GenBank Accession: SPY04274.1; Region 191-450

NmNAT nucleotide sequence

ATGTCCCCCAGCCGTTTCAATCAAACGGGCCCCAAAATCGGCCTGAGCGTACGCCTTGCAGAAACC CAAGCTGAAATCGAAGCCGCCCAAAGATTGCGCTATCAGGTTTTTGCCCAAGAGTTGGGGGCGGAA ATCGAAAGCGATGACGGCCGTGATGTCGATCCTTATGATGAGCATTGCCACCACCTGCTTGCCTTCG ACGATGCAACCGGCGAAGTTATCGGCTGCTACCGCCTGATTACCGAAGAAACCGCGAAAAAAGTCG GCGGCTGGTACAGCGAGCATGAATTCGACCATGAGCCTTTGAAAGACATTCTGCCGCAAACCGTCG AACTCGGTCGCGCCTGTACCCACCGGACTACCGCAACGGCGGCTTGGTCATGCTGTTGTGGACCG GTTTGGTCAAATTCATGAAAGACGAAAACCTGCGCTTTATGATTGGTTGCGGCAGTATCGAAATGCG CGACGGCGGCAACGATGCGGCGGGGCCTGTATCATGCTTTGAAAGACAAATACCTCGCTCCGGAACA ATGGCGCGGCAACGATGCGGCGGGGCCTGTATCATGGTTCTGCGGCGGCGCTCGAAAATCCGCCTGT ACCCGCACTGATCAAAGGCTATCTCAAAGCAGGCGCATGGTTCTGCGGCGAGCCTTGCGTCGATGA AGCATTCAACTGCGCGGGATGTGCTGATCATGATGGACATCAGCCACCTTTCCGACCGCTACTTGCAG CGTTTTGCCCCTAAAACCGATTCTCAAAG



Figure S1. Key COSY and HMBC NMR correlations for 1 and 2.

N-3-hydroxymyristoyl-L-ornithine (MH+ 359) (MeOD, 600 MHz)

	NH	αH	βH	γH	δΗ	
N-3-hydroxymyristoyl		2.35	3.95	1.48		CH2 1.23-1.37 Terminal CH3 0.88
Orn		4.45	1.74 1.99	1.49	2.94	
	CO	αC	βC	γC	δC	
N-3-hydroxymyristoyl	172.92	43.16	68.38	36.87		Terminal CH3 12.93
Orn	173.14	51.20	28.37	22.29	38.76	

N-3-hydroxymyristoyl-L-lysine (MH+ 373) (MeOD, 600 MHz)

	NH	αH	βH	γH	δΗ	
N-3-hydroxymyristoyl		2.35	3.95	1.48		CH2 1.23-1.37

Lys		4.42	1.92 1.70	1.49	1.66	Terminal CH3 0.88 εΗ 2.90
	CO	αC	βC	γC	δC	
N-3-hydroxymyristoyl	172.92	43.16	68.38	36.87		Terminal CH3 12 93
Lys	173.66	51.47	30.76	22.29	26.52	εC 39.09

Table S1. Chemical shifts of natural (bacterially-produced) **1** and **2** as a mixture identified by 1D- and 2D-NMR (gCOSY, gHSQC, gHMBC)





С



Figure S2. NMR spectra of natural 1 and 2 as a mixture (MeOD, 600 MHz)



Figure S3. Marfey's analysis of **1** and **2**. Extracted ion chromatograms (*m/z* 385.1466, *m/z* 399.1623) of the Marfey's product of L-ornithine, D-ornithine, L-lysine, D-lysine, hydrolyzed **1**, hydrolyzed **2** are shown, both (**1** and **2**) of which align in retention time with that of the *S*- configuration of their respective amino acid head groups.



Figure S4. The $\Delta \delta_{S-R}$ in ppm for the MTPA esters of compounds 1 and 2.



Figure S5. Extracted ion chromatogram of crude *Nm*NAT extract, synthetic, and co-injection of crude and synthetic 1 and 2.

