SUPPLEMENTARY FIGURES

Bioavailable iron titrations reveal oceanic *Synechococcus* ecotypes optimized for different iron availabilities

Naomi E Gilbert¹, Gary R LeCleir¹, Robert F. Strzepek^{2#}, Michael J. Ellwood³, Benjamin

S. Twining⁴, S. Roux⁵, C. Pennacchio⁵, Philip W. Boyd² and Steven W. Wilhelm^{1*}

1. Department of Microbiology, The University of Tennessee, Knoxville, TN 37996

USA

2. Institute for Marine and Antarctic Studies, University of Tasmania, Hobart, Tas.

7004, Australia.

- 3. Research School of Earth Sciences, Australian National University, Canberra, Australia
- 4. Bigelow Laboratory for Ocean Sciences, East Boothbay, Maine, USA
- 5. DOE Joint Genome Institute, Lawrence Berkeley National Laboratory, Berkeley, CA, USA

#. Current Affiliation: Australian Antarctic Program Partnership (AAPP), Institute for Marine and Antarctic Studies, University of Tasmania, Hobart, Tas. 7004, Australia.



RELATIVE FE AVAILABILITY

Supplementary Figure 1. Physiological results of the bulk photosynthetic community within the second growout experiment (GRW2). Total phytoplankton biomass estimates (chlorophyll *a* concentrations, green boxes) and photosynthetic efficiency (F_v/F_m , ratio of variable fluorescence to maximal fluorescence, black circles) as a function of relative Fe availability. Fe was reduced by the addition of 12.5 nM, 5.0 nM, or 1.25 nM DFB and Fe was increased by the addition of 2.5 nM FeCl₃. Results are shown as the means ± error of technical duplicates, averaged between duplicate bottle.



Supplementary Figure 2. Maximum likelihood placement of individual *Synechococcus* RpoB protein sequences, including all eukaryotic and prokaryotic reference RpoB/RPB1 proteins (collapsed triangles). Reference proteins are shown in bold font. Black dots represent bootstrap values > 0.5. Orange and purple blocks correspond to the heatmap shown in Figure 2B.



Supplementary Figure 3. Transcript abundance patterns of eukaryotic and cyanobacteria chlorophyll biosynthesis genes (chlB, chll, chlL, chlN, chlP) across Fe and in situ surface samples. A) Row-scaled (z-score, [Observed TPM - mean TPM]/standard deviation) normalized transcript abundance values, clustered by gene and sample using the Euclidean distance metric. Each column is a different bottle from

the GRW1 incubation, where "A" and "B" show the result for each biological replicate per treatment. B) Average transcript abundance values (TPM) of each gene across both the incubations and *in situ* samples shown within the heatmap. Row names correspond to gene IDs in Supplementary Table 4). C) Chlorophyll biosynthesis genes with transcripts overrepresented in the high-Fe/Fe-added conditions (orange shaded genes in A). Each bar represents the averaged log2(TPM) of genes within the orange-shaded region in A, with each treatment bottle shown as individual points within each bar. The p-value was calculated using the unpaired t-test with Welch's correction. D) Chlorophyll biosynthesis genes with transcripts overrepresented in the low-Fe/DFB-added conditions (purple shaded genes in A). Each bar represents the averaged log2(TPM) of genes within the purple-shaded region in A, with each treatment bottle shown as individual points the averaged log2(TPM) of genes within the purple-shaded region in A, with each treatment bottle shown as individual points to the purple-shaded region in A, with each treatment bottle shown as individual points within each bar. The p-value was calculated using the unpaired t-test with Welch's correction. D) Chlorophyll biosynthesis genes with transcripts overrepresented in the low-Fe/DFB-added conditions (purple shaded genes in A). Each bar represents the averaged log2(TPM) of genes within the purple-shaded region in A, with each treatment bottle shown as individual points within each bar. The p-value was calculated using the unpaired t-test with Welch's correction.



Supplementary Figure 4. Differentiation of high-Fe and low-Fe ferredoxin gene transcripts within the GRW1 assembly. A) Heatmap of individual ferredoxin genes assigned to *Synechococcus*. The row names show the best LAST taxonomic assignment of each gene, followed by the gene ID. Row-scaled TPM (z-score, [Observed TPM – mean TPM]/standard deviation) values are shown in the heatmap and clustered using a Euclidean distance metric. Each column is a different bottle from the GRW1 incubation, where "A" and "B" show the result for each biological replicate per treatment. Averaged transcript abundance (TPM) of each gene across the incubations and *in situ* are shown alongside. B) Ferredoxin genes with transcripts overrepresented in the high-Fe/Fe-added conditions (orange shaded genes in A). Each bar represents the averaged log2(TPM) of

genes within the orange-shaded region in A, with each treatment bottle shown as individual points within each bar. The p-value was calculated using an unpaired t-test. C) Ferredoxin genes with transcripts overrepresented in the low-Fe/DFB-added conditions (purple shaded genes in A). Each bar represents the averaged log2(TPM) of genes within the purple-shaded region in A, with each treatment bottle shown as individual points within each bar. An exact p-value was calculated using the Mann-Whitney test due to non-normal distribution of this data comparison.



Supplementary Figure 5. Maximum likelihood placement of individual *Synechococcus* (and one *Prochlorococcus*, FTN_1051508) ferritin protein sequences. Reference proteins are shown in bold font. Black dots represent bootstrap values > 0.5. Orange and purple blocks correspond to the heatmap shown in Figure 5B.



Supplementary Figure 6. Targeted genome analysis and read recruitment to Synechococcus sp. CC9311 (Genbank accession: CP000435.1) and Synechococcus sp. BL107 (Genbank accession: NZ DS022298.1). A) Heatmap displaying genes of interest involved in Fe metabolism and Fe status marker genes comparing competitive read recruitment to each strain (row annotation). Normalized transcript abundance values are scaled by row (z-score, [Observed TPM - mean TPM]/standard deviation). Each column is a different bottle from the GRW1 incubation, where "A" and "B" show the result for each biological replicate per treatment. Red stars represent genes highlighted in red within the CC9311 genome. P-values indicated by asterisk show whether or not the mean transcript values across bottles between +DFB-added and +Fe-added incubations are significantly different using either unpaired t-tests, unpaired t-test with Welch's correction, or Mann-Whitney tests (see Supplementary Methods for details). B) Averaged transcript (TPM) abundances of each gene across the incubations and in situ surface samples (March05_5m, March05_15, March07_15m, March18_15m). C) Annotated genome of Synechococcus sp. CC9311. Genes highlighted in red correspond to genes with red stars in Supplementary Figure 5A. The genome map was constructed in http://cgview.ca/.



Supplementary Figure 7. Read recruitment of a second DFB and Fe amendment incubation ("GRW2") to *Synechococcus* sp. CC9311 and *Synechococcus* sp. BL107 reference genomes. Each column is a different bottle from the GRW2 incubation, where "A" and "B" show the result for each biological replicate per treatment. Normalized transcript abundance values (TPM) are scaled by row (z-score, [Observed TPM – mean TPM]/standard deviation). Averaged transcript abundances (TPM) for each gene are shown on the left hand bar plot.