

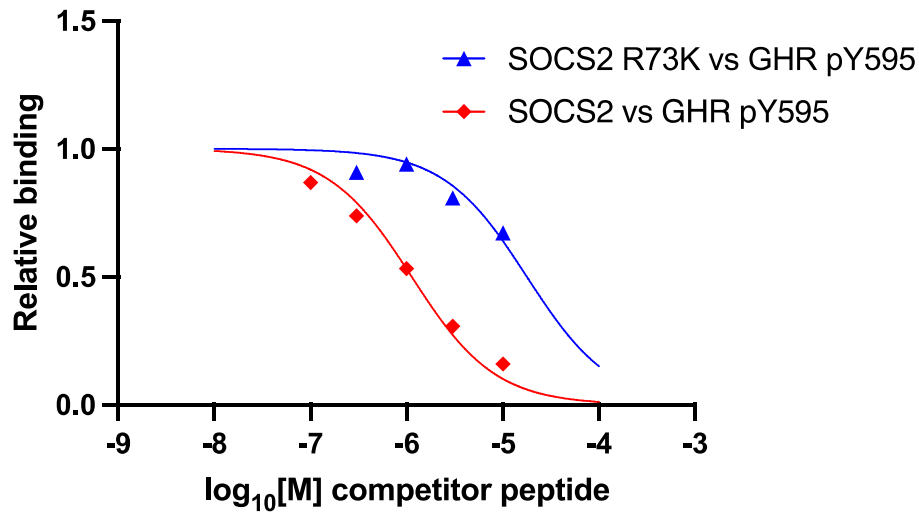
Supplementary - SOCS2 regulation of growth hormone signaling requires a canonical interaction with phosphotyrosine

Kunlun Li^{1,2}, Lizeth G. Meza Guzman^{1,2}, Lachlan Whitehead^{1,2}, Evelyn Leong¹, Andrew Kueh, Warren S. Alexander^{1,2}, Nadia J. Kershaw^{1,2}, Jeffrey J. Babon^{1,2}, Karen Doggett^{1,2}†, Sandra E. Nicholson^{1,2}†

¹The Walter and Eliza Hall Institute of Medical Research, Parkville, Australia;

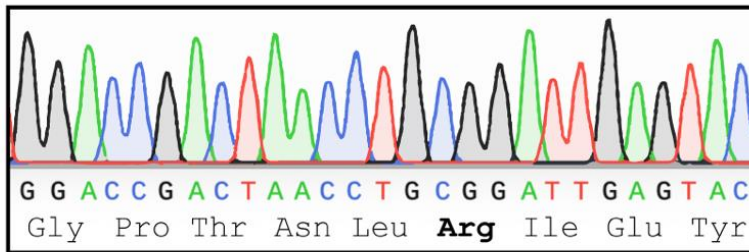
²Department of Medical Biology, University of Melbourne, Parkville, Australia.

† These authors jointly supervised this work and are corresponding authors.

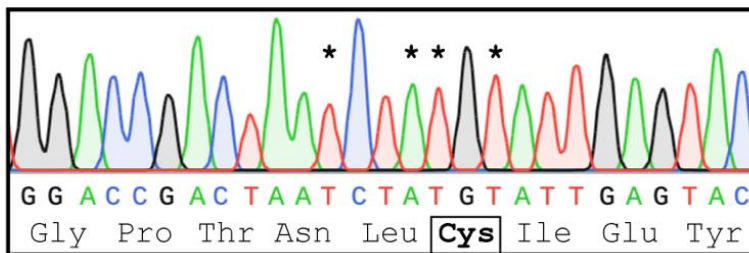


Supplementary Figure 1. Mutation of Arg73 to Lys in the SOCS2-SH2 domain results in reduced binding to phosphopeptide. A competitive surface plasmon resonance (SPR) assay was used to assess the impact of the R73K mutation. SOCS2 bound to a phosphopeptide derived from GHR pY595 with an IC_{50} $1.1 \pm 0.05 \mu\text{M}$. The SOCS2-R73K mutation reduced binding to GHR pY595, IC_{50} $26.7 \pm 1.0 \mu\text{M}$. IC_{50} values are mean \pm S.D., derived from three independent experiments.

