

Supporting Information for

Transparent, Ultra-Stretching, Tough, Adhesive Carboxyethyl Chitin/Polyacrylamide Hydrogel toward High-Performance Soft Electronics

Jipeng Zhang^{1,2}, Yang Hu^{1,2}, Lina Zhang^{1,2}, Jinping Zhou^{1,2}, and Ang Lu^{1,2,*}

¹ College of Chemistry and Molecular Sciences, Wuhan University, Wuhan 430072, People's Republic of China

² Hubei Engineering Center of Natural Polymer-based Medical Materials, Wuhan University, Wuhan 430072, People's Republic of China

*Corresponding author. E-mail: anglu@whu.edu.cn (Ang Lu)

S1 Supplementary Experimental Section

S1.1 Mechanical Property Test

Tensile and compressive measurements were tested on the hydrogels using a universal tensile-compressive tester (INSTRON instrument, Model 5576, USA). For tension, hydrogel membranes with a length of 50 mm, a width of 13 mm and a thickness of about 1.5 mm were measured at the speed of 50 mm·min⁻¹. For compression, columnar hydrogels with a height of about 10 mm and a diameter of 15.5 mm were tested at the speed of 2 mm min⁻¹. Young's modulus was calculated from the initial linear region of the stress-strain curves. The fracture energy (toughness) was calculated from full region of the stress-strain curves.

S1.2 Adhesion Performance Testing of Hydrogel Samples

The adhesion strength was determined by the lap-shear test using a universal test machine (INSTRON instrument, Model 5576, USA). The glass, plastic, wood, metal substrates and pig skin without contaminants were cut into rectangle with a length of 40 mm and a width of 15 mm. Hydrogel samples (10 × 10 × 1.5 mm³) were sandwiched between two substrates with an area of 10 × 10 mm². After preloaded by 1kg weight for 10 min, the specimens were tested by the standard lap-shear test at a velocity of 10 mm min⁻¹ under ambient conditions. The adhesion strength was calculated by dividing the maximum force by the adhesion area. Additionally, the adhesion-strip cyclic tests were also conducted to evaluate the effect of a cycle load on the adhesion strength of the hydrogels.

S1.3 Conductivity Assessment

Ionic conductivity of the hydrogels was measured by the electrochemical impedance spectroscopy (EIS) using an electrochemical workstation 165 (CHI760E, CH Instruments Ins) operated in the frequency range of 100 to 100 kHz and the amplitude of 5 mV. The hydrogels

were sandwiched between two carbon cloths for the measurement. The ionic conductivity (σ , S m⁻¹) of the hydrogels was calculated according to the following equation:

$$\sigma = \frac{L}{RS} \quad (\text{S1})$$

where L (m), S (m²), and R (Ω) was the length between two carbon cloths, the contact area of the hydrogel with carbon cloths, and resistance obtained by the intercept at the real part in Nyquist plots, respectively.

S1.4 Electrical Measurement

The electrical signals of the hydrogels were recorded by a capacitance meter (CAPACITANCE TESTER, UC2652, UCE Technologies). The change in the relative resistance/capacitance of the hydrogel sensors was examined using the above-mentioned capacitance meter at a constant voltage of 1 V, on the basis of different strains, and human motions. Relative changes in resistance and capacitance were calculated as the following equations:

$$\frac{\Delta R}{R_0} = \frac{R-R_0}{R_0} \times 100\% \quad (\text{S2})$$

$$\frac{\Delta C}{C_0} = \frac{C-C_0}{C_0} \times 100\% \quad (\text{S3})$$

where R_0 , C_0 and R , C are the original resistance, capacitance at the strain of 0% at room temperature and the real-time resistance, capacitance at a certain strain, respectively.

S1.5 Characterization

¹H NMR spectra were recorded on a Bruker Avance-III 400 MHz spectrometer at room temperature. Field emission scanning electron microscopy (FESEM, Zeiss, SIGMA, Germany) was used to characterize morphologies of lyophilized hydrogels. Fourier transform infrared spectroscopy (FT-IR) of lyophilized hydrogels were tested by a Nicolet 170-SX (Thermo Nicolet Ltd., USA) in the wavenumber range from 4000 to 400 cm⁻¹. X-ray photoelectron spectra (XPS, ESCALAB250Xi, Thermo Fisher Scientific, America) analyses were recorded using a Kratos XSAM800 X-ray photoelectron spectrometer. Optical transmittance of the hydrogel films with a thickness of 1.5 mm was observed with a UV-vis spectrometer (UV-6, Shanghai Meipuda Instrument Co., Ltd., China) at a wavelength from 900 to 200 nm. Raman spectroscopy and spatial Raman mapping were performed using a Raman imaging microscope (Thermo Scientific DXR xi, USA). The wavelength of the excitation laser was 532 nm. The collected spectra were preprocessed using cosmic ray removal, noise filtering, and normalization techniques. The multivariate curve resolution (MCR) method developed by OMNICxi software was applied for calculating the proportion of interaction domains.

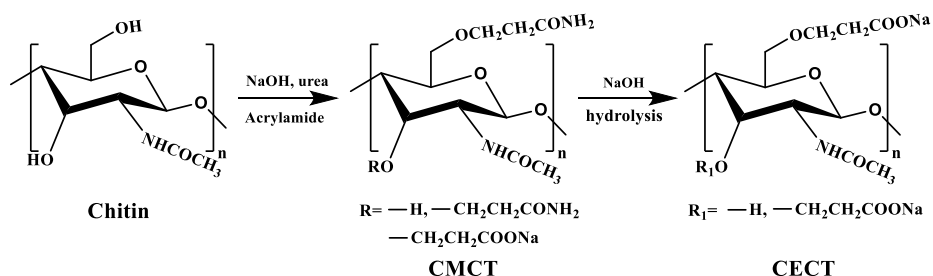
S2 Supplementary Videos

Video S1 (.mp4 format). Demonstration of the strong self-adhesive properties of the CTA hydrogel by vigorously swinging the hand.

Video S2 (.mp4 format). The proof-of-concept demonstrations of the CTA hydrogel as a human-machine interactive system.

Video S3 (.mp4 format). Demonstration of the CTA hydrogel in the field of tactile sensing as a tactile switch.

S3 Supplementary Figures



Scheme S1 Synthesis route of CECT

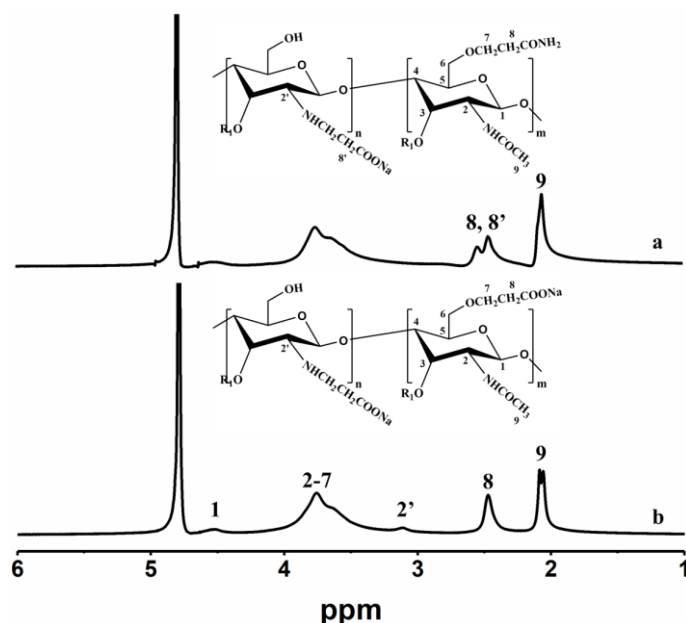


Fig. S1 ^1H NMR spectra of (a) CMCT, (b) CECT

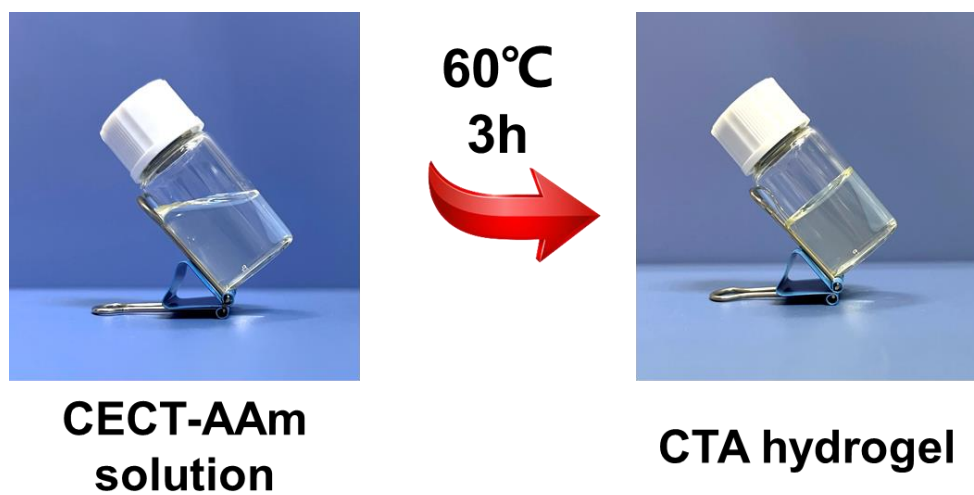


Fig. S2 Preparation of CTA hydrogel



Fig. S3 Photographs of the stretching behavior for CTA hydrogel under a weight loading of 200

