



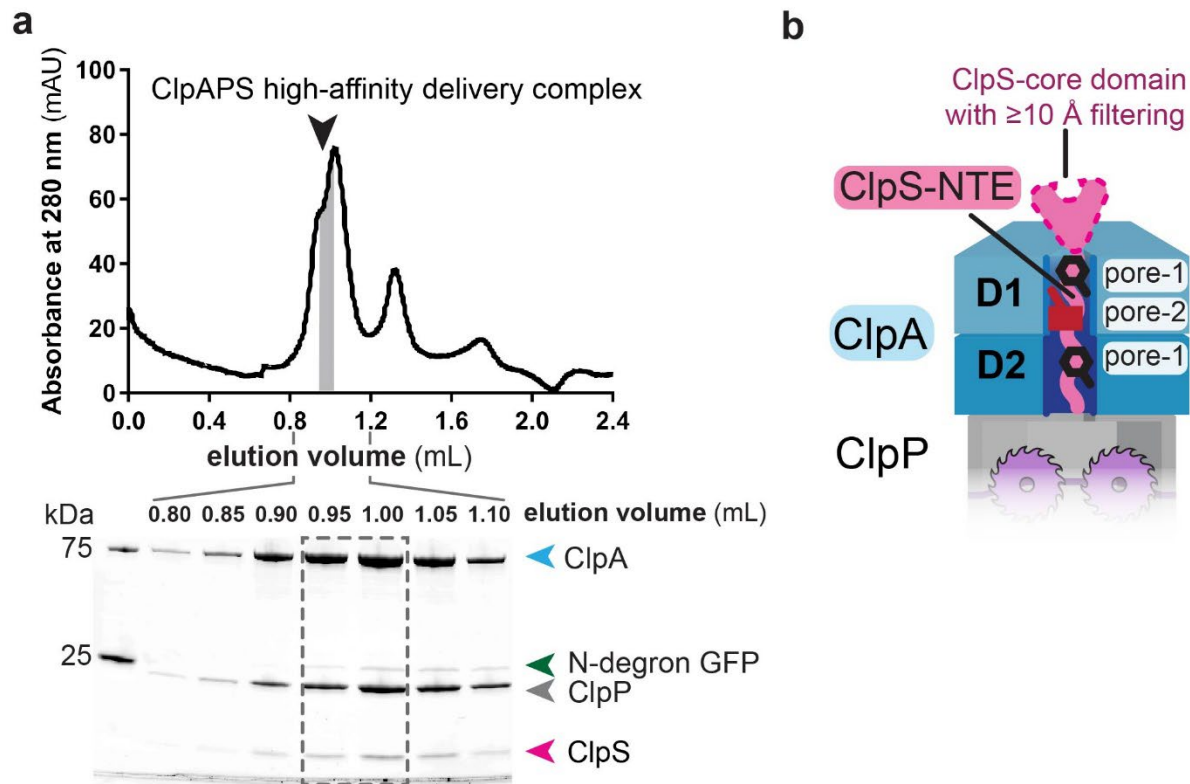
AAA+ protease-adaptor structures reveal altered conformations and ring specialization