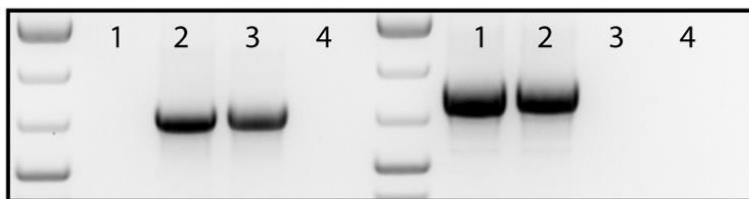
A SOCS2 WT



B SOCS2 R96C

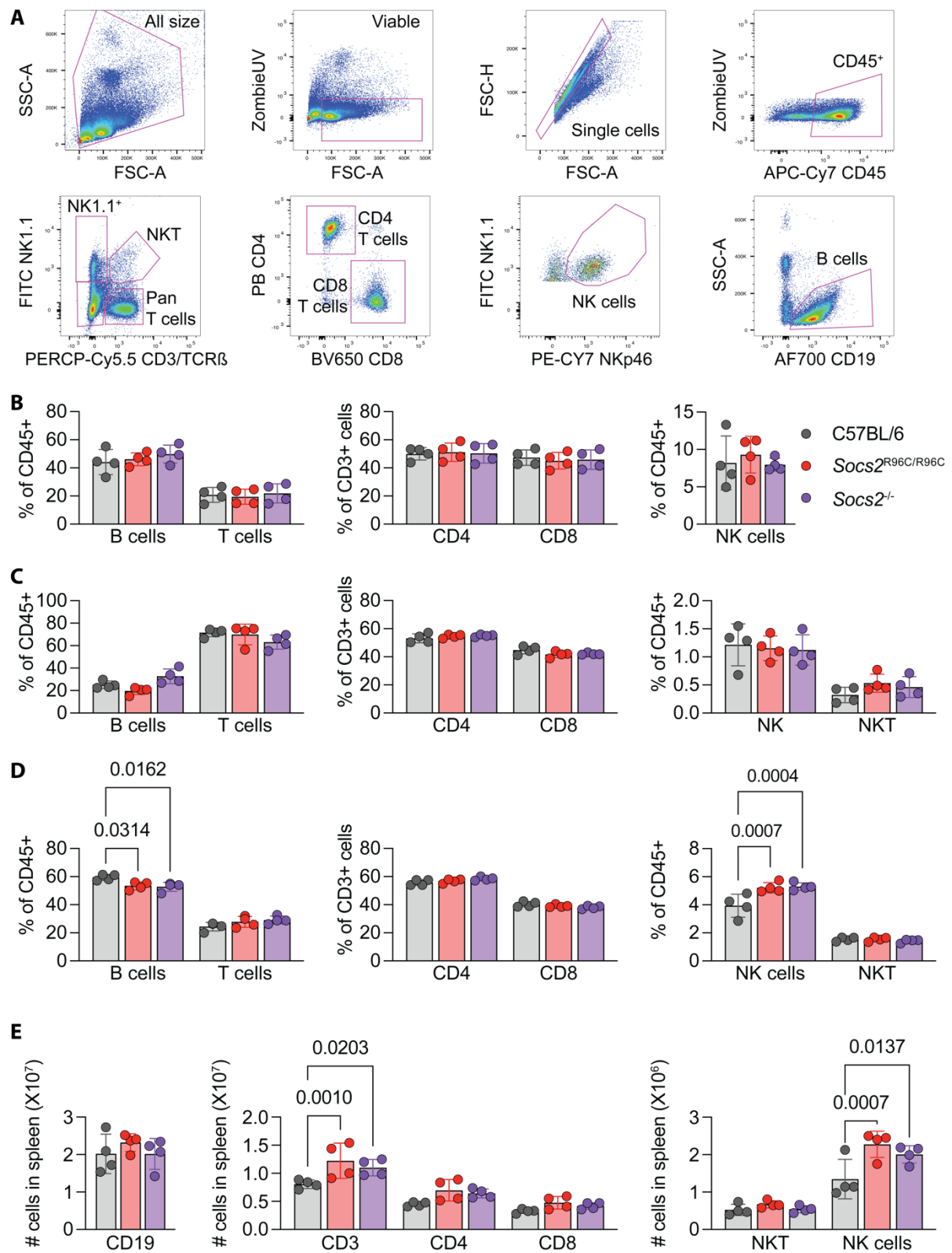


C SOCS2 R96C PCR SOCS2 WT PCR



