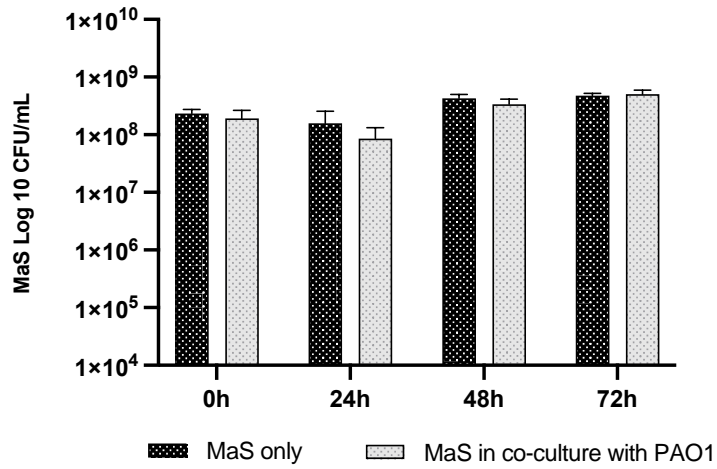


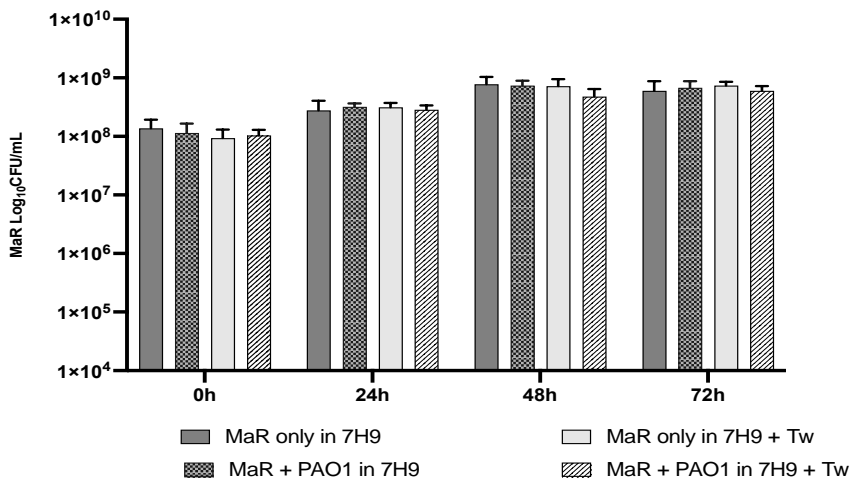
## SUPPLEMENTAL DATA

<b>Bacterial Strains</b>	<b>Locus Tag</b>	<b>Description / genotype</b>
<i>pqsA</i>	PA0996	pqsA-H05::lacZbp02q1
<i>pqsH</i>	PA2587	pqsH-F01::lacZbp03q3
<i>pqsL</i>	PA4190	pqsL-H11::lacZbp02q4
<i>lasI</i>		Deletion mutant in PAO1 background
<i>rhlI</i>		Deletion mutant in PAO1 background
<i>lasI rhlI</i>		Deletion mutant in PAO1 background
<i>vgrG</i>	PA0091	vgrG1-D02::phoAwp01q2
<i>Hcp</i>	PA0263	HcpC-E04::phoAwp04q3
<i>cplV</i>	PA0090	cplV1-F02::phoAwp02q4
<i>cdiA</i>	PA2462	PA2462-G12::lacZwp02q4
<i>cdiB</i>	PA2463	PA2463-A05::phoAwp03q4
<i>pcrV</i>	PA1706	pcrV-F02::lacZbp02q3
<i>pcrD</i>	PA1703	pcrD-A09::phoAwp01q1
<i>exoT</i>	PA0044	exoT-C09::lacZwp03
<i>fliC</i>	PA1092	fliC-G10::phoAwp05q2
<i>pilA</i>	PA4525	pilA-E01::lacZwp08q4
<i>fliC pilA</i>		Double mutant in the MPAO1 background (Parseak Lab)
<i>pchE</i>	PA4226	pchE-E06:: phoAwp07q4
<i>pvdD</i>	PA2399	pvdD-E07::lacZbp02q4
<i>xcpR</i>	PA3013	xcpR-D09:: phoAwp03q3
<i>xcpQ</i>	PA3105	xcpQ-E03:: phoAwp02q4