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SUPPLEMENTARY INFORMATION

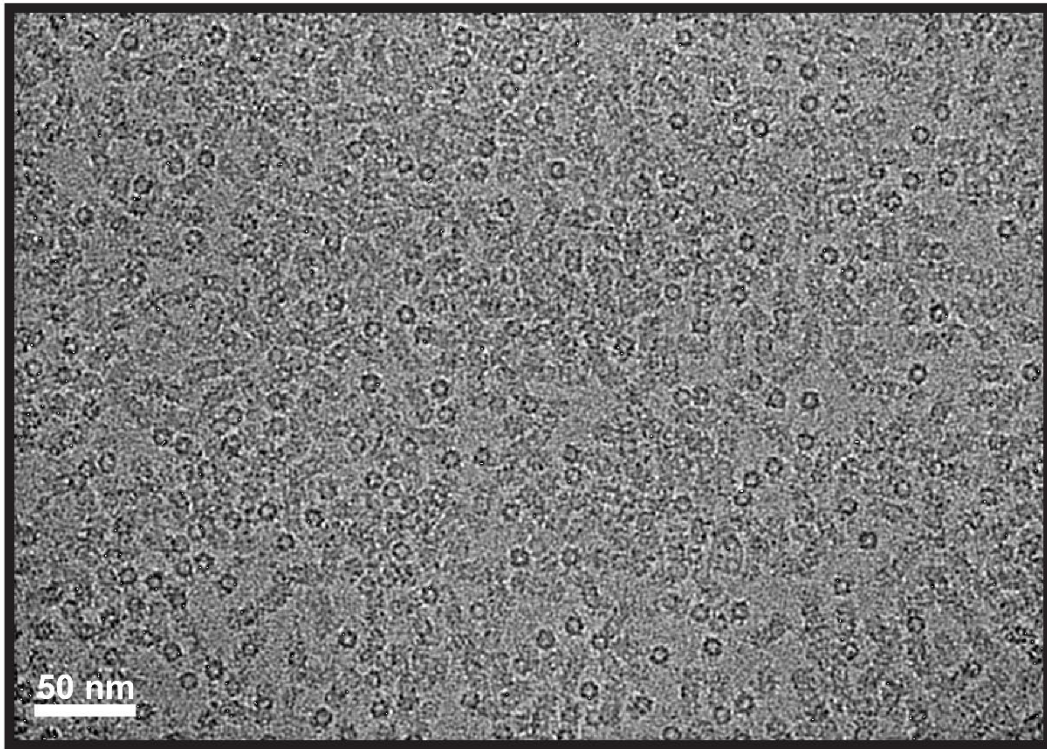
Supplementary Table 1. Oligonucleotides used in this study.

Oligonucleotide Name	Purpose	Oligonucleotide Sequence (5' to 3')
D1 pore-2 QTQ For	Round-the-horn mutagenesis of QTQClpA	CAAGGTGGTCAGGTCGATGCG
D1 pore-2 QTQ Rev	Round-the-horn mutagenesis of QTQClpA	AGTCTGACCCGCACCGATAATGG
D1 pore-2 G For	Round-the-horn mutagenesis of GGGClpA	GGAGGTGGTCAGGTCGATGCG
D1 pore-2 GG Rev	Round-the-horn mutagenesis of GGGClpA	GCCTCCACCCGCACCGATAATGG
D1 pore-2 298 For	Round-the-horn mutagenesis of $\Delta^{295-297}$ ClpA and KYRClpA	GGTGGTCAGGTCGATGCG
D1 pore-2 294 Rev	Round-the-horn mutagenesis of $\Delta^{295-297}$ ClpA	ACCCGCACCGATAATGG
D1 pore-2 KYR Rev	Round-the-horn mutagenesis of KYRClpA	ACGATATTTACCCGCACCGATAATGG



