LCK-mediated RIPK3 Activation Controls Double-Positive Thymocyte Proliferation and Restrains Thymic Lymphoma by Regulating the PP2A-ERK Axis

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KEYWORDS

RIPK3, Thymic lymphoma, Double-positive (DP) thymocytes, LCK, PP2A, ERK

Supporting Information Figure Legends

Supporting information 1. Analysis of thymus- and spleen-derived immune cells from RIPK3-deficient mice.

A) The expression level of RIPK3 protein in the thymus (left panel) and spleen (right panel). RIPK3 protein level measured by Western blot analysis.

B) Thymi from $Ripk3^{+/+}$ and $Ripk3^{-/-}$ mice are shown.

C) Populations of dendritic cells (CD11c⁺) and macrophages (F4/80⁺) in thymic T cells. (n=7 for each group).

D) Spleen from $Ripk3^{+/+}$ and $Ripk3^{-/-}$ mice is shown (left) and its weight is shown in the graph (right). (n=5 for each group).

E) Representative H&E images of spleen from both *Ripk3^{+/+}* and *Ripk3^{-/-}* mice. (R: red pulp.
W: white pulp).

F) Splenic T cell populations from $Ripk3^{+/+}$ and $Ripk3^{-/-}$ mice. Average of the splenic T cell subsets and the absolute cell number by the indicated subsets are shown in each graph. (n=7 for each group).

G) Populations of naïve (CD62L^{hi}CD44^{lo}) and effector T cell (CD62L^{lo}CD44^{hi}) in the splenic T cells. (n=7 for each group).

H) Populations of dendritic cells (CD11c⁺) and macrophages (CD11c⁻F4/80^{lo}, CD11c⁻F4/80^{hi}) in splenic T cells. (n=7 for each group).

Statistical analyses were performed using the two-tailed unpaired Student *t*-test. P values below 0.05 were considered significant in the following manner.

Supporting information 2. RIPK1, RIPK3 and MLKL expression levels in various tissues and different cell types.

A) Total thymocytes from $Ripk3^{+/+}$ (n=6) and $Ripk3^{-/-}$ (n=6) mice were treated with TNF- α (50 ng/ml), z-VAD (20 μ M) and Smac mimetic (200 nM) for 5 h and 24 h. Dead cells were identified by Annexin V and PI staining.

B-D) Distribution of Ripk1 (B), Ripk3 (C) and Mlkl (D) gene expression in various immune

cell from BioGPS website.

E, F) Expression levels of MLKL in the indicated cell lines. MLKL protein level was measured by Western blot analysis (E) and Flow Cytometry (F).

Supporting information 3. Loss of RIPK3 does not impact the population and proliferation of T cells from spleen and peripheral lymph nodes in carcinogen-induced mouse model.

A) The ratio of thymus weight from control mice and ENU-injected mice were shown in the graph.

B) Spleen (left), peripheral lymph node (pLN; middle) and mesenteric lymph node (mLN; right) of each genotype is shown.

C-H) T cell populations of spleen (C), pLN (E) and mLN (G) from the indicated genotype, displayed as 2D plots of relative fluorescence of the indicated marker. Average of the T cell subsets are shown in each graph. Relative fluorescence of Ki-67 in CD4⁺ and CD8⁺ T cells. (C, D; Spleen, E, F; pLN, G, H; mLN). *Ripk3*^{+/+} and *Ripk3*^{-/-} littermates were analyzed at 60 days after three-time injections of ENU (n=10 -12 for each group).

Statistical analyses were performed using the two-tailed unpaired Student *t*-test. P values below 0.05 were considered significant in the following manner.

Supporting information 4. Tumor progression and incidence rate according to RIPK3 expression in thymic lymphoma-bearing Trp53-deficient mice

A, B) Western blotting analysis of RIPK3 expression level in tissues lysates from $p53^{-/-}$ background thymic lymphoma.

C) Tumors found in various organs and their frequency in $Ripk3^{+/+}p53^{-/-}$ and $Ripk3^{-/-}p53^{-/-}$ animals.

Supporting information 5. Protein Interacting domains between LCK and RIPK3

A) Tandem affinity purification of the tagged RIPK3 protein and interacting proteins using

streptavidin resin followed by calmodulin resin.

B) Amino acid sequence of human LCK (upper panel) and human RIPK3 (lower panel).

C, D) Western blot analysis after immunoprecipitation of human RIPK3 and human LCK in HEK293T cells. HEK293T cells were transfected with GST-LCK and/or Flag-RIPK3 expression constructs. Cells were harvested at 24h after transfection and cell lysates were immunoprecipitated with GST antibody (left) or RIPK3 antibody (right) (C). HEK293T cells were transfected with GST-LCK and/or Flag-RIPK3 expression constructs. Cells lysates were immunoprecipitated with RIPK3 antibody (left) or phosphor-Tyrosine antibody (right) (D).