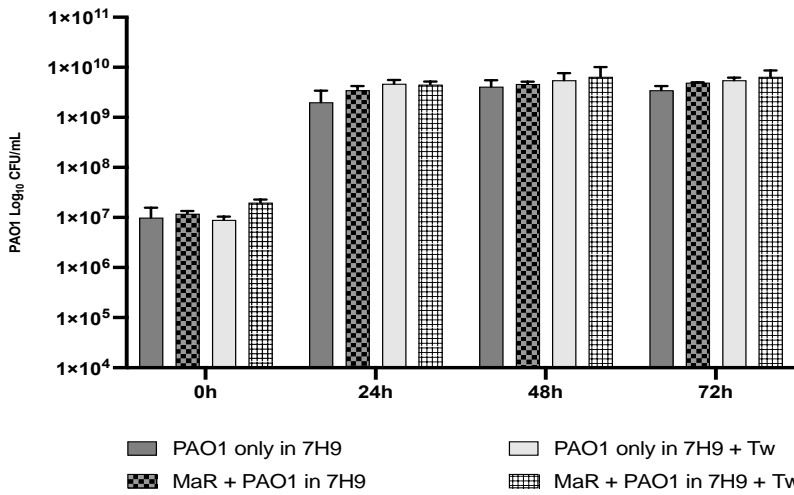
Table S1. *P. aeruginosa* mutant strain



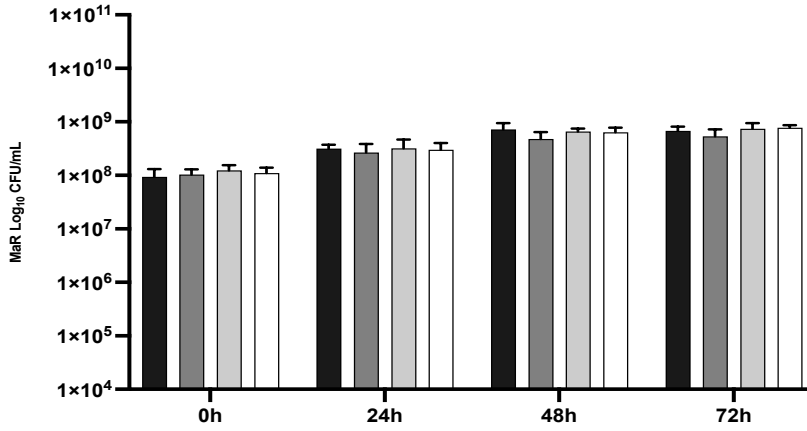
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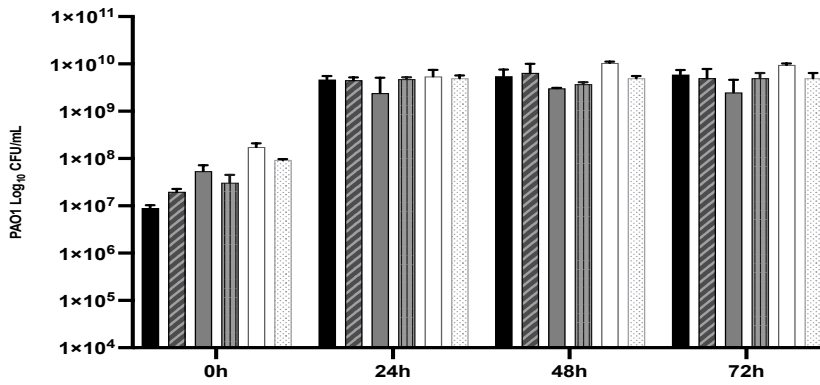
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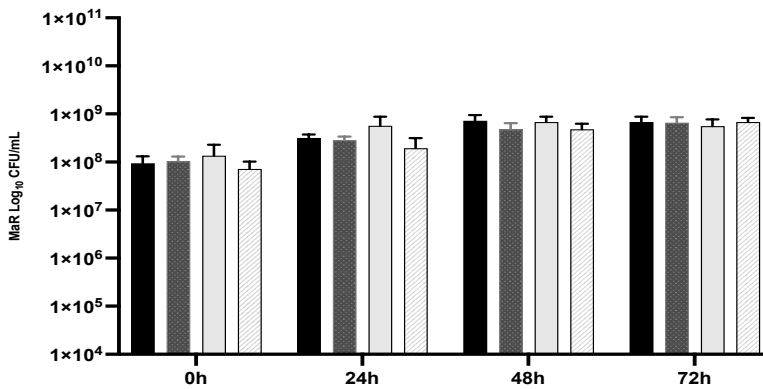
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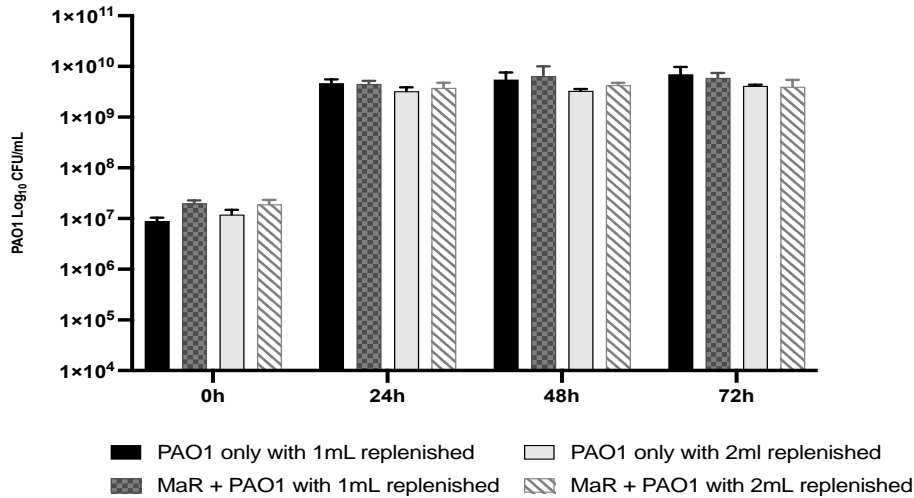
D. **MaR only** **MaR + PAO1(0.150)**  
**MaR + PAO1(0.05)** **MaR + PAO1(0.5)**