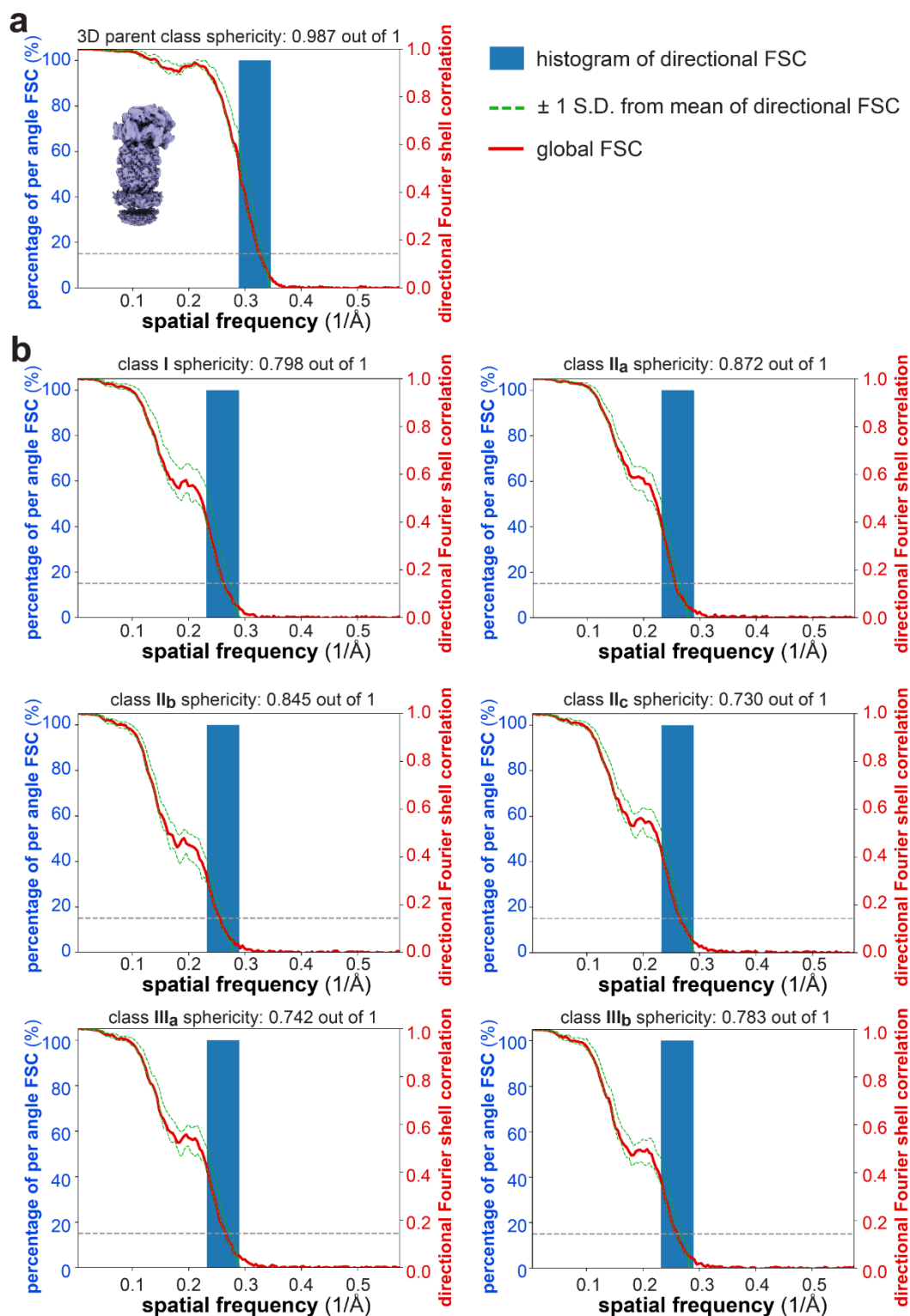
Supplementary Figure 1. Overview of ClpAPS cryo-EM structural features.

a, Size-exclusion chromatography (top panel) of a complex of ClpA, ClpP, ClpS, and YLFVQELA-GFP (an N-degron substrate) assayed by SDS-PAGE (bottom panel). Gray shaded area and boxed area indicate the fractions pooled for cryo-EM. Preparation of ClpAPS•YLFVQELA-GFP complexes by SEC was performed independently five times (during initial grid screening and optimization); only the shown SEC-purified sample was used to prepare the single grid imaged for data collection. Uncropped gel image at the end of the SI file. **b**, Cartoon of pore loops that interact with the ClpS NTE and proteins resolved in cryo-EM structures.



