## **Supplementary Information**

## Reactive Metal Boride Nanoparticles Trap Lipopolysaccharide and Peptidoglycan for Bacteria-Infected Wound Healing

Yun Meng<sup>1,2#</sup>, Lijie Chen<sup>2#</sup>, Yang Chen<sup>1,3#</sup>, Jieyun Shi<sup>1</sup>, Zheng Zhang<sup>4</sup>, Yiwen Wang<sup>4</sup>, Fan Wu<sup>2</sup>, Xingwu Jiang<sup>2</sup>, Wei Yang<sup>1</sup>, Li Zhang<sup>1</sup>, Chaochao Wang<sup>1</sup>, Xianfu Meng<sup>1</sup>, Yelin Wu<sup>1\*</sup>, Wenbo Bu<sup>1,2\*</sup>

<sup>1</sup>Tongji University Cancer Center, Shanghai Tenth Peoples Hospital, Tongji University School of Medicine, Shanghai 200072, P. R. China

<sup>2</sup>Department of Materials Science and State Key Laboratory of Molecular Engineering of Polymers, Fudan University, Shanghai 200433, P. R. China

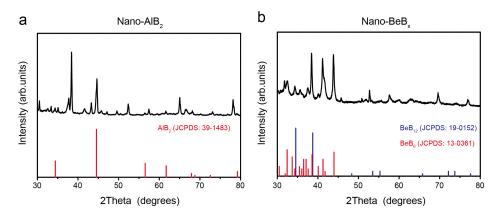
<sup>3</sup> School of Life Sciences and Technology, Tongji University, Shanghai 200092, P. R. China

<sup>4</sup>School of life Science, East China Normal University, Shanghai 200241, P. R. China

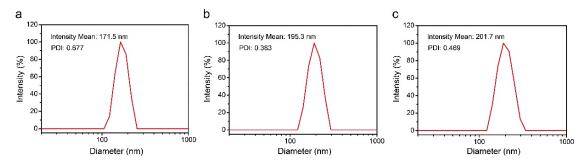
<sup>#</sup>These authors contributed equally to this work.

Email: sk\_wuyelin@tongji.edu.cn; wbbu@fudan.edu.cn;

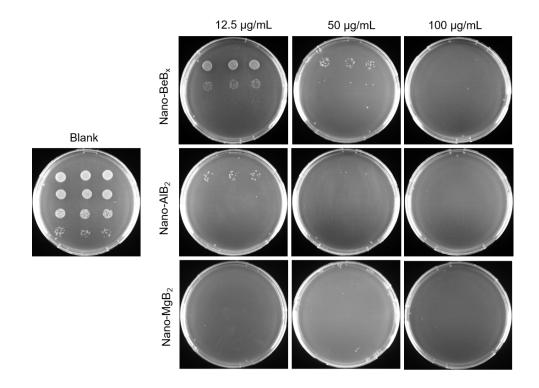
Supplementary Information: Supplementary Figures 1-23 Supplementary Tables 1, 2, 3



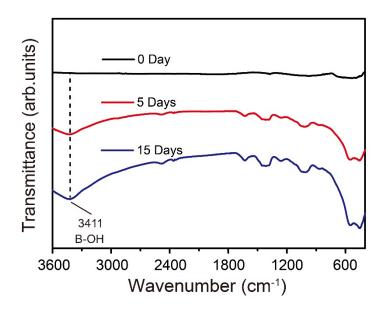
Supplementary Fig. 1 XRD patterns of  $\mathbf{a}$ , Nano-AlB<sub>2</sub> and  $\mathbf{b}$ , Nano-BeB<sub>x</sub>. Source data are provided as a Source Data file.



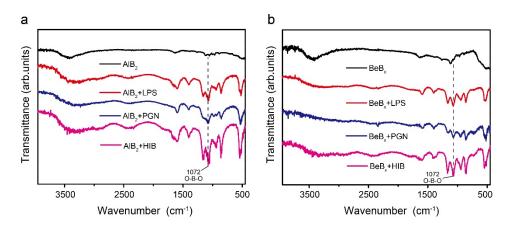
**Supplementary Fig. 2** Particle size analysis of **a**, Nano-MgB<sub>2</sub>, **b**, Nano-AlB<sub>2</sub> and **c**, Nano-BeB<sub>x</sub>. Source data are provided as a Source Data file.



