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Supplemental Information

**Condensation of the fusion focus by the
intrinsically disordered region of the
formin Fus1 is essential for cell-cell fusion**

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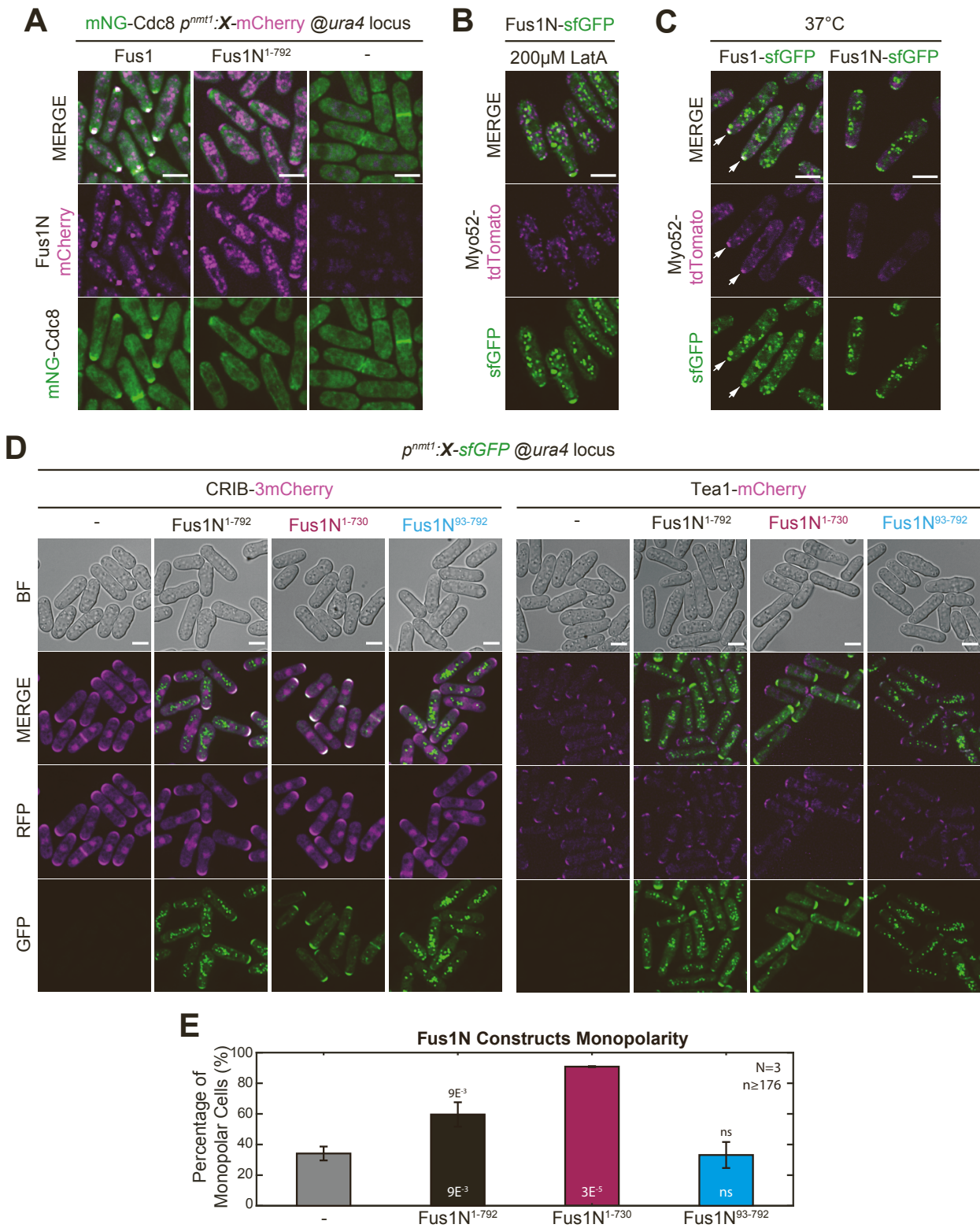


Figure S1 . Additional localization data and Fus1N tip localization effect on polarity. Related to Figure 1.