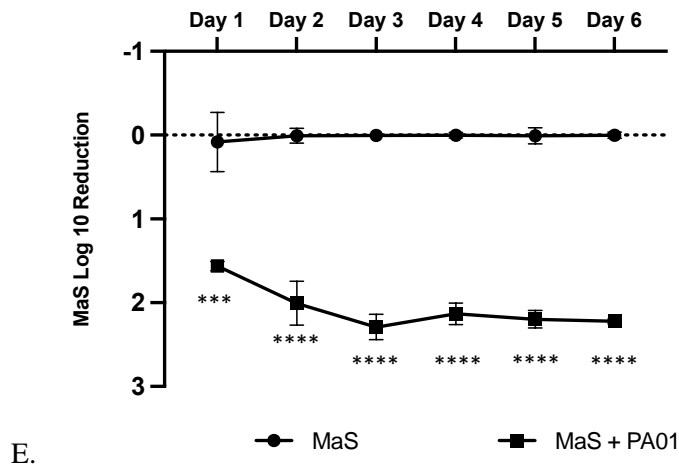
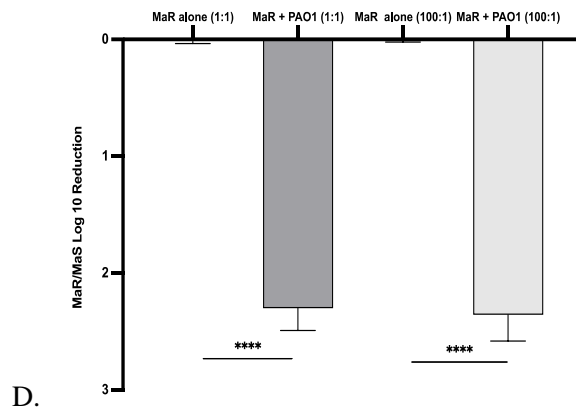
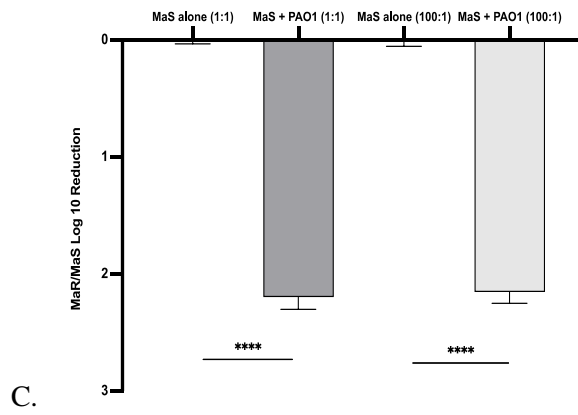
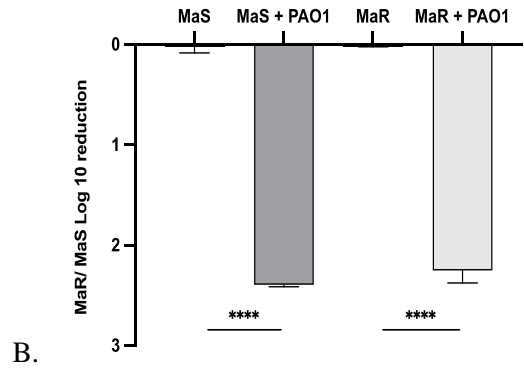
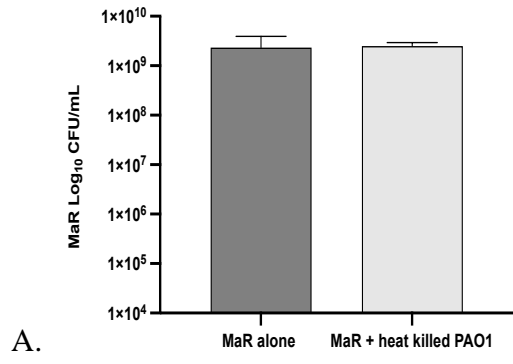
E. **PAO1(0.05) only** **PAO1(0.150) only**  
**MaR + PAO1(0.05)** **MaR + PAO1(0.150)**  
**PAO1(0.5) only** **PAO1 + MaR(0.5)**