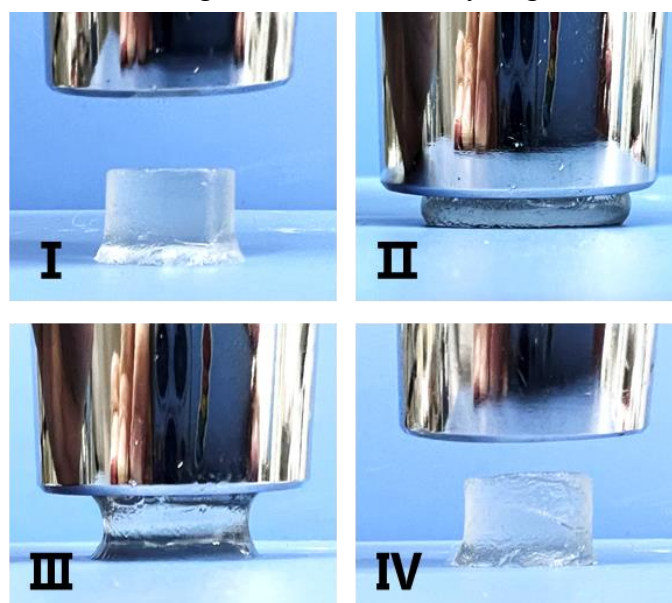


Fig. S4 Photographs of compressing the cylindrical CTA hydrogel with a heavy loading

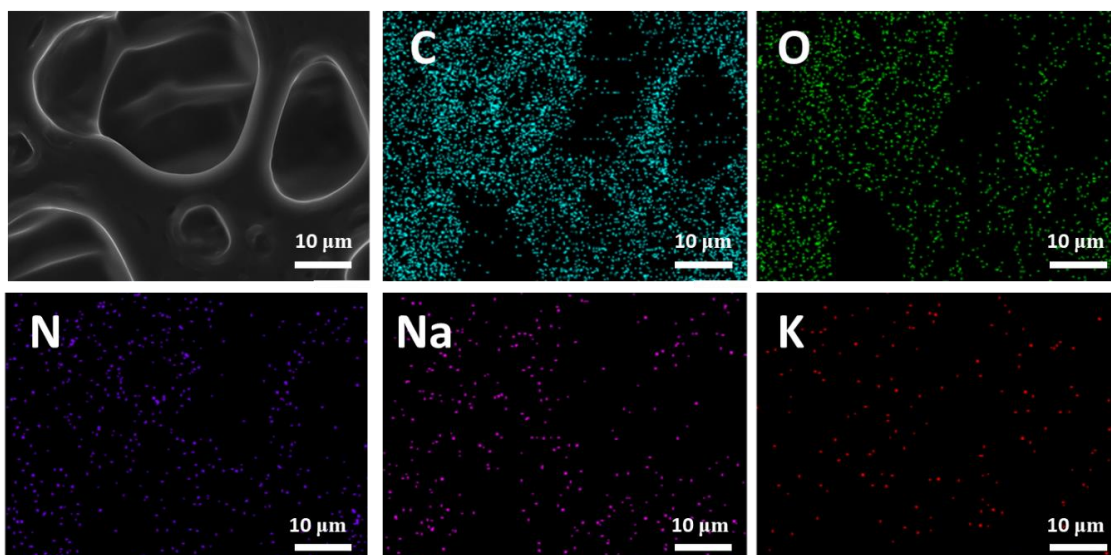


Fig. S5 Corresponding elemental mapping images of the CTA hydrogel

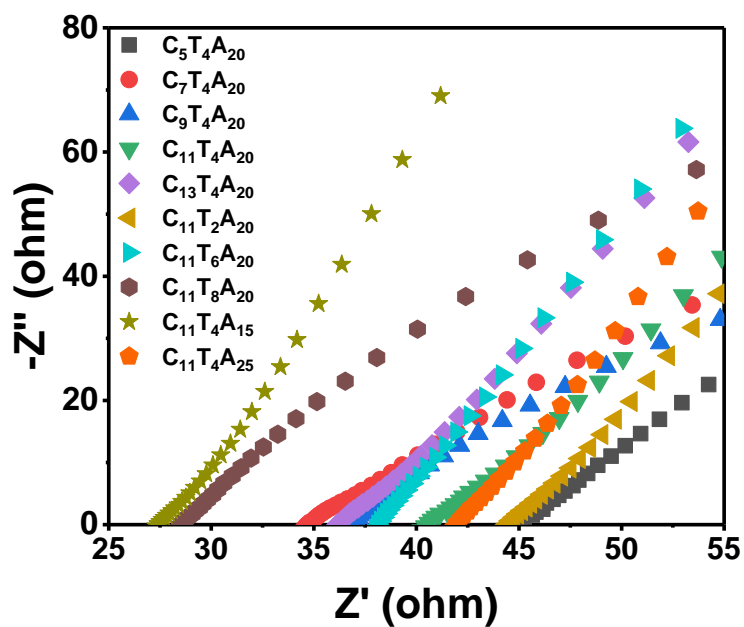


Fig. S6 EIS Nyquist plot of CTA hydrogel

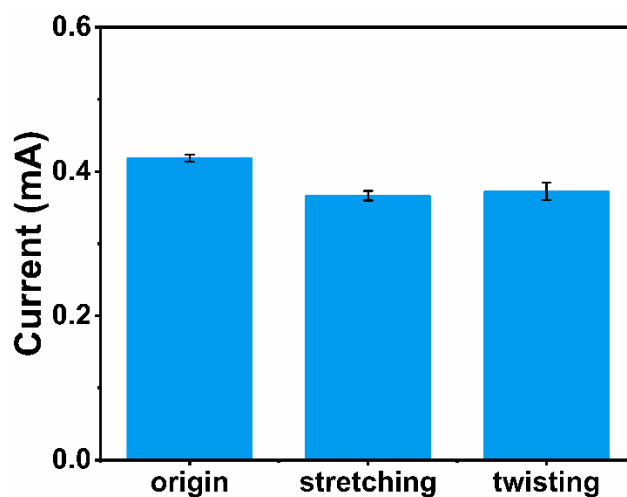


Fig. S7 Current values of $C_{11}T_4A_{20}$ hydrogel in different deformation states

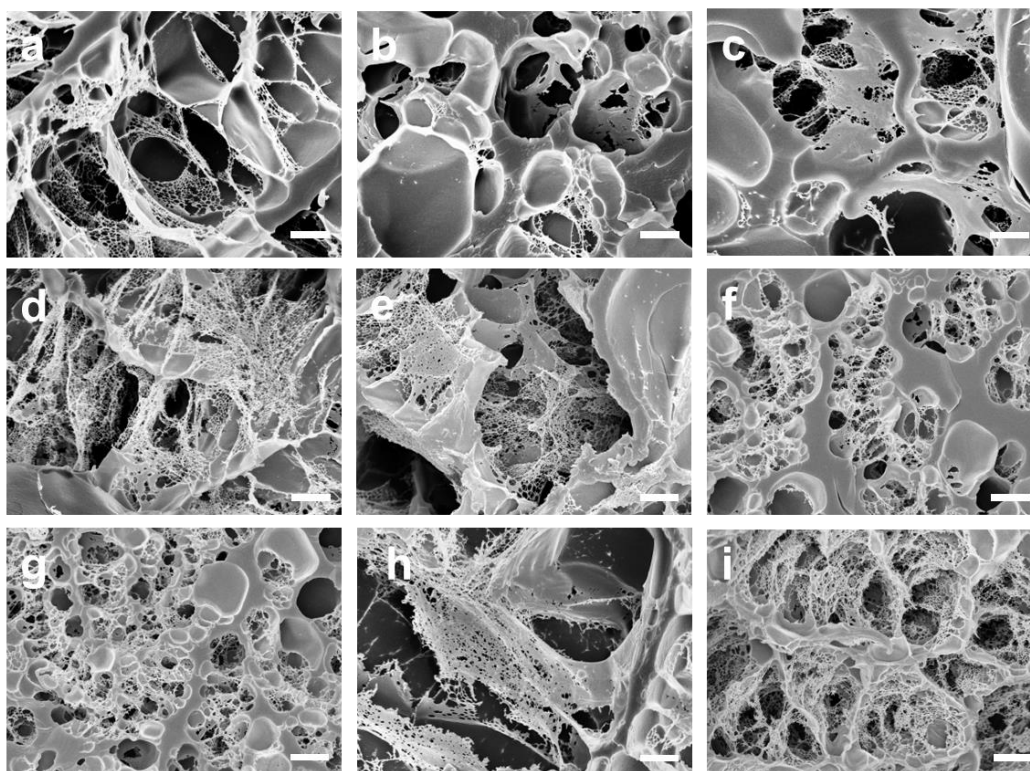


Fig. S8 SEM images of (a) $C_5T_4A_{20}$, (b) $C_7T_4A_{20}$, (c) $C_9T_4A_{20}$, (d) $C_{13}T_4A_{20}$, (e) $C_{11}T_2A_{20}$, (f) $C_{11}T_6A_{20}$, (g) $C_{11}T_8A_{20}$, (h) $C_{11}T_4A_{15}$ and (i) $C_{11}T_4A_{25}$ hydrogels. Scale bar: 2 μm