A. Interphase cells expressing mNeonGreen-Cdc8, either in combination with (left) full length Fus1-sfGFP or (middle) Fus1N-sfGFP (Fus1¹⁻⁷⁹²) or (right) alone. **B.** Interphase cells expressing Myo52-tdTomato and Fus1N-sfGFP (Fus1¹⁻⁷⁹²). Cells were treated with 200μM LatrunculinA for 5 minutes. **C.** Interphase cells expressing Myo52-tdTomato and either (left) full length Fus1-sfGFP or (right) Fus1N-sfGFP (Fus1¹⁻⁷⁹²) grown for 6h at 37°C and imaged at 40°C. White arrows mark resistant fusion focus-like structure. **D.** DIC

and fluorescence images of either (left) the polarity marker CRIB-3mCherry or (right) Tea1-mCherry in interphase WT or Fus1N¹⁻⁷⁹², Fus1N¹⁻⁷³⁰ or Fus1N⁹³⁻⁷⁹²-expressing cells. **E.** Monopolarity of the strains as in (D), assessed from the localization of CRIB on a single snapshot. Of note, a fraction of WT cells appear monopolar using this assay, because they are either before NETO or at a time point in CRIB oscillations⁵² where only one tip is decorated. All p-values are relative to WT. Bars are 5µm.

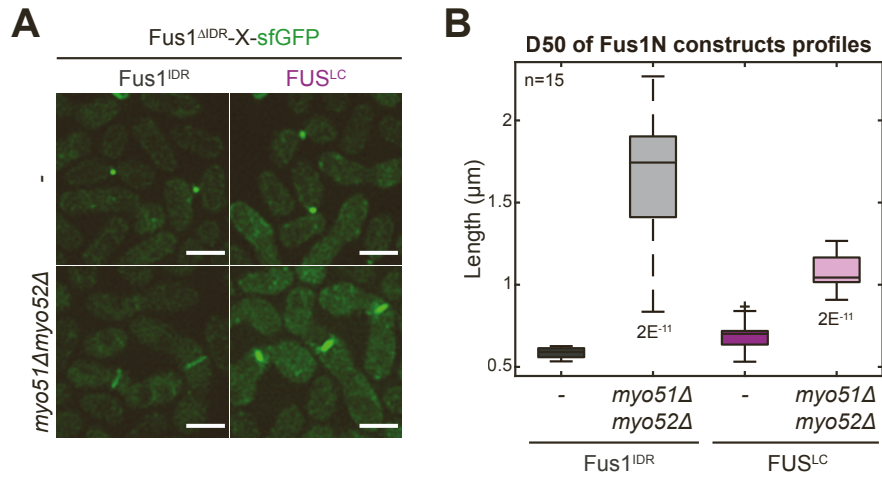


Figure S2. Type V myosins are required for fusion focus focalization in both WT cells and cells in which Fus1 IDR was replaced by FUS^{LC}. Related to Figure 4.

A. Images of (left) Fus1-sfGFP or (right) Fus1¹⁻⁴⁹¹-FUS^{LC}-Fus1⁷⁹²⁻¹³⁷²-sfGFP in mating cell pairs that are otherwise WT (top) or carry deletions of *myo51Δ* and *myo52Δ* (bottom). **B.** Width at half maximum of GFP-fluorescence profiles in strains as in (A). p-values compare WT Fus1 and Fus1¹⁻⁴⁹¹-FUS^{LC}-Fus1⁷⁹²⁻¹³⁷² in each background. Bars are 5µm.

Table S1: Strains used in this study and links to figures. Related to STAR Methods.

