

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

For Plant materials growth: PHCbi MLR-352H-PC;
For confocal microscopy: Leica TCS SP8;
For qRT-PCR: LightCycler480 II Real-Time PCR System (Roche);
For protein purification: Akta pure 25 system;
For protein gel Images: GE Amersham Imager 680(AI 680);
For viral inoculation phenotype and yeast two-hybrid images: Nikon D7100 camera;

Data analysis

The qRT-PCR datasets were performed by Microsoft Excel 2019 and Origin 8 software, the analysis using One-way ANOVA followed by two-side students's t-test and Fisher's LSD test;
The images from western blot and confocal microscope were cut using Adobe Photoshop CS6 and Adobe Illustrator 2020, and some information represented of their intensity were analyzed by Image J software;
The length of rice seedlings about the second leaf sheaths and the primary root were manually measured by ruler and then analyzed by Origin 8.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

The authors declare that all raw data supporting the findings of this study can be found within the paper and its Supplementary Files. Source data are provided with this paper.

Human research participants

Policy information about [studies involving human research participants and Sex and Gender in Research](#).

Reporting on sex and gender	N/A
Population characteristics	N/A
Recruitment	N/A
Ethics oversight	N/A

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Describe how sample size was determined, detailing any statistical methods used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.
Data exclusions	Describe any data exclusions. If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.
Replication	Experiment were independently repeated at least three times and similar results were obtained. The information has been provided in source data files.
Randomization	Related plant materials were grown on the same condition and randomly allocated in the growth chamber. The samples were collected randomly without any bias.
Blinding	For all experiments involving phenotypes with a randomized block design, the distribution of each family in the field was generated randomly before planting.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involvement
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Involvement
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Antibodies

Antibodies used

ProteinFind® Anti-DYKDDDDK Mouse Monoclonal Antibody, TransGen, Cat#HT201-01, 1:5000 dilution;
 ProteinFind® Anti-c-Myc Mouse Monoclonal Antibody, TransGen, Cat#HT101-01, 1:5000 dilution;
 Anti-Myc tag antibody - ChIP Grade, abcam, Cat#ab9132, 1:5000 dilution;
 ProteinFind® Anti-GFP Mouse Monoclonal Antibody, TransGen, Cat#HT801-01, 1:5000 dilution;
 ProteinFind® Anti-HA Mouse Monoclonal Antibody, TransGen, Cat#HT301-01, 1:5000 dilution;
 Anti-6X His tag® antibody, abcam, Cat#ab18184, 1:3000 dilution;
 GST-tag Antibody [HRP], pAb, Rabbit, Genscript, Cat#A00130, 1:5000 dilution;
 SLR1 Rabbit mAb, ABclonal, Cat#A18329, 1:5000 dilution;
 ProteinFind® Goat Anti-Mouse IgG (H+L), HRP Conjugate, TransGen, Cat#HS201-01, 1:10000 dilution;
 ProteinFind® Goat Anti-Rabbit IgG (H+L), HRP Conjugate, TransGen, Cat#HS101-01, 1:10000 dilution;
 Rabbit Anti-Goat IgG H&L (HRP), abcam, Cat#ab6741, 1:10000 dilution;
 Actin mAb for PLANTs, Cat#M20009, 1:5000 dilution;

Validation

SRBSDV P10 and RSV CP antibodies are made from Zhejiang University;
 All other antibody used in this paper were certified and validated by manufactures.
 The details about anti-DYKDDDDK is in https://www.transgen.com.cn/antibody_tag/371.html;
 The details about anti-myc is in https://www.transgen.com.cn/antibody_tag/363.html;
 The details about anti-GFP is in https://www.transgen.com.cn/antibody_tag/390.html;
 The details about anti-HA is in https://www.transgen.com.cn/antibody_tag/378.html;
 The details about anti-GST is in https://www.genscript.com.cn/antibody/A00130-40-GST_tagAntibody_HRP_pAb_Rabbit.html;
 The details about Anti-6X His tag® antibody [HIS.H8] is in <https://www.abcam.com/6x-his-tag-antibody-hish8-ab18184.html>;
 The details about anti-actin is in <http://www.ab-mart.com.cn/list.aspx?node=59>;
 The details about anti-SLR1 is in <https://abclonal.com.cn/catalog/A21231>;
 The details about Rabbit Anti-Goat IgG H&L (HRP) is in <https://www.abcam.com/rabbit-goat-igg-hl-hrp-ab6741.html>;
 The details about Anti-Myc tag antibody - ChIP Grade is in <https://www.abcam.com/myc-tag-antibody-chip-grade-ab9132.html>;
 The details about Goat Anti-Rabbit IgG (H+L), HRP is in https://www.transgen.com.cn/antibody_second/397.html;
 The details about Goat Anti-Mouse IgG (H+L), HRP is in https://www.transgen.com.cn/antibody_second/403.html;