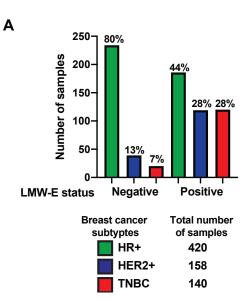
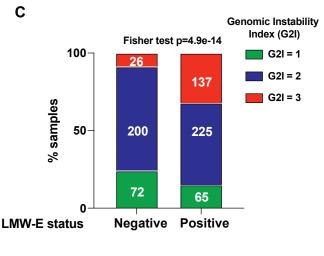
Supplementary Figure 1



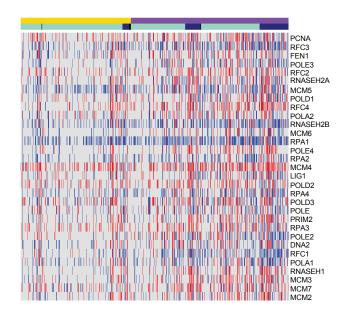


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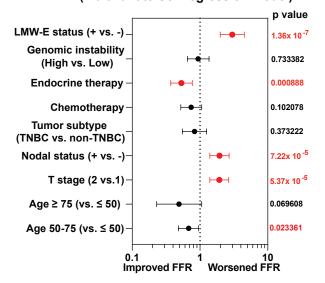
KEGG_NON_HOMOLOGOUS_END_JOINING LMW-E negative LMW-E positive POLL Non-TNBC TNBC XRCC4 DCLRE1C CN gain POLM CN loss DNTT FEN1 PRKDC RAD50 XRCC5 NHEJ1 LIG4

KEGG_DNA_REPLICATION

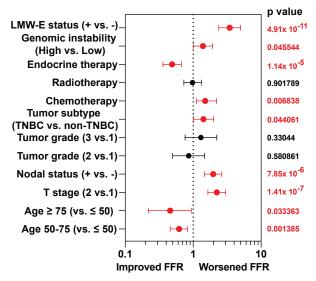


Е

Hazard ratio for FFR (multivariate Cox regression model)

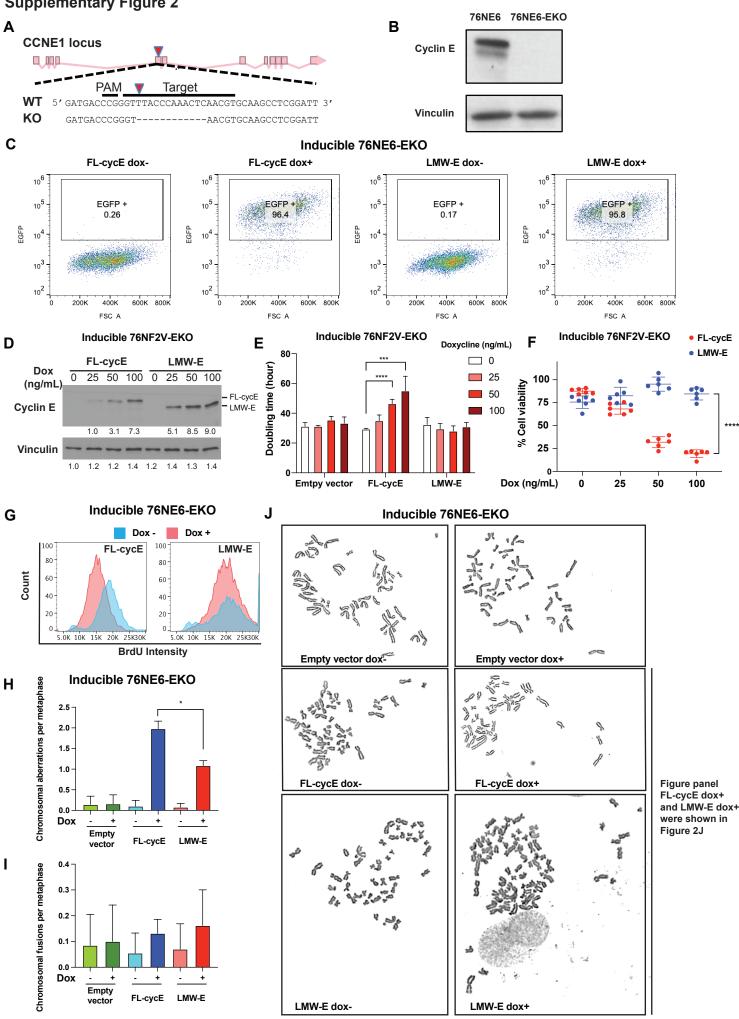


Hazard ratio for FFR (univariate *Cox* regression model)