GENOTYPE	FIGURES	STRAIN
h90 myo52-tdTomato:natMX fus1-sfGFP:kanMX ura4- leu1-32 ade6-M216	1, 4, S2	YSM3312
h90 myo52-tdTomato:natMX fus1Δ::LEU2+ ura4-294:p ^{fus1} -fus1N ¹⁻⁷⁹² -fus1C ⁷⁹³⁻¹³⁷² -sfGFP:ura4+ leu1-32	1	YSM2504
h90 myo52-tdTomato:natMX fus1Δ::LEU2+ ura4-294:p ^{fus1} -cdc12N ¹⁻⁸⁸⁷ -fus1C ⁷⁹³⁻¹³⁷² -sfGFP:ura4+ leu1-32	1	YSM2512
h90 myo52-tdTomato:natMX fus1Δ::LEU2+ ura4-294:p ^{fus1} -for3N ¹⁻⁷¹⁴ -fus1C ⁷⁹³⁻¹³⁷² -sfGFP:ura4+ leu1-32	1	YSM2510
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	1, 3, S1	YSM4002
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷³⁰ -sfGFP:term ^{nmt} leu1-32 ade6-M210	1, 3	YSM4003
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁵⁰⁰ -sfGFP:term ^{nmt} leu1-32 ade6-M210	1, 3	YSM4004
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ⁹³⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	1	YSM4005
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁴⁰⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	1	YSM4006
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁹¹⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	1, 3	YSM4007
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ⁴³¹⁻⁷⁵⁵ -sfGFP:term ^{nmt} leu1-32 ade6-M210	1	YSM4008
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1-sfGFP:term ^{nmt} leu1-32 ade6-M210	1, S1	YSM4009
h90 leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ ura4-D18 ade6-M216	S1	YSM3786
h90 ura4+:p ^{nmt1} :fus1-mCherry:term ^{nmt} leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ ade6-M216	S1	YSM4042
h90 ura4+:p ^{nmt1} :fus1N ¹⁻⁷⁹² -mCherry:term ^{nmt} leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ ade6-M216	S1	YSM4043
h+ his5+:p ^{act1} :CRIB-3mCherry:bsdMX ura4-D18	S1	YSM4010
h90 his5+:p ^{act1} :CRIB-3mCherry:bsdMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32	S1	YSM4011
h90 his5+:p ^{act1} :CRIB-3mCherry:bsdMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷³⁰ -sfGFP:term ^{nmt} ade6-M210	S1	YSM4012
h90 his5+:p ^{act1} :CRIB-3mCherry:bsdMX ura4+:p ^{nmt1} :fus1N ⁹³⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	S1	YSM4013
h90 tea1-mCherry:kanMX ura4-D18 leu1-32	S1	YSM4014
h90 tea1-mCherry:kanMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32	S1	YSM4015
h90 tea1-mCherry:kanMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷³⁰ -sfGFP:term ^{nmt} leu1-32	S1	YSM4016
h90 tea1-mCherry:kanMX ura4+:p ^{nmt1} :fus1N ⁹³⁻⁷⁹² -sfGFP:term ^{nmt} leu1-32 ade6-M210	S1	YSM4017
h90 myo52-tdTomato:natMX fus1-sfGFP:kanMX	2	YSM3888
h90 myo52-tdTomato:natMX fus1-sfGFP:kanMX acp2Δ::bleMX ura4- leu1-32 ade6-M210	2	YSM3314
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ⁹³⁻⁷⁹² -sfGFP:term ^{nmt} fus1Δ::hphMX ade6-M210 leu1-32	2	YSM4018
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1-sfGFP:term ^{nmt} fus1Δ::hphMX ade6-M210 leu1-32	2	YSM4053