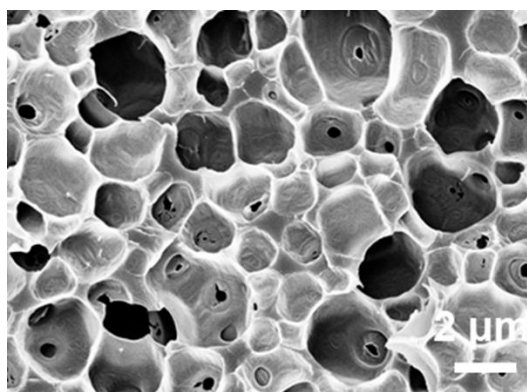


Fig. S9 SEM image of $C_0T_0A_{20}$ hydrogel

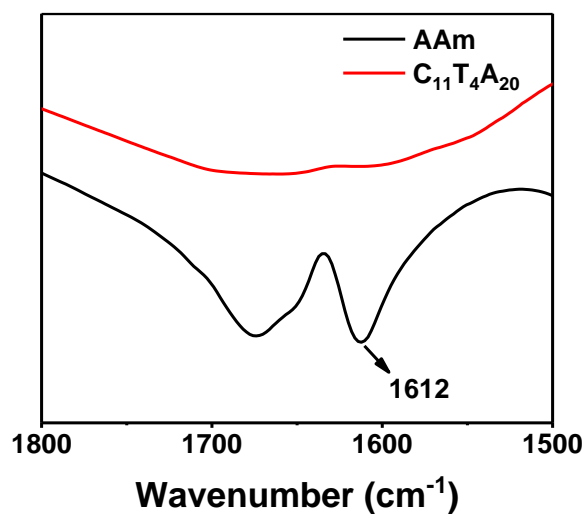


Fig. S10 FT-IR spectra of monomer AAm and $C_{11}T_4A_{20}$ hydrogel

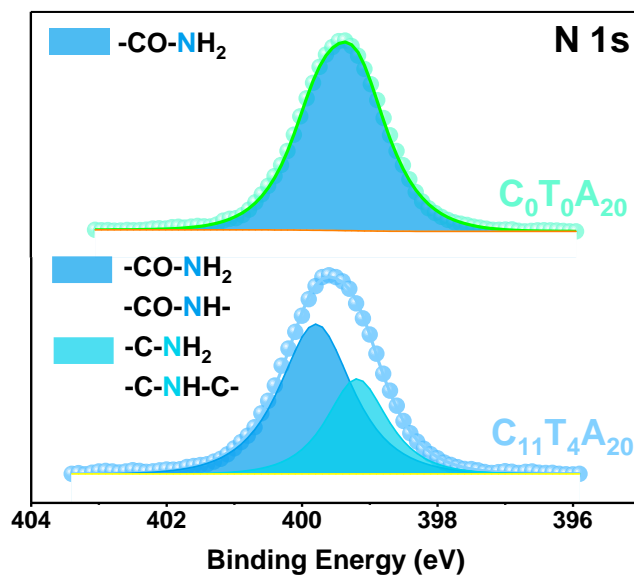


Fig. S11 N 1s XPS spectra for $C_0T_0A_{20}$ and $C_{11}T_4A_{20}$ hydrogel

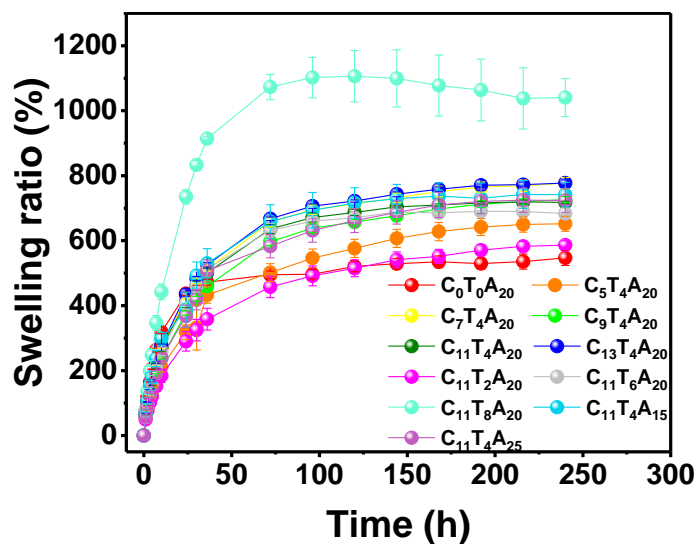


Fig. S12 Equilibrium swelling curves of CTA hydrogels in water

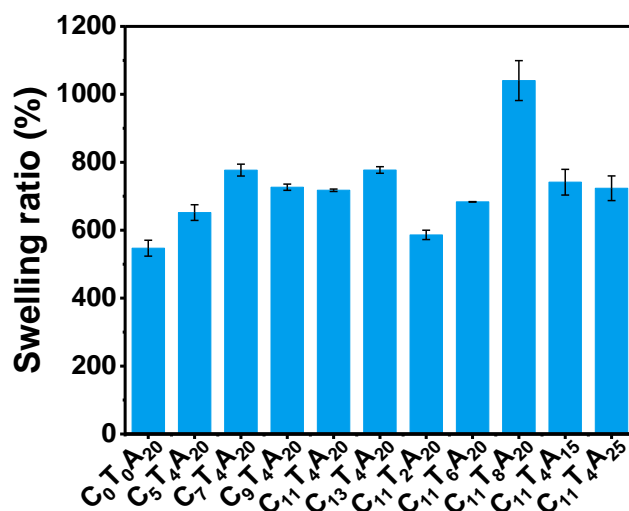


Fig. S13 Equilibrium swelling ratios of CTA hydrogels in water

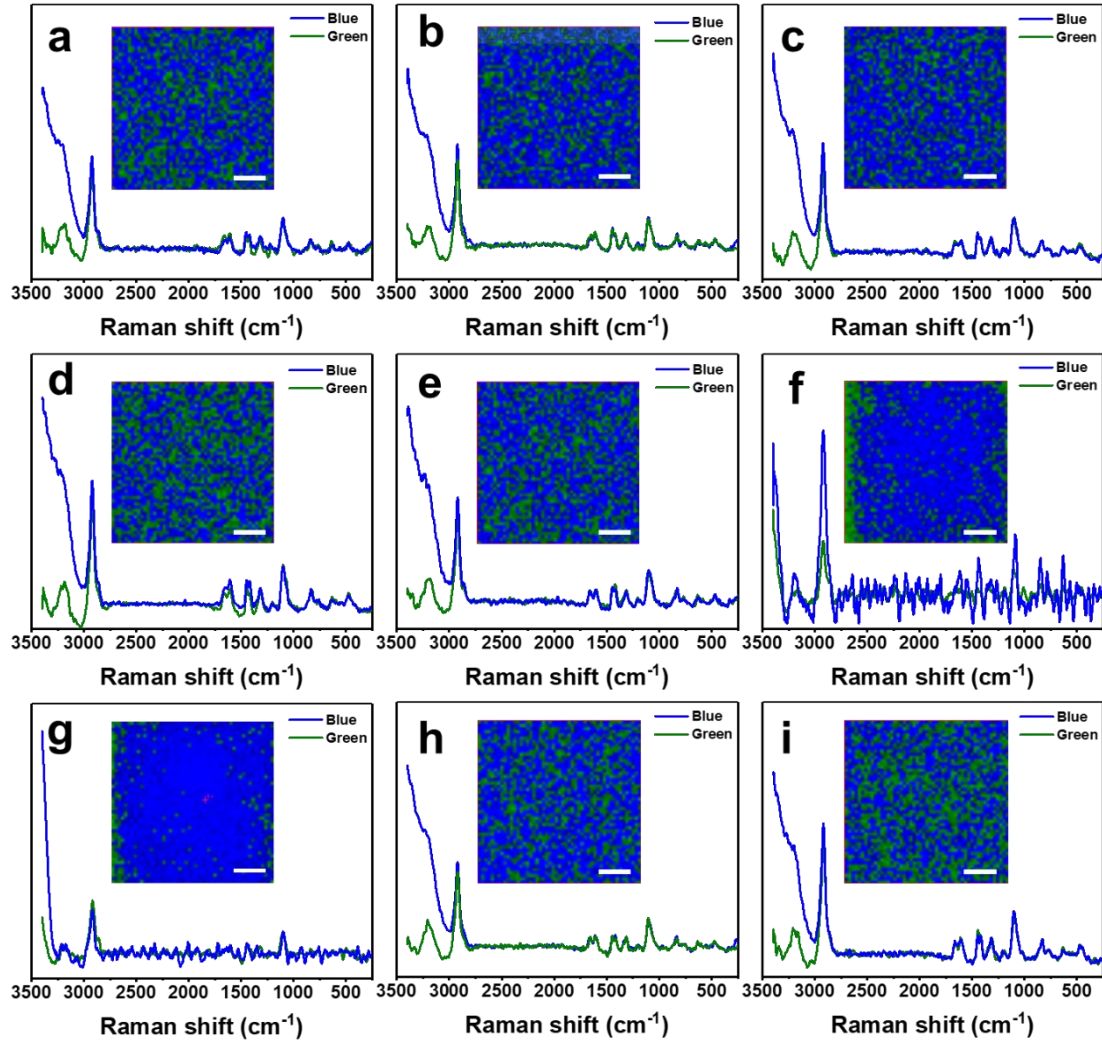


Fig. S14 Raman spectra of individual hydrophilic domains (blue) and hydrophobic domains (green) within (a) C₅T₄A₂₀, (b) C₇T₄A₂₀, (c) C₉T₄A₂₀, (d) C₁₃T₄A₂₀, (e) C₁₁T₂A₂₀, (f) C₁₁T₆A₂₀, (g) C₁₁T₈A₂₀, (h) C₁₁T₄A₁₅ and (i) C₁₁T₄A₂₅ hydrogels. The insets are the reconstructed MCR Raman mappings of hydrophilic domains (blue) and hydrophobic domains (green) obtained from -OH and -NH stretching mode intensities (3000-3400 cm⁻¹). All bars are 20 μm

