

THE LANCET

Infectious Diseases

Supplementary appendix

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Supplement to: Uraki R, Ito M, Furusawa Y, et al. Humoral immune evasion of the omicron subvariants BQ.1.1 and XBB. *Lancet Infect Dis* 2022; published online Dec 7. [https://doi.org/10.1016/S1473-3099\(22\)00816-7](https://doi.org/10.1016/S1473-3099(22)00816-7).

Humoral immune evasion of the Omicron Subvariants BQ.1.1 and XBB.

Supplementary Appendix

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Supplementary Materials and Methods

Cells.

Vero E6-TMPRSS2-T2A-ACE2 cells (provided by Dr. Barney Graham, NIAID Vaccine Research Center) were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% Fetal Calf Serum (FCS), 100 U/mL penicillin–streptomycin, and 10 µg/mL puromycin. VeroE6/TMPRSS2 (JCRB 1819) cells were propagated in the presence of 1 mg/ml geneticin (G418; Invivogen) and 5 µg/ml plasmocin prophylactic (Invivogen) in DMEM containing 10% FCS. Vero E6-TMPRSS2-T2A-ACE2 and VeroE6/TMPRSS2 cells were maintained at 37 °C with 5% CO₂. The cells were regularly tested for mycoplasma contamination by using PCR, and confirmed to be mycoplasma-free.

Viruses.

hCoV-19/Japan/TY41-796/2022 (Omicron BQ.1.1)¹, hCoV-19/Japan/TY41-795/2022 (Omicron XBB), hCoV-19/Japan/UT-NCD1288-2N/2022 (Omicron BA.2)², hCoV-19/Japan/TY41-702/2022 (Omicron BA.5)³, and SARS-CoV-2/UT-NC002-1T/Human/2020/Tokyo were propagated in VeroE6/TMPRSS2 cells.

All experiments with SARS-CoV-2 were performed in enhanced biosafety level 3 (BSL3) containment laboratories at the University of Tokyo and the National Institute of Infectious Diseases, Japan, which are approved for such use by the Ministry of Agriculture, Forestry, and Fisheries, Japan.

Clinical specimens.

After informed consent was obtained, plasma specimens were collected from COVID-19 convalescent individuals and vaccinees. The research protocol was approved by the Research Ethics Review Committee of the Institute of Medical Science of the University of Tokyo (approval numbers: 2019–71–0201 and 2020-74-0226).

Focus reduction neutralisation test.

Neutralisation activities of plasma were determined by using a focus reduction neutralisation test as previously described.⁴ After the plasma samples were incubated at 56 °C for 1 h, the samples were serially diluted five-fold with DMEM containing 2% FCS in 96-well plates and mixed with 100–400 focus-forming units (FFU) of virus/well, followed by incubation at room temperature for 1 h. The plasma-virus mixture (50 µl) was then inoculated onto Vero E6-TMPRSS2-T2A-ACE2 cells in 96-well plates in duplicate and incubated for 1 h at 37 °C. An equal volume of 1.5% Methyl Cellulose 400 (FUJIFILM Wako Pure Chemical Corporation) in culture medium was then added to each well. The cells were incubated for 14–16 h at 37 °C and then fixed with formalin. After the formalin was removed, the cells were immunostained with a mouse monoclonal antibody against SARS-CoV1/2 nucleoprotein [clone 1C7C7 (Sigma-Aldrich)], followed by a horseradish peroxidase-labeled goat anti-mouse immunoglobulin (SeraCare Life Sciences or Jackson ImmunoResearch Laboratories Inc.). The infected cells were stained with TrueBlue Substrate (SeraCare Life Sciences) and then washed with distilled water. After cell drying, the focus numbers were quantified by using an ImmunoSpot S6 Analyzer, ImmunoCapture software, and BioSpot software (Cellular Technology). The results are expressed as the 50% focus reduction neutralisation titre (FRNT₅₀). The FRNT₅₀ values were calculated by using GraphPad Prism (GraphPad Software).

Acknowledgements

We thank Susan Watson for scientific editing. We also thank Kyoko Yokota, Rie Onoue, Madoka Yoshikawa, and Naoko Mizutani for technical assistance. Vero E6-TMPRSS2-T2A-ACE2 cells were provided by Dr. Barney Graham, NIAID Vaccine Research Center.