h90 myo52-tdTomato:natMX ura4-294:p ^{fus1} :fus1N-sfGFP:ura4+ fus1Δ::LEU2+ leu1-32	3	YSM2486
h90 myo52-tdTomato:natMX ura4-294:p ^{fus1} :fus1N-sfGFP:ura4+ leu1-32	3	YSM2699
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷⁹² -sfGFP:term ^{nmt} fus1Δ::hphMX leu1-32 ade6-M210	3	YSM4054
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁹¹⁻⁷⁹² -sfGFP:term ^{nmt} fus1Δ::hphMX leu1-32 ade6-M210	3	YSM4055
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁷³⁰ -sfGFP:term ^{nmt} fus1Δ::hphMX leu1-32 ade6-M210	3	YSM4056
h90 myo52-tdTomato:natMX ura4+:p ^{nmt1} :fus1N ¹⁻⁵⁰⁰ -sfGFP:term ^{nmt} fus1Δ::hphMX leu1-32 ade6-M210	3	YSM4057
h90 myo52-tdTomato:natMX fus1 ^{Δ501-749} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3	YSM4019
h90 myo52-tdTomato:natMX fus1 ^{Δ501-791} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3	YSM4020
h90 myo52-tdTomato:natMX fus1 ^{Δ492-791} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3, 4	YSM4021
h90 myo52-tdTomato:natMX fus1 ^{Δ492-500} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3	YSM4044
h90 myo52-tdTomato:natMX fus1 ^{Δ731-791} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3	YSM4045
h90 myo52-tdTomato:natMX fus1 ^{Δ492-749} -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	3	YSM4046
h90 myo52-tdTomato:natMX fus1 ¹⁻⁴⁹¹ -FUS ^{12E} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	4	YSM4022
h90 myo52-tdTomato:natMX fus1 ¹⁻⁴⁹¹ -FUS-fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	4, S2	YSM4023
h90 myo52-tdTomato:natMX fus1 ¹⁻⁴⁹¹ -CRY2 ^{PHR} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	4	YSM4024
h90 myo52-tdTomato:natMX fus1 ¹⁻⁴⁹¹ -CRY2 ^{olig} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	4	YSM4025
h90 myo52-tdTomato:natMX fus1 ¹⁻⁴⁹¹ -FUS ^{G156E} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX ura4-294 leu1-32 ade6-M210	4	YSM4047
h90 myo52-tdTomato:natMX leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ fus1:kanMX ura4-294 ade6-M210	4	YSM4026
h90 myo52-tdTomato:natMX leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ fus1 ¹⁻⁴⁹¹ -CRY2 ^{PHR} -fus1 ⁷⁹²⁻¹³⁷² :kanMX ura4-294 ade6-M210	4	YSM4048
h90 myo52-tdTomato:natMX leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ fus1 ¹⁻⁴⁹¹ -CRY2 ^{olig} -fus1 ⁷⁹²⁻¹³⁷² :kanMX ura4-294 ade6-M210	4	YSM4049
h90 myo52-tdTomato:natMX leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ fus1 ¹⁻⁴⁹¹ -FUS-fus1 ⁷⁹²⁻¹³⁷² :kanMX ura4-294 ade6-M210	4	YSM4050
h90 myo52-tdTomato:natMX leu1-32:p ^{cdc8} :mNeonGreen-cdc8:term ^{cdc8} :term ^{ScADH1} :leu1+ fus1 ¹⁻⁴⁹¹ -FUS ^{12E} -fus1 ⁷⁹²⁻¹³⁷² :kanMX ura4-294 ade6-M210	4	YSM4051
h90 fus1-sfGFP:kanMX myo51Δ::ura4+ myo52Δ::ura4+ leu1-32	S2	YSM2543
h90 fus1 ¹⁻⁴⁹¹ -FUS-fus1 ⁷⁹²⁻¹³⁷² -sfGFP:kanMX myo51Δ::ura4+ myo52Δ::ura4+ leu1-32	S2	YSM4052

