

## Supplemental material

**Figure S1. Experimental settings used.** **A.** Workflow for CTL generation from CD8<sup>+</sup> T cells purified from buffy coats of healthy donors. **B.** Quantitative RT-PCR of human GzmB mRNA in purified CD8<sup>+</sup> T cells at days 0, 5 and 7 after stimulation with anti-CD3/CD28 mAb-coated beads in the presence of IL-2 (n=5, One-way ANOVA test). **C.** Fluorimetric analysis of cytotoxicity of CTLs (day 7) generated as depicted in panel A, using the real-time calcein release-based killing assay. The killing process was monitored in different CTL:target cell ratios over time by measuring the release of calcein fluorescence every 10 min for 4 h. Representative curves showing the kinetics of target cell lysis by CTLs at the indicated CTL:target cell ratios are shown (n=3). **D.** *Top*, Image of a representative CD8<sup>+</sup> T cell (day 7) conjugated with SAg-pulsed Raji B cells for 15 min and co-stained with anti-PCNT and anti-GzmB antibodies. The CD8<sup>+</sup> T cell was magnified to depict the parameters used for quantification in figures 3, 4, 5, S2 and S3. A line from the center of each lytic granule (marked by GzmB) to the center of the centrosome (marked by PCNT) was drawn to measure the distance ( $\mu\text{m}$ ) between lytic granules and centrosome. A line from the center of the centrosome (marked by PCNT) of the CD8<sup>+</sup> T cell to the center of the contact area with the Raji cell was drawn to measure the distance ( $\mu\text{m}$ ) between the centrosome and the T cell:APC contact area. *Bottom*, Image of a representative CD8<sup>+</sup> T cell (day 7) conjugated with SAg-pulsed Raji B cells for 15 min and stained with anti-PTyr mAb. The T cell was magnified to depict the parameters used for quantification in figures 1, 2, 4, 5, S2 and S3. Masks around both the contact area of CD8<sup>+</sup> T cell:APC and in the remaining CD8<sup>+</sup> T cell area were drawn to measure the mean grey value. The recruitment index (RI) was calculated as specified. **E.** Immunofluorescence analysis of PTyr in CTLs (day 7) pre-treated with either vehicle (PBS) or different concentrations of Spike Wuhan (Spike W), then mixed with Raji cells (APCs) either

unpulsed or pulsed with a combination of SEA, SEB and SEE (SAGs), and incubated for 15 min at 37°C. *Left*, Quantification (%) of conjugates harboring PTyr staining at the IS (100 cells/sample, n=3, One-way ANOVA test). *Right*, Relative PTyr fluorescence intensity at the IS (10 cells/sample, n=3, Kruskal-Wallis test). **F.** Flow cytometric analysis of conjugates prepared as in (**E**). Raji cells were loaded with 5  $\mu$ M CFSE prior to conjugate formation. Conjugates were stained anti-PTyr mAb followed by fluorescently labelled secondary Abs. The analysis was carried out gating on CSFE<sup>-</sup> cells (n=3, One-way ANOVA test). **G.** Fluorimetric analysis of cytotoxicity of CTLs (day 7) using the calcein release assay. CTLs were pre-treated with either vehicle (PBS) or different concentrations of Spike Wuhan (Spike W) and co-cultured with SAg-pulsed, calcein AM-loaded Raji cells at a E:T cell ratio 10:1 for 4 h. The representative curves show the kinetics of target cell lysis by CTLs at the different concentrations of Spike Wuhan (Spike W) (n=3). **H.** ELISA-based quantification of IFN $\gamma$  in 36-h supernatants of melanoma-specific CTLs, pre-treated with either vehicle (PBS) or different concentrations of Spike Wuhan (Spike W) and co-cultured with irradiated autologous APCs pulsed with 2  $\mu$ g/ml MAGE 3 (n=3, One-way ANOVA test). Data are expressed as mean $\pm$ SD. \*\*\*\* $p\leq 0.0001$ ; \*\* $p\leq 0.01$ ; \* $p\leq 0.05$ . Not significant differences are not shown. Size bar, 5  $\mu$ m.