Author Contributions

R.U.: conceptualization, formal analysis, validation, visualization, and writing the first draft. M. Ito.: data curation, formal analysis, and methodology. Y.F.: data curation. S. Yamayoshi: conceptualization and methodology. K.I-H.: resources and validation. E.A., M.S., M. Koga., T.T, S. Yamamoto, A.O, M. Kiso, Y.S-T., H.U., H.Y. : resources. M. Imai: conceptualization, supervision, formal analysis, validation, visualization, and writing the first draft. Y.K.: conceptualization, supervision, writing (review and editing), and funding acquisition. R.U., M. Ito, and Y.F. contributed equally.

Declaration of Interests

Y.K. has received unrelated funding support from Daiichi Sankyo Pharmaceutical, Toyama Chemical, Tauns Laboratories, Inc., Shionogi & Co. LTD, Otsuka Pharmaceutical, KM Biologics, Kyoritsu Seiyaku, Shinya Corporation, and Fuji Rebio. The remaining authors declare that they have no competing interests.

Table S1. Amino acid Substitutions in the clinical isolates of BA.2, BA.5, BQ.1.1, and XBB used in this study

Protein	Position	Virus (Pan-ge lineage)					
		Early SARS-CoV-2 (A)	hCoV-19/Japan/IT-NCDC1288-2022 (BA.2)	hCoV-19/Japan/1Y41-702/2022 (BA.5)	hCoV-19/Japan/1Y4-1796/2022 (BQ.1.1)	hCoV-19/Japan/1Y4-41795/2022 (XBB)	
ORF1a	47	Lys	Lys	Lys	Lys	Arg	
	135	Ser	Arg	Arg	Arg	Arg	
	556	Gln	Gln	Lys	Lys	Gln	
	842	Thr	Ile	Ile	Ile	Ile	
	1387	Gly	Ser	Ser	Ser	Ser	
	3027	Leu	Phe	Phe	Phe	Phe	
	3090	Thr	Ile	Ile	Ile	Ile	
	3201	Leu	Phe	Leu	Leu	Phe	
	3255	Thr	Ile	Ile	Ile	Ile	
	3395	Pro	His	His	His	His	
	3650	Phe	Phe	Leu (85%)	Phe	Phe	
	3675	Ser	del	del	del	del	
	3676	Gly	del	del	del	del	
	3677	Phe	del	del	del	del	
	3730	Ala	Ala	Ala	Ala	Val(22%)	
	3753	Phe	Val(20%)	Phe	Phe	Phe	
	3829	Leu	Leu	Leu	Phe	Leu	
	4391	Leu	Phe(19%)	Leu	Leu	Leu	
	ORF1b	264	Tyr	Tyr	Tyr	His	Tyr
		314	Pro	Leu	Leu	Leu	Leu
659		Met	Met	Met	Met	Ile	
662		Gly	Val	Gly	Gly	Ser	
959		Ser	Pro	Ser	Ser	Pro	
1156		Met	Met	Ile	Ile	Met	