Table S2: Primers used in this study and their usageHR stands for homologous recombination in yeast ⁴⁵ and SDM for site directed mutagenesis

NAME	SEQUENCE	ORIENTATION	PURPOSE
osm765	CAGCTCCAAATTTTGAAGTAAAACCCTAATTAGGGAATAAATAAGTAGGCAGAGCAC CTTGAAAAATACTAGATAGAATTCGAGCTCGTTTAAAC	R	HR (Myo52 3')
osm932	AATAAAAAGAGACAAACAGTCGTCCTTAAAGCTGAATGCATGCTTAAGCAGCTGGAGA ATAACAATGAACCTTAAGAGACGGATCCCCGGGTTAATTAA	F	HR (Fus1 ORF)
osm933	TTTTATTAATTATAATTTTATTATAATTTGTTTAAAGTCATTTAATTGTCATTAAGATCATT AACATTTCAAACATCAGAATTCGAGCTCGTTTAAAC	R	HR (Fus1 3')
osm1196	GATCACTGTAGGCAACGTAGCCGACAATGATGTACAGAAGCTCGAGCGACGAAGAAAAT CAAGTACCAAATGGTATTAAGTTCGGATCCCCGGGTTAATTAA	F	HR (Myo52 ORF)
osm1746	ACGGATTTTCATGAAGTTATTGGTTAAAAGCGGCCCTCTCAAATCTCCAGCTAAAGAACCA GTCCATGACAACGAAAATCGGATCCCCGGGTTAATTAA	F	HR (Tea1 ORF)
osm1747	ATGTCATCGTCGAATATTACACTATGTACAGTCTTTTCAACTAGTAAAGGAGATGCTTT CAAAATAGTTCCAAGAGGAATTCGAGCTCGTTTAAAC	R	HR (Tea1 3')
osm1772	CGTATCACGAGGCCCTTTCG	F	CLONING
osm2217	CCGGATCTCCAAGGGTGAAGAGCTATTTACTGGGG	F	CLONING
osm3005	ACTGCGGCCGCATGATGACGGCTAGTTTTAAAGG	F	CLONING
osm3006	ACTCCCGGGTCTCTTAAGTTCATTGTTATTCTCC	R	CLONING
osm3007	ACTGCGGCCGCATGGCATCTAAAATGCCTGAAG	F	CLONING
osm3009	ACTGCGGCCGCATGCGAAAATTCGTCAAAGGGAC	F	CLONING
osm3026	CTTGGATCCTCATATTTTCTATTTTAGAAAACCTC	R	CLONING
osm3027	TGAGGATCCAAGAAGTTATTGATGGGAATCC	F	CLONING
osm3028	CTGGGATCCATGGCGAAGGCGAGGAAG	R	CLONING
osm3030	TCGGGATCCTACTATTGTTGCTAACTGTTTCTGC	R	CLONING
osm3031	GTAGGATCCCGAAGCTTTGATATTCCTAATGATGC	F	CLONING
osm3091	CGGGGTACCGATCAGAAAATTATCGCCAT	F	CLONING
osm3516	ACTGCGGCCGCTGATTTAACAAAGCGACTATAAGTC	R	CLONING
osm3521	ACTCCCGGGAGTAGAAGTGTAGGAGCTTC	R	CLONING
osm4021	CTTGGATCCTATGAACCTCAAAGAATGCGTTG	R	CLONING
osm4504	CATTAAGGCCCTACTTTTATTCTGAGATCGCTATCCGGTGTATTCTTTTGTAAAGCAT TATATCATCAACTACCCGGATCCCCGGGTTAATTAA	F	HR (Acp2 5')
osm4505	CAATCTTTCTATGACTATTTTCGTTGAAGATGGAACGAATACTATGAGAAGATCACGGAA AGAAAACAAAAGCAATCGAATTCGAGCTCGTTTAAAC	R	HR (Acp2 3')
osm4577	GGAATAAGGGCGACACGG	F	ANALYTICAL
osm5452	GGCCACTAGTGGATCTGATATCGATGTATTTACTGATTACTT	R	INFUSION
osm5453	CTTCTAAACGGCTAGCTCAGCTTCATTGG	F	SDM
osm5454	CAATGAAGCTGAGCTAGCCGTTTAGAAGG	R	SDM
osm6064	CATATGGTCTGGGTATCT	R	CLONING
osm6183	GCCTTCCAACAGCTTCTCT	R	ANALYTICAL
osm6576	CTTGGATCCATCATTATTTGAATTACCAT	R	CLONING
osm6582	CTTGTTTAAACCAACATGCCTGTAAG	R	CLONING
osm6583	GAAGTTTAAACTGCTTTTGTGGTTATC	F	CLONING
osm7119	CTTCGTACGCTGCAGGTCGACACAGATGTACGCCAC	F	INFUSION
osm7122	TTCACCTTGGAGTTAATTAATCTCTTAAGTTCATTGTTAT	R	INFUSION
osm7127	ATGTACCAGGCGAAGCGCTTCTATGTCGGATGAC	F	INFUSION
osm7140	CTTCTTTGATTCTCATATCAGCTTGTAAGTAAGC	R	INFUSION
osm7141	TACTTTACAAGCTGATATGAGAATCAAAGAAGTTAT	F	INFUSION
osm7204	CTTTGTAAATCAGCGGCCGCATGTTTACCGATTTCATATGTA	F	INFUSION
osm7205	CTTGGAGTTAATTAACCCGGGGATCCTCATATTTTC	R	INFUSION
osm7254	GCTTTGTAAATCAGCGGCCGCATGATGAC	F	INFUSION
osm7255	CTTGGAGTTAATTAACCCGGGGATCCTATCATTATTTGAATTACCA	R	INFUSION
osm7256	CTTTGTAAATCAGCGGCCGCATGAAGCACACTCAAATTTCT	F	INFUSION
osm7257	CTTGGAGTTAATTAACCCGGGGATCCTAAAACCTTGTGTTTGA	R	INFUSION
osm7487	CTTCTTTGATTCTCATATCATTATTTGAATTACCAT	R	INFUSION
osm7488	TAATTCAAATAATGATATGAGAATCAAAGAAGTTAT	F	INFUSION
osm7489	AAACCTTGTGTTTGAATCAGCTTGTAAGTAAG	R	INFUSION
osm7490	TACTTTACAAGCTGATTCAAAACACAAGGTTTTTA	F	INFUSION
osm7499	GCTTTGTAAATCAGCGGCCGCATGCTCAAGTACGTGGAATCTTT	F	INFUSION