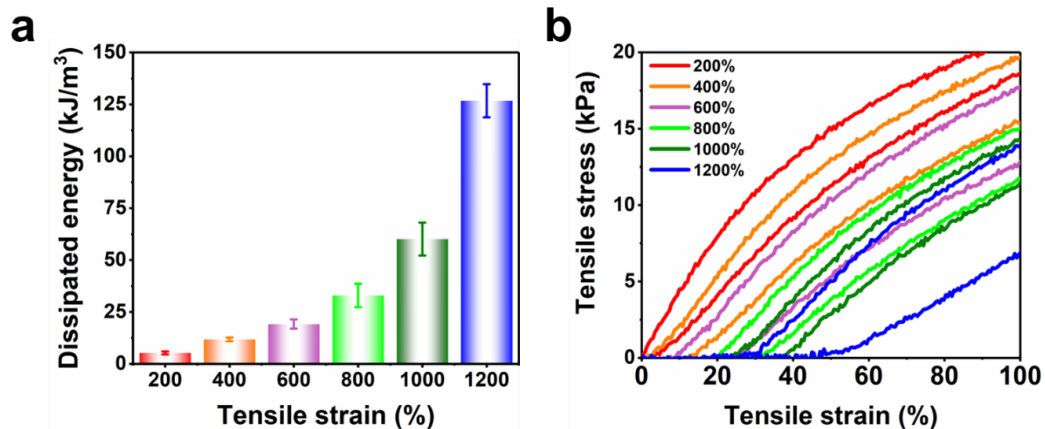


Fig. S15 (a) Dissipated energy of C₁₁T₄A₂₀ sample under various tensile strains. (b) Magnifying stress-strain curves between 0–100% for checking residual strain after stretching

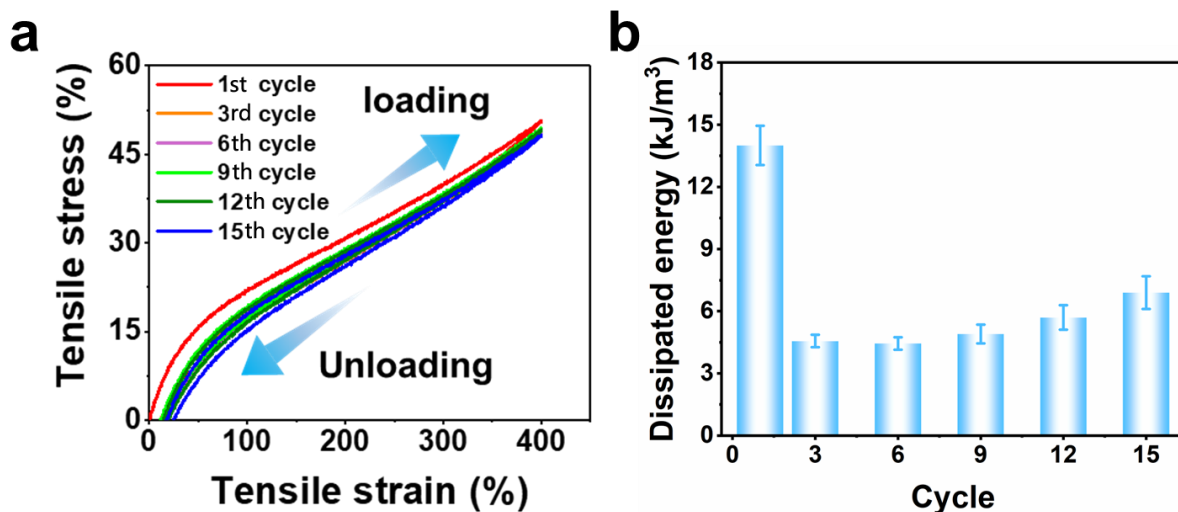


Fig. S16 (a) Successive loading–unloading curves of $C_{11}T_4A_{20}$ under 400% strain for 15 cycles. (b) Dissipated energy of $C_{11}T_4A_{20}$ sample under tensile loading-unloading cycles

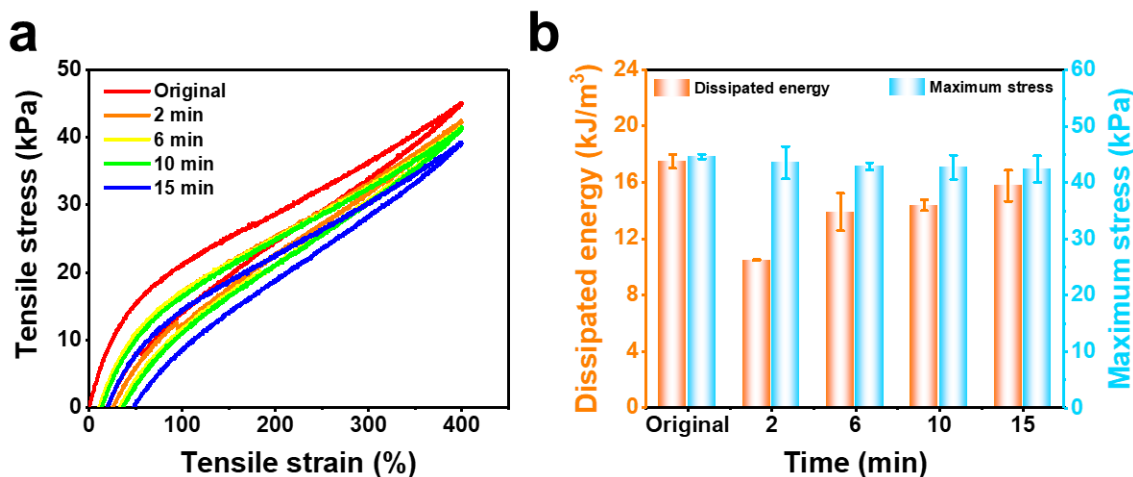


Fig. S17 (a) Tensile recovery test of $C_{11}T_4A_{20}$ sample with different recovery times. (b) Tensile dissipated energy and maximum stress during the tensile recovery test with different recovery times

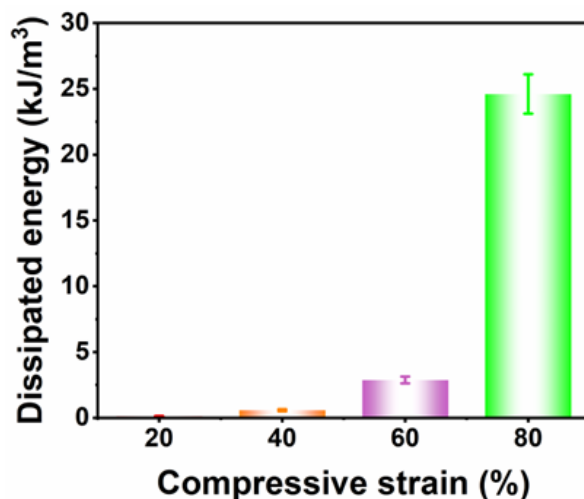


Fig. S18 Dissipated energy of $C_{11}T_4A_{15}$ sample under various compressive strains