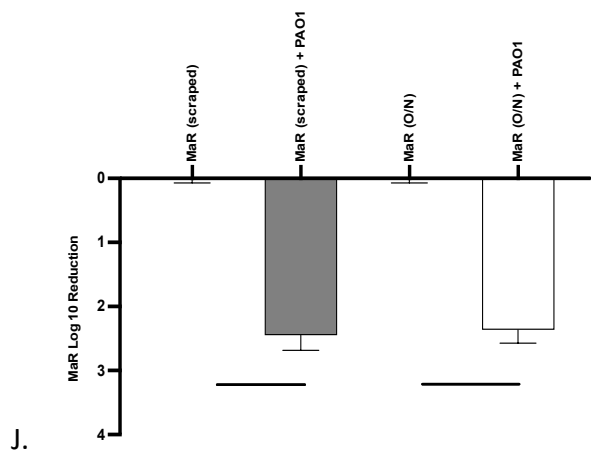
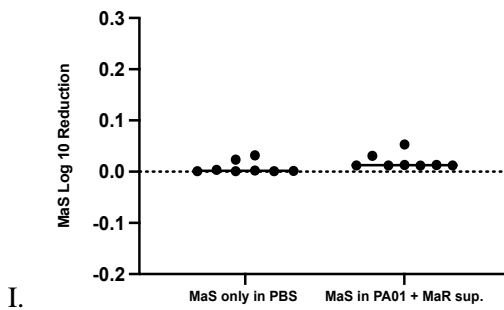
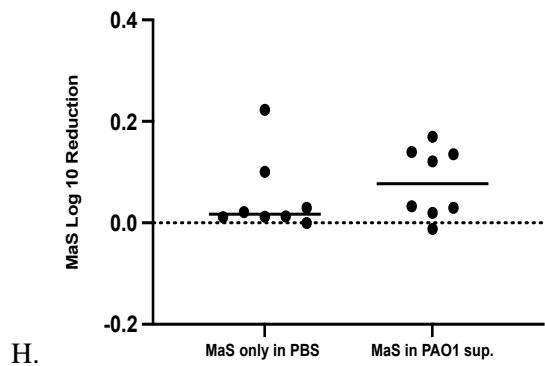
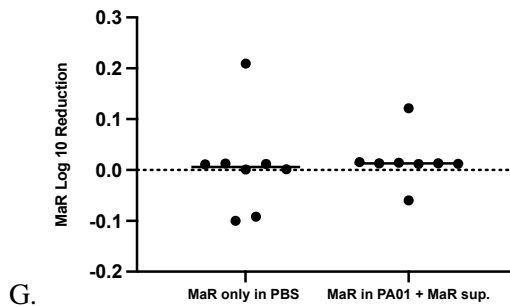
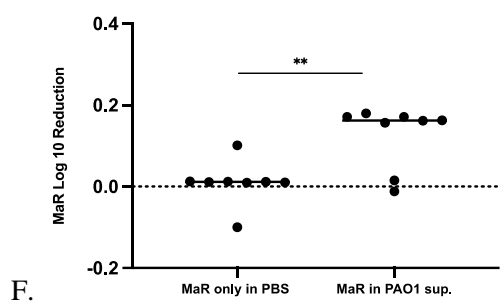


F. **MaR only with 1mL replenished** **MaR only with 2mL replenished**  
**MaR + PAO1 with 1mL replenished** **MaR + PAO1 with 2mL replenished**



