

Supporting Information for:

Synthesis and structure-activity studies of BAM complex inhibitor MRL-494

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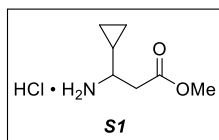
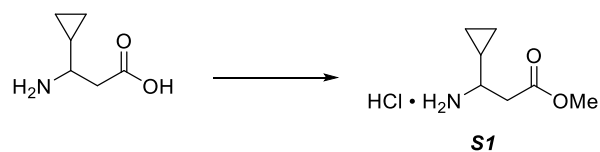
General information

HRMS analysis was performed on a Shimadzu Nexera X2 UHPLC system with a Waters Acquity HSS C18 column (2.1×100 mm, $1.8 \mu\text{m}$) at 30°C and equipped with a diode array detector. The following solvent system, at a flow rate of 0.5 mL/min, was used: solvent A, 0.1% formic acid in water; solvent B, 0.1% formic acid in acetonitrile. Gradient elution was as follows: $95:5$ (A/B) for 1 min, $95:5$ to $15:85$ (A/B) over 6 min, $15:85$ to $0:100$ (A/B) over 1 min, $0:100$ (A/B) for 3 min, then reversion back to $95:5$ (A/B) for 3 min. This system was connected to a Shimadzu 9030 QTOF mass spectrometer (ESI ionisation) calibrated internally with Agilent's API-TOF reference mass solution kit (5.0 mM purine, 100.0 mM ammonium trifluoroacetate and 2.5 mM hexakis(1H,1H,3H-tetrafluoropropoxy)phosphazine) diluted to achieve a mass count of 10000 .

HPLC analyses were performed on a Shimadzu Prominence-i LC-2030 system with a Dr. Maisch Reprosil Gold 120 C18 column (4.6×250 mm, 5 or $10 \mu\text{m}$) at 30°C and equipped with a UV detector monitoring at X and Y nm. The following solvent system, at a flow rate of 1 mL/min, was used: solvent A, 0.1% TFA in water/acetonitrile $95:5$; solvent B, 0.1% TFA in water/acetonitrile $5/95$. Gradient elution was as follows: $95:5$ (A/B) for 2 min, $95:5$ to $0:100$ (A/B) over 13 min, $0:100$ (A/B) for 2 min, then reversion back to $95:5$ (A/B) over 1 min, $95:5$ (A/B) for 2 min.

Preparative HPLC runs were performed on a BESTA-Technik system with a Dr. Maisch Reprosil Gold 120 C18 column (25×250 mm, $10 \mu\text{m}$) and equipped with a ECOM Flash UV detector monitoring at X nm. The following solvent system, at a flow rate of 12 mL/min, was used: solvent A, 0.1% TFA in water/acetonitrile $95:5$; solvent B, 0.1% TFA in water/acetonitrile $5/95$. Gradient elution was as follows: $95:5$ (A/B) for 2 min, $95:5$ to $0:100$ (A/B) over 13 min, $0:100$ (A/B) for 2 min, then reversion back to $95:5$ (A/B) over 1 min, $95:5$ (A/B) for 2 min.

Building block synthesis



(±)-Methyl 3-amino-3-cyclopropylpropanoate (**S1**). (±)-3-amino-3-cyclopropyl-propanoic acid (500 mg, 3.87 mmol, 1 eq) was dissolved in

methanol (15 mL) and cooled to 0 °C. Thionyl chloride (600 μL, 8.25 mmol, 2.1 eq) was added dropwise to the solution and stirred for 3 h before gradually warming to room temperature. The reaction was stirred for a further 18 h and monitored by TLC (99.5/0.5 DCM/NEt₃). When the reaction was complete, the solvent was removed and mixture coevaporated with toluene (3 x 10 mL) to give a white solid (quant). This was used in the next step without further purification.

¹H NMR (400 MHz, MeOD) δ 3.74 (s, 3H), 2.93 – 2.77 (m, 3H), 1.13 – 1.02 (m, 1H), 0.76 – 0.64 (m, 2H), 0.57 – 0.50 (m, 1H), 0.44 – 0.37 (m, 1H). ¹³C NMR (101 MHz, MeOD) δ 172.2, 55.1, 52.7, 37.9, 14.5, 4.8, 4.4. HRMS (ESI): calculated for C₇H₁₄NO₂ [M+H]⁺ 144.1019, found 144.1020.

Table S1. Gram-positive bacteria MIC results.

Strain	1	13	16	17
<i>MSSA</i> 29213	8	64	128	>128
<i>MRSA</i> USA 300	8	64	128	>128

Table S2. Results of **13** checkerboard assays in combination with rifampicin.

Strain	MIC ($\mu\text{g/mL}$)				FICI
	13 alone	13 in combination	Rifampicin alone	Rifampicin in combination	
<i>E. coli</i> ATCC 25922	>128	32	2	0.125	≤ 0.188
<i>E. coli</i> BW25113	64	32	4	0.125	0.281
<i>K. pneumoniae</i> ATCC 13883	>128	16	8	1	≤ 0.125
<i>A. baumannii</i> ATCC 9955	>128	16	1	0.125	≤ 0.188
<i>P. Aeruginosa</i> ATCC 27853	>128	-	16	-	-

Table S3. Results of **16** checkerboard assays in combination with rifampicin.

Strain	MIC ($\mu\text{g/mL}$)				FICI
	16 alone	16 in combination	Rifampicin alone	Rifampicin in combination	
<i>E. coli</i> ATCC 25922	128	16	2	0.125	0.188
<i>E. coli</i> BW25113	128	16	4	0.25	0.186
<i>K. pneumoniae</i> ATCC 13883	>128	16	8	1	≤ 0.125
<i>A. baumannii</i> ATCC 9955	>128	16	1	0.125	≤ 0.188
<i>P. Aeruginosa</i> ATCC 27853	128	-	16	-	-

Table S4. Results of **17** checkerboard assays in combination with rifampicin.

Strain	MIC ($\mu\text{g/mL}$)				FICI
	17 alone	17 in combination	Rifampicin alone	Rifampicin in combination	
<i>E. coli</i> ATCC 25922	>128	-	2	-	-
<i>E. coli</i> BW25113	>128	-	4	-	-
<i>K. pneumoniae</i> ATCC 13883	>128	-	8	-	-
<i>A. baumannii</i> ATCC 9955	>128	-	1	-	-
<i>P. Aeruginosa</i> ATCC 27853	>128	-	16	-	-

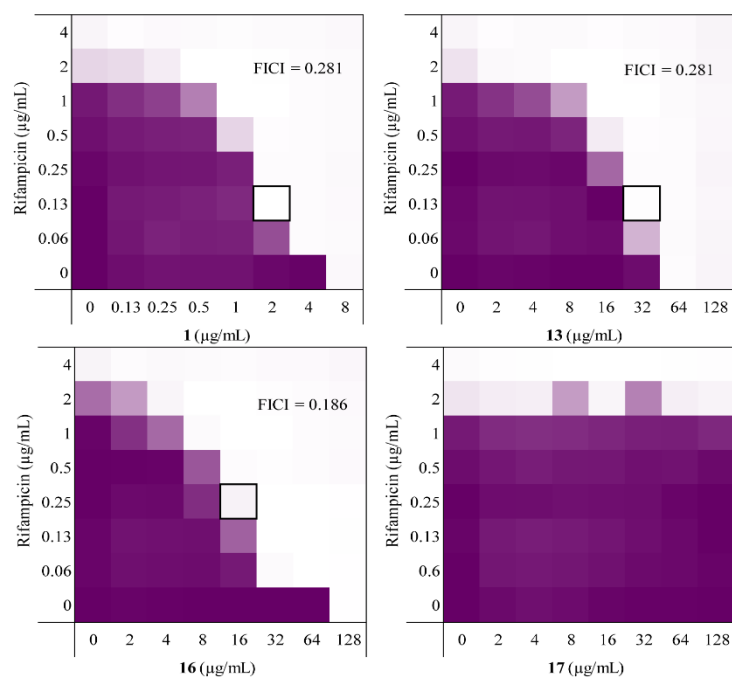


Figure S1. Checkerboard assay results for MRL-494 (**1**) and analogues (**13**, **16**, and **17**) in combination with rifampicin against *E. coli* BW25113. The combination of test compound and rifampicin which resulted in the lowest FICI is indicated by a black box. The mean optical density of the bacterial growth (OD600) is shown as a colour gradient, with purple signifying maximum bacterial growth and white as no growth.

