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Supplemental information

Protective effects of S100A8 on sepsis mortality:

Links to sepsis risk in obesity and diabetes

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Figure S1. S100A8 administration upregulated inflammatory cytokine expression in peritoneal macrophages. Related to Figure 1.

Wild-type mice at 10 weeks of age were used for in vitro and in vivo experiments.

(A) Experimental protocol for an *in vitro* study. Peritoneal macrophages from mice were incubated with PBS, S100A8 (0.3 μ g/mL), or S100A8/A9 (0.65 μ g/mL) and analyzed by quantitative PCR at 2 hours. (B) mRNA expression of the indicated inflammatory cytokine genes in peritoneal macrophages (n = 3). (C) Experimental protocol for an *in vivo* study. Mice were injected with PBS, LPS (12.5 μ g/gBW), or S100A8 (0.1 μ g/gBW), and then peritoneal cells were analyzed by quantitative PCR at 4 hours after injection.

(D) mRNA expression of the indicated inflammatory cytokine and S100 genes in peritoneal cells (n = 3). Data are represented as the mean \pm SEM. *p < 0.05, **p < 0.01.



Figure S2. S100A8 administration did not induce inflammatory response in hepatoma cells. Related to Figure 1.

(A) mRNA expression of the indicated cytokine genes in Hepa1-6 mouse hepatoma cells incubated with PBS, S100A8 (0.3 μ g/mL), or S100A8/A9 (0.65 μ g/mL) for 4 hours (n = 3).

(B and C) Wild-type mice were intraperitoneally injected with PBS, LPS (12.5 μ g/gBW), or S100A8 (0.1 μ g/gBW).

(B) mRNA expression of the indicated inflammatory cytokine genes in the mouse liver at 4 hours after injection (n = 3).

(C) AST, ALT, total cholesterol and triglyceride levels in serum samples collected from mice at 4 hours after injection (n = 3).



Figure S3. S100A8 administration did not induce sepsis. Related to Figure 1.

Wild-type mice at 10 weeks of age were used for experiments.

(A) Survival rates at 12, 24, 36, or 48 hours after PBS, LPS, or S100A8 injection (n = 3).

(B) Survival rates at 72 hours after PBS, LPS, or S100A8 injection (n = 3).

(C and D) Body temperature (C) and blood glucose levels (D) at 12, 24, 36, or 48 hours after PBS, LPS, or S100A8 injection (n = 3).

(E) Serum S100A8 levels at 1, 4, 12, or 24 hours after LPS (12.5 μ g/gBW) injection. All mice died within 48 hours of LPS injection (n = 3).

Data are represented as the mean \pm SEM. **p < 0.01; ##p < 0.01 vs. S100A8.



Figure S4. Administration of S100A8 to peritoneal macrophages *in vitro* after LPS stimulation, fails to suppress LPS-induced upregulation of inflammatory cytokine expressions in peritoneal macrophages. Related to Figure 1.

Wild-type mice at 10 weeks of age were used for *in vitro* experiments. Peritoneal macrophages from mice were pretreated with LPS (2 μ g/mL) for 30 minutes before PBS or S100A8 (0.3 μ g/mL) supplementation for 2 hours and analyzed by quantitative PCR.

(A) mRNA expression of the indicated inflammatory cytokine genes in peritoneal macrophages (n = 5). (B) mRNA expression of the indicated TLR4 signaling pathway-related genes in peritoneal macrophages (n = 5).



Figure S5. S100A8 promotes the expression of inflammatory cytokines in the liver by activating the TLR4-MyD88 pathway during LPS-induced sepsis. Related to Figure 1.

WT mice were intraperitoneally injected with either PBS or S100A8 (0.1 μ g/gBW) at 1 hour after LPS (12.5 μ g/gBW) stimulation.

(A) mRNA expression of the indicated inflammatory cytokine genes in the mouse liver at 4 hours after the first injection analyzed by quantitative PCR (n = 3).

(B) mRNA expression of the indicated TLR4 signaling pathway-related genes in the mouse liver at 4 hours after the first injection analyzed by quantitative PCR (n = 3).

(C and D) AST, ALT, total cholesterol and triglyceride levels in serum samples collected from mice at 4 hours (C) and 24 hours (D) after injection (N = 2 at 4 hours and n = 3-4 at 24 hours).



Figure S6. Intraperitoneal injection of S100A8 attenuated the upregulation of inflammatory cytokines and *Myd88* gene expression induced by CRX-527. Related to Figure 1.

C57BL/6J mice were intraperitoneally injected with either PBS or S100A8 (0.1 μ g/gBW) at 30 minutes before CRX-527 (0.5 μ g/gBW) stimulation.

(A) mRNA expression of the indicated inflammatory cytokine genes in the mouse peritoneal cells at 2 hours after the first injection analyzed by quantitative PCR (n = 8-10).

(B) mRNA expression of the indicated TLR4 signaling pathway-related genes in the mouse peritoneal cells at 2 hours after the first injection analyzed by quantitative PCR (n = 8-10).



Figure S7. S100A8 improves survival rates in mice of lethal endotoxemia and sepsis models. Related to Figure 1.

Wild-type mice at 10 weeks of age were used for experiments.

(A) The survival rates of mice injected with either PBS or S100A8 (0.1 μ g/gBW) at one hour after LPS (12.5 μ g/gBW) stimulation at 72 hours after the first injection (n = 10).

(B) The survival rates of mice injected with either PBS or S100A8 (0.1 μ g/gBW) at one hour after E. coli DH5 α (2.0*10⁹ CFU per mouse) injection were measured 72 hours later (n = 10).

(C) The survival rates of mice injected with either PBS or S100A8 (0.1 μ g/gBW) twice (at 0 hour and 24 hours) after cecal ligation and puncture (CLP) treatment were measured at 72 hours later (n = 10).