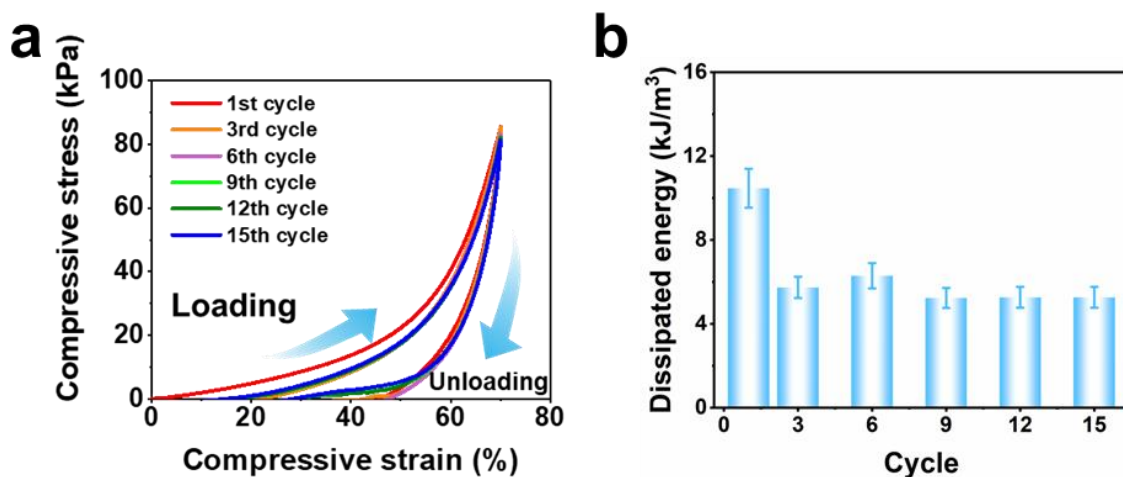


Fig. S19 (a) Successive loading–unloading curves of $C_{11}T_4A_{15}$ under 70% strain for 15 cycles. (b) Dissipated energy of $C_{11}T_4A_{15}$ sample under press loading-unloading cycles

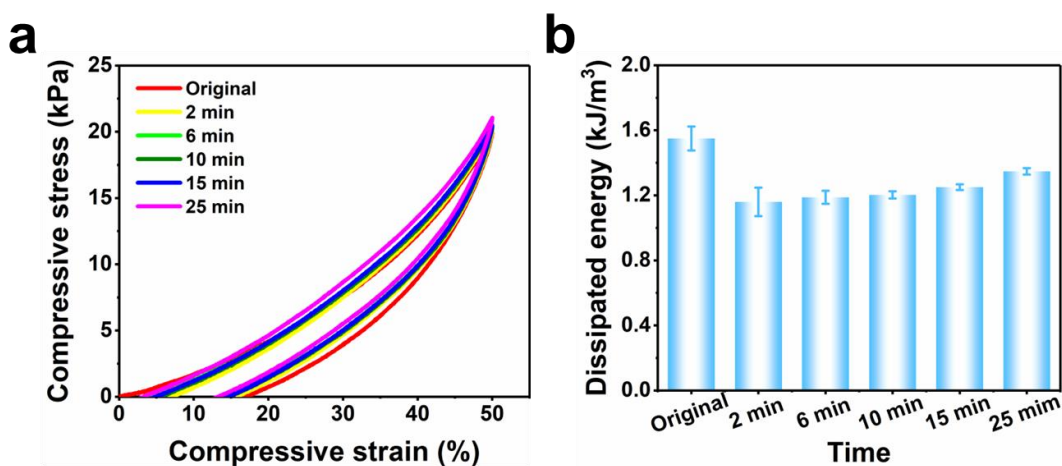


Fig. S20 (a) Press recovery test of $C_{11}T_4A_{15}$ sample with different recovery times. (b) Press dissipated energy during the press recovery test with different recovery times

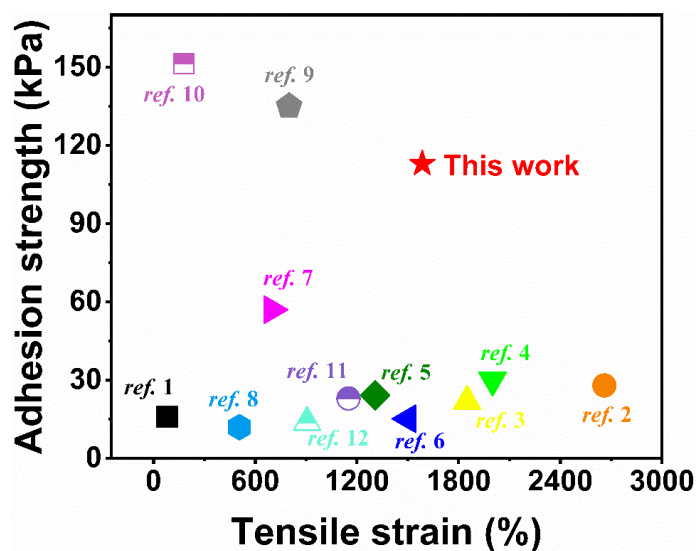


Fig. S21 Comparison of adhesion strength and tensile strain of reported conductive hydrogels [S1-S12]

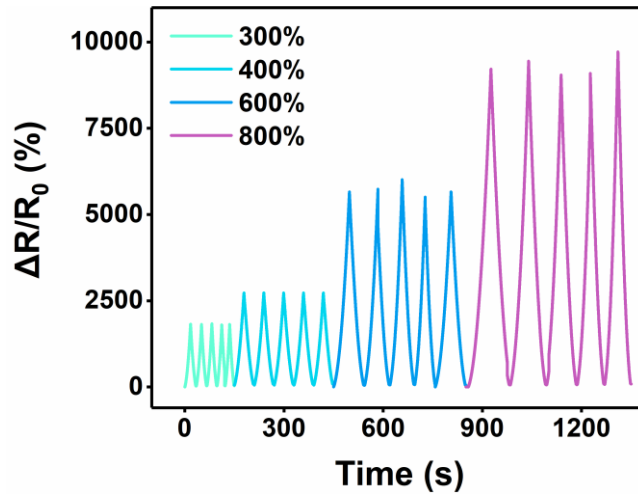


Fig. S22 Cyclic sensing behaviors of the $C_{11}T_4A_{20}$ hydrogel-based sensor upon stretching towards high strains ranging from 300% to 800%

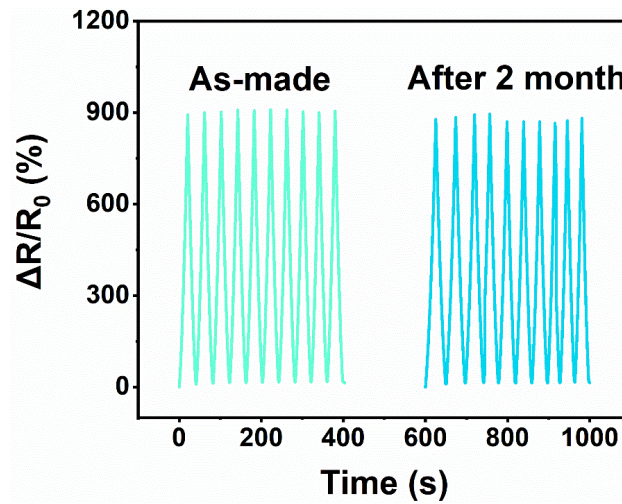


Fig. S23 Dynamic response of the encapsulated $C_{11}T_4A_{20}$ hydrogel-based sensor being placed in environment with 5 °C, 48% humidity for 2 months to cyclic loadings

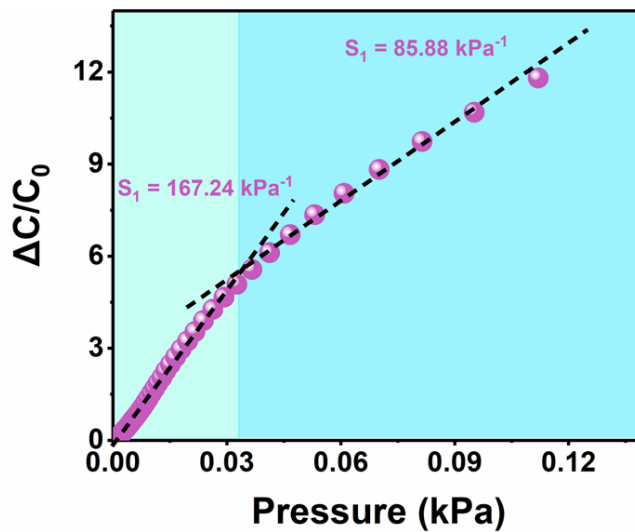


Fig. S24 Relative capacitance changes versus applied pressure within pressure range of 0–0.13 kPa of $C_{11}T_4A_{15}$