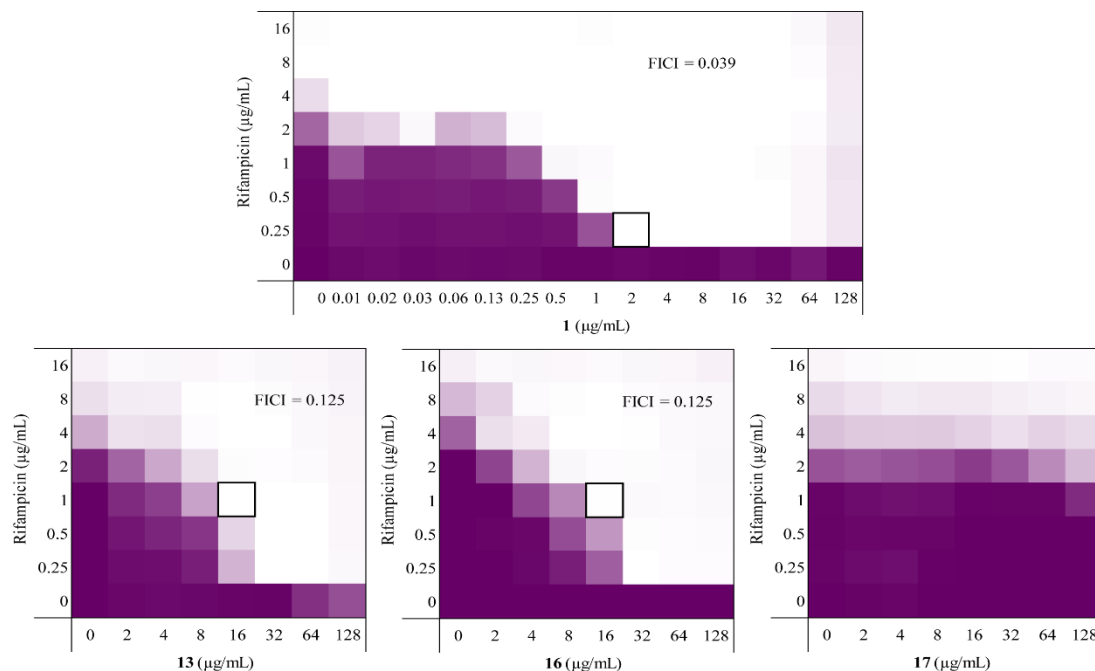


Figure S2. Checkerboard assay results for MRL-494 (**1**) and analogues (**13**, **16**, and **17**) in combination with rifampicin against *K. pneumoniae* ATCC 13883. The combination of test compound and rifampicin which resulted in the lowest FICI is indicated by a black box. The mean optical density of the bacterial growth (OD600) is shown as a colour gradient, with purple signifying maximum bacterial growth and white as no growth.

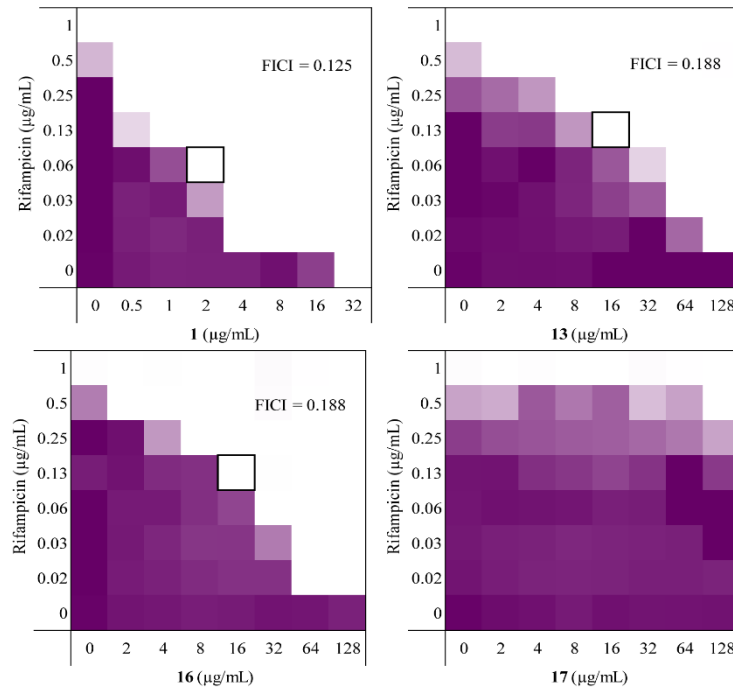


Figure S3. Checkerboard assay results for MRL-494 (**1**) and analogues (**13**, **16**, and **17**) in combination with rifampicin against *A. baumannii* ATCC 9955. The combination of test compound and rifampicin which resulted in the lowest FICI is indicated by a black box. The mean optical density of the bacterial growth (OD600) is shown as a colour gradient, with purple signifying maximum bacterial growth and white as no growth.

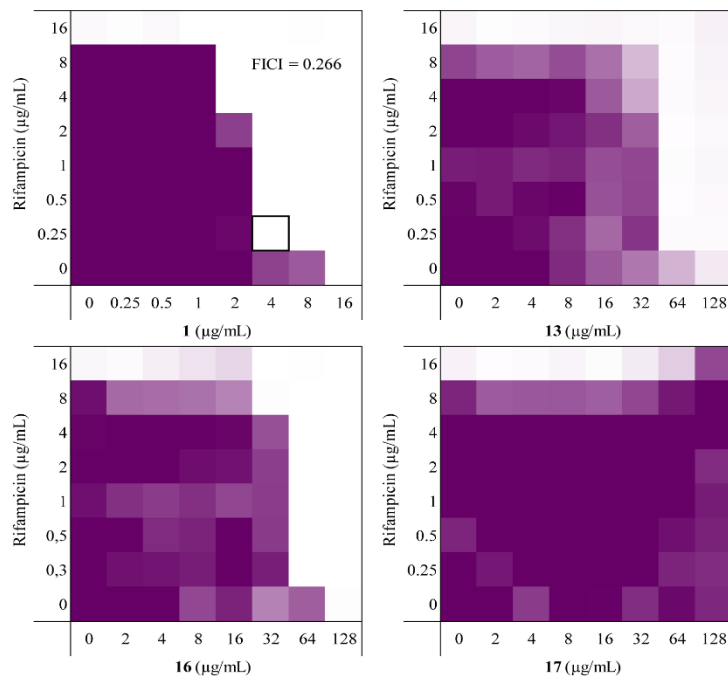


Figure S4. Checkerboard assay results for MRL-494 (**1**) and analogues (**13**, **16**, and **17**) in combination with rifampicin against *P. aeruginosa* ATCC 27853. The combination of test compound and rifampicin which resulted in the lowest FICI is indicated by a black box. The mean optical density of the bacterial growth (OD600) is shown as a colour gradient, with purple signifying maximum bacterial growth and white as no growth.