Figure S8. S100A8 administration did not alter the expression of inflammatory cytokines and TLR4 signaling pathway-related genes in peritoneal cells from db/db mice. Related to Figure 3. db/db and db/+ mice at 11 weeks of age were used for *in vivo* experiments. Mice were injected with PBS or S100A8 (0.1 μ g/gBW), and then peritoneal cells were analyzed by quantitative PCR at 4 hours after injection.

(A) mRNA expression of the indicated inflammatory cytokine and S100 genes in peritoneal cells (n = 5-7).

(B) mRNA expression of the indicated TLR4 signaling pathway-related genes in peritoneal cells (n = 5-7). Data are represented as the mean \pm SEM. *p < 0.05, **p < 0.01.



Figure S9. S100A8 improves survival rates in lethal endotoxemia with mice models of diabetes and obesity. Related to Figure 4.

(A) The survival rates of db/db and db/+ mice mice at 8 weeks of age, injected with either PBS or S100A8 (0.1 μ g/gBW) one hour after LPS (12.5 μ g/gBW) stimulation were measured at 72 hours (n = 13).

(B) The survival rates of ob/ob and wild-type (WT) mice at 8 weeks of age, injected with either PBS or S100A8 (0.1 μ g/gBW) one hour after LPS (12.5 μ g/gBW) stimulation at 72 hours after the first injection (n = 10).

(C) The survival rates of C57BL/6J mice fed Western diet (WD) or normal diet (ND) from 8 to 16 weeks of age, injected with either PBS or S100A8 (0.1 μ g/gBW) at one hour after LPS (12.5 μ g/gBW) stimulation were measured at 72 hours (n = 10).



Figure S10. Generation of myelomonocytic cell-specific S100A8 knockout mice. Related to Figure 5.

Schemes show the construction of generalized myelomonocytic cell-specific S100A8 knockout mice. See the Methods for details.

2kb



Figure S11. LPS administration did not cause liver injury in myelomonocytic cell-specific S100A8 knockout mice. Related to Figure 5.

Myelomonocytic cell-specific S100A8 knockout (Lyz2^{cre}:S100A8^{floxed/floxed}) mice or S100A8-floxed (S100A8^{floxed/floxed}) mice at 10 weeks of age were used for experiments.

AST, ALT, total cholesterol and triglyceride levels in serum samples collected at 4 hours after PBS or LPS (12.5 μ g/gBW) injection (n = 3).



Figure S12. S100A8 improves survival rates of endotoxemia in of myelomonocytic cell-specific S100A8 knockout mice. Related to Figure 5.

Myelomonocytic cell-specific S100A8 knockout (Lyz2^{cre}:S100A8^{floxed/floxed}) mice or S100A8^{floxed/floxed} mice at 10 weeks of age were injected with either PBS or S100A8 (0.1 μ g/gBW) one hour after LPS (12.5 μ g/gBW) stimulation. The survival rates were measured at 72 hours (n = 5-8).

Table S1. qPCR primer sequences, related to STAR Methods.

Gene (species)	Primers	
<i>Actb</i> (mouse)	Forward	GGCTGTATTCCCCTCCATCG
	Reverse	CCAGTTGGTAACAATGCCATGT
<i>Tnfa</i> (mouse)	Forward	CCCTCACACTCAGATCATCTTCT
	Reverse	GCTACGACGTGGGCTACAG
<i>ll-1b</i> (mouse)	Forward	GGGCCTCAAAGGAAAGAATC
	Reverse	TTGCTTGGGATCCACACTCT
<i>ll-6</i> (mouse)	Forward	TAGTCCTTCCTACCCCAATTTCC
	Reverse	TTGGTCCTTAGCCACTCCTTC
<i>Cxcl10</i> (mouse)	Forward	TGCTGGGTCTGAGTGGGACT
	Reverse	CCCTATGGCCCTCATTCTCAC
<i>Mip-1a</i> (mouse)	Forward	CCCAGCCAGGTGTCATTTTCC
	Reverse	GCATTCAGTTCCAGGTCAGTG
<i>Mcp1</i> (mouse)	Forward	AAAAACCTGGATCGGAACCAA
	Reverse	CGGGTCAACTTCACATTCAAAG
S100a8 (mouse)	Forward	AAATCACCATGCCCTCTACAAG
	Reverse	CCCACTTTTATCACCATCGCAA
S100a9 (mouse)	Forward	ATACTCTAGGAAGGAAGGACACC
	Reverse	TCCATGATGTCATTTATGAGGGC
<i>Myd88</i> (mouse)	Forward	CGATTATCTACAGAGCAAGGAATG
	Reverse	ATAGTGATGAACCGCAGGATAC
<i>lrak1</i> (mouse)	Forward	TTCCACTCCCTGTTTCCCTC
	Reverse	AACCACCCTCTCCAATCCTG
<i>lrak4</i> (mouse)	Forward	CATCGTGGCGGTGAAGAAG
	Reverse	AGCATACACTAAGCACAGGTTG
<i>Traf6</i> (mouse)	Forward	TTGGAGAGTCGCCTAGTAAG
	Reverse	GTTACACTGCTGTGCTTCC
<i>Rip1</i> (mouse)	Forward	CCTTCTTGCCCAGGAGAATGA
	Reverse	CTCTGAGGCGATCTGACGAC
<i>Trif</i> (mouse)	Forward	ACCTCCTGCATGCCATGGTTCT
	Reverse	TCAGCCAGCAGGTGGTACAA
<i>Nf-kb</i> (mouse)	Forward	ATTCTGACCTTGCCTATCTAC
	Reverse	TCCAGTCTCCGAGTGAAG