osm7638	CTTTGTAAATCAGCGGCCGCATGGTTACTCTCTCAAGAAAA	F	INFUSION
osm7677	GGAGTATTAACAACACTCGAGAAATGCGTGAAACTC	F	INFUSION
osm7690	AAATCAAGGATATGAGAATTCGAAAGAAAGTATGT	F	INFUSION
osm7738	TATAAAGCAATCAATATCAGCTTGTAAGTAAGCAC	R	INFUSION
osm7739	TACTTTACAAGCTGATATTGATTGCTTTTATAAGGAATTAAG	F	INFUSION
osm7740	GCTTATTAGAAGTGGCGCGCCTCTCTTAAGTTCATTGTTATTC	R	INFUSION
osm7875	CTTCTTTGATTCTCATATGAACCTCAAAGAATGCG	R	INFUSION
osm7876	TTCTTTGAGGTTTCATATGAGAATCAAAGAAGTTATTGAT	F	INFUSION
osm7877	CTTCTGATTTACAGTGCTAGCCTTTTTGTACTCCAGTATTAT	R	INFUSION
osm7878	TTTTGTCCATCTTCATCGTCATCATTAAACAAGCAATAG	R	INFUSION
osm7879	CTTGTTAATGATGACGATGAAGATGGACAAAAAGACTAT	F	INFUSION
osm7880	AACTAGCCGTCATCATTGCTGCTCCGATCATGATCT	R	INFUSION
osm7881	CATGATCGGAGCAGCAATGATGACGGCTAGTTTTAAAG	F	INFUSION
osm7882	GAGTTTCACGCATTTCTCGAGTTGTTTTAATACTCCTTC	R	INFUSION
osm8388	ACTGGTTCTGCTGTTTCATAGCCCTGAGGGGGATTA	R	INFUSION
osm8389	CCCTCAGGGCTATGAACAGCAGAACCAGTACAAC	F	INFUSION
osm8480	ACTTAAGAGAGGATCCCCGGGTTAATTAAC	F	INFUSION
osm8481	ATTCCTTTTACCCGGTTTACTTGACAGCTCGTCC	R	INFUSION
osm8482	CGAGCTGTACAAGTAAACCGGGTAAAAGGAATGTC	F	INFUSION
osm8483	AGGGAACAAAAGCTGGAGC	R	INFUSION
osm8484	GAAAAATATGAGGATCCCCGGGTTAATTAAC	F	INFUSION

Table S3: Plasmids used in this study and their construction. Related to STAR Methods.

For each plasmid, the column “obtained from” indicates how it was constructed, from restriction enzyme-based cloning or infusion, with the primers and restriction enzymes used. “WT” indicates that genomic DNA from a wildtype strain was used as template for PCR amplification.