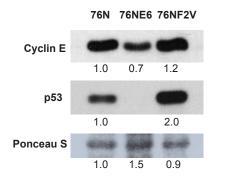


A. The distribution of breast cancer subtypes stratified according to LMW-E negative and LMW-E positive patient cohorts. **B.** Heatmaps showing the copy number (CN) gains (red) and losses (blue) of genes in KEGG non-homologous end jointing pathway (left panel) and KEGG DNA replication pathway (right panel) in LMW-E positive sample group compared with LMW-E negative sample group. TNBC status is also indicated in the horizontal bar graphs on top of the heat maps. C. The distribution of tumor samples with stable genomes (G2I-1; n = 137), intermediate stable genomes (G2I-2; n= 425), and unstable genomes (G2I-3; n= 163) in LMW-E negative and LMW-E positive patient cohorts. **D.** Univariate Cox regression analyses associated with improved or worsened freedom from recurrence (FFR) in the cohort of 725 stage I and II breast cancer patients. Hazard ratio and 95% confidence interval from univariate Cox regression model are shown in the forest plot, and the p-value for each of the analyzed variables are listed on the right and highlighted in red if p < 0.05. E. Multivariate Cox regression analyses for the factors significantly associated with improved or worsened freedom FFR (based on univariate Cox regression in panel D). Hazard ratio and 95% confidence interval from multivariate Cox regression model are shown in the forest plot, and the p-values for each of the variables are listed on the right and highlighted in red if p < 0.05.

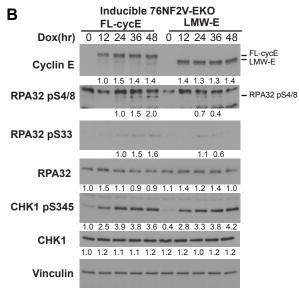




A and B. generation of CCNE1 knock-out hMEC cell line by CRISPR/gRNA. The CRISPR deletion of gRNA target in CCNE1 locus is confirmed by Sanger sequencing (A), and the depletion of endogenous cyclin E protein expression was confirmed by western blot analysis (B). C. FACS analysis for the ratio of EGFP (fused to c-terminus of inducible FL-cycE or LMW-E and expressed when FL-cycE or LMW-E were induced by doxycycline) positive inducible 76NE6-EKO cells with or without 24 hours treatment of 100ng/mL doxycycline. **D.** LMW-E and FL-cycE protein expression were examined by western blot analysis in the inducible 76NF2V- EKO cell lines after 36 hours treatment of doxycycline at the indicated concentrations. E. Cell doubling times were monitored by cell confluency mask from live cell imaging (Incucyte) in inducible 76NF2V-EKO cells after induced expression of LMW-E, FL-cycE or empty vector by doxycycline treatment at the indicated concentrations (n=4, mean with standard deviation). F. Cell viability calculated by MTT assay after induced expression of LMW-E or FL-cycE in inducible 76NF2V-EKO by treatment with doxycycline at the indicated concentrations, normalized by empty vector control (n=4, mean with standard deviation). G. DNA synthesis in inducible 76NE6- EKO cell lines with or without LMW-E or FL-cycE over-expression were assessed by bromodeoxyuridine (BrdU) incorporation and analyzed by FACS. H - I. 76NE6-EKO cells following 36 hours of induced expression of LMW-E or FL-cycE were subjected to metaphase spread assay to assess the extent of chromosomal instability-results show enumerations of total chromosomal aberration frequency (H) and chromosomal fusion frequency (I). Two biological repeats, each with 35 metaphase preparations for each condition. J. Representative metaphase spread images for inducible 76NE6-EKO empty vector, FL-cycE and LMW-E cells. Note that the representative images for FL-cycE dox+ and LMW-E dox+ were also shown in main Figure 2J. ***P<0.001 and ****P<0.0001, Student *t* test.