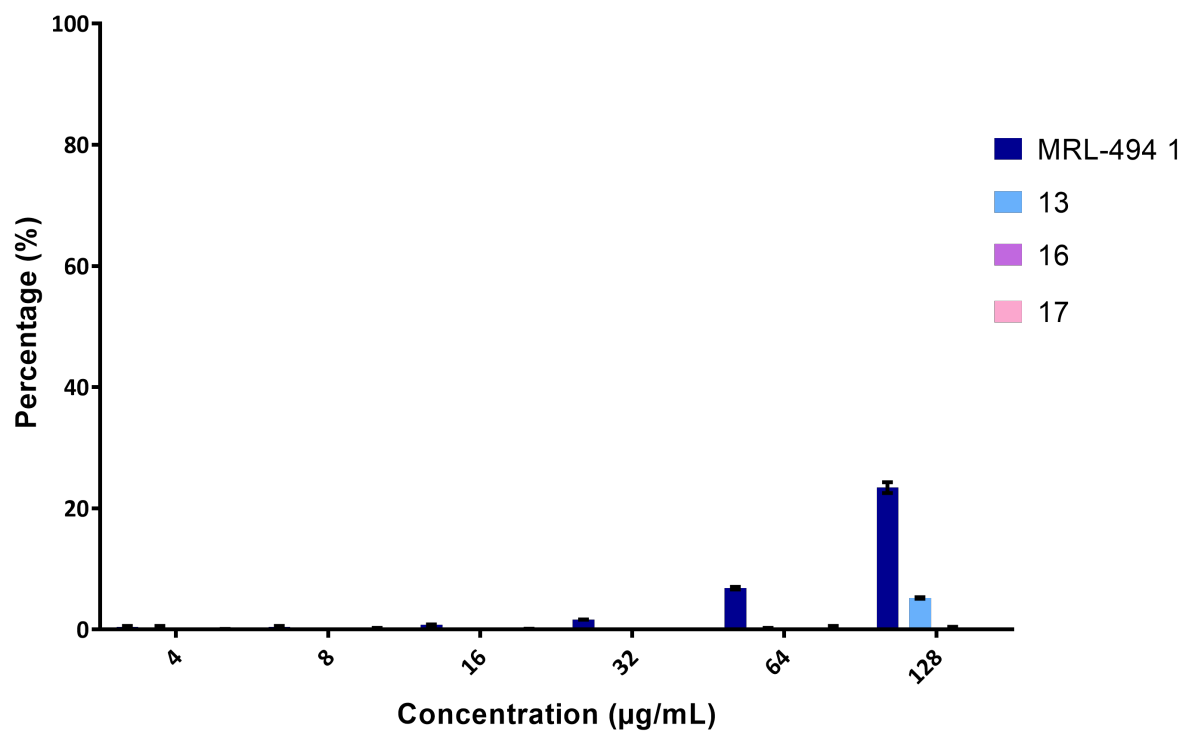


Figure S5. Hemolytic activity of all test compounds after 18 hours of incubation. A description of the hemolysis assay is found in the materials and methods. Error bars are calculated based on n=3 technical replicates.

Table S5. Hemolysis data points

Compound	Concentration (µg/mL)					
	128	64	32	16	8	4
MRL-494 1	23.4	6.8	1.6	0.8	0.4	0.4
13	5.2	0.2	-0.3	-0.4	-0.2	0.3
16	0.3	-0.4	-0.5	-0.4	-0.3	-0.3
17	0.0	0.2	-0.1	0.0	0.1	0.0

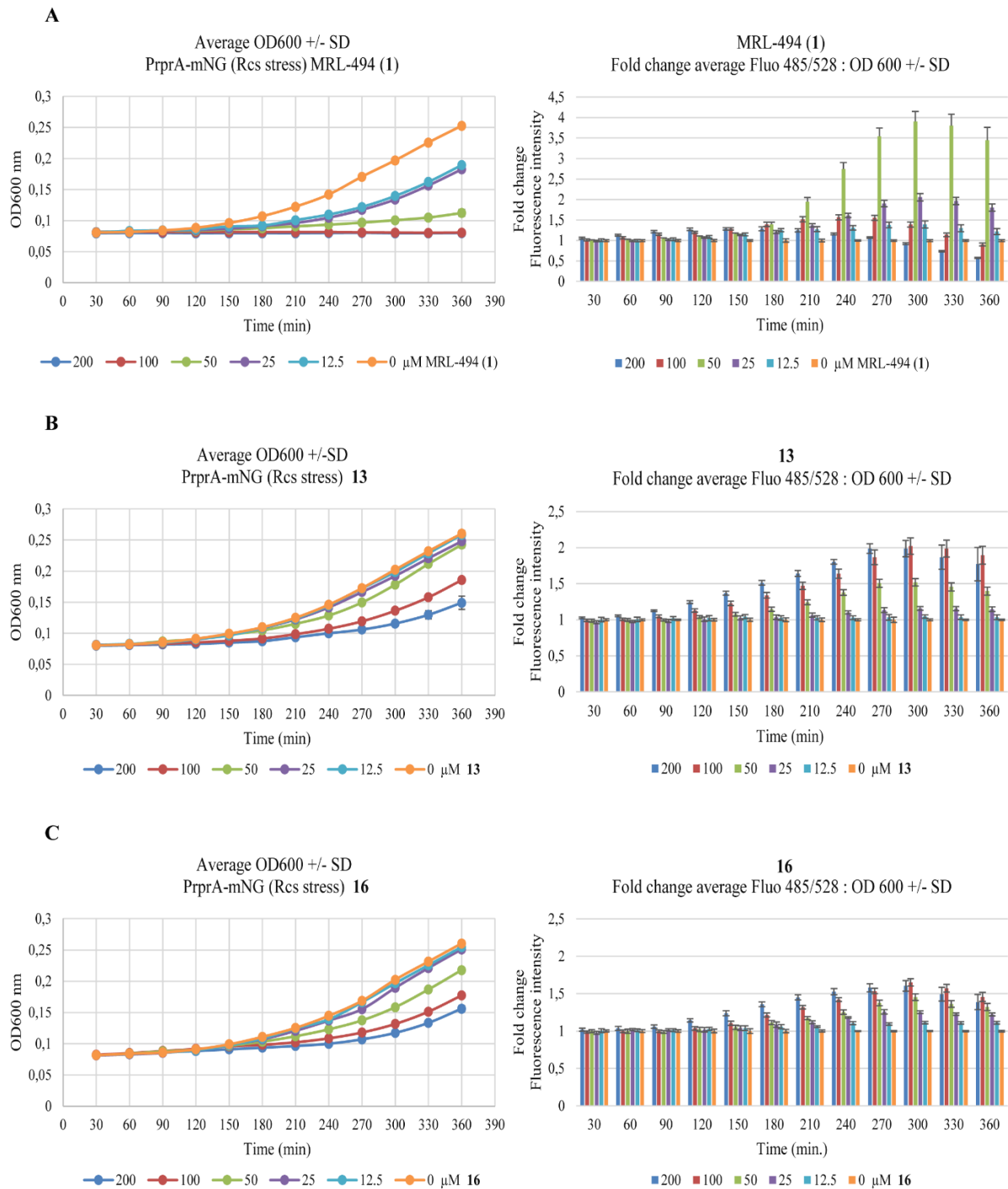


Figure S6. Real-time monitoring of bacterial growth and Rcs stress activation in response to MRL-494 1 and analogues (13 and 16). *E. coli* TOP10F⁷ cells, harboring the PrprA-mNG reporter construct, were grown in a 96-well plate and exposed to the compounds at the indicated concentration at timepoint 0. Growth (OD₆₀₀) and mNG fluorescence were measured in time. Fluorescence was corrected for growth (OD₆₀₀) and plotted as fold-change of signal compared to untreated cells (set to 1). Error bars represent the standard deviation of triplicate technical replicates.