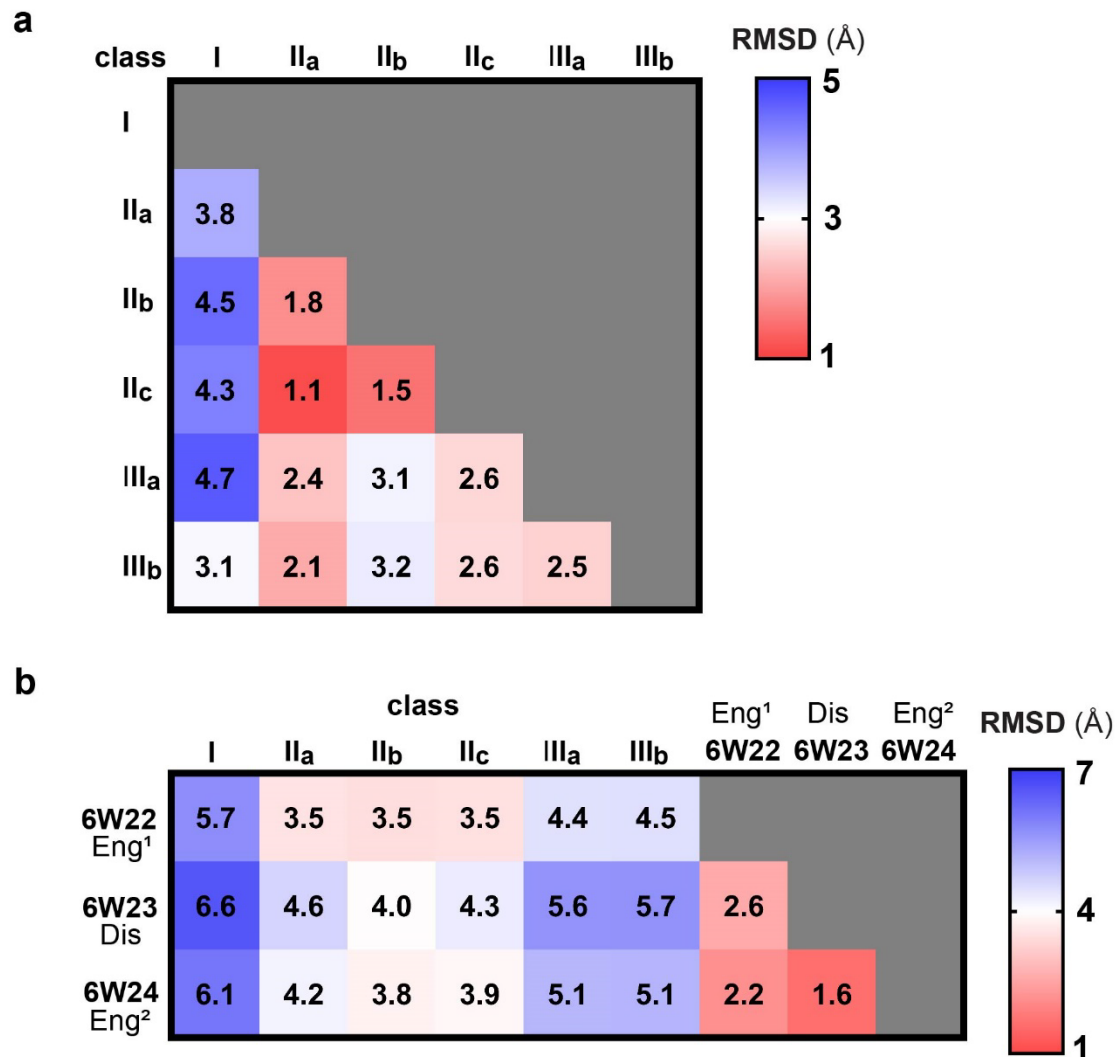
Supplementary Figure 2. ClpAPS cryo-EM micrograph.

Representative cryo-EM micrograph of ClpAPS•N-degron GFP complexes from cryo-EM data collection (9,169 micrographs collected total). The scale bar corresponds to 50 nm. The majority of particles are doubly capped with two ClpA₆ molecules per ClpP₁₄.