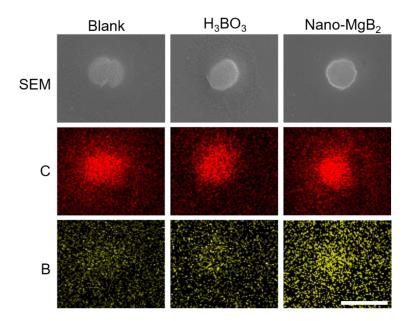
**Supplementary Fig. 3** Antibacterial activity of Nano-MgB<sub>2</sub>, Nano-AlB<sub>2</sub> and Nano-BeB<sub>x</sub>. *P. aeruginosa* treated with different concentrations of Nano-MgB<sub>2</sub>, Nano-AlB<sub>2</sub> and Nano-BeB<sub>x</sub> (n=3 biological independent cells).



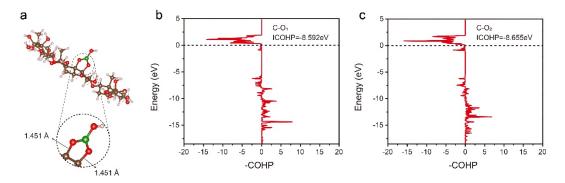
**Supplementary Fig. 4** FTIR spectrum analysis of the hydrolysis process of Nano-MgB<sub>2</sub> in 15 days. Source data are provided as a Source Data file.



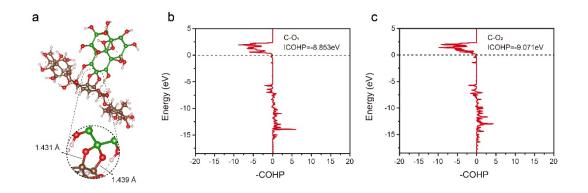
Supplementary Fig. 5 FTIR spectra of O-B-O of  $\mathbf{a}$ , Nano-AlB<sub>2</sub> and  $\mathbf{b}$ , Nano-BeB<sub>x</sub> incubated with LPS, PGN, and HIB, respectively. Source data are provided as a Source Data file.



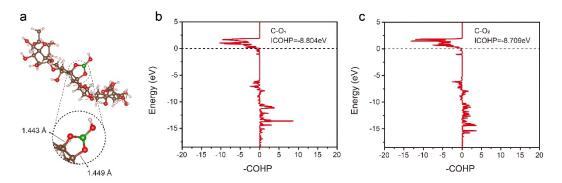
**Supplementary Fig. 6** Elemental mapping of *S. aureus* incubated with  $H_3BO_3$  and Nano-MgB<sub>2</sub> for 3 h (scale bar=500 nm). Data are representative of at least three independent experiments with similar results.



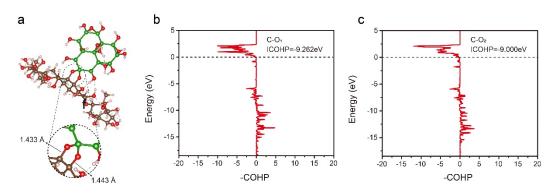
**Supplementary Fig. 7** Theoretical simulation for the  $H_3BO_3$ -saccharide complexation. **a**, DFT calculations and **b-c**, COHP analysis of C-O<sub>1</sub>, C-O<sub>2</sub> bond interactions corresponding to the complexation of  $H_3BO_3$  with the 3, 4-o-hydroxyl groups of bicyclic monosaccharide structure. Source data are provided as a Source Data file.



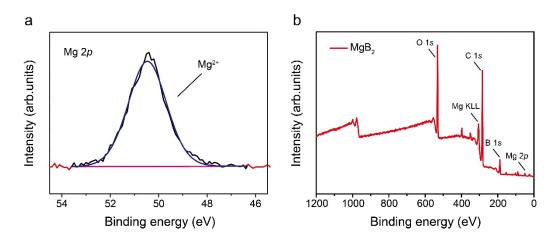
**Supplementary Fig. 8** Theoretical simulation for the Nano-MgB<sub>2</sub>-saccharide complexation. **a**, DFT calculations and **b-c**, COHP analysis of C-O<sub>1</sub>, C-O<sub>2</sub> bond interactions corresponding to the complexation of hydrolysate of Nano-MgB<sub>2</sub> with the 3, 4-o-hydroxyl groups of bicyclic monosaccharide structure. Source data are provided as a Source Data file.