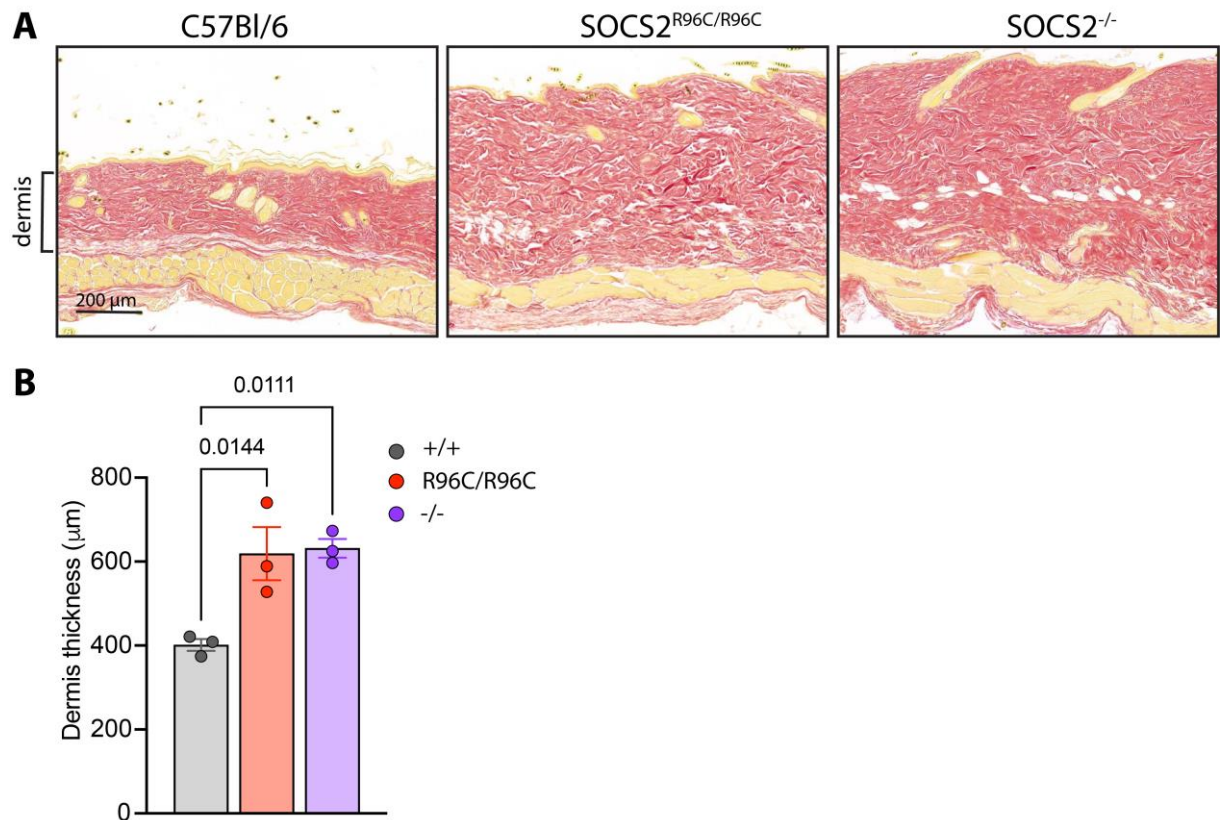
1. *Socs2*^{+/+}
2. *Socs2*^{R96C/+}
3. *Socs2*^{R96C/R96C}
4. No template

