

ceaptadn 🔻

In 1 collection

Morris Baumgardt¹, Maren Hülsemann², Maren Mieth¹, Doris Frey¹, Stefan Hippenstiel¹, Andreas C. Hocke¹, Katja Hönzke¹

¹Charité – Universitätsmedizin Berlin, corporate member of Freie Universität Berlin and Humboldt Univ ersität zu Berlin, Department of Infectious Diseases and Respiratory Medicine, Charitéplatz 1, 10117 B erlin, Germany.;

²Berlin Institute of Health at Charité (BIH), BIH QUEST Center for Responsible Research, Berlin, Germa ny.

1 Works for me

Reserved DOI:

10.17504/protocols.io.14egn7z8zv5d/v3



morris.baumgardt

DISCLAIMER

Informed written consent was obtained from all volunteers and the study was approved by the Charité Ethics Committee (project 451, EA2/079/13).

ABSTRACT

This protocol describes the RNA extraction from human alveolar-like organoids followed by the perfomance of a RT-qPCR to quantify the amount of hostfactors (ACE2, TMPRSS2, and FURIN) of SARS-CoV-2. The protocol contains detailed steps for the lysis of human alveolar-like organoids, as wells as the RNA extraction followed by the reverse transcription into cDNA. In the next section the protocol describes in detail how to quantify the gene expression with the TaqMan gene expression kit to obtain donor dependent C_t values.

PROTOCOL INFO

Morris Baumgardt, Maren Hülsemann, Maren Mieth, Doris Frey, Stefan Hippenstiel, Andreas C. Hocke, Katja Hönzke . RNA Extraction and RT-qPCR of Human Lung Organoids . **protocols.io**

https://protocols.io/view/rna-extraction-and-rt-qpcr-of-human-lung-organoids-ceaptadn

Version created by morris.baumgardt



COLLECTIONS (i)



Collection: State-of-the-Art Analytical Methods of Viral Infections in Human Lung Organoids **KEYWORDS**

RNA extraction, human lung organoids, TaqMan, RT-qPCR, Sars-CoV-2 hostfactors

CREATED

Jul 26, 2022

LAST MODIFIED

Sep 27, 2022

PROTOCOL INTEGER ID

67631

PARENT PROTOCOLS

Part of collection

Collection: State-of-the-Art Analytical Methods of Viral Infections in Human Lung Organoids

GUIDELINES

This protocol describes the processing of human alveolar-like organoids which have been grown according to Youk et al., 2020. https://doi.org/10.1016/j.stem.2020.10.004.

MATERIALS TEXT

Α	В	С	D
Substance	Company	Order number	Concentration
GlutaMax 100x	invitrogen	35050-038	5 mL/500 mL medium
Hepes	invitrogen	15630-056	5 mL/500 mL medium
Advanced	invitrogen	12634-034	1x
DMEM/F12			

Composition of base medium

Α	В	С
Substance	Company	Order number
High Capacity cDNA Reverse	Applied biosciences	4368814
Transcription Kit		
RNeasy Mini Kit (50)	Qiagen	74104
Gene Expression Master Mix	AppliedBiosystems by	4369510
	ThermoFisher Scientific	
Ambion™ Nuclease Free Water	Invitrogen By ThermoFisher	AM 9937
	Scientific	
RLT buffer	Qiagen	7921
Bioanalyzer RNA Analysis	Agilent	G2939BA
Agilent RNA 6000 Nano Kit	Agilent	5067-1511
Biometra TRIO PCR Cycler	analytikjena	846-2-070-720
β-Mercaptoethanol	Merck	M6250

Reagents, kits and devices for RNA extraction and qPCR

A	В	С
TaqMan Gene Expression Assay	ThermoFisher Scientific	4331182
	ACE2	Hs01085333_m1
	TMPRSS2	Hs01122322_m1
	FURIN	Hs00965485_g1
	GAPDH	Hs02758991_g1
	ACTIN	Hs01060665_g1

TaqMan Gene Expression Assay and primers

SAFETY WARNINGS

If working with SARS-CoV-2 infected material please be sure to work at biosafety level 3 (BSL3) until step 7.

DISCLAIMER:

Informed written consent was obtained from all volunteers and the study was approved by the Charité Ethics Committee (project 451, EA2/079/13).

BEFORE STARTING

To avoid contaminations RNAase/DNAase free reagents, consumables and filter tips must be used at all steps.

Please use RNA low bind tubes.

Organoid lysis for RNA isolation

1 Grow your 3D model as described.

Youk J, Kim T, Evans KV, Jeong YI, Hur Y, Hong SP, Kim JH, Yi K, Kim SY, Na KJ, Bleazard T, Kim HM, Fellows M, Mahbubani KT, Saeb-Parsy K, Kim SY, Kim YT, Koh GY, Choi BS, Ju YS, Lee JH (2020). Three-Dimensional Human Alveolar Stem Cell Culture Models Reveal Infection Response to SARS-CoV-2.. Cell stem cell. https://doi.org/10.1016/j.stem.2020.10.004

- 2 Remove the entire organoid medium from the wells.
- 3 Add 11 mL ice-cold base medium and collect Cultrex with organoids in a 2 mL tube and flush well with additional 11 mL base medium and collect also.
- 4 To disolve the Cultrex place the tube at $\& 4 \degree C$ for & 00:05:00.