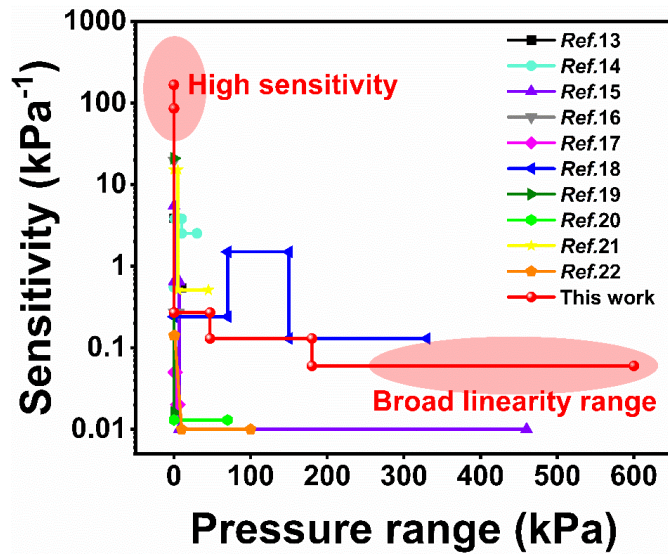


Fig. S25 Comparison of sensitivity and pressure range with literatures [S13-S22]

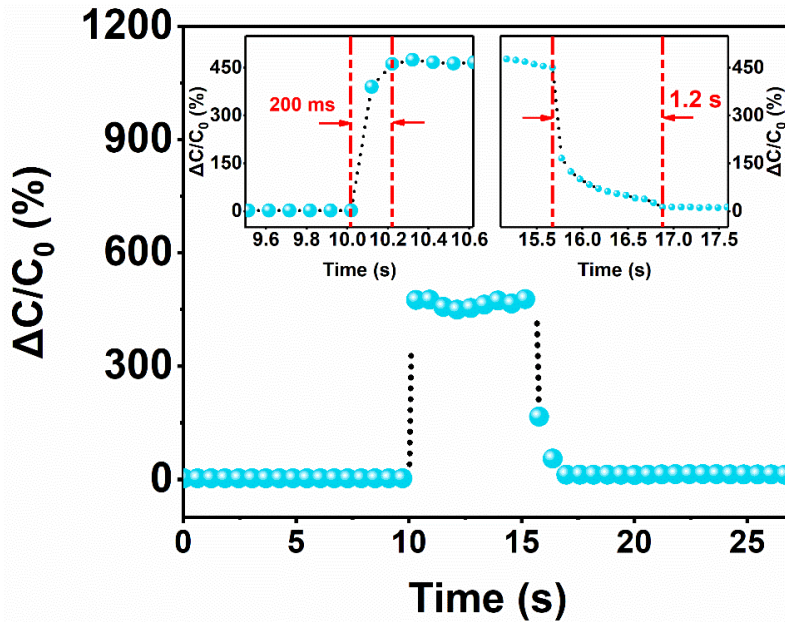


Fig. S26 The response time of the C₁₁T₄A₁₅ hydrogel-based sensor to the applied pressure

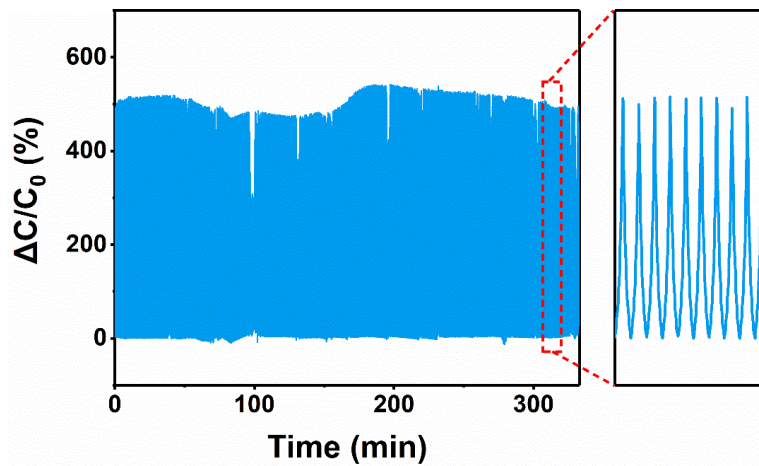


Fig. S27 Cyclic stability test of the C₁₁T₄A₁₅ hydrogel-based sensor under 50% compressive strain for 2000 cycles

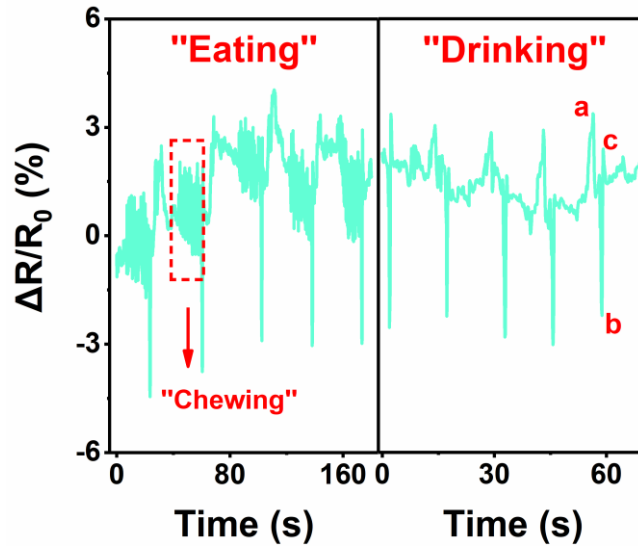


Fig. S28 Responsive signals of $C_{11}T_4A_{20}$ hydrogel-based sensor in monitoring tiny muscle movement of eating, chewing and drinking

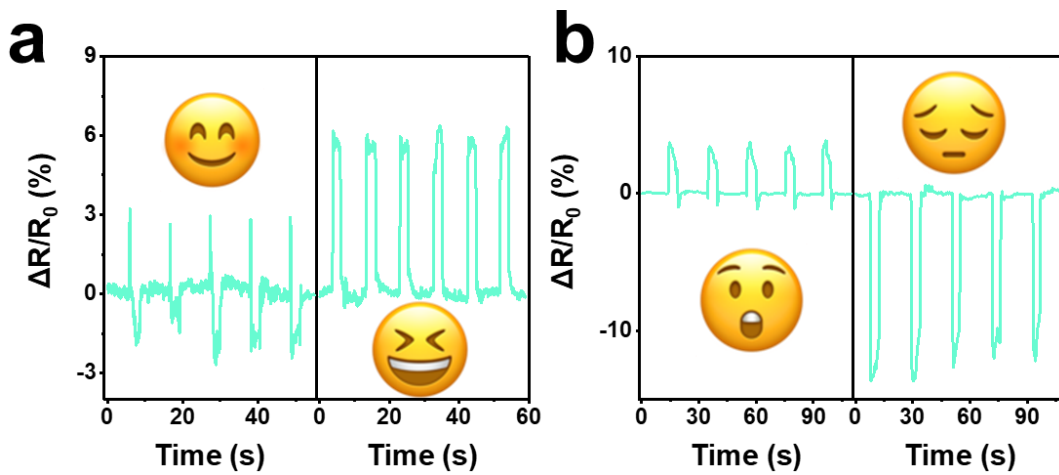


Fig. S29 Responsive signals of $C_{11}T_4A_{20}$ hydrogel-based sensor in monitoring tiny muscle movement of (a) smile, laughing and (b) raise, frown

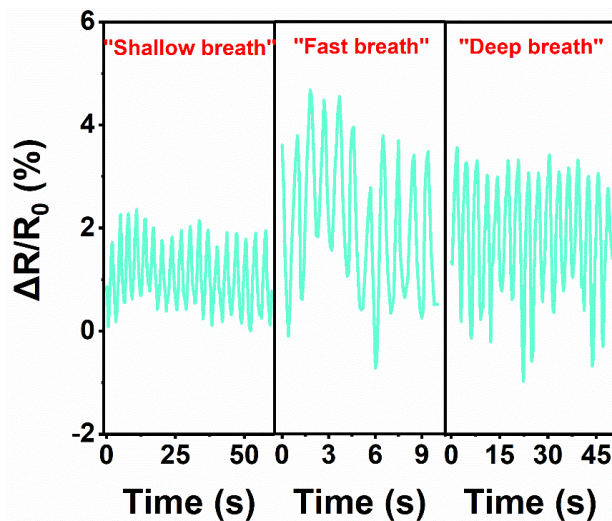


Fig. S30 Responsive signal of three different respiration modes: shallow breath, fast breath and deep breath

