<u>Materials Design Analysis Reporting (MDAR)</u> Checklist for Authors

The MDAR framework establishes a minimum set of requirements in transparent reporting applicable to studies in the life sciences (see Statement of Task: doi:10.31222/osf.io/9sm4x.). The MDAR checklist is a tool for authors, editors, and others seeking to adopt the MDAR framework for transparent reporting in manuscripts and other outputs. Please refer to the MDAR Elaboration Document for additional context for the MDAR framework.

For all that apply, please note where in the manuscript the required information is provided.

Materials:

Newly created materials	indicate where provided: page no/section/legend)	n/a
The manuscript includes a dedicated "materials		
availability statement" providing transparent	A data availability statement was provided on p.321.	
disclosure about availability of newly created	All new materials are freely available on reasonable	
materials including details on how materials can be	request.	
accessed and describing any restrictions on access.		

Antibodies	indicate where provided: page no/section/legend)	n/a
For commercial reagents, provide supplier name, catalogue number and RRID, if available.		х

DNA and RNA sequences	indicate where provided: page no/section/legend)	n/a
Short novel DNA or RNA including primers, probes:	See Materials and Methods: p. 15, Cloning strategies; p.	
Sequences should be included or deposited in a	17, Viral RNA isolation and 3CL ^{pro} sequencing	
public repository.	See Supplemental file S2	
	See Supplemental Material tables S12 and S13, p. 20	

Cell materials	indicate where provided: page no/section/legend	n/a
Cell lines: Provide species information, strain. Provide accession number in repository OR supplier name, catalog number, clone number, OR RRID.	See Materials and Methods p. 16, Cell lines BHK21 (ATCC CCL-10 C-13) 293T (ATCC CRL-3216) 293VSV is a laboratory stock (Panda et al., see section Cell lines) and available upon request via MTA. Vero E6 (ATCC CRL-1586) A549 hACE2 (Biomedical Resource Ontology NR-53821)	.,, u
Primary cultures: Provide species, strain, sex of origin, genetic modification status.		х

Experimental animals	indicate where provided: page no/section/legend)	n/a
Laboratory animals or Model organisms: Provide species, strain, sex, age, genetic modification status. Provide accession number in repository OR supplier name, catalog number, clone number, OR RRID.		x
Animal observed in or captured from the field: Provide species, sex, and age where possible.		х

Plants and microbes	indicate where provided: page no/section/legend)	n/a
Plants: provide species and strain, ecotype and cultivar where relevant, unique accession number if available, and source (including location for collected wild specimens).		х
Microbes: provide species and strain, unique accession number if available, and source.	See Materials and Methods: pp. 15, 16, 17, 21, 22 Chimeric VSVs generated for this work and previous variants are laboratory stocks and are available upon request via MTA. See Supplemental file S2 for VSV sequence 10-β competent <i>E. coli</i> NEB C3019	

Human research participants	indicate where provided: page no/section/legend) or state if these demographics were not collected	n/a
If collected and within the bounds of privacy constraints report on age, sex and gender or		Х

ethnicity for all study participants.	

Design:

Study protocol	indicate where provided: page no/section/legend)	n/a
If study protocol has been pre-registered, provide DOI. For clinical trials, provide the trial registration		
number OR cite DOI.		Х

Laboratory protocol	indicate where provided: page no/section/legend)	n/a
Provide DOI OR other citation details if detailed step- by-step protocols are available.	See Materials and Methods p. 22: Nanopore sequencing, "Midnight protocol" (dx.doi.org/10.17504/protocols.io.bwyppfvn)	

For in vivo studies: State whether and how the following have been done	indicate where provided: page no/section/legend. If it could have been done, but was not, write not done	n/a
Sample size determination		х
Randomisation		Х
Blinding		Х
Inclusion/exclusion criteria		Х

Sample definition and in-laboratory replication	indicate where provided: page no/section/legend	n/a
State number of times the experiment was replicated in laboratory.	Experiments were repeated at least twice. For further details see, Study design, Materials and Methods section and figures for replicates number.	
Define whether data describe technical or biological replicates.	Both technical (TCID ₅₀) and biological (all other) replicates were performed. See Materials and Methods for more details.	

Ethics	indicate where provided: page no/section/legend	n/a
Studies involving human participants: State details of authority granting ethics approval (IRB or equivalent committee(s), provide reference number for approval.		х
Studies involving experimental animals: State details of authority granting ethics approval (IRB or equivalent committee(s), provide reference number for approval.		Х
Studies involving specimen and field samples: State if relevant permits obtained, provide details of authority approving study; if none were required, explain why.		х

Dual Use Research of Concern (DURC)	indicate where provided: page no/section/legend	n/a
If study is subject to dual use research of concern		
regulations, state the authority granting approval		Х
and reference number for the regulatory approval.		

Analysis:

Attrition	indicate where provided: page no/section/legend	n/a
Describe whether exclusion criteria were		
preestablished. Report if sample or data points were		
omitted from analysis. If yes report if this was due to		Х
attrition or intentional exclusion and provide		
justification.		

Statistics	indicate where provided: page no/section/legend	n/a
Describe statistical tests used and justify choice of tests.	See Materials and Methods: p.23, Statistical analyses for further details.	

Data availability	indicate where provided: page no/section/legend	n/a
For newly created and reused datasets, the manuscript includes a data availability statement that provides details for access or notes restrictions on access.	See Data availability section, p. 32	
If newly created datasets are publicly available, provide accession number in repository OR DOI OR URL and licensing details where available.	See Materials and Methods: p. 15, Cloning strategies for used plasmids. See Supplemental file S2 for chimeric VSV-3CLpro sequence.	х
If reused data is publicly available provide accession number in repository OR DOI OR URL, OR citation.	See Supplemental material file S1 See Supplemental figure S2 Public datasets: GISAID (gisaid.org) and NCBI (https://www.ncbi.nlm.nih.gov/labs/virus/vssi/#/scov2 _snp)	

Code availability	indicate where provided: page no/section/legend	n/a
For all newly generated custom computer code/software/mathematical algorithm or re-used code essential for replicating the main findings of the study, the manuscript includes a data availability statement that provides details for access or notes restrictions.		х
If newly generated code is publicly available, provide accession number in repository, OR DOI OR URL and licensing details where available. State any restrictions on code availability or accessibility.		х
If reused code is publicly available provide accession number in repository OR DOI OR URL, OR citation.	See Materials and Methods: p. 22, Nanopore sequencing.	

Reporting

MDAR framework recommends adoption of discipline-specific guidelines, established and endorsed through community initiatives. Journals have their own policy about requiring specific guidelines and recommendations to complement MDAR.

Adherence to community standards	indicate where provided: page no/section/legend	n/a
State if relevant guidelines (e.g., ICMJE, MIBBI, ARRIVE) have been followed, and whether a checklist (e.g., CONSORT, PRISMA, ARRIVE) is provided with the manuscript.		х