NAME	DESCRIPTION	OBTAINED FROM	USAGE
pAV133	pUra4 ^{AfeI}	46	Single integration at <i>ura4</i>
pSM617	pREP3x	Lab Stock	Pombe expression
pSM677	pFA6a-mCherry-kanMX	Lab Stock	template for PCR-based HR
pSM684	pFA6a-mCherry-natMX	Lab Stock	template for PCR-based HR
pSM685	pFA6a-tdTomato-natMX	Lab Stock	template for PCR-based HR
pSM694	pFA6a-bleMX	Lab Stock	template for PCR-based HR
pSM1538	pFA6a-sfGFP-kanMX	Lab Stock	template for PCR-based HR
pSM1638	pRIP-p ^{fus1} -sfGFP	Lab Stock	Multiple integration at <i>ura4</i>
pSM1650	pRIP-p ^{fus1} -fus1N-sfGFP	Regular cloning : pSM1638 ^{NotI/BamHI} +(WT ^{osm3005-osm3026}) ^{NotI/BamHI}	Multiple integration at <i>ura4</i>
pSM1656	pRIP-p ^{fus1} -fus1-sfGFP	Regular cloning : pSM1638 ^{NotI/XmaI} +(WT ^{osm3005-osm3006}) ^{NotI/XmaI}	Multiple integration at <i>ura4</i>
pSM1659	pRIP-p ^{fus1} -fus1N-fus1C-sfGFP	3-point ligation cloning : pSM1638 ^{NotI/XmaI} +(WT ^{osm3005-osm3026}) ^{NotI/BamHI} +(WT ^{osm3027-osm3006}) ^{BamHI/XmaI}	Multiple integration at <i>ura4</i>
pSM1662	pRIP-p ^{fus1} -for3N-fus1C-sfGFP	3-point ligation cloning : pSM1638 ^{NotI/XmaI} +(WT ^{osm3007-osm3028}) ^{NotI/BamHI} +(WT ^{osm3027-osm3006}) ^{BamHI/XmaI}	Multiple integration at <i>ura4</i>
pSM1663	pRIP-p ^{fus1} -cdc12N-fus1C-sfGFP	3-point ligation cloning : pSM1638 ^{NotI/XmaI} +(WT ^{osm3009-osm3030}) ^{NotI/BamHI} +(WT ^{osm3027-osm3006}) ^{BamHI/XmaI}	Multiple integration at <i>ura4</i>
pSM1823	pRIP-p ^{nmt41} -sfGFP	Lab Stock	Multiple integration at <i>ura4</i>
pSM1826	pRIP-p ^{nmt41} -fus1N-sfGFP	Subcloning : pSM1650 ^{KpnI/NotI} +pSM1823 ^{KpnI/NotI}	Multiple integration at <i>ura4</i>
pSM2229	pUra4 ^{AfeI} -p ^{nmt41} -fus1-sfGFP	Lab Stock, Derived from pAV133	Single integration at <i>ura4</i>
pSM2251	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ501-749} -sfGFP-kanMX-fus1 ^{3'UTR}	SDM : pSM2827 ^{osm5453/5454}	Single integration at endogenous <i>fus1</i> locus
pSM2390	pRIP-p ^{fus1} -CRY2olig-For3N-fus1C-sfGFP	Lab Stock, Derived from pAV133	Multiple integration at <i>ura4</i>
pSM2475	pUra4 ^{AfeI} -p ^{fus1} -CRY2PHR-fus1C-sfGFP	Lab Stock, Derived from pSM1662	Single integration at <i>ura4</i>
pSM2478	pUra4 ^{PmeI} -p ^{nmt41} -fus1-sfGFP	3-point ligation cloning : pSM2229 ^{AatII/StuI} +(pSM2229 ^{osm4577-osm6582}) ^{AatII/PmeI} +(pSM2229 ^{osm6583-osm6183}) ^{PmeI/StuI}	Single integration at <i>ura4</i>
pSM2507	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ501-749} -sfGFP-kanMX-fus1 ^{3'UTR}	3-point ligation cloning : pSM2251 ^{Sall/PacI} +(pSM2251 ^{osm1772-osm6576}) ^{Sall/BamHI} +(pSM2229 ^{osm3031-osm3521}) ^{BamHI/PacI}	Single integration at endogenous <i>fus1</i> locus
pSM2600	pUra4 ^{PmeI} -p ^{nmt1} -fus1N-sfGFP	3-point ligation cloning : pSM2478 ^{KpnI/SacI} +(pSM617 ^{osm3091-osm3516}) ^{KpnI/NotI} +pSM1826 ^{NotI/SacI}	Single integration at <i>ura4</i>
pSM2601	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ¹⁻⁷³⁰ -sfGFP	3-point ligation cloning: pSM2600 ^{NotI/MscI} +(pSM2600 ^{osm3005-osm4021}) ^{NotI/BamHI} +(pSM2600 ^{osm2217-osm6064}) ^{BamHI/MscI}	Single integration at <i>ura4</i>
pSM2602	pUra4 ^{PmeI} -p ^{nmt1} -fus1-sfGFP	Subcloning : pSM2600 ^{NotI/XmaI} +pSM1656 ^{NotI/XmaI}	Single integration at <i>ura4</i>
pSM2625	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ492-791} -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2507 ^{Sall/PacI} +WT ^{osm7119-osm7140} +WT ^{osm7141-osm7122}	Single integration at endogenous <i>fus1</i> locus
pSM2630	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ⁹³⁻⁷⁹² -sfGFP	Infusion cloning : pSM2600 ^{NotI/XmaI} +pSM2600 ^{osm7204-osm7205}	Single integration at <i>ura4</i>
pSM2644	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ¹⁻⁵⁰⁰ -sfGFP	Infusion cloning : pSM2600 ^{NotI/XmaI} +pSM2600 ^{osm7254-osm7255}	Single integration at <i>ura4</i>
pSM2645	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ⁴³¹⁻⁷⁵⁵ -sfGFP	Infusion cloning : pSM2600 ^{NotI/XmaI} +pSM2600 ^{osm7256-osm7257}	Single integration at <i>ura4</i>