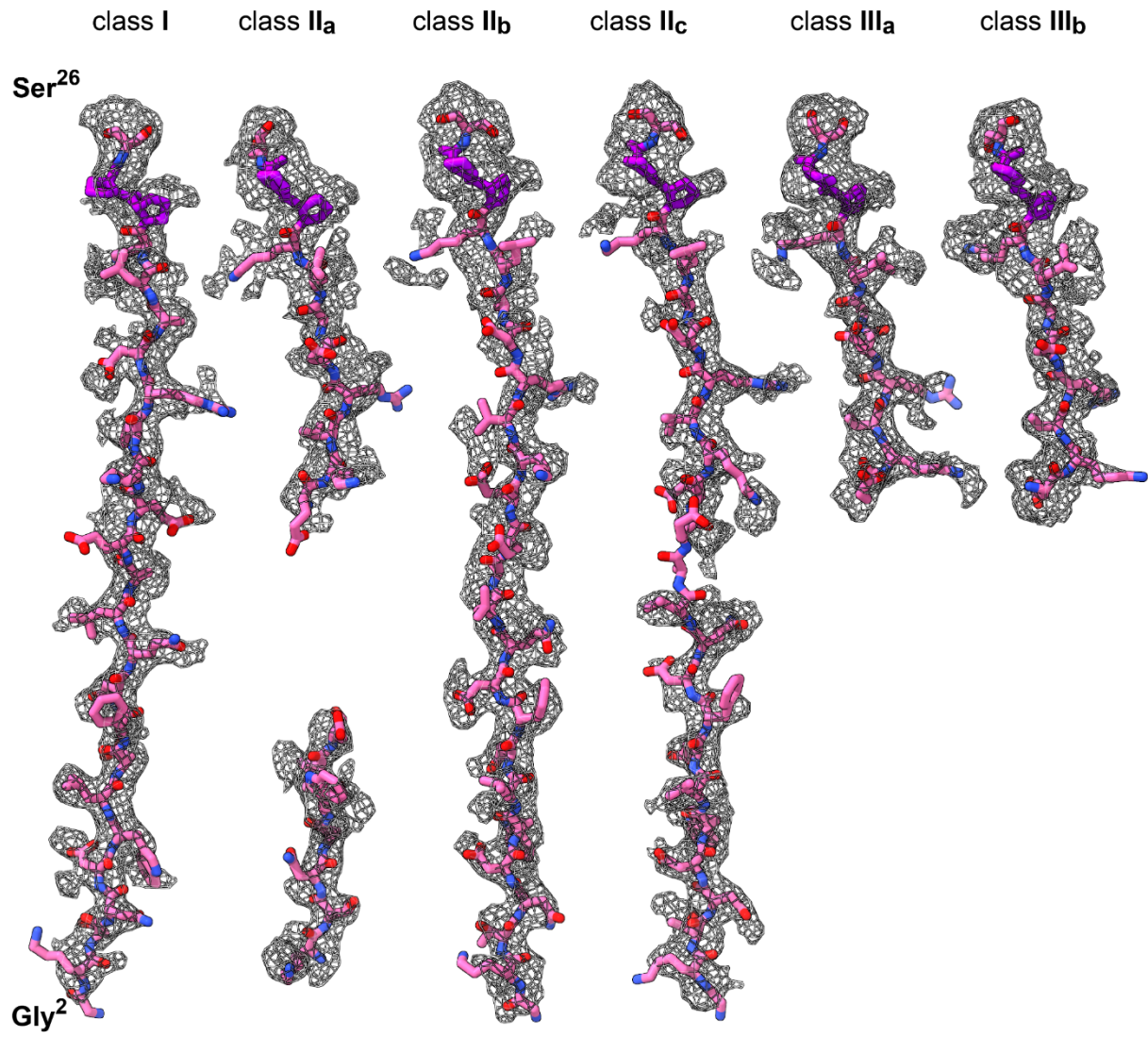
Supplementary Figure 3. 3DFSC sphericity

3DFSC plots of classes I, II, and III and parent class prior to focused classification. Due to non-uniform box dimensions following density modification, intermediate maps from RELION-3 (from 3D auto-refine) were used as 3DFSC inputs for class I, II, and III structures.