**Figure S2. Spike suppresses IS assembly and function in CTLs but not in resting CD8<sup>+</sup> T cells.** **A,B.** ELISA-based quantification of IFN $\gamma$  in 36-h supernatants of melanoma-specific CTLs derived from 3 patients, pre-treated with either vehicle (PBS) or Spike Wuhan (Spike W) (**A**), or with 1.2x10<sup>9</sup> liposomes or MiniVs (**B**), and co-cultured with irradiated autologous APCs pulsed with 2  $\mu$ g/ml MAGE 3 (n=3). **C.** ELISA-based quantification of IFN $\gamma$  in 36-h supernatants of melanoma-specific CTLs

derived from 3 patients, co-cultured with A375 melanoma cells transiently transfected with either a construct encoding Spike-W or the empty control vector. (n=3, unpaired two-tailed Student's *t*-test). **D-F. Top**, Immunofluorescence analysis of PTyr in freshly purified CD8<sup>+</sup> T cells (day 0) pre-treated with either vehicle (PBS) or 0.05μg/μl Spike Wuhan (Spike W), then mixed with Raji cells (APCs) either unpulsed or pulsed with a combination of SEA, SEB and SEE (SAGs), and incubated for 15 min at 37°C. The histograms show the quantification (%) of conjugates harboring PTyr (**D**), CD3ζ (**E**) or PCNT (**F**) staining at the IS (≥50 cells/sample, n=3, One-way ANOVA test). *Bottom*, Relative PTyr (**D**) and CD3ζ (**E**) fluorescence intensity at the IS (recruitment index) (10 cells/sample, n=3, Kruskal-Wallis test). Measurement of the distance (μm) of the centrosome (PCNT) from the T cell-APC contact site (**F**) (10 cells/sample, n=3, Kruskal-Wallis test). The data are expressed as mean±SD. \*\*\*\**p*≤0.0001; \*\*\* *p*≤0.001; \*\**p*≤0.01; \**p*≤0.05.

**Figure S3. ACE2 suppresses IS assembly and function in CTLs but not in resting CD8<sup>+</sup> T cells.** **A.** ELISA-based quantification of IFN<sub>γ</sub> in 36-h supernatants of melanoma-specific CTLs derived from 3 patients, pre-treated with either vehicle (PBS) or 2 μg/ml anti-ACE2 Ab (ACE2), and co-cultured with irradiated autologous APCs pulsed with 2 μg/ml MAGE 3 (n=3, unpaired two-tailed Student's *t*-test). **B-D. Top**, Quantification (%) of 5-min and 15-min conjugates harboring PTyr (**B**), CD3ζ (**C**) or P-ZAP-70 (**D**) staining at the IS (≥50 cells/sample, n=3, One-way ANOVA test). CTLs (day 7), pre-treated with vehicle (PBS) or 2 μg/ml anti-ACE2 Ab (ACE2), were conjugated with Raji cells (APCs) in the absence or presence of SAGs and either an anti-ACE2 Ab, or angiotensin II (AngII), or the peptide angiotensin 1-7 (Ang 1-7) (≥50 cells/sample, n=3, One-way ANOVA test). *Bottom*, Relative PTyr (**B**), CD3ζ (**C**) or ZAP-70 (**D**) fluorescence intensity at the IS (recruitment index) (10 cells/sample, n=3,

Kruskal-Wallis test). **E,F.** Immunofluorescence analysis of PTyr and CD3 $\zeta$  in freshly purified CD8<sup>+</sup> T cells (day 0) pre-treated with either vehicle (PBS) or 2  $\mu$ g/ml anti-ACE2 Ab (ACE2), then mixed with Raji cells (APCs) either unpulsed or pulsed with a combination of SEA, SEB and SEE (SAGs), and incubated for 15 min at 37°C. *Left*, Quantification (%) of conjugates harboring PTyr (**E**), CD3 $\zeta$  (**F**) ( $\geq 50$  cells/sample, n=3, One-way ANOVA test). *Right*, Relative PTyr (**E**) and CD3 $\zeta$  (**F**) fluorescence intensity at the IS (recruitment index) (10 cells/sample, n=3, Kruskal-Wallis test). **G.** Immunofluorescence analysis of PCNT in conjugates formed as described in (**E,F**). *Left*, Quantification (%) of conjugates harboring PCNT at the IS ( $\geq 50$  cells/sample, n=3, One-way ANOVA test). *Right*, Measurement of the distance ( $\mu$ m) of the centrosome (PCNT) from the T cell-APC contact site (10 cells/sample, n=3, Kruskal-Wallis test). **H.** Quantitative RT-PCR of human MAS1 mRNA in purified CD8<sup>+</sup> T cells at days 0, 5 and 7 after stimulation with anti-CD3/CD28 mAb-coated beads in the presence of IL-2 (n=3). \*\*\*\* $p \leq 0.0001$ ; \*\*\*  $p \leq 0.001$ ; \*\* $p \leq 0.01$ ; \* $p \leq 0.05$ . Non significant differences are not shown.