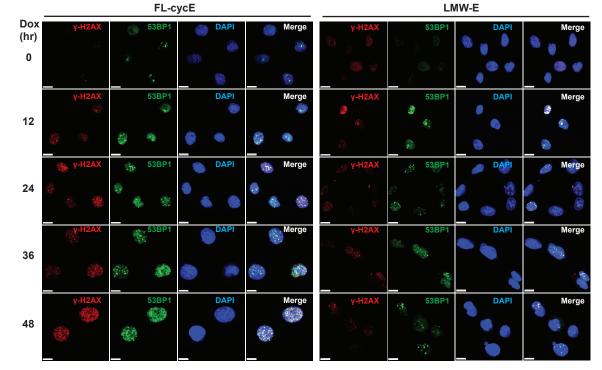


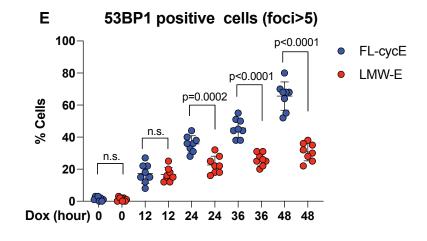
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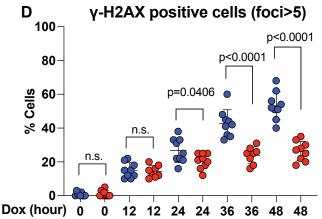


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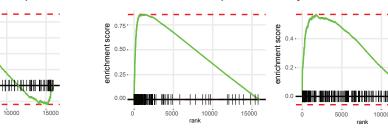






A. Western blot analysis of cyclin E and p53 in 76N (mortal cell strains), 76NE6 and 76NF2V cell lines. **B.** Western blot analysis of the indicated replication stress markers in 76NF2V-EKO cells following induced expression of LMW-E or FL-cycE. Cells were treated with 100ng/ml doxycycline in a time course dependent manner. Uninduced control (Dox 0 hour) were treated with DMSO for 48 hours. **C.** Representative images of immunofluorescent analysis of γ -H2AX and 53BP1 foci in 76NF2V-EKO cells following induced expression of LMW-E or FL-cycE (scale bar=10 µm). **D and E.** Quantitation of time course analysis of γ -H2AX (panel D) and 53BP1(panel E) positive cells (nuclear foci>5) in 76NF2V-EKO cell lines induced for FL-cycE or LMW-E expression (100ng/mL doxycycline for induction, DMSO for uninduced control, cell number>600, Mean with standard deviation, p values shown in graph, n.s.: not significant, Student *t* test).