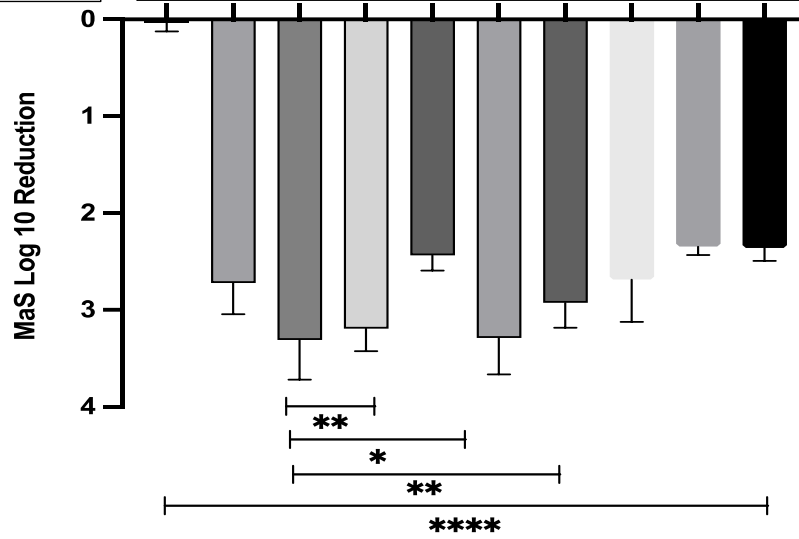
**Figure S1.** (A) Log<sub>10</sub> CFU/mL of MaS grown alone versus in co-culture with PAO1 for 72 hrs. There was no statistical significance between the two conditions. (B) Log<sub>10</sub> CFU/mL of MaR grown alone versus in co-culture with PAO1 for 72 hrs. in 7H9 vs 7H9 + Tween. There was no statistical significance between the two conditions. (C) Log<sub>10</sub> CFU/mL of PAO1 grown alone versus in co-culture with MaR for 72 hrs. in 7H9 vs 7H9 + Tween. There was no statistical significance between the two conditions. (D) Log<sub>10</sub> CFU/mL of MaR at OD<sub>600</sub> of 0.150 grown alone versus in co-culture with PAO1 at different starting OD<sub>600</sub> values of 0.05, 0.150 and 0.5 for 72 hrs. There was no statistical significance between the two conditions. (E) Log<sub>10</sub> CFU/mL of PAO1 at different starting OD<sub>600</sub> values of 0.05, 0.150 and 0.5 grown alone versus in co-culture with MaR at OD<sub>600</sub> of 0.150 for 72 hrs. There was no statistical significance between the two conditions. (F) Log<sub>10</sub> CFU/mL of MaR grown alone versus in co-culture with PAO1 for 72 hrs. with 1mL vs. 2mL media replenished every 24hrs. There was no statistical significance between the two conditions. (G) Log<sub>10</sub> CFU/mL of PAO1 grown alone versus in co-culture with MaR for 72 hrs. with 1mL vs. 2mL media replenished every 24hrs. There was no statistical significance between the two conditions. N=2 for all data shown.





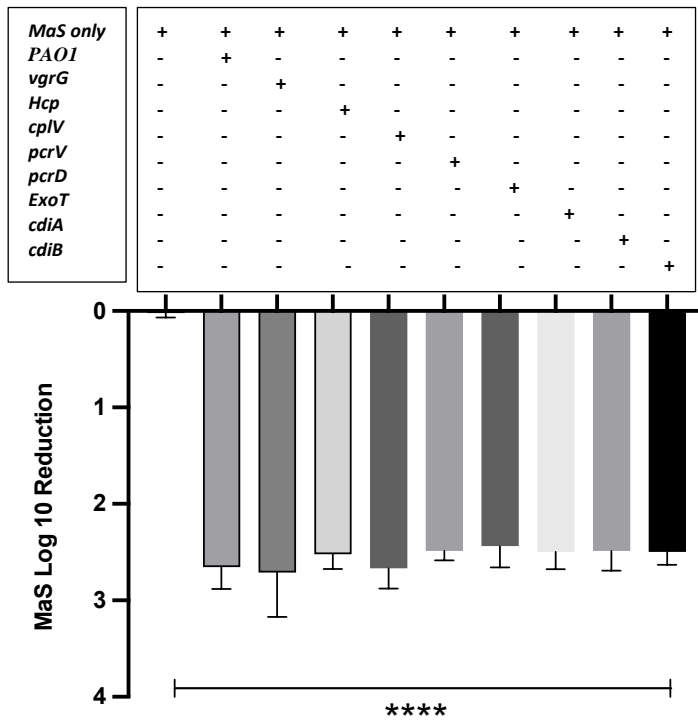
**Figure S2.** (A) Log<sub>10</sub> CFU graph of heat-killed *P. aeruginosa* biofilm with MaR and MaS. Two filters were placed on each plate with two biological replicates. There was no statistical significance between the two conditions. (B). Log<sub>10</sub> reduction graph of MaR, MaS with *P. aeruginosa* biofilm in six-well