7*; Green, *HPCAL1*; Light blue, *NPY*; Blue, DAPI. Scale bar, 10 μm . This staining and Fig. 4c, d were captured from the V1 of 3 different macaques. $n = 3$ macaques.
- e) The statistical result showing the quantification of RNAscope data represented as mean \pm s.d. from three macaques. The randomly selected positive cells are in the same slice as the Exc *NPY* cells. The statistical results are not significant, as determined by two-tailed Mann-Whitney U-test. The *P*-value for *SLC17A7* is 0.933; for *HPCAL1*, 0.776; for *NPY*, 0.602. Dots represent single cells. $n = 3$ macaques.
- f) Statistical analyses of RNAscope data represented as mean \pm s.d. from four macaques. The randomly selected positive cells are in the same slice as the Exc *NPY* cells. The statistical results are not significant, as determined by two-tailed Mann-Whitney U-test. The *P*-value for *HPCAL1* is 0.600, for *NPY*, 0.596; for *DRD3*, 0.537. Dots represent single cells. $n = 4$ macaques. Source data are provided as a Source Data file.



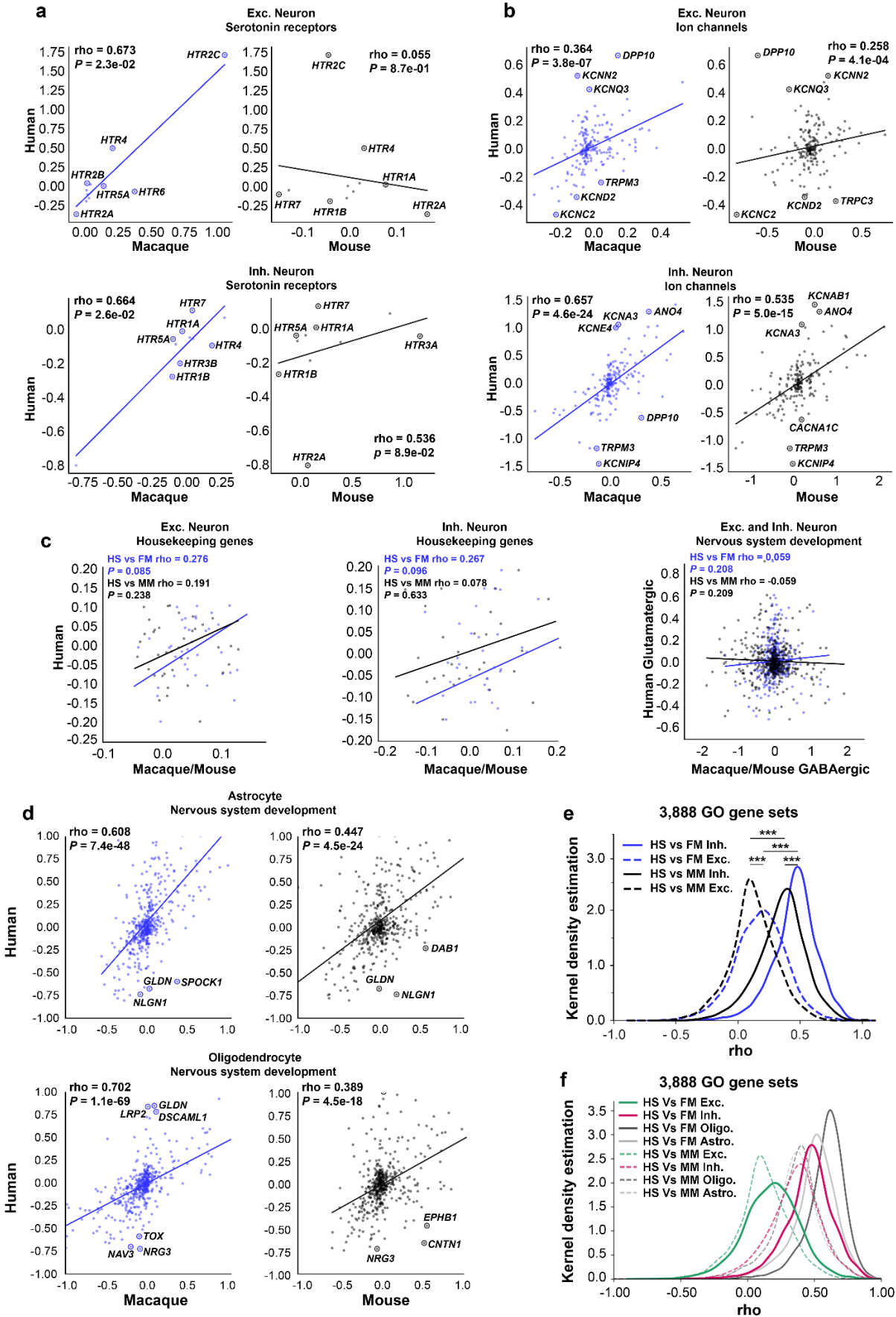
Supplementary Figure 9. Patch-seq protocol and physiological data in the macaque V1.