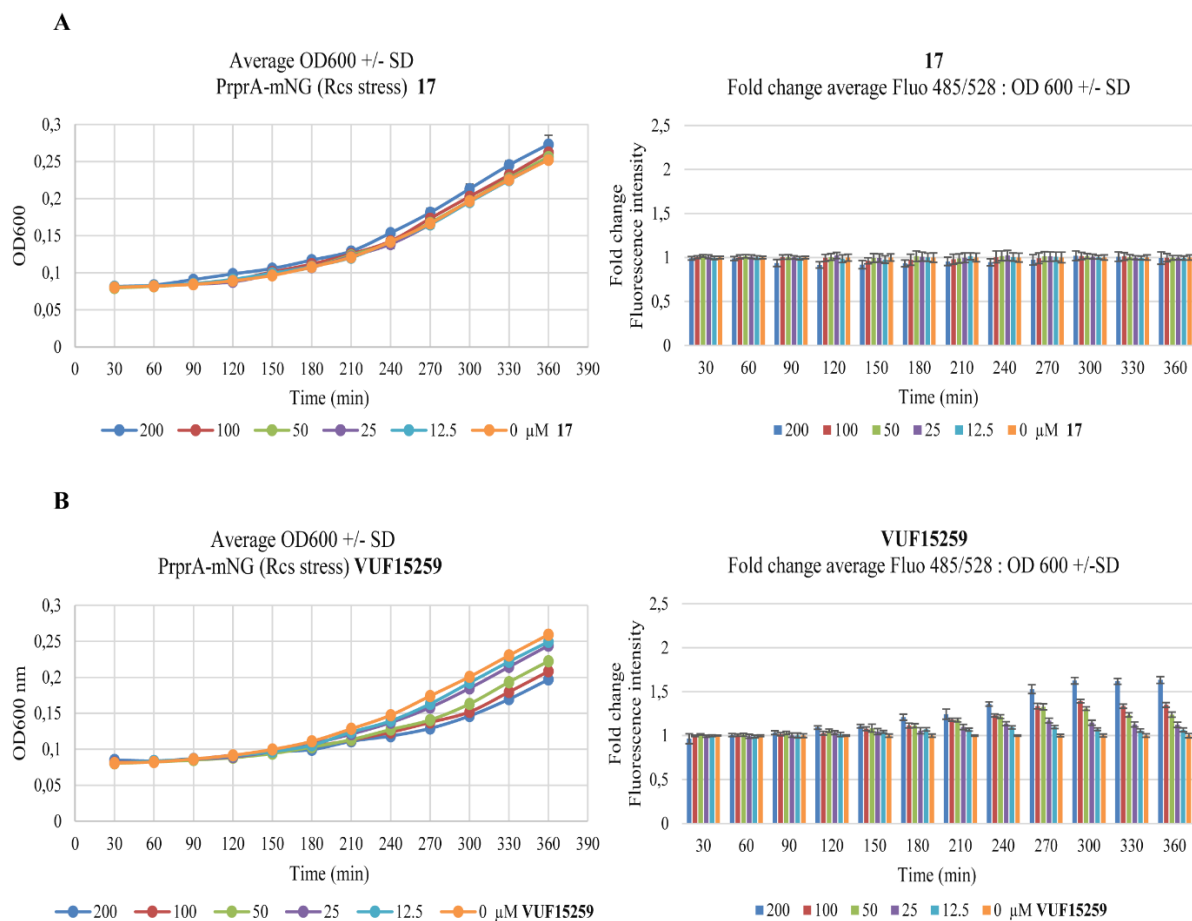
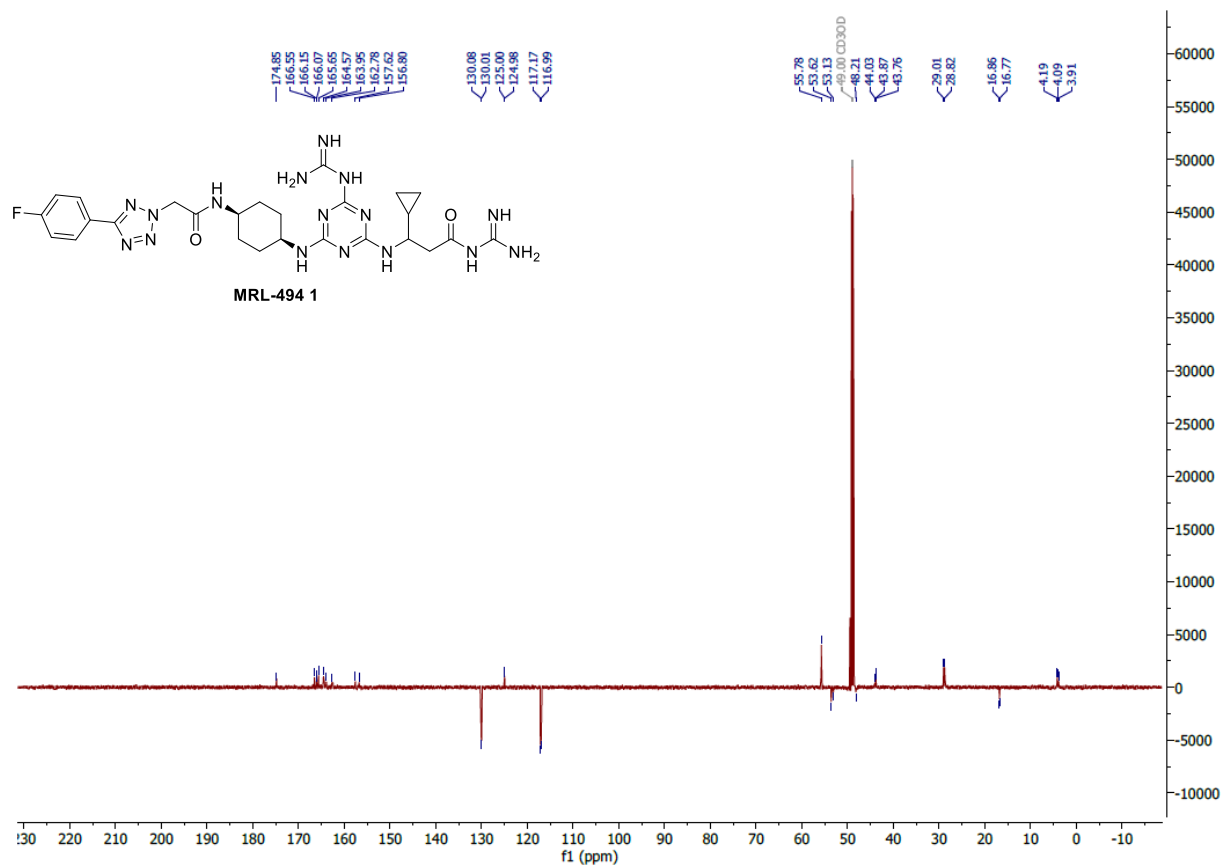
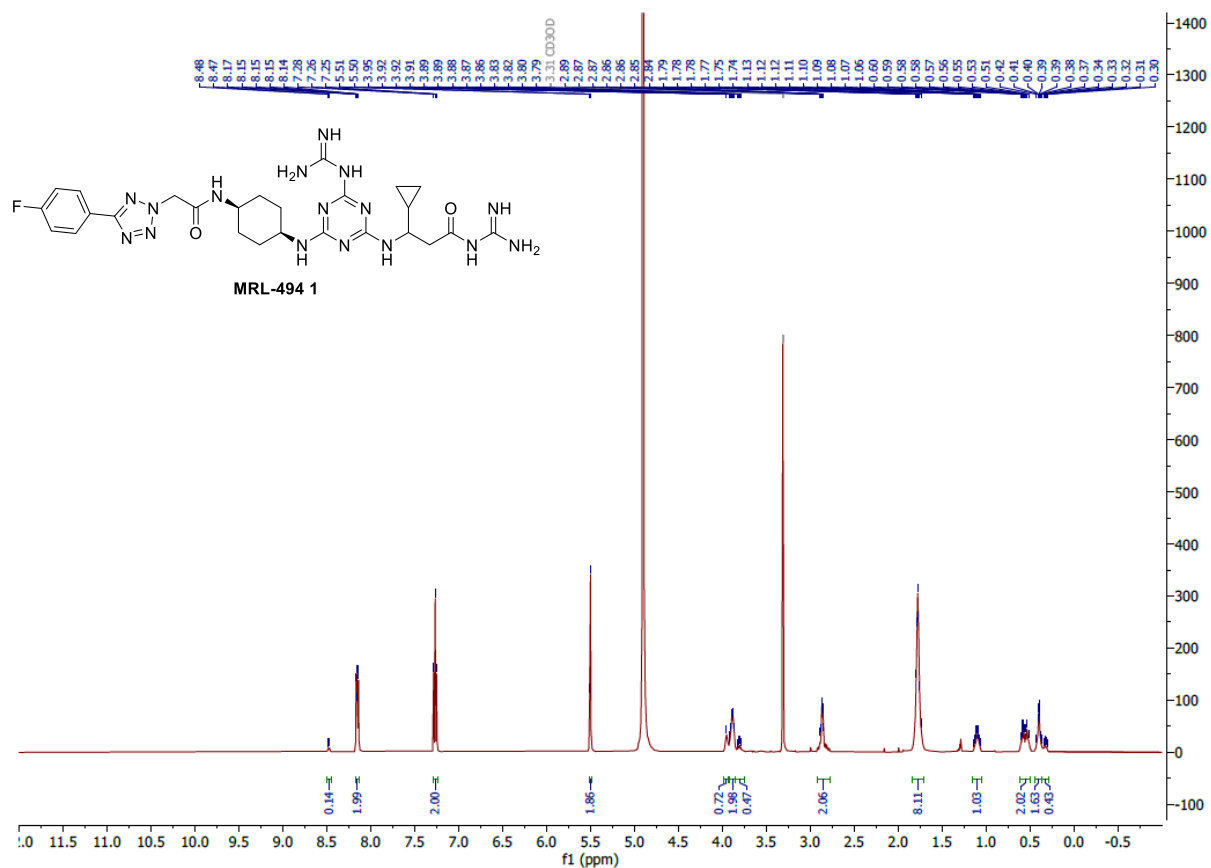