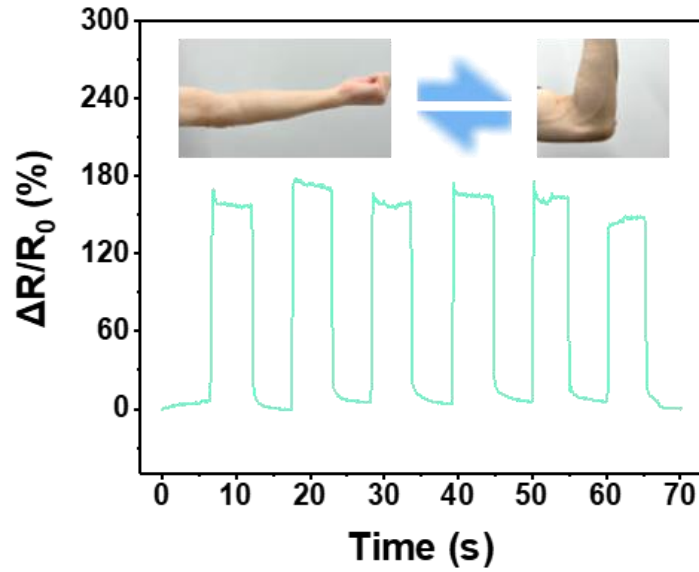


Fig. S31 Relative resistance changes with bending of the elbow

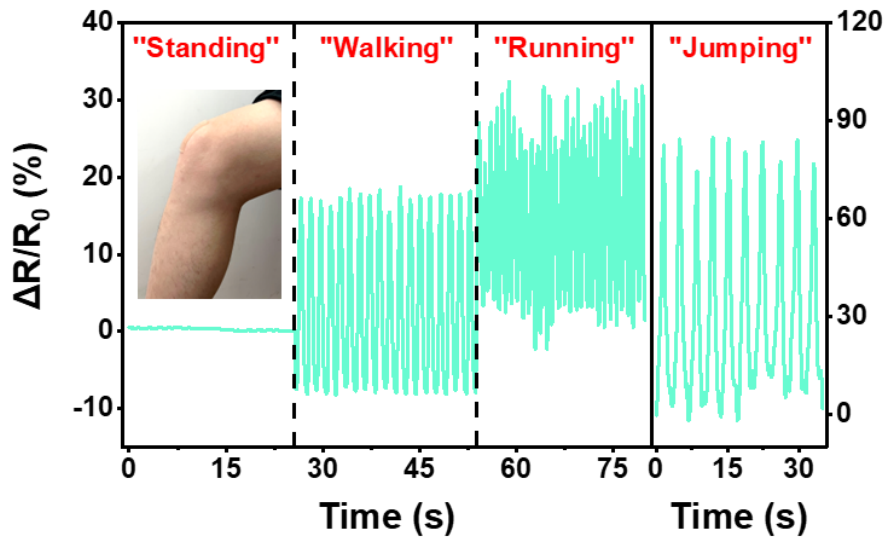


Fig. S32 Photograph of sensor attached on a knee joint, and corresponding signals of standing, walking, running, and jumping

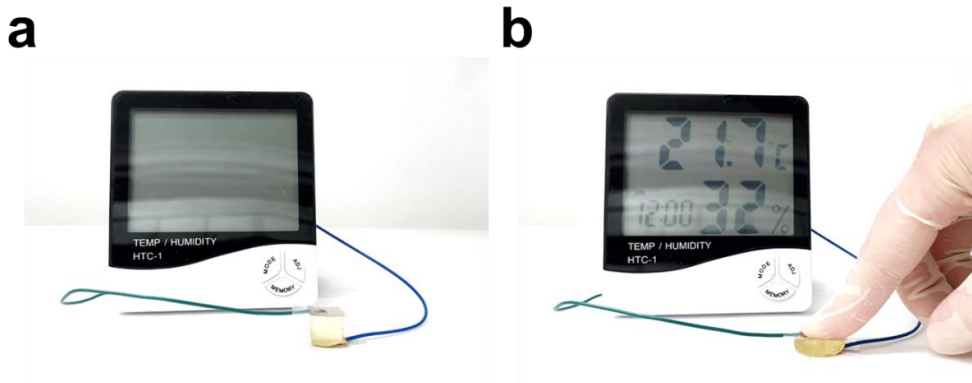


Fig. S33 (a) Photograph of a flexible tactile switch and a digital thermometer/hygrometer. (b) Photograph of a volunteer turning on the digital thermometer/hygrometer by pressing the flexible tactile switch

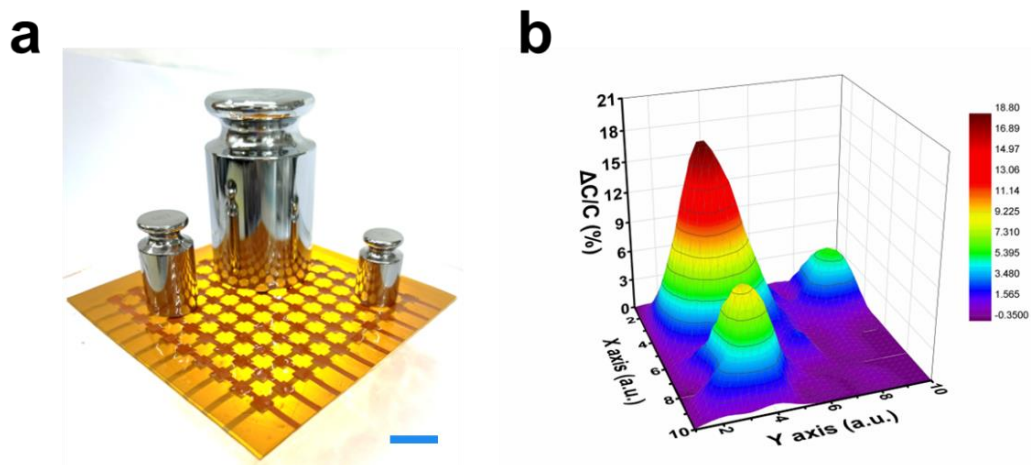


Fig. S34 (a) Photograph showing three weights distributed on the surface of an integrated 10×10 pressure sensor array (scale bar: 3 cm). (b) Corresponding signal map showing the precise pressure distribution in (a)

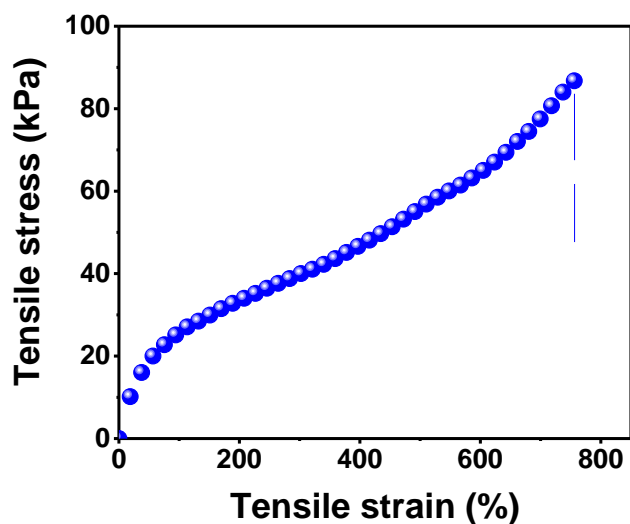


Fig. S35 The stress-strain curve of $C_{11}T_4A_{20}$ -VHB based single-electrode CTA-TENG

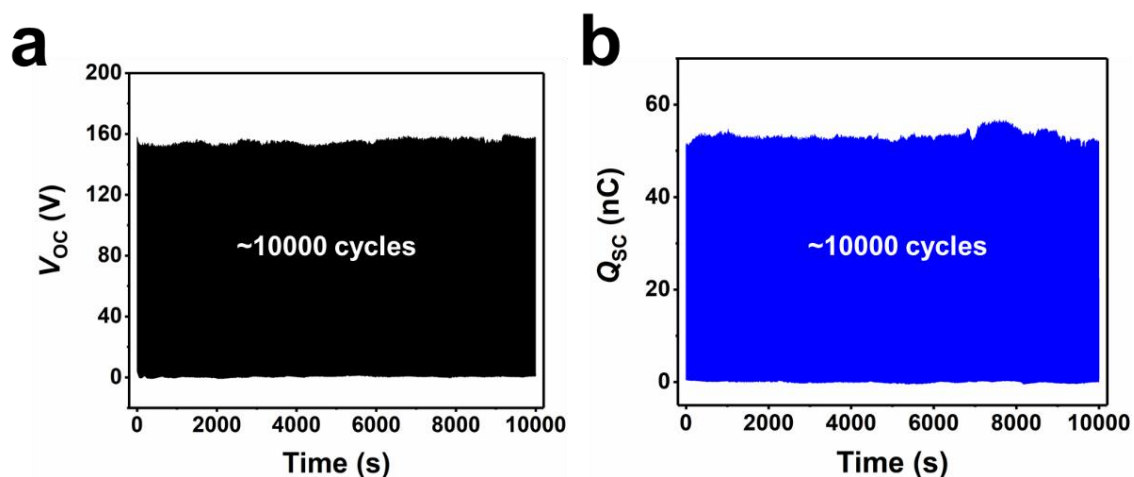


Fig. S36 V_{oc} and Q_{sc} of CTA-TENG that lasted for ~ 10000 cycles of contact-separation motions

