Supplementary Figures

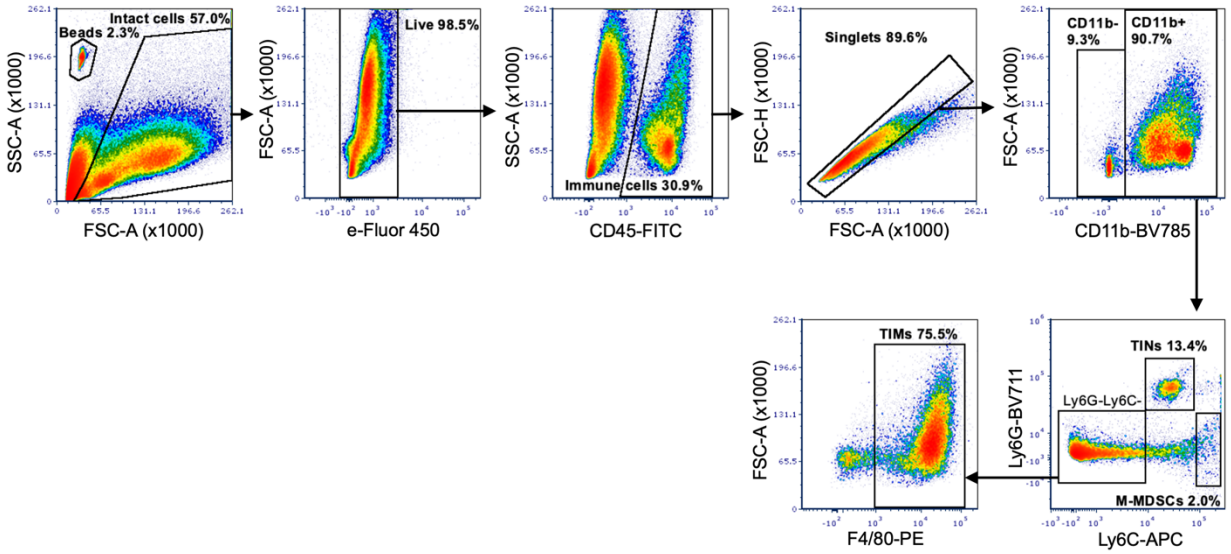

Figure S1. Flow cytometry gating strategy for tumor-infiltrating macrophages (TIMs), tumor-infiltrating neutrophils (TINs)/polymorphonuclear myeloid-derived suppressor cells (PMN-MDSCs), and monocytic myeloid-derived suppressor cells (M-MDSCs). Live cells were eFluor 450-negative. All other gates were set using isotype controls and FMO (fluorescence minus one) controls. Single-stained compensation controls were used to calculate the proper compensation matrix.

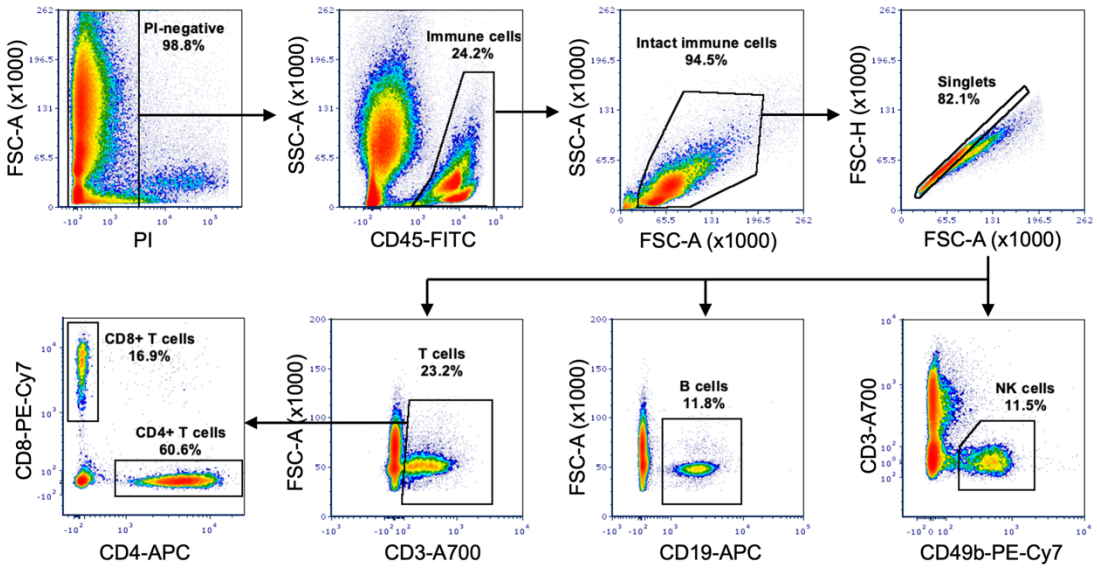

Figure S2. Flow cytometry gating strategy for tumor-infiltrating T cells (CD4+ and CD8+), B cells, and NK cells. Live cells were propidium iodide (PI)-negative. All other gates were set using isotype controls and FMO (fluorescence minus one) controls. Single-stained compensation controls were used to calculate the proper compensation matrix.

