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Supplementary Materials for

Parental mutations influence wild-type offspring via transcriptional adaptation

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Figs. S1 to S9 Legends for tables S1 and S2

Other Supplementary Materials for this manuscript includes the following:

Tables S1 and S2



Fig. S1. Overexpression of *act-5(ptc)* transgene induces transcriptional adaptation and transgenerational inheritance of transcriptional adaptation in *C. elegans.* (A) Relative mRNA levels of *act-5* and *act-3* in wild-type and *Si[eft-3p::act-5(ptc)](tg/tg)* nematodes. n = 6 biologically independent samples. Data are mean \pm s.d., and a two-tailed Student's *t*-test was used to calculate *P* values. *Ct* values are listed in Table S1. (B) Representative picture of RFP expression in the pharynx and spermatheca of *Ex1000[act-3p::rfp]* adult control nematodes. (C) Representative picture of RFP expression in the pharynx, spermatheca, and uterus of *Ex1001[act-3p::rfp]* adult wild-type offspring from *Si[eft-3p::act-5(ptc)](tg/+);Ex1001[act-3p::rfp]* nematodes that have lost the ectopic uterus expression. Scale bars: 200 µm. Illustrations in panels B to D were created with BioRender.com.



Fig. S2. $aldh1a2^{ptc}$ and $aldh1a2^{floxed}$ alleles are new zebrafish models of transcriptional adaptation and transgenerational inheritance of transcriptional adaptation. (A) Relative mRNA levels of aldh1a2 and aldh1a3 in 24 hpf $aldh1a2^{ptc/ptc}$ embryos when compared with $aldh1a2^{+/+}$ siblings. (B) Relative mRNA levels of aldh1a3 in 24 hpf $aldh1a2^{ptc/ptc}$ embryos when compared with $aldh1a2^{ptc/ptc}$ embryos injected with wild-type aldh1a2 mRNA. (C) Schematic view of $aldh1a2^{floxed}$ allele based on Burg et al. (34), with the previously integrated loxP site shown as a green triangle and the additional loxP site shown as a cyan triangle in intron 8 of the aldh1a3 in 24 hpf $aldh1a2^{floxed/+}$ embryos injected with Cre mRNA

compared with $aldh1a2^{+/+}$ siblings injected with *Cre* mRNA. (E) Relative mRNA levels of aldh1a2 and aldh1a3 mRNA in 24 hpf wild-type embryos from TL incrosses (inx) and from $aldh1a2^{ptc/+}$ intercrosses (intx). (F) Relative mRNA levels of aldh1a2 and aldh1a3 in 24 hpf wild-type embryos obtained from TL incrosses and F1 $aldh1a2^{+/+}$ (i.e., wild types from $aldh1a2^{ptc/+}$ intercrosses) incrosses. $n \ge 2$ biologically independent samples. Control expression levels were set at 1. Data are mean \pm s.d., and a two-tailed Student's *t*-test was used to calculate *P* values. *Ct* values are listed in Table S1.



Fig. S3. Intergenerational inheritance of transcriptional adaptation is not observed in wild-type offspring obtained from RNA-less alleles. (A) Relative mRNA levels of *alcama* and *alcamb* in 28 hpf wild-type embryos from AB incrosses and *alcama^{RNA-less/+}* intercrosses. (B) Relative mRNA levels of *egfl7*, *emilin2a*, and *emilin3a* mRNA in 24 hpf wild-type embryos from AB incrosses and *egfl7^{RNA-less/+}* intercrosses. (C) Schematic view of *vcla^{RNA-less}* allele. (D) Representative image of gel providing evidence for the genomic deletion in the *vcla^{RNA-less}* allele. (E) Relative mRNA levels of *vcla* and *vclb* in 24 hpf *vcla^{RNA-less/RNA-less</sub> embryos when* compared with *vcla^{+/+}* and *vcla^{RNA-less/+}* siblings. (F) Relative mRNA levels of *vcla* and *vclb* mRNA in 24 hpf wild-type embryos from AB incrosses and *vcla^{RNA-less/+}* intercrosses. (G) Relative mRNA levels of *alcamb* in 24 hpf wild-type embryos from AB incrosses and F1 *egfl^{+/+}* (i.e., wild types from *egfl7^{ptc/+}* intercrosses) incrosses. *n* ≥ 2 biologically independent samples. Control expression levels were set at 1. Data are mean ± s.d., and a two-tailed Student's *t*-test was used to calculate *P* values. *Ct* values are listed in Table S1.}