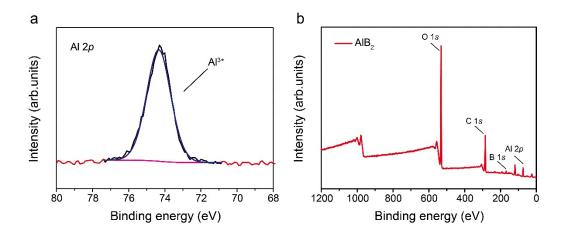
**Supplementary Fig. 9** Theoretical simulation for the  $H_3BO_3$ -saccharide complexation. **a**, DFT calculations and **b-c**, COHP analysis of C-O<sub>1</sub>, C-O<sub>2</sub> bond interactions corresponding to the complexation of  $H_3BO_3$  with the 4, 5-o-hydroxyl groups of bicyclic monosaccharide structure. Source data are provided as a Source Data file.



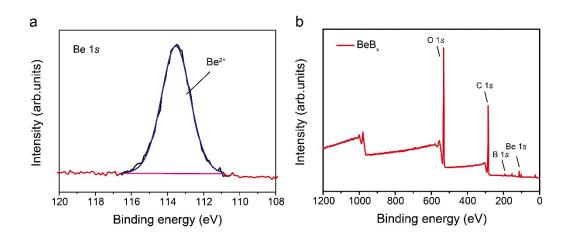
**Supplementary Fig. 10** Theoretical simulation for the Nano-MgB<sub>2</sub>-saccharide complexation. **a**, DFT calculations and **b-c**, COHP analysis of C-O<sub>1</sub>, C-O<sub>2</sub> bond interactions corresponding to the complexation of hydrolysate of Nano-MgB<sub>2</sub> with the 3, 4-o-hydroxyl groups of bicyclic monosaccharide structure. Source data are provided as a Source Data file.



Supplementary Fig. 11 XPS characterization of Nano-MgB<sub>2</sub>. **a**, XPS fine spectra of Mg 2p signal and **b**, XPS survey spectra of Nano-MgB<sub>2</sub> deconvoluted by the multi-Gaussian function.

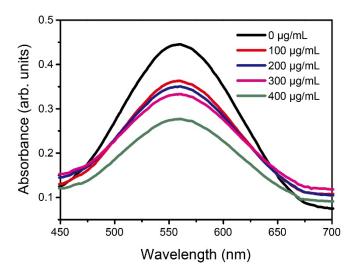


Supplementary Fig. 12 XPS characterization of Nano-AlB<sub>2</sub>. **a**, XPS fine spectra of Al 2p signal and **b**, XPS survey spectra of Nano-AlB<sub>2</sub> deconvoluted by the multi-Gaussian function.

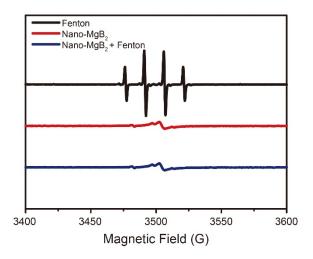


Supplementary Fig. 13 XPS characterization of Nano-BeB<sub>x</sub>. a, XPS fine spectra of Be 1s signal

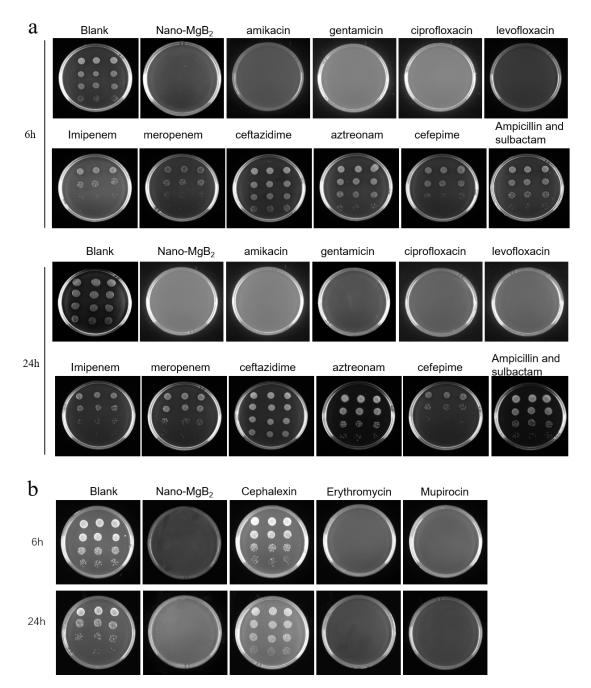
and **b**, XPS survey spectra of Nano-BeB $_x$  deconvoluted by the multi-Gaussian function.