pSM2697	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ501-791} -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2507 ^{Sall/Pacl} +WT ^{osm7119-osm7487} +WT ^{osm7488-osm7122}	Single integration at endogenous <i>fus1</i> locus
pSM2698	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ492-749} -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2507 ^{Sall/Pacl} +WT ^{osm7119-osm7489} +WT ^{osm7490-osm7122}	Single integration at endogenous <i>fus1</i> locus
pSM2703	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ¹⁹¹⁻⁷⁹² -sfGFP	Infusion cloning : pSM2600 ^{NotI/XmaI} +pSM2600 ^{osm7499-osm7205}	Single integration at <i>ura4</i>
pSM2825	pUra4 ^{PmeI} -p ^{nmt1} -fus1N ¹⁴⁰⁻⁷⁹² -sfGFP	Infusion cloning : pSM2600 ^{NotI/XmaI} +pSM2600 ^{osm7638-osm7205}	Single integration at <i>ura4</i>
pSM2827	pFA6a-fus1 ^{5'UTR} -fus1-sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM1538 ^{Sall/EcoRV} +IBC180 ^{osm7119-osm5452}	Single integration at endogenous <i>fus1</i> locus
pSM2912	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ492-500} -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2507 ^{AfeI/Pacl} +pSM2507 ^{osm7127-osm7738} +pSM2507 ^{osm7739-osm7122}	Single integration at endogenous <i>fus1</i> locus
pSM2913	pFA6a-fus1 ^{5'UTR} -fus1-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2827 ^{EcoRI/AscI} +pSM2827 ^{osm7690-osm7740}	Single integration at endogenous <i>fus1</i> locus
pSM2937	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -CRY2PHR-fus1 ⁷⁹²⁻¹³⁷² -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2625 ^{Sall/XhoI} +pSM2625 ^{osm7119-osm7878} +pSM2475 ^{osm7879-osm7880} +pSM2625 ^{osm7881-osm7882}	Single integration at endogenous <i>fus1</i> locus
pSM2938	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -CRY2olig-fus1 ⁷⁹²⁻¹³⁷² -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2625 ^{Sall/XhoI} +pSM2625 ^{osm7119-osm7878} +pSM2390 ^{osm7879-osm7880} +pSM2625 ^{osm7881-osm7882}	Single integration at endogenous <i>fus1</i> locus
pSM2939	pFA6a-fus1 ^{5'UTR} -fus1 ^{Δ731-791} -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2827 ^{XhoI/NheI} +pSM2827 ^{osm7677-osm7875} +pSM2827 ^{osm7876-osm7877}	Single integration at endogenous <i>fus1</i> locus
pSM2940	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -FUS-fus1 ⁷⁹²⁻¹³⁷² -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2625 ^{XhoI/SwaI} +fus1 ^{XhoI} site-491-FUS-fus1 ⁷⁹² -SwaI site ordered as a gBlock	Single integration at endogenous <i>fus1</i> locus
pSM2941	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -FUS ^{12E} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2625 ^{XhoI/SwaI} +fus1 ^{XhoI} site-491-FUS ^{12E} -fus1 ⁷⁹² -SwaI site ordered as a gBlock	Single integration at endogenous <i>fus1</i> locus
pSM3032	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -FUS ^{G156E} -fus1 ⁷⁹²⁻¹³⁷² -sfGFP-kanMX-fus1 ^{3'UTR}	Infusion cloning : pSM2827 ^{XhoI/NheI} +pSM2940 ^{osm7677-osm8388} +pSM2940 ^{osm8389-osm7877}	Single integration at endogenous <i>fus1</i> locus
pSM3034	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -CRY2PHR-fus1 ⁷⁹²⁻¹³⁷² -kanMX-fus1 ^{3'UTR}	Subcloning : pSM2913 ^{Sall/NheI} +pSM2937 ^{NotI/XmaI}	Single integration at endogenous <i>fus1</i> locus
pSM3035	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -CRY2olig-fus1 ⁷⁹²⁻¹³⁷² -kanMX-fus1 ^{3'UTR}	Subcloning : pSM2913 ^{Sall/NheI} +pSM2938 ^{NotI/XmaI}	Single integration at endogenous <i>fus1</i> locus
pSM3036	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -FUS-fus1 ⁷⁹²⁻¹³⁷² -kanMX-fus1 ^{3'UTR}	Subcloning : pSM2913 ^{Sall/NheI} +pSM2940 ^{NotI/XmaI}	Single integration at endogenous <i>fus1</i> locus
pSM3037	pFA6a-fus1 ^{5'UTR} -fus1 ¹⁻⁴⁹¹ -FUS ^{12E} -fus1 ⁷⁹²⁻¹³⁷² -kanMX-fus1 ^{3'UTR}	Subcloning : pSM2913 ^{Sall/NheI} +pSM2941 ^{NotI/XmaI}	Single integration at endogenous <i>fus1</i> locus
pSM3055	pUra4 ^{PmeI} -p ^{nmt1} -fus1-mCherry	Infusion cloning : pSM2602 ^{XmaI/SacI} +pSM684 ^{osm8480-osm8481} +pSM2602 ^{osm8482-osm8483}	Single integration at <i>ura4</i>
pSM3056	pUra4 ^{PmeI} -p ^{nmt1} -fus1-mCherry	Infusion cloning : pSM2600 ^{XmaI/SacI} +pSM684 ^{osm8484-osm8481} +pSM2602 ^{osm8482-osm8483}	Single integration at <i>ura4</i>