Fig. S4. Gender regulation of intergenerational inheritance of transcriptional adaptation. (A) Relative mRNA levels of *alcama* and *alcamb* in 28 hpf wild-type embryos from AB incrosses (inx) and *alcama*^{*ptc/+*} male outcrosses (outx). (B) Relative mRNA levels of *alcama* and *alcamb* in 28 hpf wild-type embryos from AB incrosses and *alcama*^{*ptc/+*} female outcrosses. (C) Relative pre-mRNA levels of *alcama* and *alcamb* in 6 hpf wild-type embryos from AB incrosses and *alcama*^{*ptc/+*} female outcrosses. (D) Relative pre-mRNA levels of *alcama* and *alcamb* in 6 hpf wild-type embryos from AB incrosses and *alcama*^{*ptc/+*} female outcrosses. (E) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} male outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) Relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) relative mRNA levels of *egfl7* and *emilin2a* in 24 hpf wild-type embryos from AB incrosses and *egfl7*^{*ptc/+*} female outcrosses. (F) relative mRNA l



Fig. S5. Transgenerational inheritance of transcriptional adaptation models display increased transcription of the adapting gene. (A) Genetic crosses used to obtain F2 wild-type offspring from heterozygous zebrafish. Black arrows indicate heterozygous intercrosses (intx); red arrow indicates subsequent wild-type incrosses (inx). (B) qPCR analysis of 4sU-labelled *alcamb* mRNA levels in 28 hpf wild-type embryos from AB incrosses and from F1 *alcama*^{+/+} (i.e., wild types from *alcama*^{ptc/+} intercrosses) incrosses. (C) qPCR analysis of 4sU-labelled *emilin2a* mRNA levels in 24 hpf wild-type

embryos from AB incrosses and from F1 $egfl7^{+/+}$ (i.e., wild types from $egfl7^{ptc/+}$ intercrosses) incrosses. (D) qPCR analysis of 4sU-labelled *vclb* mRNA levels in 24 hpf wild-type embryos from AB incrosses and from F1 $vcla^{+/+}$ (i.e., wild types from $vcla^{ptc/+}$ intercrosses) incrosses. (E) qPCR analysis of 4sU-labelled *aldh1a3* mRNA levels in 24 hpf wild-type embryos from TL incrosses and from F1 $aldh1a2^{+/+}$ (i.e., wild types from $aldh1a2^{ptc/+}$ intercrosses) incrosses. n = 3 biologically independent samples. Control expression levels were set at 1. A two-tailed paired *t*-test was used to calculate *P* values. *Ct* values are listed in Table S1.



alcama pre-mRNA

alcamb pre-mRNA

Fig. S6. Mutated genes in transgenerational inheritance of transcriptional adaptation models are expressed in germ cells. (A) qPCR analysis of *alcama*, *egfl7*, *aldh1a2*, and *vcla* mRNA levels from oocytes of wild-type zebrafish.
(B) qPCR analysis of *alcama*, *egfl7*, *aldh1a2*, and *vcla* mRNA levels from sperm of wild-type zebrafish.
(C) qPCR analysis of *alcama*, *egfl7*, *aldh1a2*, and *vcla* mRNA levels from sperm of wild-type zebrafish.
(D) Transcripts per million for *rpl13a*,

alcama, alcamb, egfl7, emilin2a, aldh1a2, aldh1a3, vcla, and *vclb* in the indicated tissues, as obtained from the GSE111882 transcriptome dataset. **(E)** Gel electrophoresis of qPCR products of the housekeeping gene *rpl13a* from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from 6 hpf wild-type embryos as control. **(F)** Gel electrophoresis of qPCR products of *alcama pre-mRNA* and *alcamb pre-mRNA* from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from total RNA sample obtained from total RNA samples of testes, oocytes and sperm obtained from wild-type and *alcama^{ptc/+}* zebrafish with total RNA sample obtained from 6 hpf wild-type embryos as control.