Supporting information 6. Protein Interacting domains between RIPK3 and PP2A

A) Venn diagram represents the overlap of proteins and unique proteins identified by LC/MS/MS among TAP-purified samples as indicated. Total 503 proteins were identified as specific RIPK3 binding proteins.

B) Amino acid sequence of human RIPK3 (upper panel) and human PPP2R2A (lower panel).

C) In vitro kinase activity of RIPK3 toward PPP2R2A with ³²P-labeled ATP.

D) HEK293T cells were transfected with GFP-PPP2R2A and/or Flag-RIPK3 expression constructs. Cells were harvested at 24h after transfection.

Supporting information 7. Treatment of PP2A inhibitor or activator alters ERK phosphorylation status in thymic tumorigenesis.

A) Phosphorylation of ERK increased in response to PMA (1μ M) and ionomycin (3μ M) stimulation, which was further increased by LB-100 treatment.

B) Representative images of thymus from LB-100 or PBS injected mice (upper panel). LB-100 did not significantly change the thymus phenotype. Spleen, mesenteric lymph node and peripheral lymph node (lower) of each group were shown. (n=4 for each group)

C, D) Western blotting analysis of PPP2R2A and PPP2CA expression level in thymus tissues lysates from ENU-injected mice. The PP2A expression pattern was not affected by injection of LB-100 or SMAP. E) Representative images of thymus from SMAP or PBS injected mice (upper panel). Injection of SMAP decreased thymus size in ENU-injected $Ripk3^{-/-}$ mice. Spleen, mesenteric lymph node and peripheral lymph node (lower) of each group were shown. (n=4 - 6 for each group)

Statistical analyses were performed using the two-tailed unpaired Student *t*-test. P values below 0.05 were considered significant in the following manner: P<0.05, P<0.01, P<0.01, P<0.001.

















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> Recepter-interacting serine/threonine-protein kinase 3 (RIPK3) [Homo sapiens (human)]

MSCVKLWPSG	APAPLVSIEE	LENQELVGKG	GFGTVFRAQH	RKWGYDVAVK	IVNSKAISRE	VKAMASLDNE
FVLRLEGVIE	KVNWDQDPKP	ALVTKFMENG	SLSGLLQSQC	PRPWPLLCRL	LKEVVLGMFY	LHDQNPVLLH
R DLKPSNVLL	DPELHVKLAD	FGLSTFOGGS	OSGTGSGEPG	GTLGYLAPEL	FVNV <mark>N</mark> RKAST	ASDVYSFGIL
MWAVLAGREV	ELPTEPSLVY	EAVC <mark>N</mark> RONRP	SLAELPOAGP	ETPGLEGLKE	LMOLCWSSEP	KDRPSFOECL
PKTDEVFQMV	ENNMNAAVST	VKDFLSQLRS	SNRRFSIPES	GQGGTEMDGF	RRTIENQHSR	NDVMVSEWLN
KLNLEEPPSS	VPKKCPSLTK	RSRAQEEQVP	QAWTAGTSSD	SMAQPPQTPE	TSTFRNQMPS	PTSTGTPSPG
PRGNQGAERQ	GMNWSCRTPE	PNPVTGRPLV	NIYNCSGVOV	GDNNYLTMQQ	TTALPTWGLA	PSGKGRGLQH
PPPVGSQEGP	KDPEAWSRPQ	GWYNHSGK				

<mark>.</mark> : PP2A-Bα551 binding sites 🔛 : Kinase domain 🧾 : RHIM domain

> Serine/threonine-protein phosphatase 2A 55kDa regulatory subunit B alpha isoform 1 (PP2A-B55Bα) [Homo sapiens (human)]

MAGAGGGNDI	QWCFSQVKGA	VDDDVAEADI	ISTVEFNHSG	ELLATGDKGG	RVVIFQQEQE	NKIQSHSRGE
YNVYSTFQSH	EPEFDYLKSL	EIEEKINKIR	WLPQKNAAQF	LLSTNDKTIK	LWKISERDKR	PEGYNLKEED
GRYRDPTTVT	TLRVPVFRPM	DLMVEASPRR	IFANAHTYHI	NSISINSDYE	TYLSADDLRI	NLWHLEITDR
SFNIVDIKPA	NMEELTEVIT	AAEFHPNSCN	TFVYSSSKGT	IRLCDMRASA	LCDRHSKLFE	EPEDPSNR <mark>S</mark> F
FSEIISSISD	VKFSHSGRYM	MTRDYLSVKI	WDLNMENRPV	ETYQVHEYLR	SKLCSLY <mark>E</mark> ND	CIFDKFECCW
NGSDSVVMTG	SYNNFFRMFD	RNTKRDITLE	ASRENNKPRT	VLKPRKVCAS	GKRKKDEISV	DSLDFNKKIL
HTAWHPKENI	IAVATTNNLY	IFQDKVN				

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: RIPK3 binding sites
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