1161		Pro	Ser(27%)	Pro	Pro	Pro	
1191		Asn	Asn	Asn	Ser	Asn	
1315		Arg	Cys	Cys	Cys	Cys	
1566		Ile	Val	Val	Val	Val	
2163		Thr	Ile	Ile	Ile	Ile	
2513		Ala	Val(18%)	Ala	Ala	Ala	
NTD (13-304)		19	Thr	Ile	Ile	Ile	Ile
		24	Leu	del	del	del	del
		25	Pro	del	del	del	del
	26	Pro	del	del	del	del	
	27	Ala	Ser	Ser	Ser	Ser	
	68	His	His	del	del	His	
	70	Val	Val	del	del	Val	
	83	Val	Val	Val	Val	Ala	
	142	Gly	Asp	Asp	Asp	Asp	
	145	Tyr	Tyr	Tyr	Tyr	del	
	146	His	His	His	His	Gln	
	183	Gln	Gln	Gln	Gln	Gln	
	213	Val	Gly	Gly	Gly	Gln	
	252	Gly	Gly	Gly	Gly	Val	
	RBD (331-527)	339	Gly	Asp	Asp	Asp	His
		346	Arg	Arg	Arg	Thr	Thr
		368	Leu	Leu	Leu	Leu	Ile
		371	Ser	Phe	Phe	Phe	Phe
		373	Ser	Pro	Pro	Pro	Pro
		375	Ser	Phe	Phe	Phe	Phe
376		Thr	Ala	Ala	Ala	Ala	
405		Asp	Asn	Asn	Asn	Asn	
408		Arg	Ser	Ser	Ser	Ser	
417		Lys	Asn	Asn	Asn	Asn	
440		Asn	Lys	Lys	Lys	Lys	
444		Lys	Lys	Lys	Thr	Lys	
445		Val	Val	Val	Val	Pro	
446		Gly	Gly	Gly	Gly	Ser	
452		Leu	Leu	Arg	Arg	Leu	
460		Asn	Asn	Asn	Lys	Lys	
477		Ser	Asn	Asn	Asn	Asn	
478		Thr	Lys	Lys	Lys	Lys	
484		Gln	Ala	Ala	Ala	Ala	
486		Phe	Phe	Val	Val	Ser	
490	Phe	Phe	Phe	Phe	Ser		
493	Gln	Arg	Gln	Gln	Gln		
498	Gln	Arg	Arg	Arg	Arg		
501	Asn	Tyr	Tyr	Tyr	Tyr		
SD1/2 (528-684)	505	Tyr	His	His	His	His	
	614	Asp	Gly	Gly	Gly	Gly	
	655	His	Tyr	Tyr	Tyr	Tyr	
	679	Asn	Lys	Lys	Lys	Lys	
S2 (685-1210)	681	Pro	His	His	His	His	
	764	Asn	Lys	Lys	Lys	Lys	
	796	Asp	Tyr	Tyr	Tyr	Tyr	
	954	Gln	His	His	His	His	
969	Asn	Lys	Lys	Lys	Lys		
ORF3a	223	Thr	Ile	Ile	Ile	Ile	
E	9	Thr	Ile	Ile	Ile	Ile	
	11	Thr	Thr	Thr	Thr	Ala	
M	3	Asp	Asp	Asn	Asn	Asp	
	19	Gln	Gln	Gln	Gln	Gln	
ORF6	63	Ala	Thr	Thr	Thr	Thr	
	61	Asp	Leu	Asp	Asp	Leu	
ORF7b	10	Pro	Ser	Pro	Pro	Pro	
	27	Gln	del	Gln	Gln	Gln	
	28	Asn	del	Asn	Asn	Asn	
	29	Ala	del	Ala	Ala	Ala	
N	13	Pro	Leu	Leu	Leu	Leu	
	31	Gln	del	del	del	del	
	32	Arg	del	del	del	del	
	33	Ser	del	del	del	del	
	136	Gln	Gln	Asp	Asp	Gln	
	203	Arg	Lys	Lys	Lys	Lys	
	204	Gly	Arg	Arg	Arg	Arg	
413	Ser	Arg	Arg	Arg	Arg		