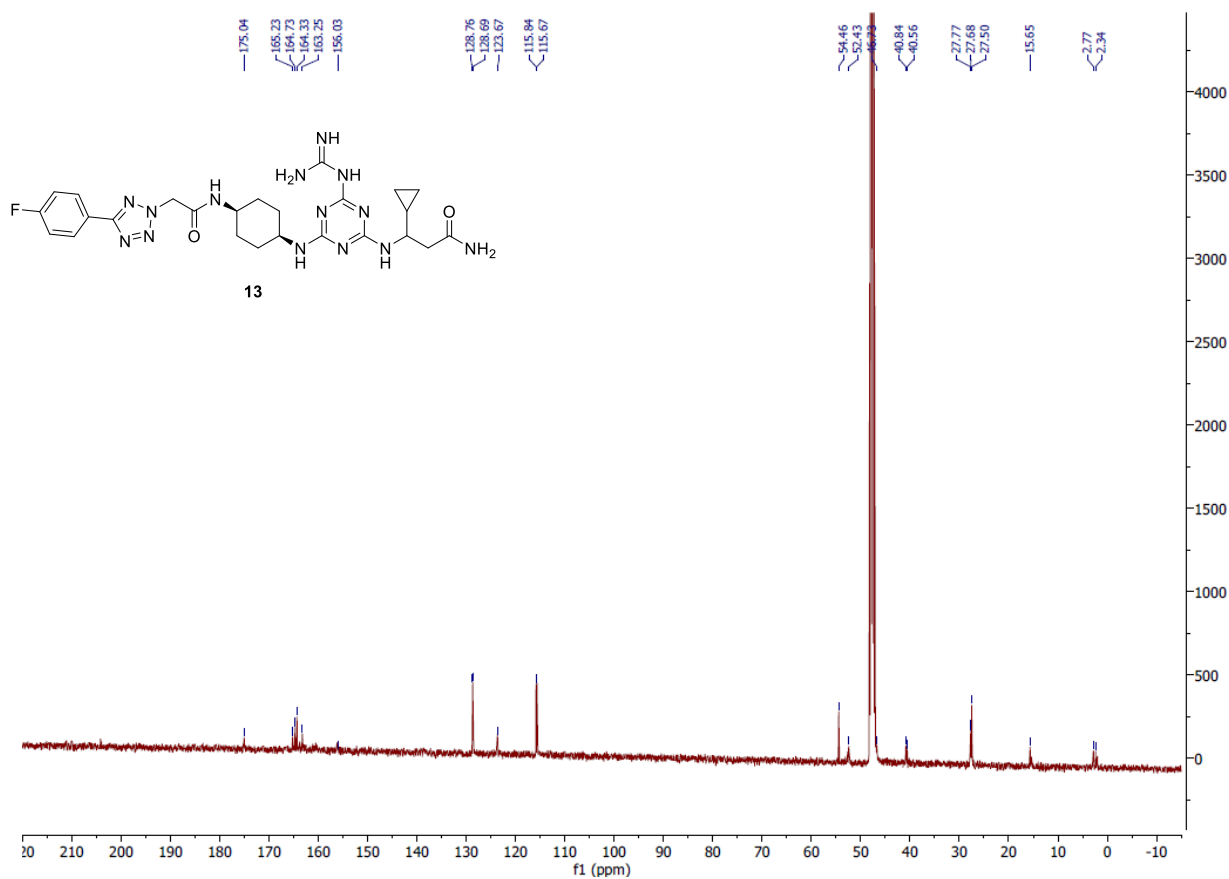
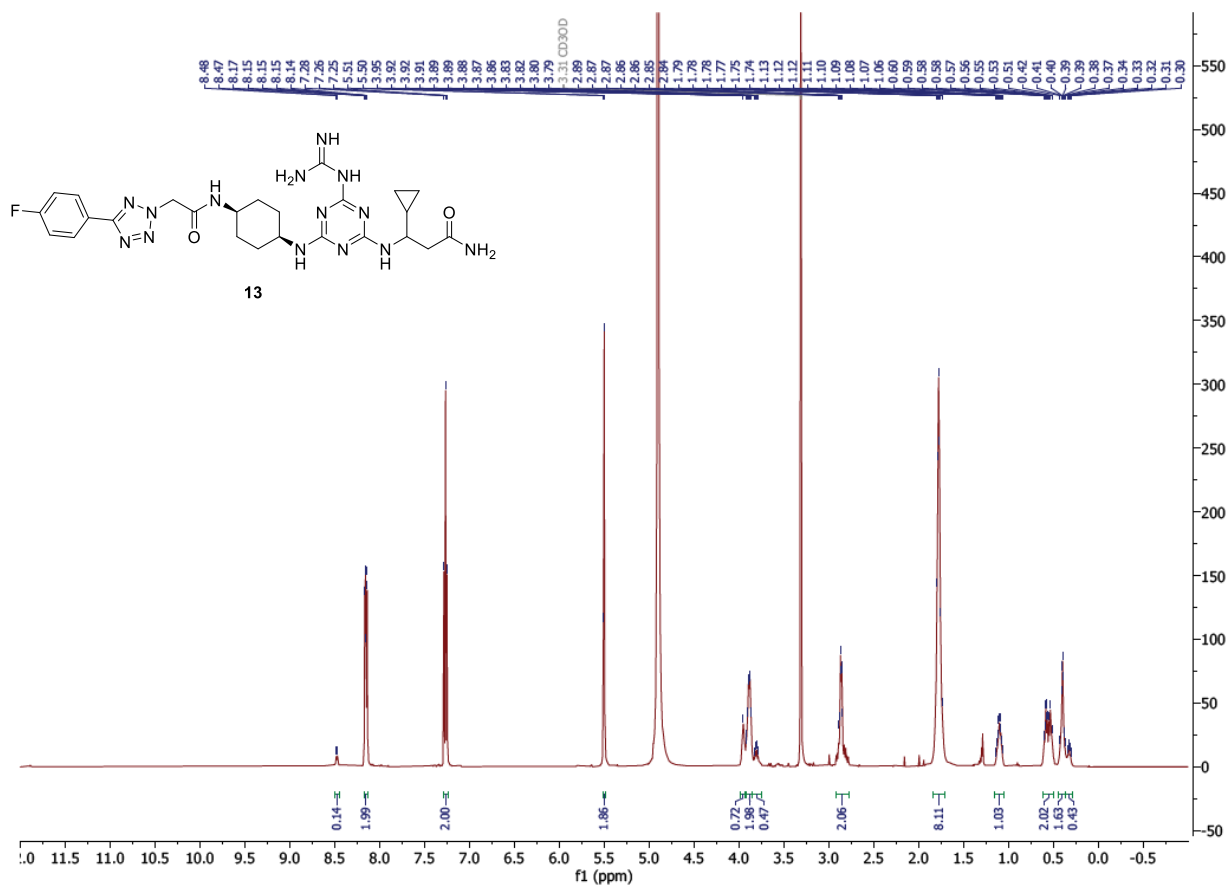


