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Supplementary appendix

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Supplement to: Zhang, X, Chen L-L, Ip JD, et al. Omicron sublineage recombinant XBB evades neutralising antibodies in recipients of BNT162b2 or CoronaVac vaccines. *Lancet Microbe* 2022; published online December 6. https://doi.org/10.1016/S2666-5247(22)00335-4.





- **Supplementary Figure S1.** Comparison of live virus neutralising antibody titers against
- 5 ancestral strain and Omicron sublineages BA.5.2, XBB.1 and XBB.3 for individuals with
- 6 different history of vaccination and infection. Dotted horizontal lines represent the lower
- 7 limit of detection. P values were shown if <0.05. NT_{50} , 50% neutralisation titer



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Patient	Collection date (Days after symptom onset)		No. of vaccine doses received	Type of vaccine
	Acute serum	Convalescent serum		
BA.5.2 infected	3	39	2	BNT162b2
XBB.1 infected	5	18	3	BNT162b2
XBB.3 infected	3	18	4	BNT162b2

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Supplementary Figure S2. Comparison of live virus neutralising antibody titers against ancestral strain and Omicron sublineages BA.5.2, XBB.1 and XBB.3 for the 3 patients with acute and convalescent serum specimens available. Details of the 3 infected patient are shown in the table. The fold increase 50% neutralisation titer between acute and convalescent sera is shown. Dotted horizontal lines represent the lower limit of detection. NT50, 50% neutralisation titer

Supplementary Table S1. Characteristics of study participants

	2x-vaccinated/ BA.2 infected (n=7)	3x-vaccinated/ BA.2 infected (n=7)	3x-vaccinated/ non-infected (n=9)	4x-vaccinated/ non-infected (n=7)	Total (n=30)
Demographics					
Age in years, median (range)	52 (44-59)	51 (41-59)	58 (53-73)	72 (70-86)	57 (49-71)
Female sex	6 (85.7)	5 (71.4)	6 (66.7)	4 (57.1)	21 (70)
Vaccine type					
BNT162b2	4 (57.1)	4 (57.1)	6 (66.7)	3 (42.9)	17 (56.7)
CoronaVac	3 (42.9)	3 (42.9)	3 (33.3)	4 (57.1)	13 (43.3)
Interval between vaccination/infection and serum collection date					
1 st vaccine dose and serum collection	350 (336-365)	350 (344-372)	364 (315-391)	491 (450-494)	N/A
2 nd vaccine dose and serum collection	322 (311-344)	329 (311-344)	339 (287-366)	464 (429-470)	N/A
3 rd vaccine dose and serum collection	N/A	116 (85-148)	126 (92-146)	215 (197-232)	N/A
4 th vaccine dose and serum collection	N/A	N/A	N/A	89 (82-111)	N/A
Infection ^a and serum collection	140 (135-150)	104 (21-130)	N/A	N/A	N/A

Data are expressed as no. (%) unless otherwise specified N/A, not applicable ^a Defined as the date of diagnosis

23 METHODS

24 Serum specimens

Serum specimens were collected in our ongoing prospective cohort study which enrolled
COVID-19 vaccine recipients and COVID-19 patients in Hong Kong¹. Details these
demographics, vaccine type, interval between vaccine doses and serum collection, and interval
between infection and serum collection were documented. The study has been approved by the
Institutional Review Board of the University of Hong Kong/Hospital Authority Hong Kong West
Cluster (UW 21-214 and UW 13-265). Written informed consent were obtained from all study
participants.

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33 Viral culture

34 Viral culture was performed in a biosafety level 3 facility as we described previously with slight modifications^{2,3}. Briefly, TMPRSS2-expressing VeroE6 (VeroE6/TMPRSS2) cells 35 (JCRB Cat#JCRB1819) were seeded with 1 mL of minimum essential medium (MEM) (Gibco®, 36 37 Thermo Fisher Scientific) with 10% fetal bovine serum (FBS) and 5 mg/mL G418 (Gibco®, Thermo Fisher Scientific) at 1×10^5 cells in a shell vial (Diagnostic Hybrids, Inc). The plates 38 39 were incubated at 37°C in a carbon dioxide incubator until confluence for inoculation. Each well was inoculated with 100 µL of clinical specimen. One hour after incubation, the clinical 40 specimen was removed and cells were replenished with 1 mL of MEM medium with 1% FBS, 41 42 100 U/ml penicillin-streptomycin, 100 U/ml of nystatin, and 25 mM HEPES (Gibco®, ThermoFisher Scientific). The cells were incubated at 37° C with 5% CO₂ and observed 2-3 43 times per day for virus-induced cytopathic effect (CPE). Cultures with more than 50% virus-44 induced CPE were expanded to large volume in VeroE6/TMPRSS2 cells with the same culture 45 condition. The 50% tissue culture infective doses (TCID₅₀) were determined in 46

47 VeroE6/TMPRSS2 cells. The whole genome sequence of the culture isolates was determined
48 using nanopore sequencing (Oxford Nanopore Technologies).

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50 Live virus neutralising antibody assay

Live virus neutralising antibody (nAb) assay was performed as we described previously ⁴⁻ 51 ⁶. The SARS-CoV-2 BA.5.2 isolate (GISAID accession number EPI ISL 13777658), XBB.1 52 isolate (GISAID accession number EPI_ISL_15602393), and XBB.3 isolate (GISAID accession 53 number EPI_ISL_15602394) were isolated in this study. The ancestral strain (GISAID 54 accession number: EPI ISL 434571) was isolated from previous studies ³. Briefly, serum 55 samples were heat-inactivated at 56°C for 30 min and were serially diluted in 2-folds with MEM 56 containing 1% FBS. Duplicates of each diluted serum were mixed with a SARS-CoV-2 virus 57 isolate to reach a final concentration of 100 TCID₅₀ and were incubated at 37°C for 1 hour. After 58 59 incubation, 100 µL of the serum-virus mixture was then added to VeroE6/TMPRSS2 cells that were seeded in 96-well plates 24 hours before infection. The cells were incubated with the 60 mixture at 37°C. After incubation for 3 days, CPE was visually scored for each well by two 61 62 independent observers. The 50% neutralisation titer (NT₅₀) was determined by using log (inhibitor) vs normalized response- variable slope in GraphPad PRISM version 9.4.0. For 63 statistical analysis, a value of 5 was assigned if the live virus neutralising antibody titer is <10. 64 65

66 Statistical analysis

67 We did statistical analysis using PRISM 9.4.0. NT₅₀ against ancestral strain, BA.5.2,
68 XBB.1 and XBB.3 were compared using one way ANOVA and corrected for multiple

comparison using Dunn's multiple comparisons test. A *P* value of less than 0.05 was judgedstatistically significant.

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