N-3-hydroxymyristoyl-L-ornithine (MH+ 359) (DMSO, 600 MHz)

	NH	αΗ	ßH	vH	δΗ	
N-3-hydroxymyristoyl		2.16	3.72	1.30,		CH2 1.20-1.32 Terminal CH3 0 82
Orn	7.84	4.02	1.58, 1.72	1.55	2.74	
	CO	αC	βC	γC	δC	
N-3-hydroxymyristoyl	171.13	44.10	67.95	37.34		Terminal CH3 14.40
Orn	173.92	52.51	29.27	24.26	38.99	

N-3-hydroxymyristoyl-L-lysine (MH+ 373) (DMSO, 600 MHz)

	NH	αH	βH	γH	δΗ	
N-3-hydroxymyristoyl		2.17	3.73			CH2 1.20-1.32 Terminal CH3 0.82
Lys	7.95	4.11	1.66, 1.52	1.30	1.49	εН 2.72
	СО	αC	βC	γC	δC	
N-3-hydroxymyristoyl	171.33	43.98		36.87		Terminal CH3 14.45
Lys	174.20	52.05	31.13	22.66	26.91	εC 39.11

Table S2. Chemical shifts of synthetic 1 and 2 identified by 1D- and 2D- NMR (gCOSY, gHSQC, gHMBC)





В



С



D



Figure S6. NMR spectra of synthetic compound 1 (DMSO, 600 MHz)







С





Figure S7. NMR spectra of synthetic compound 2 (DMSO, 600 MHz)

А



В



С



D



Figure S8. Extracted ion chromatograms of all identified *N*-acyl ornithines and lysines from *Nm*NAT expression in *E. coli*

					<u>.</u>	
Head			Retention Time	Theoretical	Observed	
Group	Acyl Chain	Compound	(min)	m/z	m/z	Ppm error
Orn						
	OH-C14:0	1	17.177	359.2904	359.2934	8.349792
	OH-C15:0	3a	18.294	373.3061	373.30857	6.616554
	OH-C16:0	4a	19.411	387.3217	387.3246	7.487316
	OH-C17:0	5a	20.678	401.3374	401.33806	1.644502
Lys						
	OH-C14:0	2	17.31	373.3061	373.3086	6.696917
	OH-C15:0	3b	18.41	387.3217	387.3234	4.389116
	OH-C16:0	4b	19.511	401.3374	401.33929	4.709255
	OH-C17:0	5b	20.778	415.353	415.3536	1.444554
Orn						
	C14	6a	19.027	343.2955	343.29833	8.243627
	C16	7a	21.561	371.3268	371.3279	2.96235
Lys						
	C14	6b	19.261	357.3112	357.3132	5.597362
	C16	7b	21.778	385.3425	*385.34231	-0.519019

Orn						
	OH-C14:1	8a	16.093	357.2748	357.2746	-0.559793
	OH-C16:1	9a	18.027	385.3061	385.3071	2.595339
	OH-C17:1	10a	19.011	399.3217	399.3218	0.250425
	OH-C18:1	11a	20.111	413.3374	413.3387	3.14513
Lys						
	OH-C14:1	8b	16.193	371.2904	*371.2915	2.96241
	OH-C16:1	9b	18.127	399.3217	399.3224	1.752973
	OH-C17:1	10b	19.111	413.3374	*413.3376	0.483866
	OH-C18:1	11b	20.211	427.353	427.3548	4.211975
Orn						
	C14:1	12a	17.177	341.2799	341.28111	3.545477
	C15:1	13a	18.294	355.2955	355.2959	1.125823
	C16:1	14a	19.411	369.3112	369.3113	0.270774
	C17:1	15a	21.111	383.3268	383.3278	2.60874
	C18:1	16a	22.195	397.3425	397.3426	0.251672
Lys						
	C14:1	12b	17.293	355.2955	355.2962	1.970191
	C15:1	13b	18.41	369.3112	*369.31151	0.8394
	C16:1	14b	19.527	383.3268	383.3275	1.826118
	C17:1	15b	21.311	397.3425	*397.3427	0.503344
	C18:1	16b	22.395	411.3581	*411.3589	1.944778