Figure S7. Real-time monitoring of bacterial growth and Rcs stress activation in response to MRL-494 analogue **17** and known BAM complex inhibitor VUF15259 **3**. *E. coli* TOP10F⁺ cells, harboring the PrprA-mNG reporter construct, were grown in a 96-well plate and exposed to the compounds at the indicated concentration at timepoint 0. Growth (OD₆₀₀) and mNG fluorescence were measured in time. Fluorescence was corrected for growth (OD₆₀₀) and plotted as fold-change of signal compared to untreated cells (set to 1). Error bars represent the standard deviation of triplicate technical replicates.

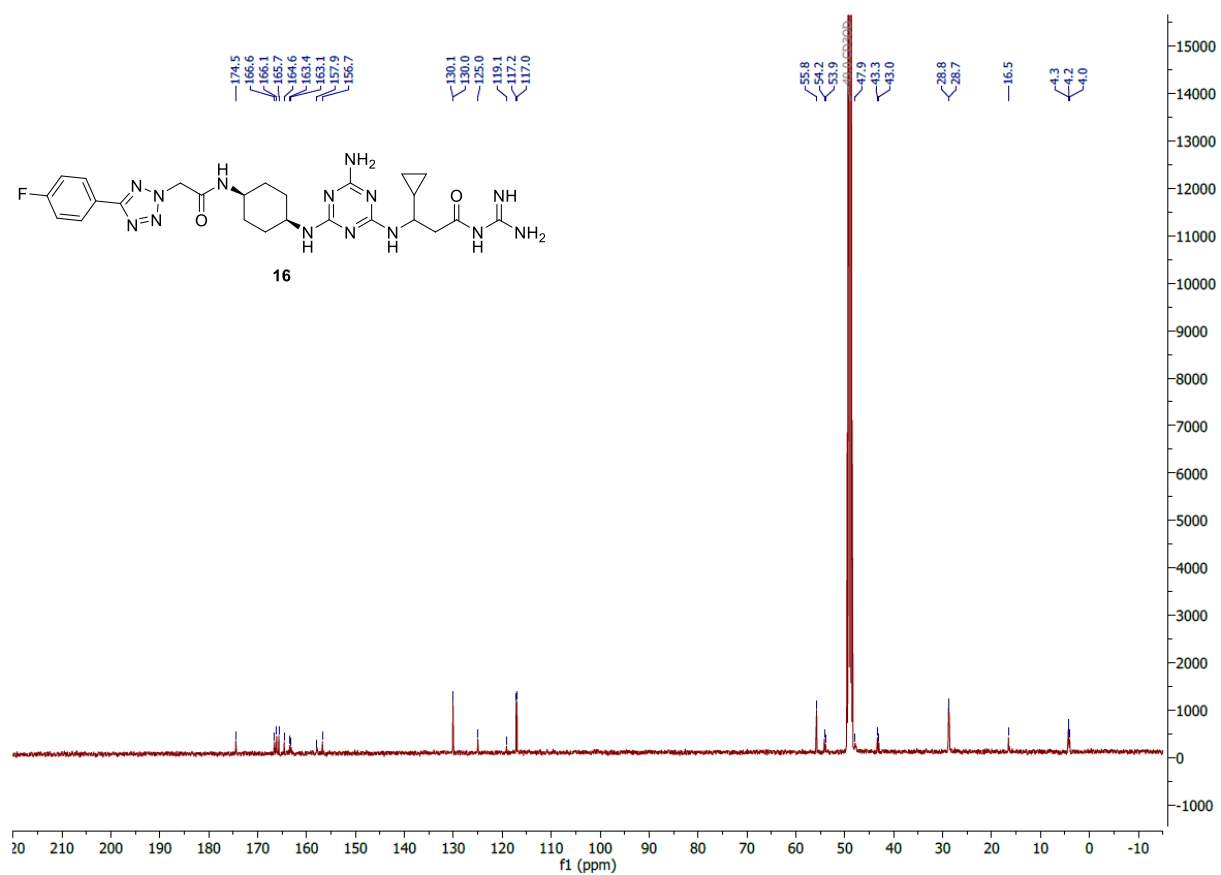
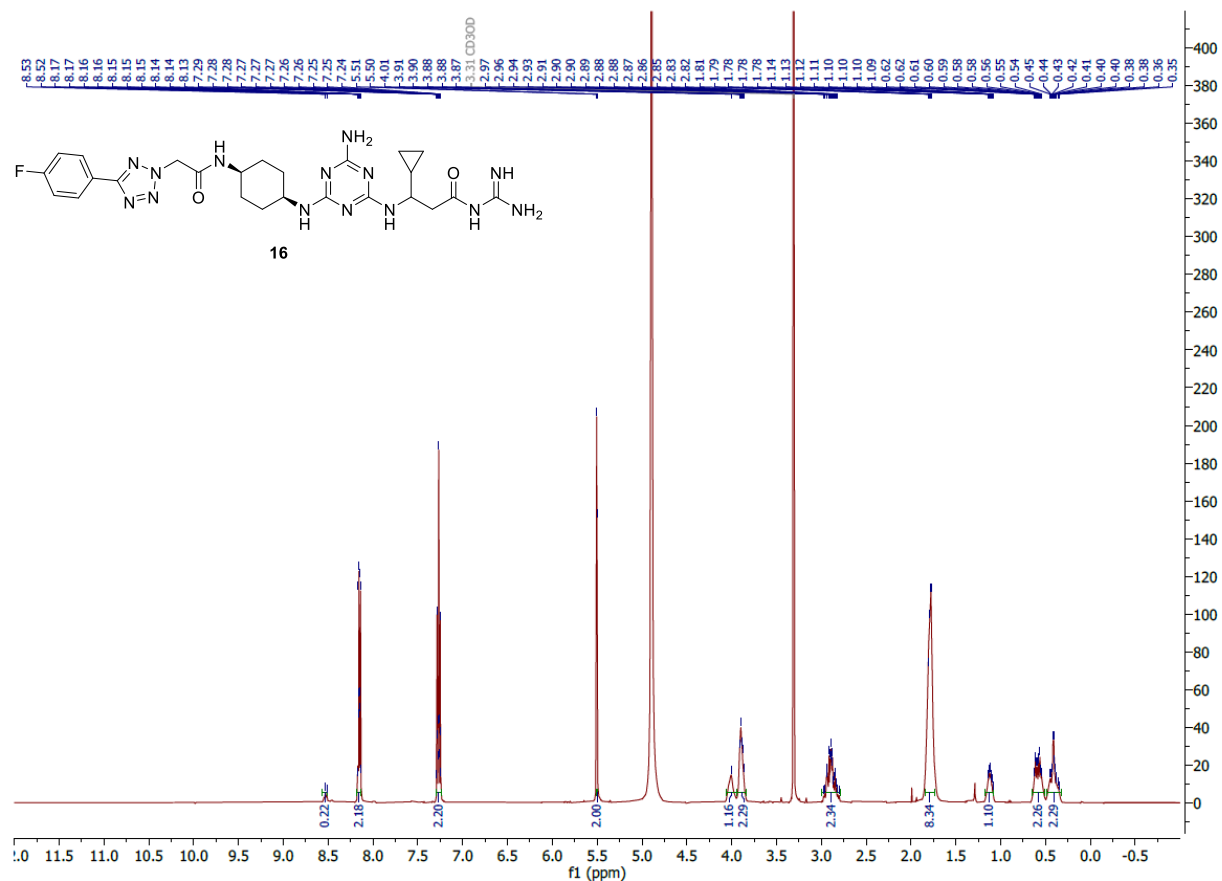
^1H and ^{13}C NMR spectra for MRL-494 (1)