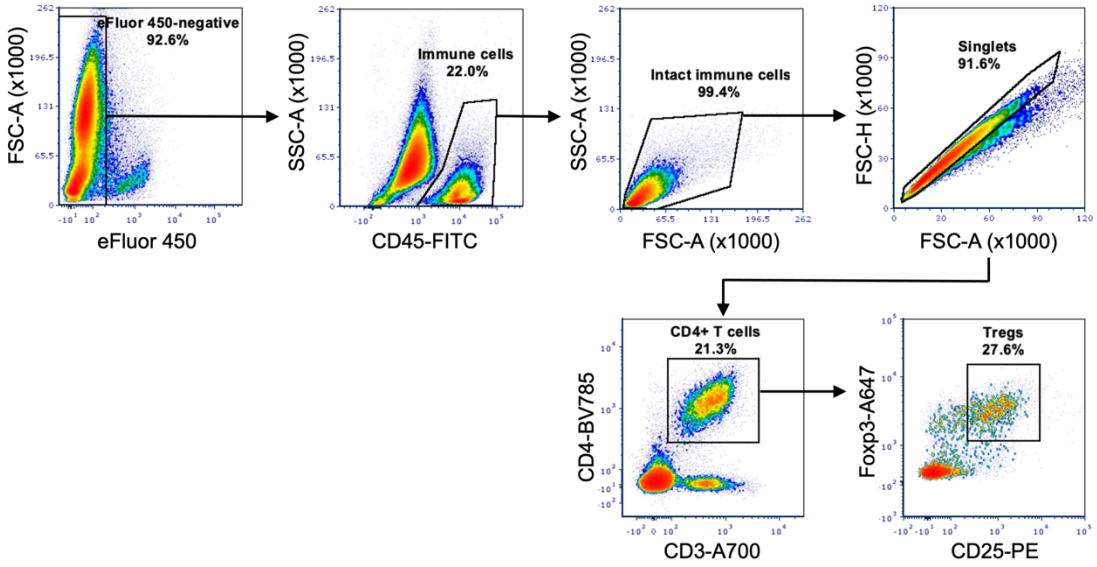

Figure S3. Flow cytometry gating strategy for tumor-infiltrating regulatory T cells (Tregs). Isolated cells were first incubated with fixable eFluor 450 viability dye, followed by staining of cell surface markers, fixation and permeabilization, and staining of intracellular Foxp3. Live cells were eFluor 450-negative. All other gates were set using isotype controls and FMO (fluorescence minus one) controls. Single-stained compensation controls were used to calculate the proper compensation matrix.

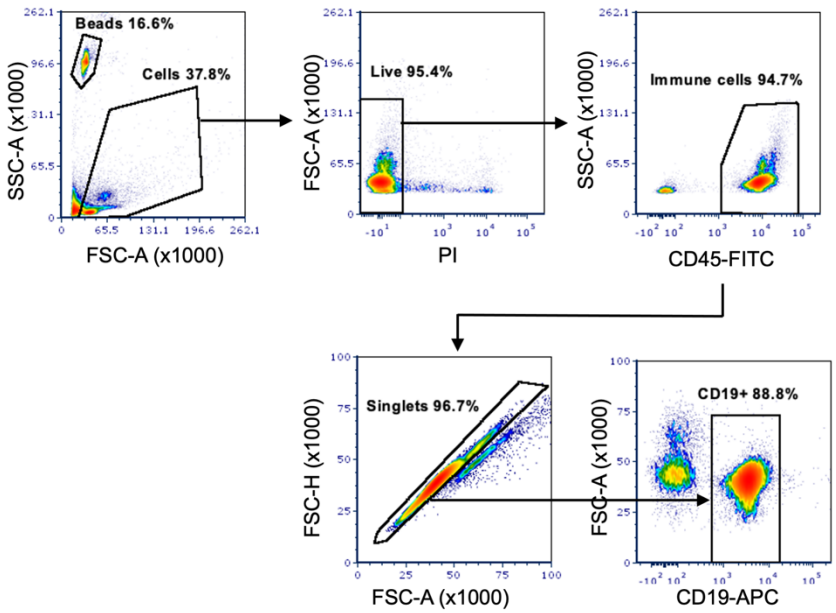

Figure S4. Flow cytometry gating strategy for splenic B cells. Live cells were propidium iodide (PI)-negative. All other gates were set using isotype controls and FMO (fluorescence minus one) controls. Single-stained compensation controls were used to calculate the proper compensation matrix.

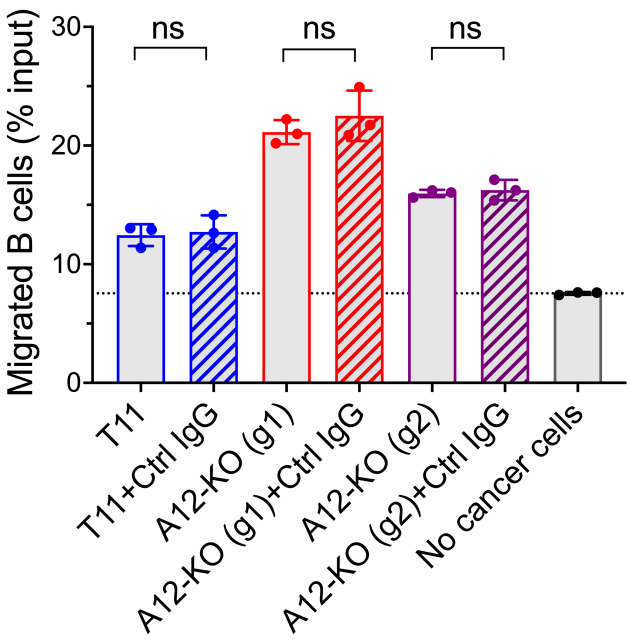

Figure S5. B cell migration toward T11, A12-KO (g1), or A12-KO (g2) cells, in the absence or presence of 2.5 μg/ml rat IgG2a,κ isotype control antibody added to the lower transwell chambers. This is a control experiment for Fig. 5l-n showing the effect of anti-CXCL12-blocking antibody on B cell migration.