plate in the absence of a membrane filter. There were two technical and biological replicates for each condition (N=2). Unpaired t-test \*\*\*\* P<0.0001. (C) Log<sub>10</sub> reduction graph of MaS with PAO1 biofilm at MOI of 1:1 and 100:1 respectively. Two filters were placed on each plate with two biological replicates (N=2). Unpaired t-test \*\*\*\* P<0.0001. (D) Log<sub>10</sub> reduction graph of MaR with PAO1 biofilm at MOI of 1:1 and 100:1 respectively. Two filters were placed on each plate with two biological replicates (N=2). Unpaired t-test \*\*\*\* P<0.0001. (E) Log<sub>10</sub> reduction CFU graph of kinetic biofilm over 6 days comparing *P. aeruginosa* growth in single-species biofilm to dual-species biofilm with MaS. Two filters were placed on each plate with three biological replicates. Unpaired T-test, \*\*\* P <0.001, \*\*\*\* P<0.0001 (N=2). (F) Log<sub>10</sub> reduction graph of MaR grown in a single species biofilm compared to growth in the presence of diluted PAO1 supernatant (N=2). Unpaired t-test \*\* P=00034. (G) Log<sub>10</sub> reduction graph MaR grown in a single species biofilm compared to growth in the presence of diluted PAO1 and MaR supernatant (N=2). There is no statistical significance. (H) Log<sub>10</sub> reduction graph of MaS grown in a single species biofilm compared to growth in the presence of diluted PAO1 supernatant (N=2). There was no statistical significance. (I) Log<sub>10</sub> reduction graph MaS grown in a single species biofilm compared to growth in the presence of diluted PAO1 and MaS supernatant (N=2). There was no statistical significance. (J) Antagonism of MaR was observed by either scraping an agar plate to dilute to the desired OD or preparing an overnight culture diluted to the desired OD. Two filters were placed on each plate with two biological replicates (N=2). Unpaired t-test \*\*\*\* P<0.0001.