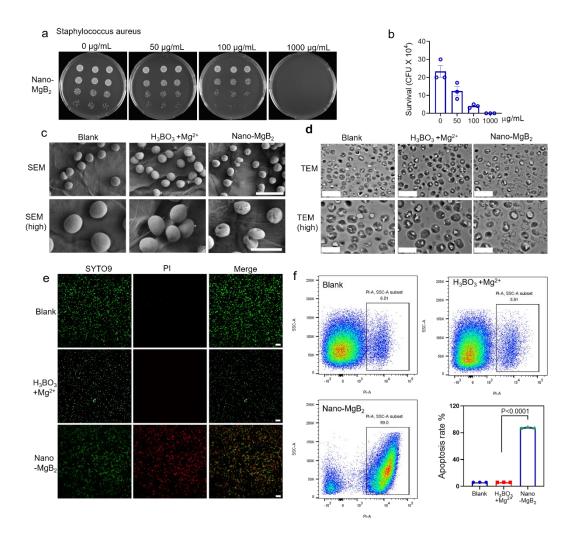
**Supplementary Fig. 14** UV-vis spectra of PTIO• scavenging ability with different concentrations of Nano-MgB<sub>2</sub>. Data are representative of at least three independent experiments. Source data are provided as a Source Data file.



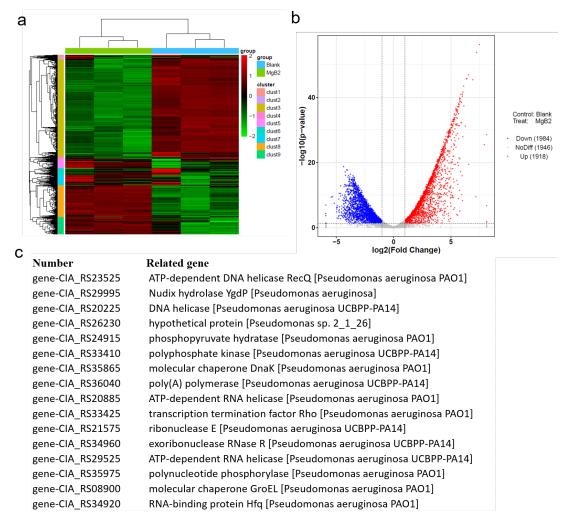
**Supplementary Fig. 15** ESR spectra of Nano-MgB<sub>2</sub> under different conditions. Source data are provided as a Source Data file.



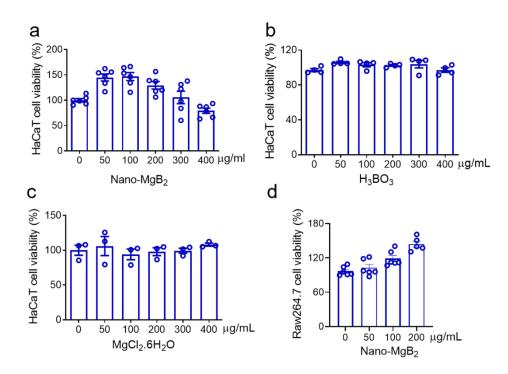
**Supplementary Fig. 16** Antibacterial activity of Nano-MgB<sub>2</sub> and antibiotics. **a**, *P. aeruginosa* treated with Nano-MgB<sub>2</sub> (12.5  $\mu$ g/mL) and different kinds of antibiotics (16  $\mu$ g/mL) for 6 h and 24 h. **b**, *S. aureus* treated with Nano-MgB<sub>2</sub> (1 mg/mL) and different kinds of antibiotics (1 mg/mL) for 6 h and 24 h (n=3 biological independent cells).