A KEGG pathways significantly enriched by FL-cycE overexpression В HALLMARK pathways significantly enriched by FL-cycE overexpression HOMOLOGOUS_RECOMBINATION P53_SIGNALING_PATHWAY CEL_CYCLE OCCYTE_MEISIS SPLICETSOME ENDOCYTOSIS PATHWAYS_IN_CANCER DILATED_CARDIOMYOPATHY ACUTE MYELOID_LEUKEMA WMT_SIGNALING_PATHWAY B_CELL_RECEPTOR_SIGNALING_PATHWAY CHEMOKINE_SIGNALING_PATHWAY CHEMOKINE_SIGNALING_PATHWAY CLEUCINE_AND_ISOLULAR CARDIOMYOPATHY COCORECTAL_CANCER VALINE_LEUCINE_AND_ISOLULAR CARDIOMYOPATHY LEUKOCYTE_TRANSENDOTHELIA_MICRATION LEUKOCYTE_TRANSENDOTHELIA_MICRATION COCORECTAL_CANCER VALINE_LEUCINE_AND_ISOLULAR CARDIOMYOPATHY STARCH_AND_SUCROSE_METABOLISM HEDGEHOG_SIGNALING_PATHWAY STARCH_AND_SUCROSE_METABOLISM T_CELL_RECEPTOR_SIGNALING_PATHWAY STARCH_AND_SUCROSE_METABOLISM T_CELL_RECEPTOR_SIGNALING_PATHWAY GLYCOSAMINOGLYCAN_BIOSYNTHESIS_HEPARAN_SULFATE NATURAL_KLIER CELL_MEDIATED_CYTOTOXICITY GLYCOSAMINOGLYCAN_BIOSYNTHESIS_HEPARAN_SULFATE NATURAL_KLIER CELL_MEDIATED_CYTOTOXICITY GLYCOSAMINOGLYCAN_BIOSYNTHESIS_CHODROITIN_SULFATE NATURAL_KLIER CELL_MEDIATED_CYTOTOXICITY GLYCOSAMINOGLYCAN_BIOSYNTHESIS_CHODROITIN_SULFATE NATURAL_KLIER CELL_MEDIATED_CYTOTOXICITY NEUROTROPHIN_SIGNALING_PATHWAY ECM_RECEPTOR_INTERAL_CANCER ADDERENS_JUNCTION NEUROTROPHIN_SIGNALING_PATHWAY ECM_RECEPTOR_INTERACION REGULATION_OF_ACTIN_CYTOSKELETON E2F_TARGETS p=0.0019 TNFA_SIGNALING_VIA_NFKB p=0.0019 G2M_CHECKPOINT p=0.0019 SPERMATOGENESIS p=0.0020 UV_RESPONSE_UP p=0.0019 INFLAMMATORY_RESPONSE p=0.0038 KRAS_SIGNALING_UP p=0.0039 MITOTIC SPINDLE p=0.00<u>3</u>7 IL2_STAT5_SIGNALING =0.0075 ESTROGEN_RESPONSE _EARLY p=0.0106 p=0.0143 UV_RESPONSE_DN ADIPOGENESIS p=0.0086 p=0.0083 COAGULATION GLYCOLYSIS p=0.0043 **HYPOXIA** p=0.0022 EPITHELIAL_MESENCHYMAL p=0.0021 TRANSITION APICAL_JUNCTION p=0.0021 Normalized Enrichment Score 2 (FL-cycE dox+ vs. dox-, p.adj<0.05) Normalized Enrichment Score (FL-cycE dox+ vs. dox-, p.adj<0.05) С D KEGG_CELL_CYCLE HALLMARK_E2F_TARGETS LMW-E dox+ vs. doxp=0.0015 FL-cycE dox+ vs. doxp=0.002 LMW-E dox+ vs. dox- p=0.0014 FL-cycE dox+ vs. dox- p=0.0019 0.8