Supplementary Figure 2. Genotyping confirmation of the correct sequence change in the *Socs2*^{R96C} mouse. Sanger sequencing chromatograms confirming (A) the WT *Socs2* sequence and (B) the mutated *Socs2*^{R96C} sequence (homozygous mutant allele). *Highlights the CRISPR targeted base residue changes, two of which are synonymous and were introduced to enable primer specificity for standard gDNA genotyping. (C) Example 2% agarose gel confirming genotyping specificity by PCR. Primer sequences are available in **Supplementary Table 1**.

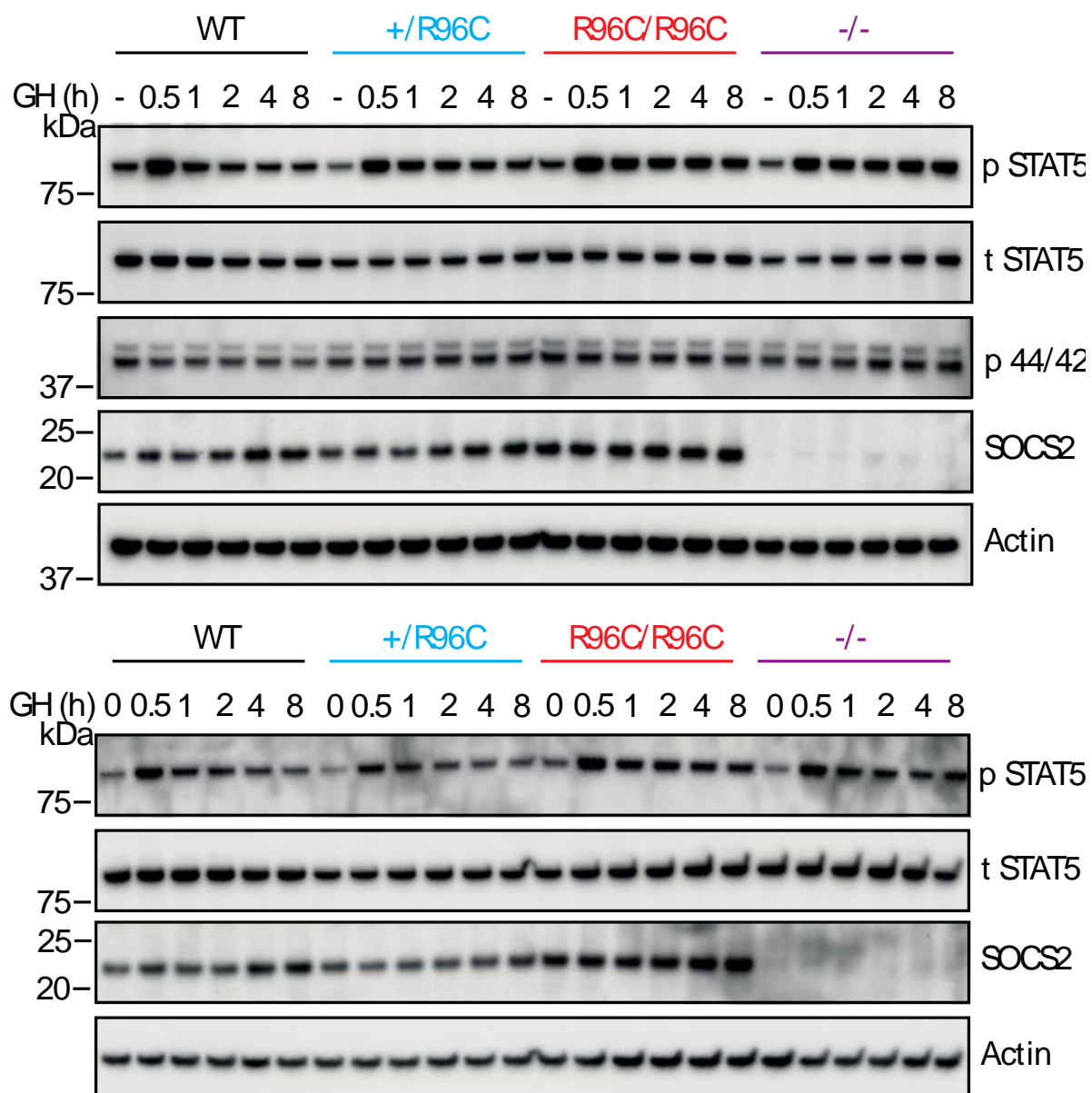


Supplementary Figure 3. Homozygous *Socs2*^{R96C/R96C} and *Socs2*^{-/-} mice display increased numbers of splenic NK cells. Single cell suspension peripheral blood mononuclear cells (PBMCs), splenocytes and lymphocytes from 8-12-week-old control C57BL/6 (black),

Socs2^{R96C/R96C} (red) and *Socs2*^{-/-} (purple) mice were stained with fluorescently conjugated antibodies to various immune markers and analysed by flow cytometry. **(A)** Example gating for the analysis of viable leukocytes (ZombieUV⁻CD45⁺), NKT (CD3⁺TCRβ⁺NK1.1⁺), Pan T cells (CD3⁺TCRβ⁺NK1.1⁻), CD4 T cells (CD3⁺TCRβ⁺NK1.1⁻CD4⁺), CD8 T cells (CD3⁺TCRβ⁺NK1.1⁻CD8⁺), Pan B cells (CD3⁻TCRβ⁻NK1.1⁻CD19⁺) and NK cells (CD3⁻TCRβ⁻NK1.1⁺NKp46⁺). Percentage of various immune populations in the **(B)** blood, **(C)** axial and inguinal lymph nodes, and **(D)** spleen. **(E)** Enumeration of immune populations in the spleen using counting beads (123count eBeads). **(B-E)** Each dot represents cells from an individual mouse. Significance determined by two-way ANOVA with Sidak's multiple comparisons test.