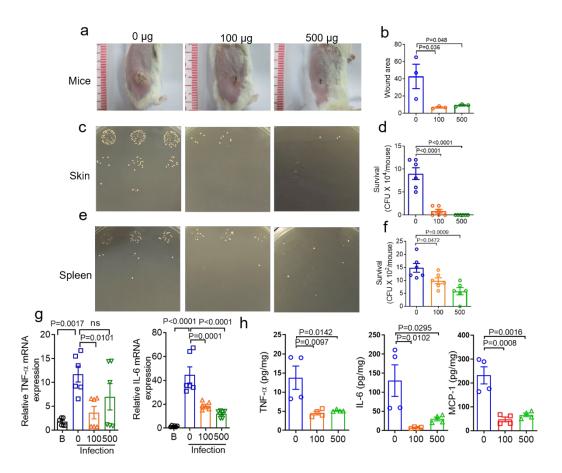
**Supplementary Fig. 17** Antibacterial activity of Nano-MgB<sub>2</sub> against *S. aureus*. **a**, *S. aureus* treated with different concentrations of Nano-MgB<sub>2</sub> using colony-forming units counting method. **b**, Survival rates of *S. aureus* taken as in (**a**) (n=3 biologically independent cell). **c**, SEM images of *S. aureus* subjected to Nano-MgB<sub>2</sub> and H<sub>3</sub>BO<sub>3</sub>+Mg<sup>2+</sup> Scale bar (up)=2 µm, scale bar (down)=1 µm. **d**, TEM images of *S. aureus* subjected to Nano-MgB<sub>2</sub> and H<sub>3</sub>BO<sub>3</sub>+Mg<sup>2+</sup> Scale bar (up)=2 µm, scale bar (up)=2 µm, scale bar (down)=1 µm. **e**, Cell membrane permeability of *S. aureus* treated with Nano-MgB<sub>2</sub> (500 µg/mL) and H<sub>3</sub>BO<sub>3</sub>+Mg<sup>2+</sup> by SYTO9 and PI staining. All cells were labelled by the membrane-permeable SYTO9 (green), whereas only cells with damaged membrane were positive for PI (red). Scale bar=20 µm. **f**, FACS analysis of PI-positive bacteria treated as in (**e**) (n=3 biologically independent cells). Data are representative of at least three independent experiments with similar results. Values are the mean ± SEM, one-way ANOVA with Bonferroni post-test was used to analyze multiple groups. Source data are provided as a Source Data file.



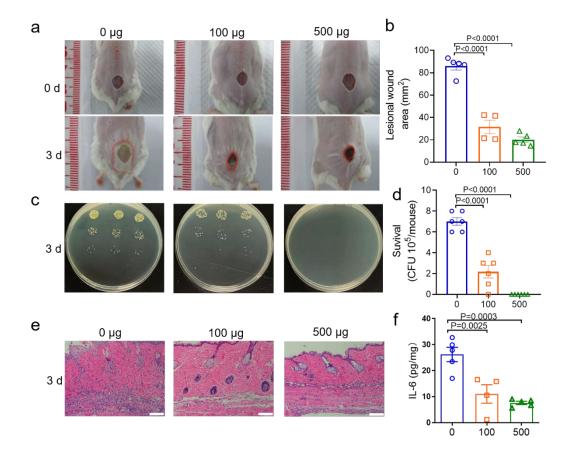
**Supplementary Fig. 18.** RNA seq of *P. aeruginosa* treated with Nano-MgB<sub>2</sub>. **a**, Heat map of differential expressed genes after *P. aeruginosa* treated with Nano-MgB<sub>2</sub>. **b**, Volcano plots comparing gene expression after *P. aeruginosa* treated with Nano-MgB<sub>2</sub>. The P values were calculated using the negative binomial distribution. **c**, The related gene name of Fig 4j.