15000

10000

rank

Ε FL-cycE dox+ vs. dox-

-

rank

10000

15000

enrichment score

0.6

0.4

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Volcano plot

EnhancedVolcano

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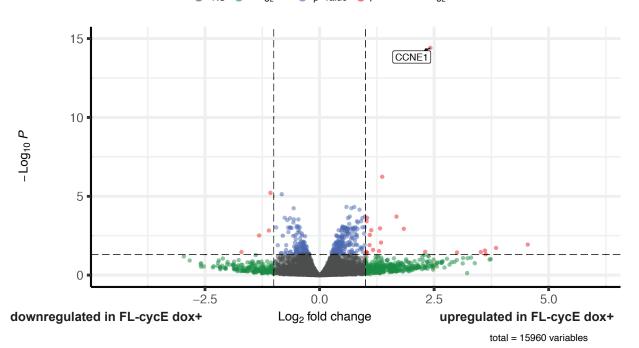
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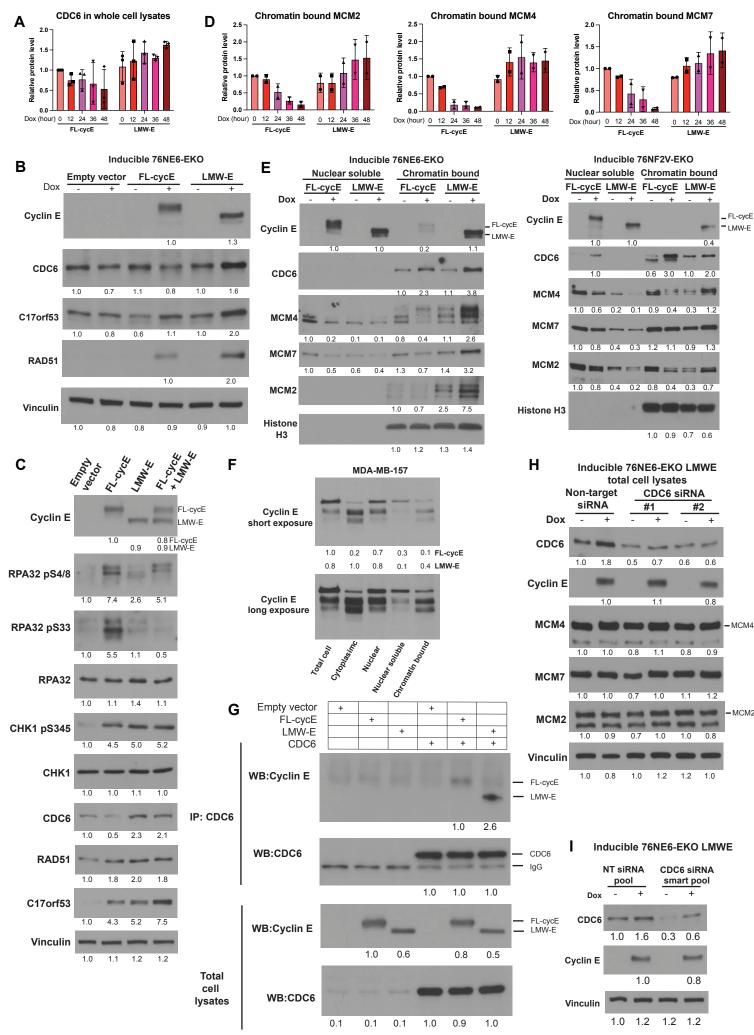
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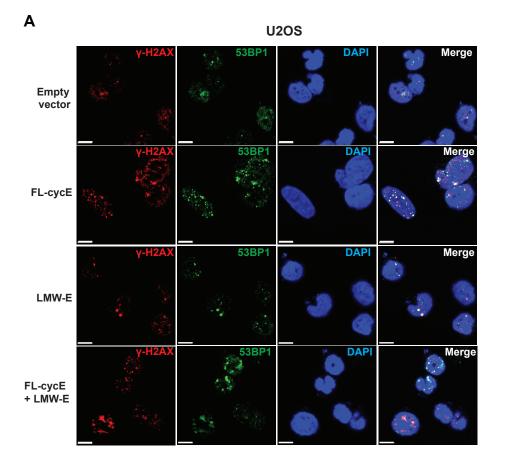


NS Log₂ FC p-value p - value and log₂ FC

A. Waterfall plots of the KEGG pathways significantly enriched in inducible 76NE6-EKO FLcycE cells treated with 100ng/ml doxycycline for 36 hours, followed by RNA-seq and transcriptional profiling analysis. Inducible 76NE6-EKO FL-cycE cells cultured without doxycycline (dox-, DMSO added) were served as reference (adjusted p<0.05). **B.** Waterfall plots of HALLMARK pathways significantly enriched in inducible 76NE6-EKO FL-cycE dox+ group, compared to FL-cycE dox- group (adjusted p <0.05). **C.** Enrichment plots of KEGG cell cycle gene sets in LMW-E dox+ group compared to LMW-E dox- group (left panel) and FL-cycE dox+ group compared to FL-cycE dox- group (right panel). **D** Enrichment plots of HALLMARK E2F targets gene set in LMW-E dox+ group compared to LMW-E dox- group (left panel) and FL-cycE dox+ group compared to FL-cycE dox- group (right panel). **D** Enrichment plots of to compare the expression profiles between FL-cycE dox+ and FL-cycE dox- 76NE6-EKO cell lines. *CCNE1* is the top ranked gene overexpressed in FL-cycE dox+ cells.

