# nature portfolio

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## **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
×	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
×	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

### Software and code

Policy information about availability of computer code

Data collection Metamorph 6.3 image acquisition software

Data analysis Analysis was conducted with a custom-wri

Analysis was conducted with a custom-written Python script (Python version 3.8, SVM algorithms are a component of the scikit-learn Python library) https://github.com/rqi14/PhaseScan

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our <u>policy</u>

Source data are provided with this paper and are available in the FigShare repository under accession code 10.6084/m9.figshare.21405129. LLPSDB: A Database of Proteins Undergoing Liquid-Liquid Phase Separation in Vitro is available at http://bio-comp.org.cn/llpsdb/home.html

earch participants				
about studies involving human	research participants and Sex and Gender in Research.			
and gender N/A				
teristics N/A				
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ation on the approval of the study p	protocol must also be provided in the manuscript.			
ecific reporting				
one below that is the best fit for	your research. If you are not sure, read the appropriate sections before making your selection.			
Behavioural & so	ocial sciences Ecological, evolutionary & environmental sciences			
the document with all sections, see <u>natu</u>	ure.com/documents/nr-reporting-summary-flat.pdf			
the number of analyzable data po for each experiment. This was fou	on was a key parameter for the methodology presented in the manuscript, sample size was determined by oints (see data exclusions) after ~5 min of droplet generation. This resulted in data sets of >1500 data points und to be a sufficient sample size since increasing the sample size further did not result in a significant change			
In image analysis of trapped micr	ereported phase behaviour (e.g. comparison of Fig. S2 and Fig. S4).  Ange analysis of trapped microdroplets, data were excluded for overlapping or touching droplets since these effects prevent quantitative			
	droplet analysis. Examples of this process are shown in Fig. S10.  We confirm that experiment replicates were successful. Example replicate experiments are shown in Fig. S2 and Fig. S4			
Droplet collection took place over a significantly longer timescale than the flow profile required to produce the phase diagram, so that microdroplets generated according to the flow profile were sampled evenly. All trapped microdroplets were imaged, and imaged and analysed equally, with no sub-sampling bias. There was thus no requirement for randomization.				
Quantification was performed using an automated analysis pipeline applied equally to all conditions and replicates for a given experiment. A subset of the data was analysed in a blinded experiment, which produced the same output as in the non-blinded analysis.				
	about studies involving human  Ind gender  N/A  N/A  N/A  Ation on the approval of the study of			

Palaeontology and archaeology Animals and other organisms

**x** Dual use research of concern

Clinical data

## Eukaryotic cell lines

Policy information about <u>cell lines and Sex and Gender in Research</u>				
Cell line source(s)	sf9 cell line (Expression Systems)			
Authentication	Authenticated at source			
Mycoplasma contamination	All cell lines tested negative for mycoplasma contamination			
Commonly misidentified lines (See ICLAC register)	None			