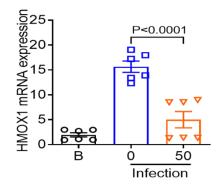
**Supplementary Fig. 19** Biocompatibility of Nano-MgB<sub>2</sub>. **a-c**, HaCaT cells treated with different concentrations of Nano-MgB<sub>2</sub> (n=6 biologically independent cells), H<sub>3</sub>BO<sub>3</sub> (n=4 biologically independent cells) and MgCl<sub>2</sub>. (n=3 biologically independent cells). **d**, Raw 264.7 cells treated with different concentrations of Nano-MgB<sub>2</sub> (n=5 biologically independent cells). Data are representative of at least three independent experiments with similar results. Values are the mean  $\pm$  SEM. Source data are provided as a Source Data file.



Supplementary Fig. 20 Nano-MgB<sub>2</sub> inhibited *S. aureus*-induced skin infection and inflammation. **a**, Photographs of *S. aureus*-infected mouse skin treated with or without 100 µg and 500 µg/mouse Nano-MgB<sub>2</sub> (n=3 biologically independent mice). **b**, The wound healing rate of mice treated as in (**a**) (n=3 biologically independent mice). **c**, **d**, Survival of bacteria from *S. aureus*-infected mouse skin or (**e**, **f**,) spleen taken as in (**a**) (n=6 biologically independent mice). **g**, QPCR of TNF- $\alpha$  and IL-6 in *S. aureus*-infected mouse skin taken as in (**a**) (n=6 biologically independent mice). **h**, TNF- $\alpha$ , IL-6 and MCP-1 protein expression in *S. aureus*-infected mouse skin taken as in (**a**) (n=4 biologically independent mice). Data are representative of at least three independent experiments with similar results. Values are the mean ± SEM, one-way ANOVA with Bonferroni post test was used to analyze multiple groups. Source data are provided as a Source Data file.

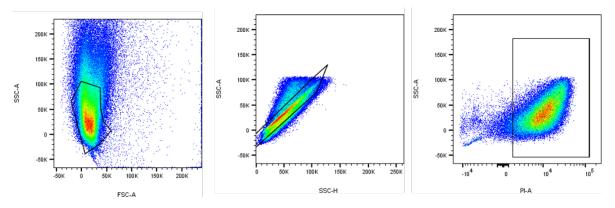


**Supplementary Fig. 21.** Nano-MgB<sub>2</sub> enhanced *S. aureus*-infected skin wound healing. **a**, Photographs of *S. aureus*-infected skin wounds topically treated with 100  $\mu$ g and 500  $\mu$ g/mouse Nano-MgB<sub>2</sub> for 3 days. **b**, The wound healing rate of mice treated as in (**a**) (n=5 biologically independent mice). **c**, **d**, The survival of *S. aureus* in *S. aureus*-infected skin wounds (n=6 biologically independent mice). **e**, The H&E staining of the infected wound areas (Scale bar=100  $\mu$ m). **f**, The Inflammatory factor IL-6 protein expression in the lesional skins from mice treated as in (**a**) (n=5 biologically independent mice). Data are representative of at least three independent experiments with similar results. Values are the mean ± SEM, one-way ANOVA with Bonferroni post test was used to analyze multiple groups. Source data are provided as a Source Data file.



Supplementary Fig. 22. Nano-MgB2 inhibited oxidative stress-related gene hmox1 expression in

*P. aeruginosa*-infected skin wound (n=6 biologically independent mice). Data are representative of at least three independent experiments with similar results. Values are the mean  $\pm$  SEM, one-way ANOVA with Bonferroni post test was used to analyze multiple groups. Source data are provided as a Source Data file.



Supplementary Fig. 23 Gating strategy for bacterial Flow Cytometry.

	MB NPs	LPS-binding peptides	Metal nanomaterials	2D antibacterial materials
Materials	$\begin{array}{l} Nano-MgB_2\\ Nano-AlB_2\\ Nano-BeB_2 \end{array}$	Cathelicidins	Silver MgO ZnO	BP MoS <sub>2</sub> hBN
Structure	Hydrolytic 2D materials	Peptide	Low-dimensional materials	2D materials
Function	Anti-microbial Anti-inflammation	Anti-microbial Anti-inflammation	Anti-microbial	Anti-microbial
Mechanisms	<ul> <li>(1)Trap LPS or PGN by forming stable borate ester bond</li> <li>(2)Trap LPS or PGN to enhance the local cation concentration, leading to cell membrane damage</li> <li>(3) Trap LPS or PGN to inhibit excessive inflammation</li> </ul>	(1) Bind to LPS through electrostatic interaction, leading to cell membrane damage		Cell membrane damage Charge transfer ROS production Oxidative stress