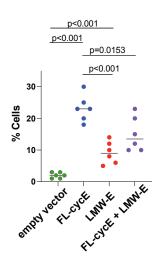


A. Densitometry and quantitation of CDC6 protein levels (see Figure 5B) in whole cell lysates collected from inducible 76NE6-EKO cells with time-course expression (0-48 hours) of FL-cycE or LMW-E (3 biological repeats). B. Western blot analysis of cyclin E, CDC6, C17orf53, and RAD51 in inducible 76NE6-EKO cell lines cultured with or without doxycycline (100 ng/mL, 24 hours) to induce the expression of LMW-E, FL-cycE, or empty vector control. C. Western blot analysis of the effect of LMW-E and FL-cycE on the indicated replication stress markers phospho-CHK1 and phospho-RPA32 in U2OS cells. The levels of CDC6, RAD51 and C17orf53 were also examined by western blot analysis in the same experiment. **D.** Densitometry and quantitation of of MCM2, MCM4 and MCM7 protein levels (see Figure 5C) in chromatin bound fractions collected from inducible 76NE6-EKO cells with time-course expression (0-48 hours) of FL-cycE or LMW-E (2 biological repeats). E. Western blot analysis of cyclin E (FL-cycE or LMW-E) and the indicated DNA pre-replication complex proteins in the inducible 76NE6-EKO cells (left panel) and 76NF2V-EKO cells (right panel) with or without treatment with doxycycline to induce LMW- E or FL-cycE expression. Western blot analysis was performed using protein lysates from cell nuclear soluble fraction and non-soluble (chromatin bound) fraction. F. Western blot analysis of FL-cycE or LMW-E in MDA-MB-157 in total cell lysates as well as the indicated fractionated lysates. G. Immunoprecipitation (IP) followed by western blot analysis to show the binding between cyclin E (FL-cycE or LMW-E) and CDC6 in HEK293T cells transfected with the indicated plasmids. Top panel shows the IP/westerns blot analysis and bottom panels show the input by western blot analysis. H and I. Western blot analysis of CDC6 and indicated proteins in inducible 76NE6- EKO LMW-E cells transfected with two individual siRNAs (H) and siRNA smart pool (I) targeting CDC6 or non-target siRNA controls. These cells were transfected with indicated siRNAs, followed by 24 hours treatment of 100ng/mL doxycycline to induce LMW-E expression.



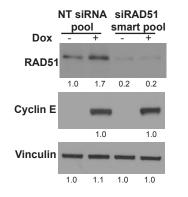
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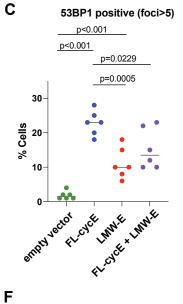
γ-H2AX positive (foci>5)