<i>MaS only</i>	+	+	+	+	+	+	+	+	+	+
<i>PAO1</i>	-	+	-	-	-	-	-	-	-	-
<i>pqsA</i>	-	-	+	-	-	-	-	-	-	-
<i>pqsH</i>	-	-	-	+	-	-	-	-	-	-
<i>pqsL</i>	-	-	-	-	+	-	-	-	-	-
<i>lasI</i>	-	-	-	-	-	+	-	-	-	-
<i>rhII</i>	-	-	-	-	-	-	+	-	-	-
<i>lasIrhII</i>	-	-	-	-	-	-	-	+	-	-
<i>XcpR</i>	-	-	-	-	-	-	-	-	+	-
<i>XcpQ</i>	-	-	-	-	-	-	-	-	-	+

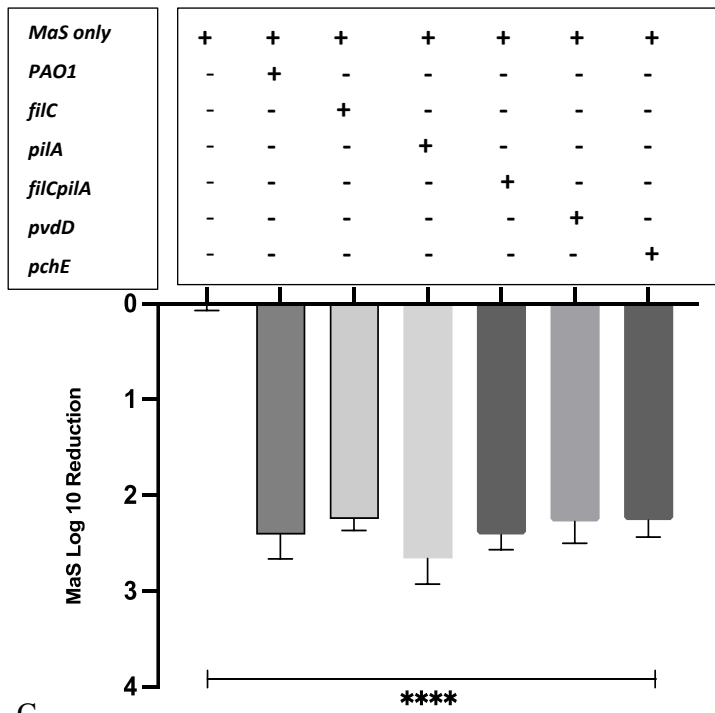


A.