Substitutions are shown relative to the reference strain Wuhan/Hu-1/2019. The spike protein is composed of two subunits, S1 and S2. ORF, open reading frame; SP, signal peptide; NTD, N-terminal domain; RBD, receptor-binding domain; SD1/2, subdomain 1 and 2; E, Envelope; M, Membrane; and N, Nucleocapsid. del; deletion

Table S2. Neutralising antibody titres of human plasma obtained from individuals immunized with three doses of BNT162b2 or mRNA-1273 vaccine

Sample ID	Gender	Age	Documented SARS-CoV-2 infection	Vaccine type	Days post-vaccination	FRNT50: 50% focus reduction neutralisation titre				
						(A)	Omicron/BA.2	Omicron/BA.5	Omicron/BQ.1.1	Omicron/XBB
						SARS-CoV-2/UT-NC002-1T/Human/2020/Tokyo	hCoV-19/Japan/UT-NCD1288-2N-2022	hCoV-19/Japan/TY41-702/2022	hCoV-19/Japan/TY41-796/2022	hCoV-19/Japan/TY41-795/2022
HP(H)- 004	F	50	No	BNT162b2x 3	183	426.5	32.8	28.7	<10	<10
HP(H)- 019	F	57	No	BNT162b2x 3	182	147.7	32	10.2	<10	<10
HP(H)- 026	F	25	No	BNT162b2x 3	181	335.9	30.5	15.1	<10	<10
HP(H)- 065	F	40	No	BNT162b2x 3	181	166.9	11	<10	<10	<10
HP(H)- 106	F	29	No	BNT162b2x 3	182	620.1	156.3	49.3	<10	<10
HP(H)- 115	F	48	No	BNT162b2x 3	182	162.1	17.2	<10	<10	<10
HP(H)- 131	M	61	No	BNT162b2x 3	183	172.2	19.4	<10	<10	<10
HP(H)- 168	M	49	No	BNT162b2x 3	182	86.6	<10	<10	<10	<10
HP(H)- 173	M	39	No	BNT162b2x 3	180	74.3	<10	<10	<10	<10
HP(H)- 181	M	54	No	BNT162b2x 3	182	607.9	15.4	<10	<10	<10
HP(H)- 182	F	51	No	BNT162b2x 3	176	411.9	64.9	13.5	<10	<10
HP(H)- 194	M	49	No	BNT162b2x 3	182	86.6	30.7	14.4	<10	<10
HP(H)- 221	F	55	No	BNT162b2x 3	181	66	<10	<10	<10	<10
HP(H)- 303	M	50	No	BNT162b2x 3	188	135.4	15.8	<10	<10	<10
AG- 001	F	54	No	mRNA-1273x 3	189	843.2	284.8	235.2	33.7	73.7
AG- 002	F	48	No	mRNA-1273x 3	187	353.1	<10	<10	<10	<10
AG- 007	F	53	No	mRNA-1273x 3	183	721.6	60.9	38.7	<10	<10
AG- 008	F	57	No	mRNA-1273x 3	183	250.7	22.6	27.7	<10	<10
AG- 010	M	61	No	mRNA-1273x 3	184	585.2	297.8	278.6	122.3	42.2
AG- 011	F	55	No	mRNA-1273x 3	185	647.5	74.7	48.1	13.1	<10

Table S3. Neutralising antibody titres of human plasma obtained from individuals immunized with four doses of BNT162b2 or mRNA-1273 vaccine

Sample ID	Gender	Age	Documented SARS-CoV-2 infection	Vaccine type	Days post-vaccination	FRNT50: 50% focus reduction neutralisation titre				
						(A)	Omicron/BA.2	Omicron/BA.5	Omicron/BQ.1.1	Omicron/XBB
						SARS-CoV-2/UT-NC002-1T/Human/2020/Tokyo	hCoV-19/Japan/UT-NCD1288-2N-2022	hCoV-19/Japan/TY41-702/2022	hCoV-19/Japan/TY41-796/2022	hCoV-19/Japan/TY41-795/2022
HP(H)-032	M	39	No	BNT162b2x 3, mRNA-1273 x 1	53	629	58.9	47.2	10	<10
HP(H)-058	F	58	No	BNT162b2x 3, mRNA-1273 x 1	48	4108.2	836.7	632.9	158.9	97.5
HP(H)-088	F	46	No	BNT162b2x 3, mRNA-1273 x 1	43	844.2	125.8	59.2	24.7	21.6
HP(H)-113	M	62	No	BNT162b2x 3, mRNA-1273 x 1	41	972.2	63	64.9	<10	<10
HP(H)-158	F	29	No	BNT162b2x 3, mRNA-1273 x 1	41	356.3	19.9	<10	<10	<10
HP(H)-182	F	52	No	BNT162b2x 4	54	1581.7	191.7	86.6	29.8	14.9
HP(H)-183	F	47	No	BNT162b2x 3, mRNA-1273 x 1	44	728.2	108.4	72	<10	<10
HP(H)-185	F	48	No	BNT162b2x 3, mRNA-1273 x 1	50	1275.5	203.8	129.4	35.8	13.7
HP(H)-189	F	57	No	BNT162b2x 4	33	598.8	83.7	60	12.7	<10
HP(H)-198	F	33	No	BNT162b2x 3, mRNA-1273 x 1	45	590.3	39.5	21.6	<10	<10
HP(H)-220	M	34	No	BNT162b2x 3, mRNA-1273 x 1	44	360.4	41.7	13	<10	<10
HP(H)-228	F	49	No	BNT162b2x 3, mRNA-1273 x 1	51	595.6	178.2	102.6	21.8	<10
HP(H)-241	F	62	No	BNT162b2x 4	57	659.2	116.2	91	<10	10
HP(H)-250	F	35	No	BNT162b2x 3, mRNA-1273 x 1	49	1741.7	300.5	335.6	42.9	32.7
HP(H)-255	F	51	No	BNT162b2x 3, mRNA-1273 x 1	42	892.1	78.1	39.9	<10	<10
HP(H)-264	M	43	No	BNT162b2x 3, mRNA-1273 x 1	41	901	34.7	65.7	<10	<10
HP(H)-282	M	43	No	BNT162b2x 3, mRNA-1273 x 1	48	256.6	36.5	35.5	<10	<10
HP(H)-297	M	56	No	BNT162b2x 3, mRNA-1273 x 1	45	1184.8	253.9	154.7	64.9	72.8
HP(H)-299	F	53	No	BNT162b2x 4	53	202.9	21	25.4	<10	<10
HP(H)-303	M	51	No	BNT162b2x 4	49	349.6	52.6	41.4	<10	<10