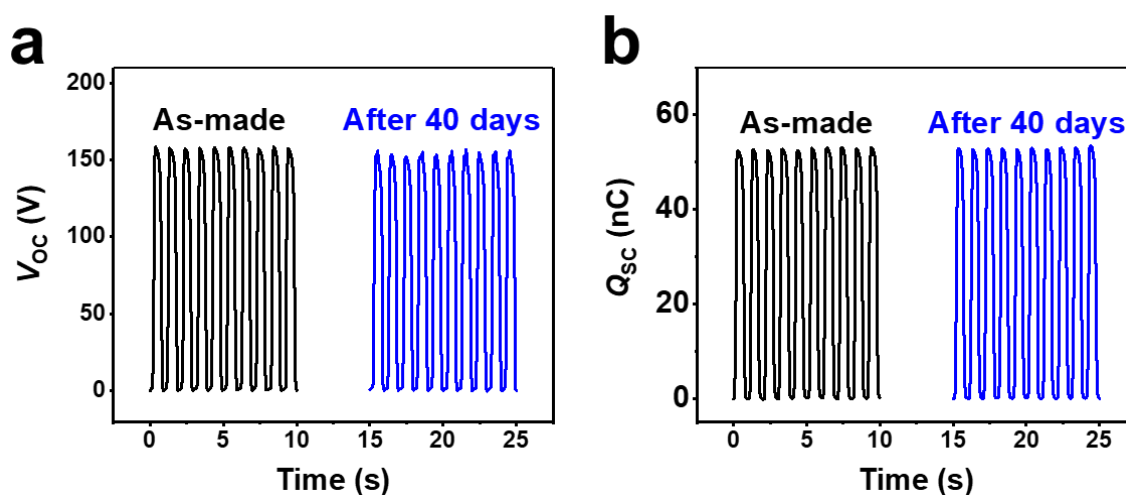


Fig. S37 Comparison of V_{OC} and Q_{SC} of CTA-TENG before and after storage in ambient environment for at least 40 days

S4 Supplementary Tables

Table S1 Effect of molar ratio on the properties of CECT

Sample	Feed ratio ^{a)}	DS_{COONa} ^{b)}	Water-solubility
CECT-5	5:1	0.51	soluble
CECT-7	7:1	0.58	soluble
CECT-9	9:1	0.69	soluble
CECT-11	11:1	0.75	soluble
CECT-13	13:1	0.86	soluble

a) Molar ratio of acrylamide to glucose units; b) Calculated from 1H NMR

Table S2 Preparation and physical properties of CTA hydrogels

Sample	DS _{COONa} ^{a)}	c _{CECT} (wt%)	c _{AAm} (wt%)	Water content (wt%)	Tension				Compression			
					σ_b (kPa)	ϵ_b (%)	E (kPa)	W _f (kJ m ⁻³)	σ_b (MPa)	ϵ_b (%)	E (kPa)	W _f (kJ m ⁻³)
C ₀ T ₀ A ₂₀	0	0	20	74.8	57.65	735.43	30.93	227.72	0.68	90	16.75	44.98
C ₅ T ₄ A ₂₀	0.51	4	20	67.4	213.93	1157.73	60.69	1031.74	2.93	90	43.59	156.54
C ₇ T ₄ A ₂₀	0.58	4	20	68.7	213.78	1291.52	47.59	873.73	2.87	90	30.19	175.62
C ₉ T ₄ A ₂₀	0.69	4	20	70.3	186.41	1252.12	45.15	739.70	2.76	90	28.15	157.85
C ₁₁ T ₄ A ₂₀	0.75	4	20	69.7	192.67	1585.77	43.26	1299.71	1.31	90	19.35	77.43
C ₁₃ T ₄ A ₂₀	0.86	4	20	69.6	191.15	1307.11	53.75	1030.17	2.89	90	33.09	173.32
C ₁₁ T ₂ A ₂₀	0.75	2	20	73.58	121.26	1109.54	39.16	498.93	0.97	90	13.45	59.48
C ₁₁ T ₆ A ₂₀	0.75	6	20	69.60	226.84	1090.83	54.67	838.47	2.38	90	49.82	139.54
C ₁₁ T ₈ A ₂₀	0.75	8	20	66.50	140.74	835.57	66.62	505.57	2.96	90	52.92	153.45
C ₁₁ T ₄ A ₁₅	0.75	4	15	76.60	195.32	1341.41	24.98	924.08	0.38	90	9.76	24.65
C ₁₁ T ₄ A ₂₅	0.75	4	25	66.10	184.71	852.57	90.28	671.33	3.32	90	57.77	248.94

a) Calculated from ¹H NMR

σ_b is the stress. ϵ_b is the strain. E is the Young's modulus. W_f is the fracture energy

Table S3 Comparison of conductivity of CTA hydrogel and the reported conductive composites

Materials	Conductive medium	Conductivity (S m ⁻¹)	Refs.
A-MXene/D-PDMS	A-MXene	5.27×10 ⁻²	[S23]
TPU/CNCTT	CNT	1×10 ⁻²	[S24]
HSAH/PHEAA	DESs	0.19	[S25]
ELO/PANI	PANI	8.64×10 ⁻⁴	[S26]
AHS	SBMA/HEMA	0.39	[S12]
PEDOT:PSS-PAAm	PEDOT:PSS-PAAm	1.08×10 ⁻²	[S27]
NAGA-co-AAm/LiCl	Li ⁺ , Cl ⁻	0.69	[S28]
XSBR/SSCNT	CNT	7.08×10 ⁻²	[S29]
ICE	[Emim][OAc]	0.77×10 ⁻²	[S30]
PTCM-Gly5	MXene	0.19	[S31]
Ionogel-HPC	[Bmin]Cl	10.2×10 ⁻³	[S32]
MMCOH	CNT	1×10 ⁻³	[S33]
MASTA-PANI ₅	PANI	7.8×10 ⁻⁴	[S34]
Ionogel-4050	[EMIM][TFSI]	0.29	[S35]
CPH	BzMe ₃ NOH	0.46	[S36]
CTAs	Na⁺, K⁺, OH⁻	0.62	This work

Table S4 A table lists the letters corresponding to International Morse code

Code	Letter	Code	Letter	Code	Letter	Code	Letter
•—	A	••••	H	— — —	O	•••—	V
—•••	B	••	I	•— —•	P	•— —	W
—• —•	C	• — — —	J	— — • —	Q	—•• —	X
—••	D	—• —	K	• —•	R	—• — —	Y
•	E	• —••	L	•••	S	— — —•	Z
•• —•	F	— —	M	—	T		
— —•	G	—•	N	•• —	U		

Table S5 Comparison of the electrical output of the TENG device with previous literature

Literature	Friction materials	Fre (Hz)	P	Refs.
Yao et al.	CNFs-FEP	10	140 mW/m ²	[S37]
Gao et al.	PEDOT:PSS-PLA	5	211 mW/m ²	[S38]
Yeh et al.	Ecoflex-Liquid metal	/	19.04 mW/m ²	[S39]
Kim et al.	PTFE-Al/Tin	/	147 mW/m ²	[S40]
Zheng et al.	PLGA-PCL	1	32.6 mW/m ²	[S41]
Wang et al.	Chitosan-Ecoflex	0.5	17.5 μW/m ²	[S42]
Pang et al.	Alginate-Al	1	9.5 μW	[S43]
Pan et al.	PLA-Gelatin	5	5 W/m ²	[S44]
Liang et al.	SA-PVA	1	3.8 mW/m ²	[S45]
S. Parandeh et al.	Paper-PCL/GO	3	72.5 mW/m ²	[S46]
Sriphan et al.	Ti ₂ NbO ₇ NSs-BC	/	28 μW	[S47]
Lu et al.	Sugar-Nickel	4	4.21 W/m ²	[S48]
Wu et al.	PBS-SR	5	1.25 W/m ²	[S50]
Chi et al.	Rice paper-PVC	5	376 mW/m ²	[S51]
Zhang et al.	Chitin-VHB	3	1.25 W/m ²	[S52]
Yang et al.	Weighing paper-PTFE	4	13 mW	[S53]
/	Latex-VHB	1	1.17 W/m²	This work

Supplementary References

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