5m

5m

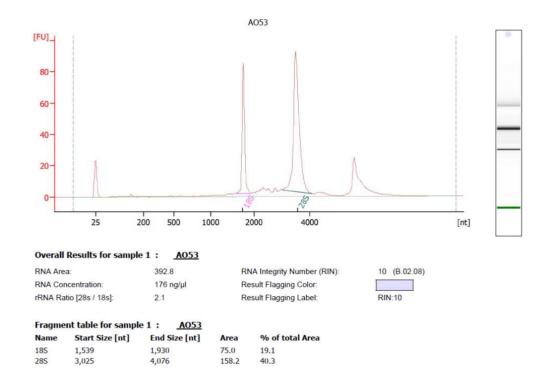
- 5 Centrifuge at **\$900 x g, 4°C, 00:05:00**.
- 6 Carefully remove supernatant and add $\blacksquare 350~\mu L$ RLT Puffer + β -Mercaptoethanol to the organoids (pellet).
- 7 Place supernatant in a 1.5 mL safelock tube.
 - A

In case the organoids were infected with a virus they can now be transported out of the biosafety level 3 (BSL3) laboratory

Isolate RNA using "RNeasy Mini Kit" from Qiagen (Cat No./ID 74104).

This step represents the RNA isolation protocol based on the "RNeasy Mini Kit" from Qiagen (Cat No./ID 74104) - no changes have been made to the protocol included in the kit

After isolation, measure RNA quality via RIN (RNA Integrity Number) using 2100 Bioanalyzer instrument (manufacturer's instructions, no changes have been made to the protocol included in the kit "Agilent RNA 6000 Nano Kit"). High-quality RNA will contain an RIN of at least 8. All samples used in this protocol had a RIN of 10.



Examplary RNA integrity check using a Bioanalyzer. This RIN example contains a RIN of 10 (completely intact).

Preparation of 2x RT master mix and cDNA reverse transcription

2h 15m

9



All steps & On ice.

- To quantify the amount of RNA before reverse transcription use $\blacksquare 1.5 \, \mu L$ of your RNA solution using the NanoDrop.
- 11 Allow the kit components to thaw on ice (high capacity cDNA reverse transcription kit, Applied

biosciences).

- 12 Calculate the volume of components needed to prepare the required number of reactions.
- 13 Prepare the RT master mix on ice.
- 14 One reaction needs:

Α	В
Reagent	Amount (µL)
10x RT Buffer	2
25x dNTP Mix (100mM)	0.8
10x RT Random Primers	2
MultiScribe™ RT	1
Nuclease-free H2O	4.2
TOTAL	10

Master mix (reverse transcription)

- According to NanoDrop-measurement prepare an RNA/water solution containing $\Box 1 \mu g$ RNA/ $\Box 10 \mu L$ water in a thin wall PCR tube. If RNA concentration is low, $\Box 500 ng$ RNA/ $\Box 10 \mu L$ water is also sufficient.
- 16 Include RT control: use water instead of RNA, run one control with each primer.
- 17 Add **10 μL** of 2x reverse transcription master mix to each tube.
- 18 Use the following program for reverse transcription:

Α	В	С
Step	Temperature	Duration
1	25°C	10 min
2	37°C	2 h
3	85°C	5 min
4	4°C	infinite

Program for Reverse Rranscription in a Thermal Cycler

- 19 Add \blacksquare 80 µL H₂O to each tube (cDNA).
- 20 Use the cDNA directly for qPCR or store at $\, \& \, -20 \, ^{\circ} \text{C} \,$.

TaqMan - quantitative PCR 14m 15s

21 Prepare the qPCR master mix. Calculate the volume of components needed to prepare the required number of reactions for each Gene expression assay.

Α	В
Reagent	Amount (µL)
Gene Expression Master Mix	10
H20	4
TaQMan Gene Expression Assay	1
Total	15

TaqMan Gene Expression Assay Master Mix (1 Reaction)

- 22 Perform qPCR, pipetting $\Box 15 \, \mu L$ of respective TaqMan gene expression assay master mix to the bottom of the well, then pipet $\Box 5 \, \mu L$ of cDNA on the upper wall of the well.
- 23 Seal the plate with a Clear Adhesive Film.
- 24 Centrifuge the plate for $\$1200 \times g$, 00:00:30.

30s

25



Important quality controls:

Run each sample in duplicates.

Always run house keeping genes (e.g., GAPDH and B-ACTIN) and gene of interest on the same plate.

Include water control for each primer.

Include RT control from previous step.

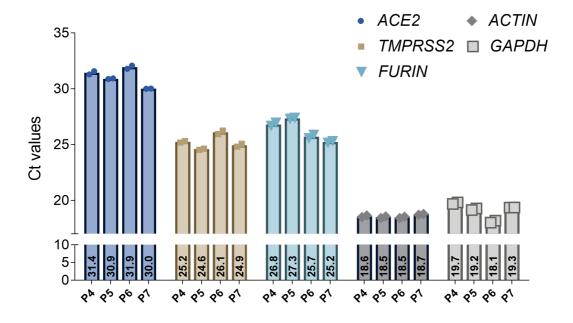
26 Run the qPCR reaction in a Thermal Cycler:

Α	В	С	D
Stage	Temperature	Duration	Repetitions
1	50°C	2 min	1
2	95°C	10 min	1
3	95°C	15 s	40
4	60°C	1 min	40

qPCR Program (PCR Volume = 20 μL)

Analysis

27 Exemplary result:



Examplary mRNA expression (Ct values) of ACE2, TMPRSS2, and FURIN in human alveolar-like organoids (GAPDH and B-ACTIN serve as reference genes, 4 donors)