Table S4. Neutralising antibody titres of human plasma obtained from individuals who were infected with Omicron BA.2 after three doses of COVID-19 vaccines

Sample ID	Gender	Age	Documented SARS-CoV-2 infection	Vaccine type	Days post-onset	FRNT50: 50% focus reduction neutralisation titre				
						(A)	Omicron/BA.2	Omicron/BA.5	Omicron/BQ.1.1	Omicron/XBB
						SARS-CoV-2/UT-NC002-1T/Human/2020/Tokyo	hCoV-19/Japan/UT-NCD1288-2N-2022	hCoV-19/Japan/TY41-702/2022	hCoV-19/Japan/TY41-796/2022	hCoV-19/Japan/TY41-795/2022
HPCo-383	F	56	Yes (BA.2)	mRNA-1273 x 2, BNT162b2 x 1	29	425.6	46.4	51.8	<10	<10
HP-S(H)0377	M	29	Yes (BA.2)	BNT162b2x 3	44	499.4	227.7	103.2	30.2	13.6
HP-S(H)0380	M	22	Yes (BA.2)	BNT162b2x 3	42	1452.2	262.9	194.2	16.9	13.9
HP-S(H)0381	M	22	Yes (BA.2)	BNT162b2x 3	44	650.4	218.6	161.5	<10	10
HP-S(H)0382	M	21	Yes (BA.2)	BNT162b2x 3	44	4222.7	656.6	399.2	94.4	25.1
HP-S(H)0383	M	18	Yes (BA.2)	BNT162b2x 3	42	648.9	377.7	258.4	76.9	14.9
HP-S(H)0882	M	23	Yes (BA.2)	BNT162b2x 3	89	601.7	156.5	92.7	<10	<10
HP-S(H)0883	M	24	Yes (BA.2)	BNT162b2x 3	84	3532.7	595.1	219.5	66.4	28.6
HP-S(H)0888	M	21	Yes (BA.2)	BNT162b2x 3	89	3813.5	734.8	300	72.5	36.7
HP-S(H)1056	M	22	Yes (BA.2)	BNT162b2x 3	42	3448.3	1141.6	417.2	288.1	252.7

Supplementary References

1. Imai M, Ito M, Kiso M, et al. Efficacy of Antiviral Agents against Omicron BQ.1.1 and XBB Subvariants. *N Engl J Med. in press*
2. Takashita E, Kinoshita N, Yamayoshi S, et al. Efficacy of Antiviral Agents against the SARS-CoV-2 Omicron Subvariant BA.2. *N Engl J Med. 2022;386:1475-1477.*
3. Takashita E, Yamayoshi S, Simon V, et al. Efficacy of Antibodies and Antiviral Drugs against Omicron BA.2.12.1, BA.4, and BA.5 Subvariants. *N Engl J Med. 2022;387:468-470.*
4. Vanderheiden A, Edara VV, Floyd K, et al. Development of a Rapid Focus Reduction Neutralization Test Assay for Measuring SARS-CoV-2 Neutralizing Antibodies. *Current Protocols in Immunology. 2020;131:e116.*