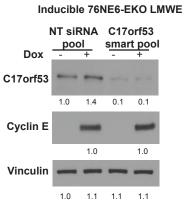
Ε

Inducible 76NE6-EKO LMWE



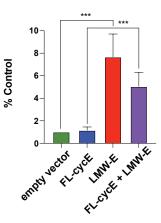


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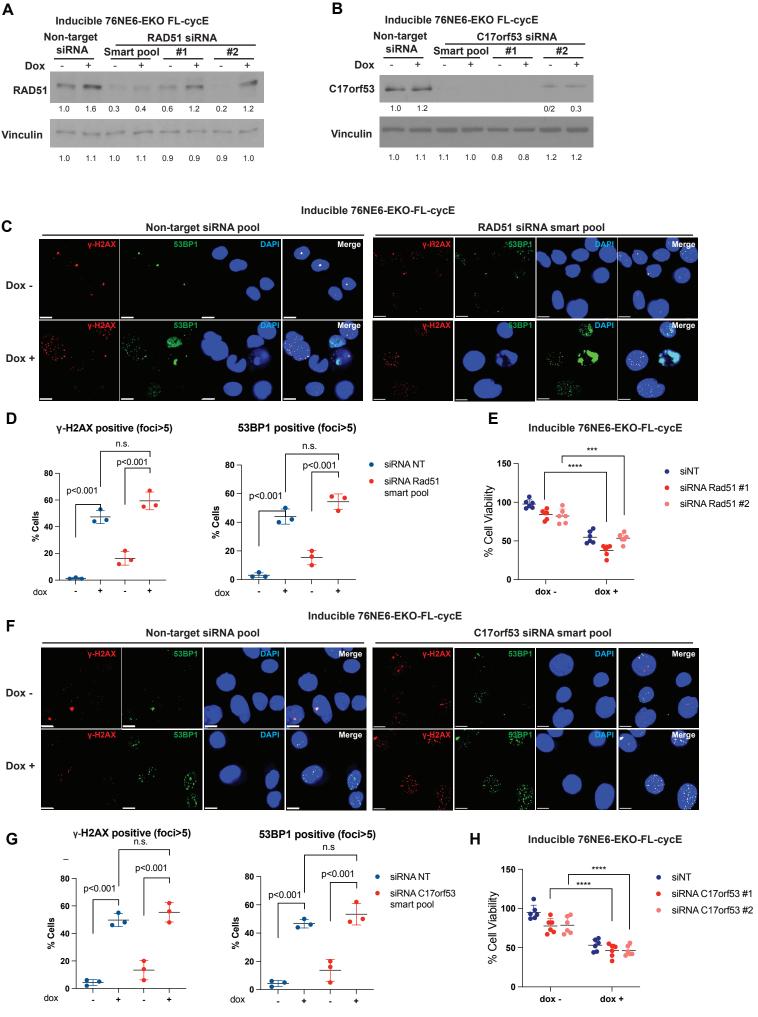




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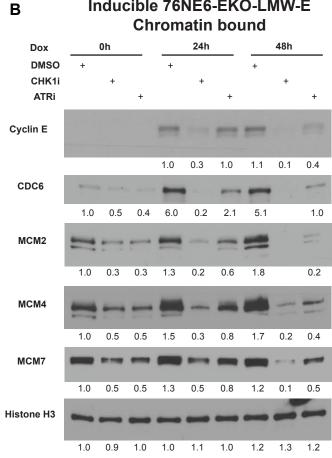


A - C. Assessment of the effect of LMW-E and FL-cycE on DNA damage intensity by IF for γ -H2AX and 53BP1 foci. Representative immunofluorescence images are shown in panel A and quantification for positive cells (foci>5) are illustrated in dot-plots (panel B and C; cell number>300, mean with standard deviation, scale bar=10 µm). **D.** EJ5-GFP DNA repair reporter assay. U2OS cells engineered to express EJ5-GFP DNA repair reporter and pCBASce plasmid (an I- SceI expression vector to induce DNA double strand break), were transfected with empty vector, FL-cycE, LMW-E or FL- cycE + LMW-E followed by FACS analysis to detect and quantitate for GFP positive cells. Values are normalized with the control group. Error bars represent mean standard deviation (n=3 independent experiments, ***P<0.001) **E.** Western blot analysis of RAD51 knock-down efficiency in inducible 76NE6-EKO LMWE cells transfected with or without siRNA smart pool targeting RAD51 in cells cultured in the presence or absence of doxycycline. **F.** Western blot analysis of C17orf53 knock-down efficiency in inducible 76NE6-EKO LMWE cells transfected in the presence or absence of doxycycline.