**Supplementary Table 1** The functional and mechanistic differences among MB NPs, LPS-binding peptides, Metal nanomaterials, and 2D antibacterial materials.

mGAPDH-F	CTTAGCCCCCTGGCCAAG
mGAPDH-R	TGGTCATGAGCCCTTCCACA
mTNF-α-F	TCAAGGACTCAAATGGGCTTTC
mTNF-α-R	TGCAGAACTCAGGAATGGACAT
mIL-6-F	CTGCAAGAGACTTCCATCCAGTT
mIL-6-R	GGGAAGGCCGTGGTTGTC
mIL-1β-F	TAACCTGTGGCCTTGG
mIL-1β-R	TGTGCTCTGCTTGTGAG
mMCP-1-F	CAGGTCCCTGTCATGCTTCT

mMCP-1-R	GTGGGGCGTTAACTGCATCT
mHmox1-1-F	CAGAACCCAGTCTATGCCCC
mHmox1-1-R	GTGAGGCCCATACCAGAAGG

Supplementary Table 2 QPCR primers for mice.

Pseudomonas aeruginosa UCBPP-PA14 16S F (96bp)	TGCCTGGTAGTGGGGGGATAA
Pseudomonas aeruginosa UCBPP-PA14 16S R(96bp)	TCTGATAGCGTGAGGTCCGA
gene-CIA_RS23525 F	GAAGAGCGGGAAATGTGGGA
gene-CIA_RS23525 R	GGGGAAGATCACATAGGGCG
gene-CIA_RS29995 F	ATCAGGAAGCCTGGCAGTTC
gene-CIA_RS29995 R	GCTTGAAGGTCACCACCTGT
gene-CIA_RS20225 F	AGTCCAAGGACGGCCTCTAT
gene-CIA_RS20225 R	CATCAGCCAGGGTATCGACC
gene-CIA_RS26230 F	ATCCACATCGACGGCATCAG
gene-CIA_RS26230 R	ATACGGTGCACGTAGTCGTC
gene-CIA_RS24915 F	AGATCGTCGACATCAAGGGC
gene-CIA_RS24915 R	CAGGATCACGTCCGCTTCAA
gene-CIA_RS33410 F	GACCGATCCGAAGGTCATCC
gene-CIA_RS33410 R	ATCGAGCGCACGTGGATATT
gene-CIA_RS35865 F	GTGATCGAGATCGCCGAAGT
gene-CIA_RS35865 R	GGTAGTCGATCAGGCGGATG
gene-CIA_RS36040 F	GCGACTTCACCATCAATGCC
gene-CIA_RS36040 R	CGATTGCGGATGTCGTGAAC
gene-CIA_RS20885 F	CACAGATCCTCGTCGCTACC
gene-CIA_RS20885 R	TTCCAGGTCGTCCATGAAGC
gene-CIA_RS33425 F	TCTTACGATTCTCGCCACGG
gene-CIA_RS33425 R	TTGATGTTGATGGCCGGGAA
gene-CIA_RS21575 F	CTGAACTTCATCCGCCAGGT
gene-CIA_RS21575 R	CTGGAAGCGATTGAACAGCG
gene-CIA_RS34960 F	CTGGAGAAGATCCACGACCG
gene-CIA_RS34960 R	GAGGTGAAGTGGGTGTAGGC
gene-CIA_RS29525 F	ATCCATGGCAACAAGAGCCA
gene-CIA_RS29525 R	CATGGGGCAACTGGTCGATA
gene-CIA_RS35975 F	CATCAAGCAGGACCGCTACA
gene-CIA_RS35975 R	CAGGCCGAATACGTCCTTGA
gene-CIA_RS08900 F	GTTAAGTTCGGCGATTCCGC
gene-CIA_RS08900 R	ACGACTTGTCCAGAACCACG
gene-CIA_RS34920 F	AGGGCATTCGCTACAAGACC
gene-CIA_RS34920 R	GTCGAAGGACTCGATCTGGC

Supplementary Table 3 QPCR primers for *Pseudomonas aeruginosa*.