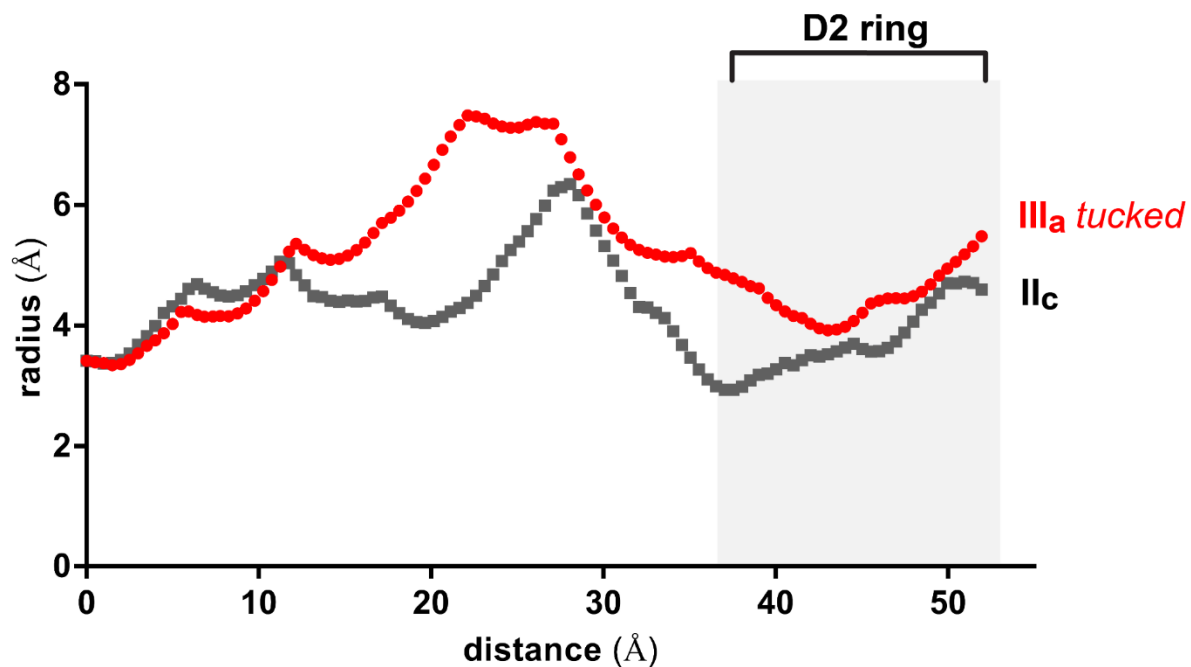
Supplementary Figure 4. C α RMSD analysis of ClpAP structures.

a, Pairwise RMSD (Å) calculations of class I, II, and III subclasses between ClpA C α atoms in subunits A through F using PyMOL “align”. **b**, Pairwise RMSD comparisons of ClpA C α atoms in ClpAPS cryo-EM structures and ClpAP•RepA-GFP structures (PDB 6W22, 6W23, and 6W24), using PyMOL “align”.