Supplementary Figure 4. Homozygous *Socs2*^{R96C/R96C} and *Socs2*^{-/-} mice display thickening of the skin. (A) Van Gieson-stained dorsal skin section from 10-week-old male WT, *Socs2*^{R96C/R96C} and *Socs2*^{-/-} mice. *Socs2* mutant and null mice show increased collagen deposits and a thickened dermis. Scale bar = 200 μm. Representative images of 3 mice of each genotype. (B) Quantification of dermis thickness. n=3 mice/genotype. Data were analyzed using a one-way ANOVA.



Supplementary Figure 5. *Socs2*^{R96C/R96C} and *Socs2*^{-/-} MEFs display prolonged growth hormone signal activation. *Socs2*^{+/+}, *Socs2*^{R96C/+}, *Socs2*^{R96C/R96C} and *Socs2*^{-/-} MEFs were treated with 50 ng/mL of GH, lysed and analysed by immunoblotting with antibodies to the indicated proteins. P: phosphorylated, T: total. Two additional independent experiments related to **Figure 4**.

Supplementary Table 1. *Socs2*^{R96C} genotyping primers.

	Allele	Forward Primer (5'-3')	Reverse Primer (5'-3')	PCR Product (bp)
Standard genotyping	<i>Socs2</i> ^{R96C}	AGCTTTCCAACCTTGTCCCCTA	ATCTGAATTTCCCATCTTGGTACTCAAT ACAT	766
	<i>Socs2</i> ^{+/+}	GCTGGACCGACTAACCTGC	AGCATGGTCAGCTTAACGGAA	901
NGS*		GTGACCTATGAACTCAGGAGTCTGAC TGTTAATGAAGCCAAAGAG	CTGAGACTTGACATCGCAGCGTGAACA GTCCCATTCGGTG	290

*Next generation sequencing