Table S3. HRMS of all identified *N*-acyl ornithines and lysines from expression of *Nm*NAT in *E. coli*. Ions marked with * are detected but have inconclusive tandem MS spectra



OH-C14:0 Ornithine

OH-C15:0 Ornithine



OH-C16:0 Ornithine















OH-C15:0 Lysine















C14:0 Ornithine





239.2369



C16:0 Ornithine







OH-C14:1 Ornithine





II.

h



OH-C17:1 Ornithine

-H₂0



OH-C18:1 Ornithine







OH-C18:1 Lysine





C14:1 Ornithine













C17:1 Ornithine





251.236







C16:1 Lysine



Figure S9. Tandem MS of 24 of the 30 *N*-acyl amides detected from *Nm*NAT expression. Corresponding structures show deduced structures, where regiochemistry of hydroxyl groups and unsaturation, as well as stereochemistry of points of unsaturation are assumed. The straight acyl chain is also a simplification, as they may occur as branched chain fatty acids.

Structural elucidation of 3 and 4

Compound	Molecular Formula	Theoretical m/z	Observed m/z	Ppm error
3	C ₁₆ H ₃₁ NO ₃	286.2377	286.2381	1.39744
4	C ₁₉ H ₂₉ NO ₃	320.222	320.2229	2.81055

Table S4. Theoretical and observed HRMS values of compounds 3 and 4



Figure S10. HRMS of compounds 3 and 4



Figure S11. Key COSY and HMBC correlations of natural 3 and 4 identified from a mixture



Figure S12. Marfey's analysis for stereochemical elucidation of compounds 3 and 4 (left). Co-injection with synthetic 3 and 4 for validation of structure (right).

	NH	αH	βH	γH	δΗ	
<i>N</i> -decanoyl		2.26	1.64			CH2 1.25-1.35 Terminal CH3 0.89
Leu	8.05	4.62	1.74 1.60	1.73	0.97	
	CO	αC	βC	γC	δC	
<i>N</i> -decanoyl	174.17	36.50	25.61			CH2 21.86, 29.19- 31.86 Terminal CH3 14.07
Leu	176.39	50.93	41.16	24.91	22.65 22.83	

N-decanoyl-L-leuci	ne (MH+ 286)	(CDCl3, 4	00 MHz)
--------------------	--------------	-----------	---------

Table S5. Chemical shifts of synthetic 3 as identified by 1D- (¹H and ¹³C) and 2D- (gCOSY) NMR

	NH	αH	βH			
N-decanoyl		2.03	1.39	CH2 1	.08-1.32	
				Termir	nal CH3 0.87	
Phe	8.07	4.43	2.84	Aroma	itic	12.65
			3.06	7.17-7	.29	COOH
	CO		αC	βC		
N-decanoyl	172.57	7	35.53	25.63	CH2 22.55-31	.69
					Terminal CH3	14.39
Phe	173.69)	53.77	37.24	Aromatic	
					126.77-138.2	6

N-decanoyl-L-phenylalanine (MH+ 320) (DMSO, 400 MHz)

Table S6. Chemical shifts of synthetic 3 as identified by 1D- (1H and 13C) and 2D- (gCOSY) NMR





Figure S13. NMR spectra of synthetic compound 3 (CDCI3, 400 MHz)

В



В



А



Figure S14. NMR spectra of synthetic compound 4 (DMSO, 600 MHz)



B. subtilis inhibition

Figure S15. Antibacterial assay of compounds **3** and **4** against *Bacillus subtilis*. IC₅₀ was calculated as 301.2 μ M and 153 μ M for **3** and **4**, respectively, through a variable slope (four parameters) fitting on GraphPad Prism.



Figure S16. Cytotoxicity (LDH readout) measurements of compounds **1** and **2** at varying concentrations, and of compounds **3** and **4** at fixed concentration of $30 \ \mu$ M.