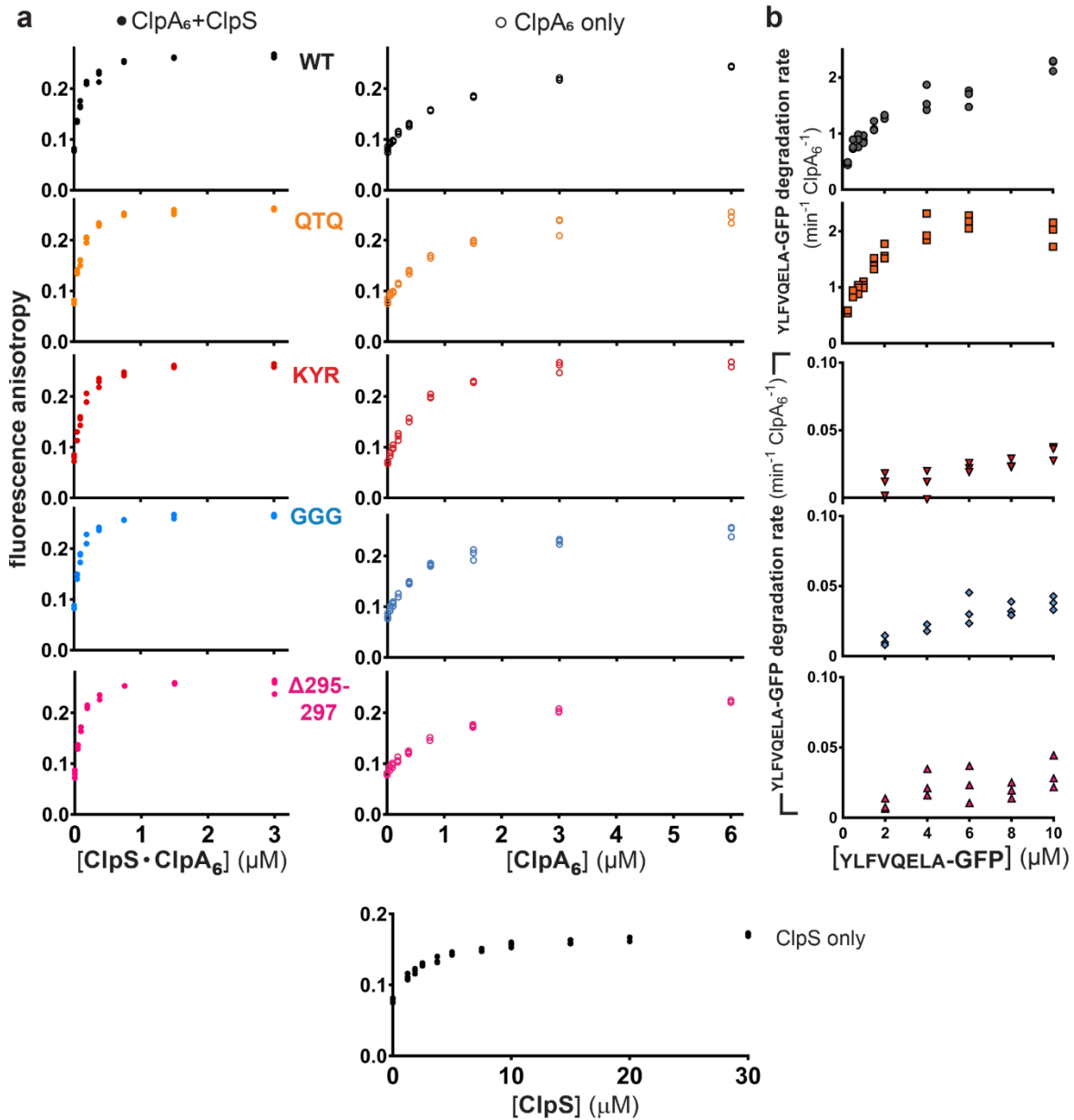


Supplementary Figure 5. ClpS NTE map-model comparisons.

ClpS NTE density (mesh at contour level 0.25) and atomic models (shown in sticks) in the ClpA axial channel in classes I, II, and III. The junction sequence residues, Pro²⁴ and Pro²⁵, are colored purple, whereas the rest of the NTE is colored pink.