**Videos 1-3.** 3-D reconstruction of a representative CTL-APC conjugate formed with either SAg-unpulsed (video 1) or SAg-pulsed (videos 2 and 3) APCs and co-stained for PTyr (orange) and CD3 $\zeta$  (green). CTLs used in SAg-specific conjugates were pre-treated with either vehicle (video 2) or Spike-W (video 3).

**Videos 4-6.** 3-D reconstruction of representative CTL-APC conjugate formed with either SAg-unpulsed (video 4) or SAg-pulsed (videos 5 and 6) APCs and co-stained for PNCT (orange) and GzmB (green). CTLs used in SAg-specific conjugates were pre-treated with either vehicle (video 5) or Spike-W (video 6).

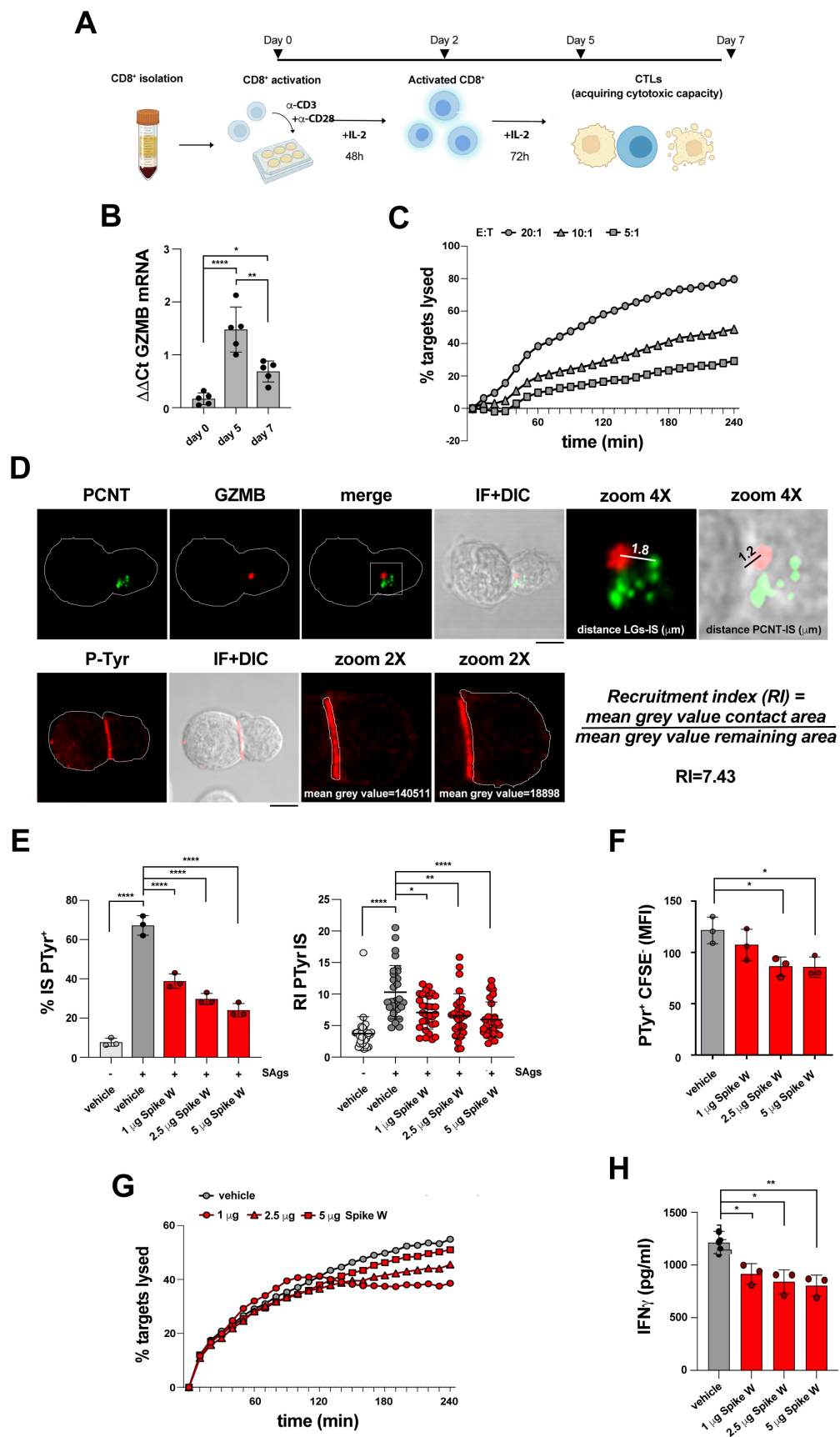
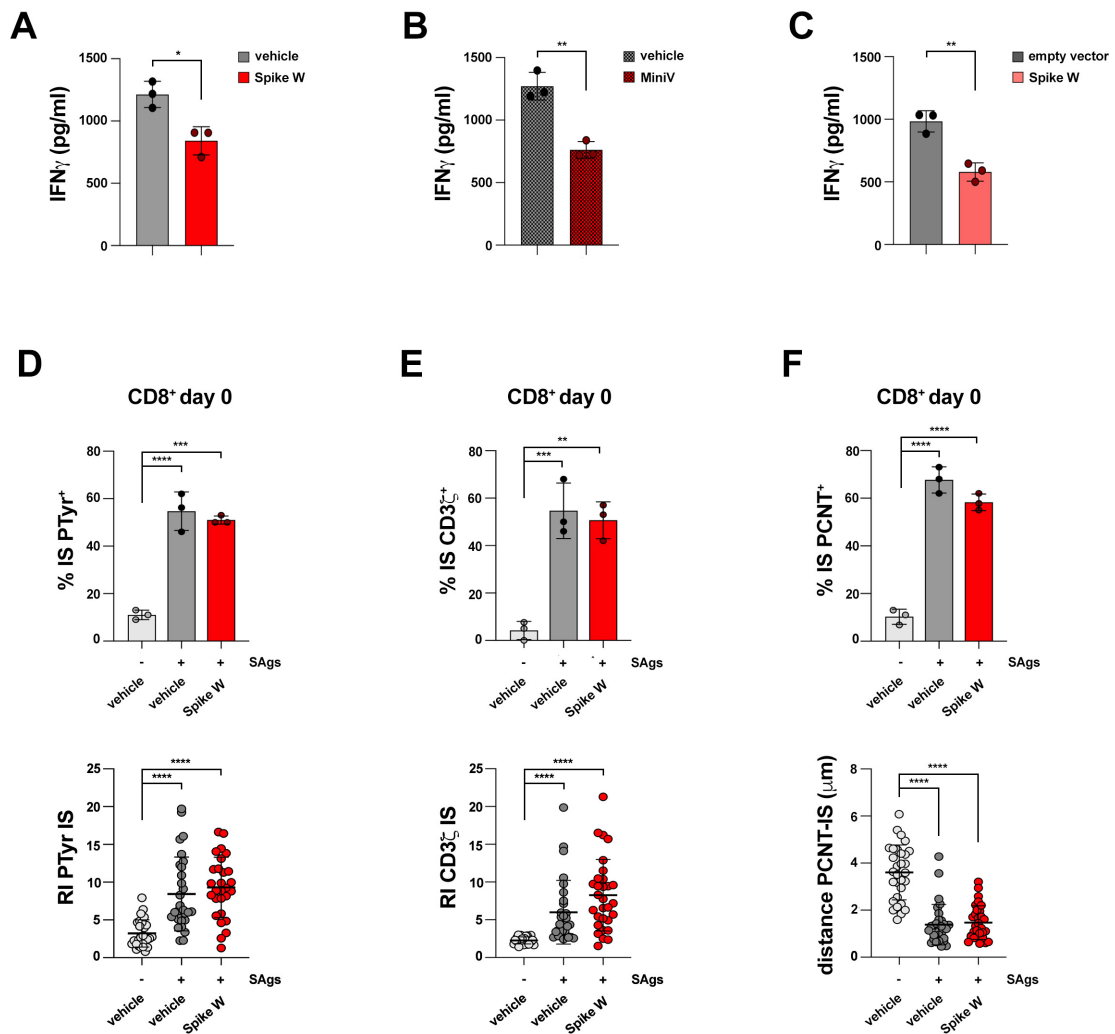
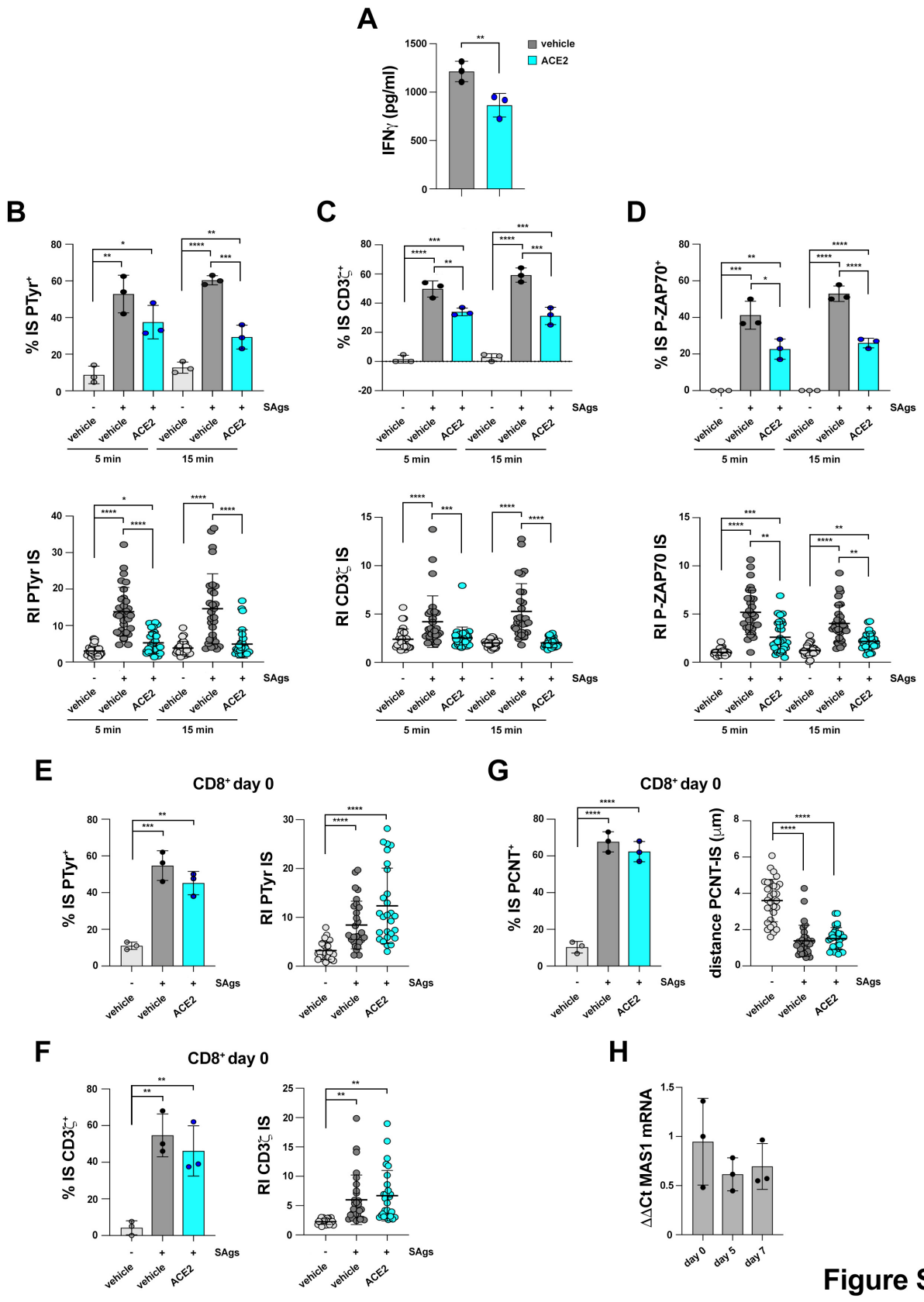


Figure S1



**Figure S2**



**Figure S3**