- a) Patch-seq involves the collection of morphological, physiological and transcriptomic data from the same neuron. Following electrophysiological recording and cell filling with biocytin via whole-cell patch clamp, the RNA of the cell is aspirated and processed for scRNA-seq. Drawing of macaque was created with BioRender.com.
- b) Spiking responses to step currents for the three morphologically defined cell types in Fig. 4 i.
- c) Plots show additional automatically extracted electrophysiological features. Circles represent single cells. Bars represent mean \pm s.d.. AP, action potential; AHP, afterhyperpolarization. The significance of differences in AP width, AP rising time and AP decay time was determined using two-tailed Mann-Whitney test (** $P < 0.01$; * $P < 0.05$). The P -value for AP is 0.008; for AP rising time, 0.029; for AP decay time, 0.019; for Adaptation index, 0.852; for Rebound, 0.662; for AP delay time, 0.223; for Threshold, 0.755; for AP amplitude, 0.999; for AHP, 0.662; for Rheobase, 0.087. The $n = 8$ for *HPCAL1* L2/3; $n = 6$ for Exc *NPY*.



Supplementary Figure 10. Comparison of cell types and gene enrichment between human and macaque.

- a) The express pattern of the canonical makers of human and macaque.
- b) River plots to compare the annotation of macaque neurons with predicted labels based on human dataset ⁵⁶.
- c) Comparison of the gene enrichment between human and macaque for the clusters HPCAL1, RORB, FEZF2, THEMIS, PVALB, SST, VIP and LAMP5. Solid line, equal expression; dashed lines: 2-fold enrichment. Red dots, high-diversity genes that are > 2-fold enrichment in humans. Top five genes labeled by texts. Gray dots, mean expression of each neuron differentiation-related gene ($n = 2,006$ genes) of all cells of each population. Blue dots, high-diversity genes that are > 2-fold enrichment in macaques with the top five genes labeled by text.



Supplementary Figure 11. The gene expression architecture is evolutionarily diverse for excitatory neurons between primates and mice.

- a) The serotonin receptor-related gene expression level of human excitatory (top row) and inhibitory (bottom row) neurons are plotted against that of macaque (left, blue) and mouse (right, black), using the median z-score of each cell type within species. The Spearman's Correlation's correlation coefficient (ρ) and significance were calculated and shown in each plot. The correlation is stronger between human and macaque than between human and mouse, for both excitatory neurons and inhibitory neurons.
- b) The ion channel-related gene expression level of human excitatory (top row) and inhibitory (bottom row) neurons are plotted against that of macaque (left panel, blue) and mouse (right panel, black).
- c) The housekeeping genes express level across species and the nervous system related-genes between excitatory and inhibitory neurons did not have significant correlation.
- d) The nervous system development-related gene expression of human astrocyte (top row) and oligodendrocyte (bottom row) are plotted against that of macaque (left panel, blue) and mouse (right panel, black).
- e) The distribution of Spearman's correlation coefficient (ρ) between human (HS) and macaque (FM, blue lines) or mouse (MM, black lines) for both excitatory (Exc., dashlines) and inhibitory neurons (Inh., solid lines), for 3888 GO gene sets. The correlations between human and macaque are stronger compared with between human and mouse, and are stronger between inhibitory neurons than between excitatory neurons. **** $P < 0.0001$ (P -value is $10e-30$) with Kruskal-wallis test followed by Dunn's test.
- f) The distribution of Spearman's correlation coefficient (ρ) between human (HS) and macaque (FM, solid lines) or mouse (MM, dashlines) for excitatory (Exc., green) and inhibitory neurons (Inh., red), and for oligodendrocytes (dark gray lines) and astrocytes (light gray lines) for 3888 GO gene sets. The correlations for glias (oligodendrocytes and astrocytes) are stronger compared with the neurons.