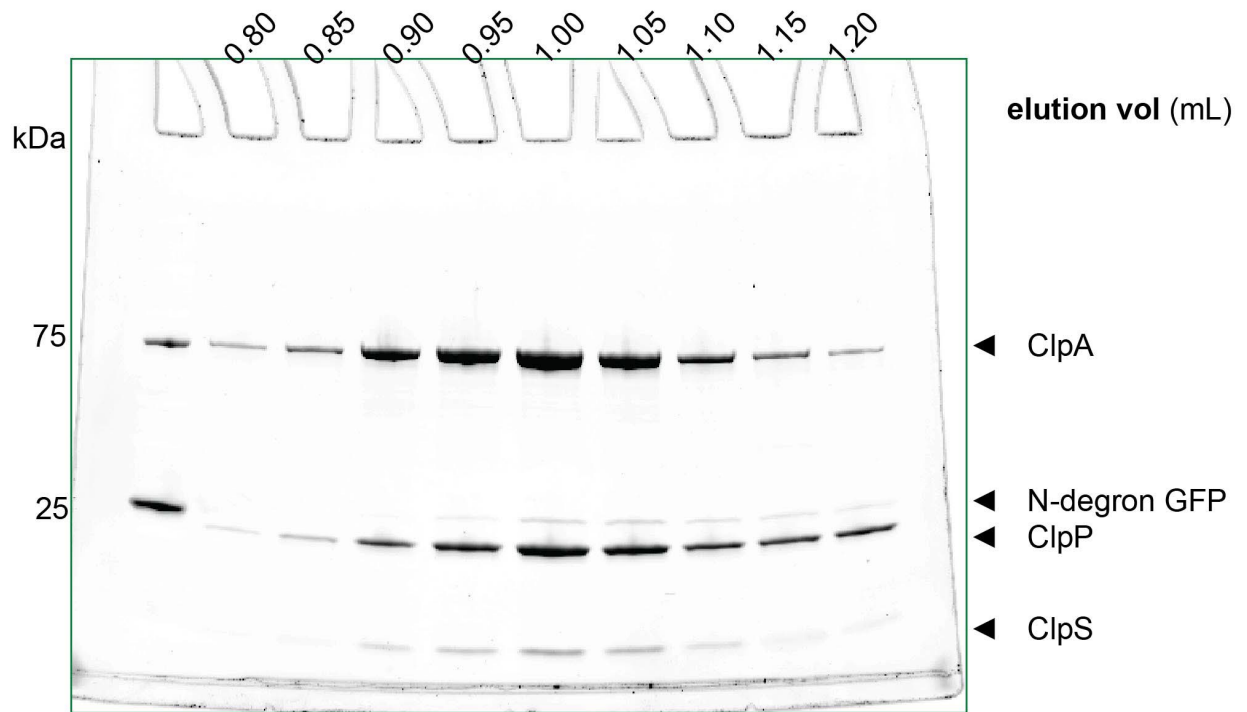
Supplementary Figure 6. Axial channel radius comparison. Plot of channel radius vs. channel distance (length of channel from top to bottom of ClpA) calculated by CAVER¹ in class-II_c (dark gray squares) and class-III_a (red circles) atomic models. Portion of axial channel in D2 ring is indicated by the light gray box.



Supplementary Figure 7. Individual fluorescence anisotropy and YLFVQELA-GFP degradation rate values from Fig. 5a–b. **a**, Fluorescence anisotropy of ClpA pore-2 variants alone (ClpA₆, open circles), with an equimolar mixture with ClpS (+ClpS, filled circles), or ClpS only (bottom right panel), titrated in increasing concentrations against a fixed concentration of fluorescein-labeled N-degron peptide. **b**, Kinetic analysis of YLFVQELA-GFP degradation by ClpA D1 pore-2 variants (see *Methods* for concentrations). The graphed y-axis of WT and the KYR variant ranges from 0–2.5 min⁻¹ ClpA₆⁻¹, whereas the y-axis shown for KYR, GGG, and Δ295–297 variants ranges from 0–0.10 min⁻¹ ClpA₆⁻¹.

SUPPLEMENTARY REFERENCES

1. Jurcik, A., Bednar, D., Byska, J., Marques, S. M., Furmanova, K., Daniel, L., Kokkonen, P., Brezovsky, J., Strnad, O., Stourac, J., Pavelka, A., Manak, M., Damborsky, J., & Kozlikova, B. CAVER Analyst 2.0: analysis and visualization of channels and tunnels in protein structures and molecular dynamics trajectories. *Bioinformatics (Oxford, England)*, 34(20), 3586–3588 (2018).



Unprocessed gel image for Supplementary Fig. 1a. SDS-PAGE gel (stained with SYPRO Red) of ClpAPS•YLFVQELA-GFP complexes prepared by SEC.