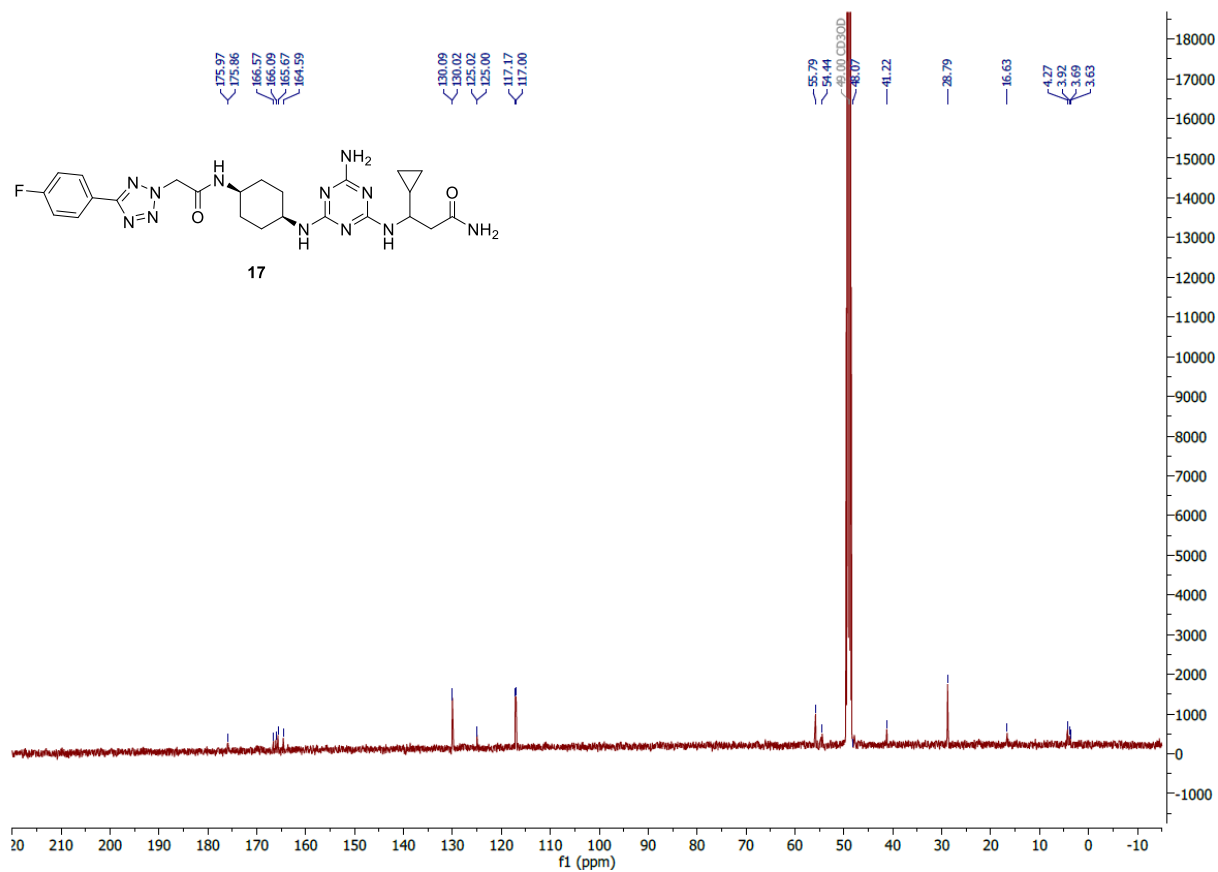
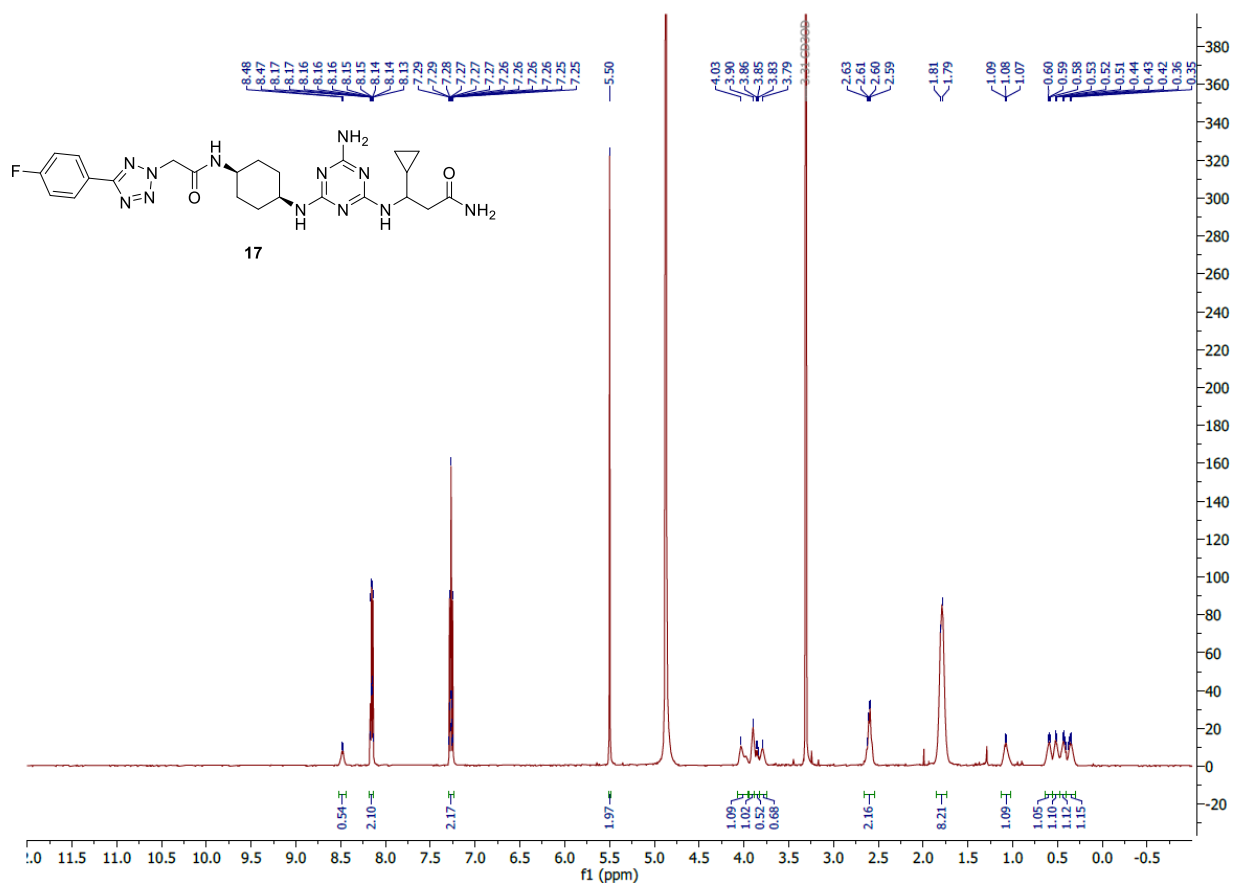
^1H and ^{13}C NMR spectra for compound **13**