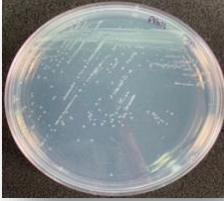

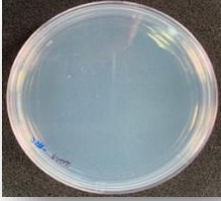
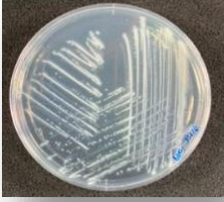




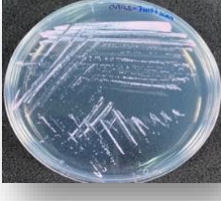


B.

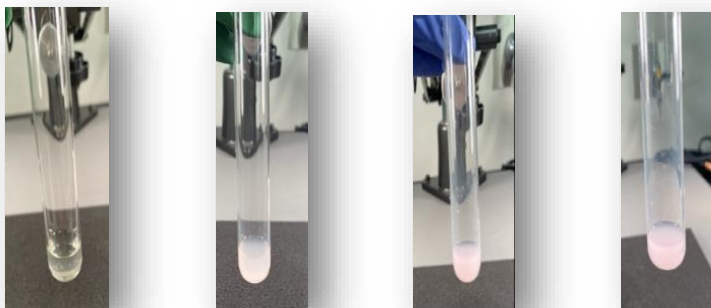


C.

**Figure S3.** (A) Log<sub>10</sub> reduction graph of MaS single-species biofilm growth compared to dual-species biofilm with *P. aeruginosa* QS (*lasI*, *rhII*, and *lasIRhII*), PQS biosynthesis mutants (*pqsA*, *pqsL*, and *pqsH*) and T2SS mutants (*xcpR* and *xcpQ*). (N=3). Unpaired t-test \*\*\*\* P<0.0001 (Unpaired t-test comparing PAO1 to *pqsA*, *pqsH* and, *lasI* mutants\* P=0.0112, \*\* P=0.0083, and \*P=0.0105 respectively). (B) Log<sub>10</sub> reduction graph of MaS single-species biofilm growth compared to dual-species biofilm with *P. aeruginosa* T6SS mutants (*vgrG*, *hcp*, and *cplV*), T3SS (*pcrV*, *pcrD*, and *exoT*) and CDI mutants ( $\Delta cdiA_{PA0041}$  and  $\Delta cdiB_{PA2463}$ ). (N=2). Unpaired t-test \*\*\*\* P<0.0001. (C) Log<sub>10</sub> reduction of MaS single-species biofilm growth compared to dual-species biofilm with *P. aeruginosa* motility mutants (*fliC*, *pilA*, and *fliCpilA*) and iron sequestering mutants (*pvdD* and *pchE*) (N=2). Unpaired t-test \*\*\*\* P<0.0001. All experiments evaluated two technical replicates and two biological replicates except for with three biological replicates.

	PAO1	MaR	MaS
PIA			
7H10			
7H10 + Kanamycin			

A.



B

0h	24h	48h	72h
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**Figure S4.** (A) MaS, MaR and PAO1 streaked on PIA, 7H10 plate and 7H10 + Kan plates. PIA plates incubated for 1 day since CFUs are counted on PIA plates are after 1 day of incubation and 7H10 and 7H10 + Kan plates incubated for 4 days. (D) Image showing the growth of MaR in liquid culture over 72hrs. Images were taken every 24hrs.