A. Western blot analysis of RAD51 knock-down efficiency in inducible 76NE6-EKO FL-cycE cells. The cells were treated with 100 ng/mL doxycycline (or DMSO control) for 24 hours after indicated siRNA transfection to induce FL-cycE expression. Non-target (NT) siRNA was used as a control. B. Western blot analysis of C17orf53 knock-down efficiency in inducible 76NE6-EKO-FL-cycE cells. The cells were treated with 100 ng/mL doxycycline (or DMSO control) for 24 hours after siRNA transfection. Non-target (NT) siRNA was used as a control. C. Analysis of DNA damage intensity by immunofluorescence assay for γ -H2AX and 53BP1 foci in inducible 76NE6-EKO-FL-cycE cells (scale bar=10 µm). Cells were treated with siRNA targeting RAD51, followed by 24 hours of treatment with 100 ng/mL doxycycline to induce FL-cycE expression. Non-target siRNA and DMSO were used as controls. **D.** Quantification of γ -H2AX and 53BP1 foci in panel C (cell number > 150, mean with standard deviation). E. Analysis of cell viability by MTT assay in inducible 76NE6-EKO-FL-cycE cells after transfection with specific siRNAs targeting RAD51, followed by treatment with 100 ng/mL doxycycline to induce FL-cycE expression for 48 hours Non-target siRNA and DMSO (dox-) were used as controls. F. Analysis of DNA damage intensity by immunofluorescence assay for γ -H2AX and 53BP1 foci in inducible 76NE6-EKO-FL-cycE cells (scale bar=10 µm). Cells were treated with siRNA targeting C17orf53, followed by 24h hours of treatment with 100 ng/mL doxycycline to induce FL-cycE expression. Non-target siRNA and DMSO were used as controls. G. Quantification of γ -H2AX and 53BP1 foci in panel F (cell number > 150, mean with standard deviation). **H.** Analysis of cell viability by MTT assay in inducible 76NE6-EKO cells after transfection with specific siRNAs targeting C17orf53, followed by 100 ng/mL doxycycline to induce FL-cycE expression for 48 hours. Nontarget siRNA and DMSO (dox-) were used as controls. For all statistical analyses, ***p < 0.001 and ****p < 0.0001, Student *t* test.

| Α | Inducible 76NE6-EKO-L | | | | | | | /W- | E |
|-----------------------|-----------------------|-----|-----|-----|-----|-----|-----|------------|-----|
| Dox | 0h | | | 24h | | | 48h | | |
| DMSO CHK1i ATRi | + | + | + | + | + | + | + | + | + |
| Cyclin E | | | | - | - | - | - | - | = |
| | | | | 1.0 | 0.9 | 0.8 | 1.0 | 0.7 | 0.8 |
| CDC6 | - | | | - | | - | - | - | |
| | 1.0 | 1.6 | 2.7 | 4.1 | 3.6 | 4.8 | 3.4 | 3.3 | 1.1 |
| RAD51 | | - | - | - | | - | - | | - |
| | 1.0 | 2.1 | 2.3 | 2.3 | 2.0 | 2.4 | 2.4 | 0.9 | 0.6 |
| C17orf53 | - | - | - | - | - | | = | - | 1 |
| | 1.0 | 1.7 | 1.6 | 2.0 | 2.1 | 2.1 | 2.0 | 1.7 | 1.6 |
| MCM2 | - | - | - | - | - | - | - | - | - |
| | 1.0 | 1.1 | 0.9 | 0.9 | 0.9 | 0.8 | 0.9 | 0.6 | 0.5 |
| MCM4 | - | | - | - | - | - | - | - | 1 |
| I | 1.0 | 1.3 | 1.1 | 1.1 | 1.0 | 1.1 | 1.1 | 0.6 | 0.6 |
| MCM7 | - | - | - | - | - | - | - | - | - |
| | 1.0 | 1.1 | 1.0 | 1.1 | 1.0 | 1.2 | 1.0 | 0.5 | 0.2 |
| Vinculin | - | - | - | - | - | - | - | - | - |
| | 1.0 | 0.9 | 0.9 | 1.0 | 0.9 | 0.8 | 0.9 | 0.9 | 1.0 |



Inducible 76NE6-EKO-LMW-E

A. Western blot analysis for the levels of LMW-E, CDC6, C17orf53, RAD51, MCM2, MCM4 and MCM7, using whole cell lysates collected from inducible 76NE6-EKO-LMW-E cells with CHK1 inhibitor rabusertib (70nM), or ATR inhibitor ceralasertib (125nM; both at their respective IC50 concentrations), with or without LMW-E expressing for 24 and 48 hours. **B.** Western blot analysis for chromatin bound LMW-E, CDC6, MCM2, MCM4 and MCM7 in inducible 76NE6-EKO-LMW-E cells treated using the same strategy as panel (A).