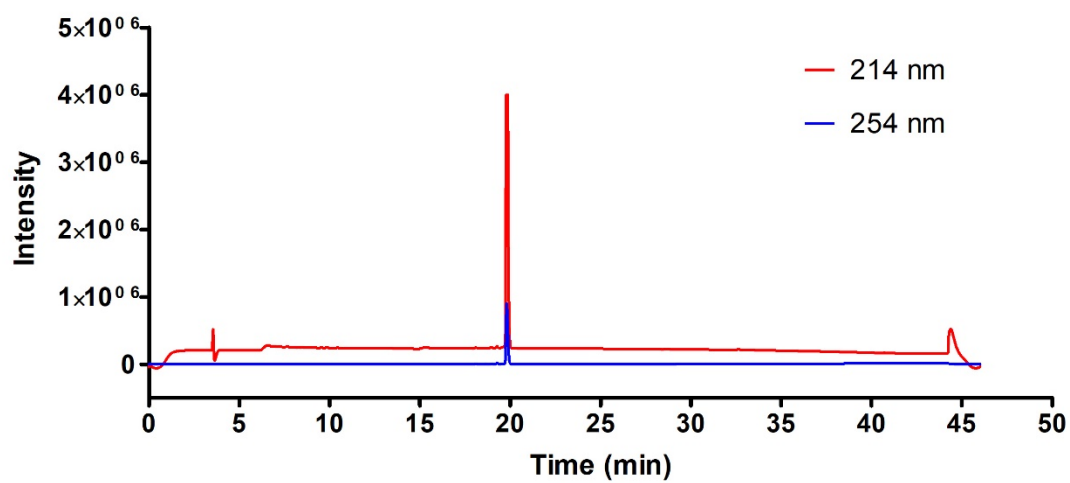
^1H and ^{13}C NMR spectra for compound **16**



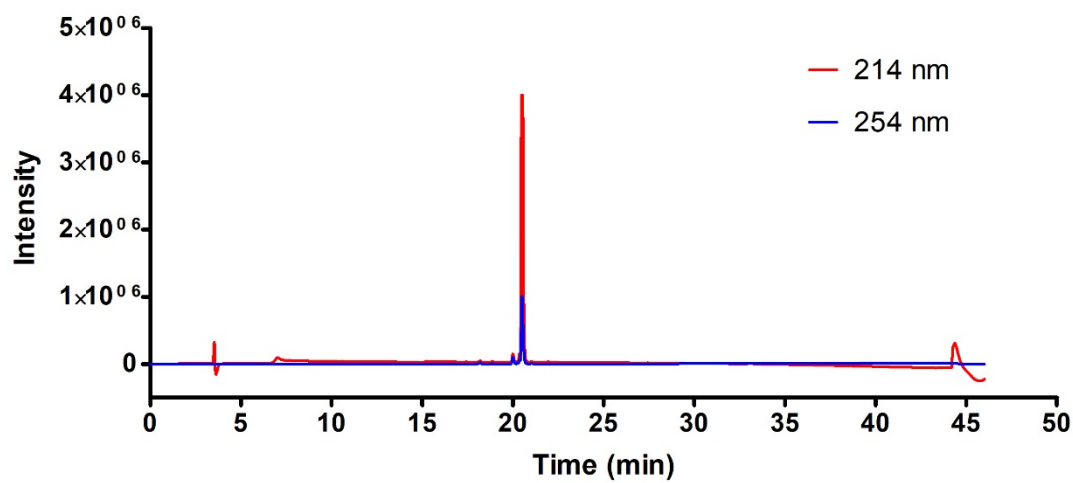
^1H and ^{13}C NMR spectra for compound **17**



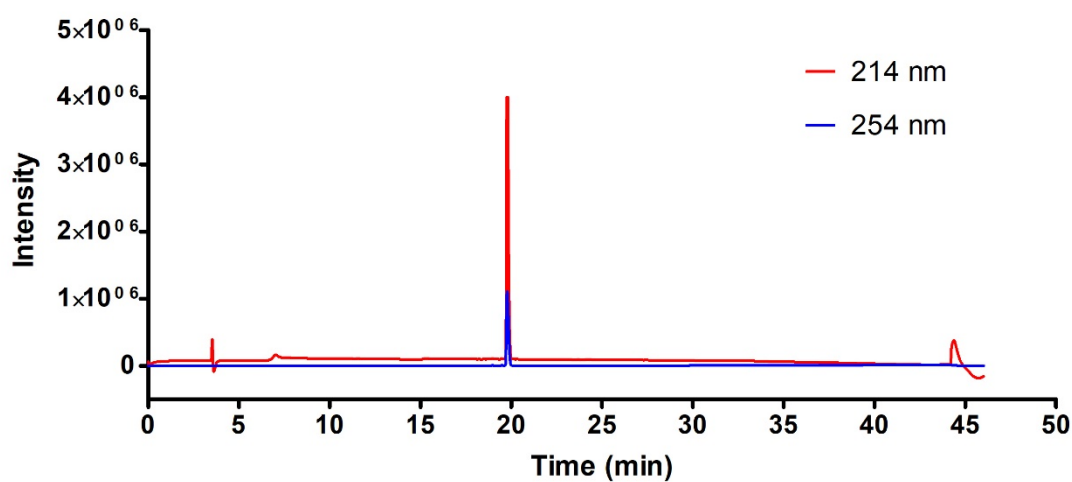
Analytical RP-HPLC data for compound MRL-494 (1)



Analytical RP-HPLC data for compound 13



Analytical RP-HPLC data for compound **16**



Analytical RP-HPLC data for compound **17**