Fig. S7. *aldh1a2*^{floxed} allele shows germline-specific recombination upon injection of *vasa-Cre-nos1 3'UTR* mRNA. (A to C) Lateral views of 24 hpf wild-type embryos injected with *vasa-GFP-nos1 3'UTR* mRNA. Scale bars: 1000 μ m. (A' to C') Higher magnification images of red boxed area in panels A, B and C, respectively. Scale bars: 100 μ m. (D) FACS contour plots of cells isolated from 24 hpf embryos from *aldh1a2*^{floxed/+} intercrosses (intx) injected with *vasa-GFP-nos1 3'UTR* mRNA; GFP+/DAPI- cells were isolated. (E) Negative control FACS contour plots of cells isolated from 24 hpf embryos from *aldh1a2*^{floxed/+} intercrosses. (F) *aldh1a2*^{floxed} allele displays germline-specific recombination upon injection of *vasa-Cre-nos1 3'UTR* mRNA. Green arrowheads point to wild-type allele; yellow arrowhead points to *aldh1a2*^{floxed} allele; red arrowhead points to recombined *aldh1a2*^{floxed} allele.



Fig. S8. Intergenerational inheritance of transcriptional adaptation is detected in somatic cells. (A, B) Lateral views of 24 hpf wild-type embryos from AB incrosses (inx) and from $alcama^{ptc/+}$ intercrosses (intx) injected with 100 pg *vasa-GFP-nos1 3'UTR* mRNA plus 0.6 ng control MO. Scale bars: 100 µm. (**C**, **D**) Lateral views of 24 hpf wild-type embryos from AB incrosses injected with 100 pg *vasa-GFP-nos1 3'UTR* mRNA plus 0.6 ng *dnd1* MO. Scale bars: 100 µm. (**E**) Relative mRNA levels of *alcamb* in 28 hpf AB embryos and F1 *alcama^{+/+}* (i.e., wild types from *alcama^{ptc/+}* intercrosses) embryos injected with 100 pg *vasa-GFP-nos1 3'UTR* mRNA plus 0.6 ng *dnd1* MO. n = 3 biologically independent samples. Control expression levels were set at 1. Data are mean \pm s.d., and a two-tailed Student's *t*-test was used to calculate *P* values. *Ct* values are listed in Table S1.



Fig. S9. Transgenerational inheritance of transcriptional adaptation in zebrafish is associated with H3K4me3 histone marks. (A) Peaks of H3K4me3 ChIP-seq at the *alcamb* locus at different stages (obtained from DANIO-CODE). Red boxed area was enlarged in (C) to show the relative location of the ChIP-qPCR primers marked by black arrows. (B) Genetic crosses used to obtain F2 wild-type offspring from heterozygous zebrafish. Black arrows indicate heterozygous intercrosses (intx); red arrow indicates subsequent wild-type incrosses (inx). (C) Chromatin immunoprecipitation coupled with qPCR analysis of H3K4me3 occupancy near exon 1 of *alcamb* in high stage (3.3 hpf) wild-type embryos from incrosses of F1 *alcama*^{+/+} (i.e., wild types from *alcama*^{ptc/+} intercrosses), compared with wild-type embryos from AB incrosses. Green peaks in *alcamb* locus represent prim-5 stage (24 hpf) H3K4me3 ChIP-seq data obtained from DANIO-CODE. Scale bar: 1 kb. *Ct* values are listed in Table S1.

Table S1: Ct values for Fig. 2-5 and Fig. S1-S9.

Table S2: Used oligos for qPCR primers, genotyping primers, gRNA for mutagenesis, and primers for cloning.