

Fig S1. The expression pattern of MULTI-seq barcodes in the E18\_P1 dataset. The red signal represents high enrichment, and the black signal represents low or negative signal.

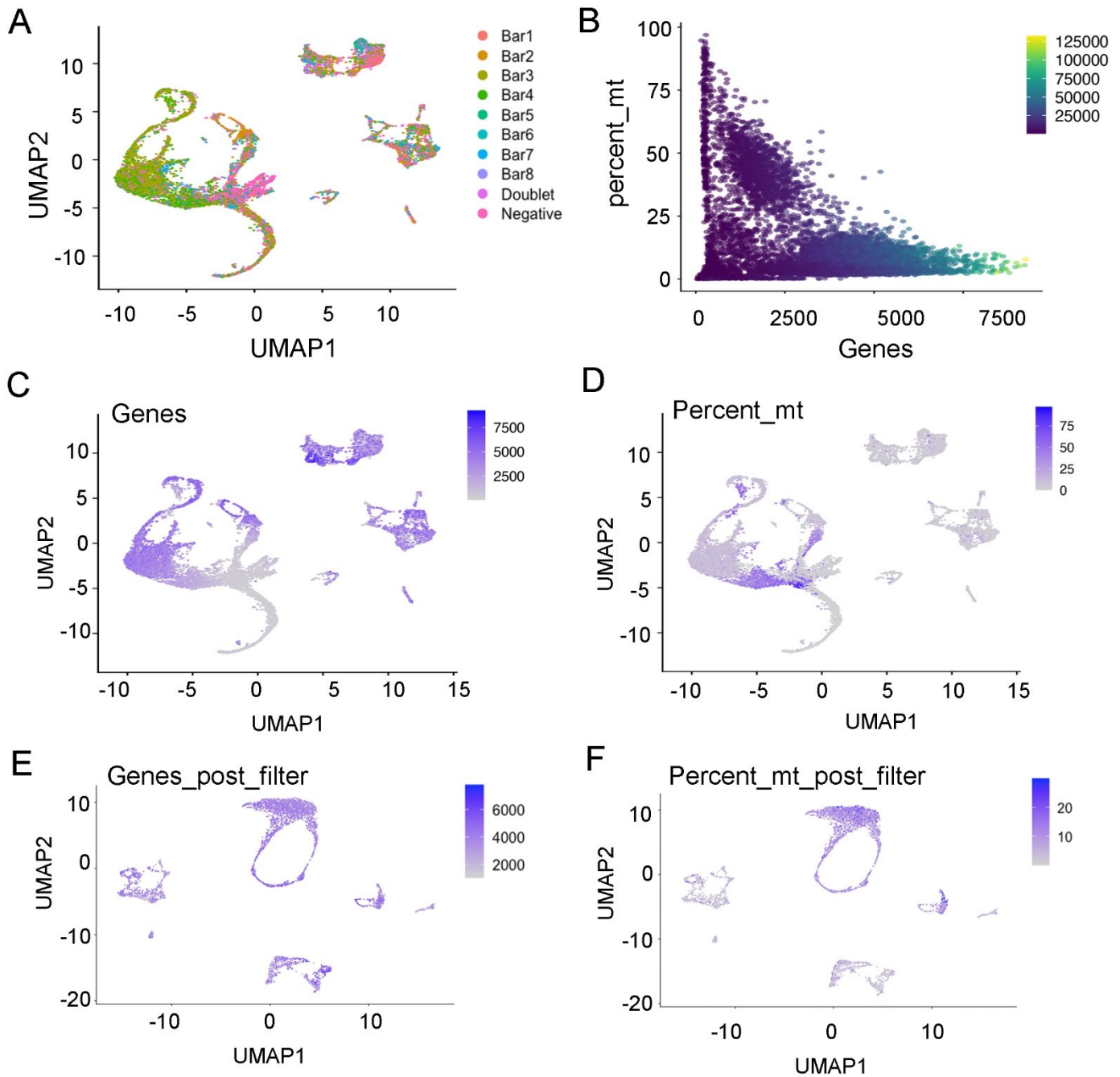


Fig S2. Quality control analysis of the E18\_P1 dataset. (A) Identification of singlets, doublets, and negative cells in the E18\_P1 dataset. (B) Analysis of the number of expressed genes (x-axis), the percentage of mitochondria genes (y-axis), and the total number of molecules detected in each cell (color). (C, D) UMAP plots of the gene numbers and mitochondria gene percentages in each single cell before filtering. (E, F) UMAP plots of the gene numbers and mitochondria gene percentages in each single cell after filtering.

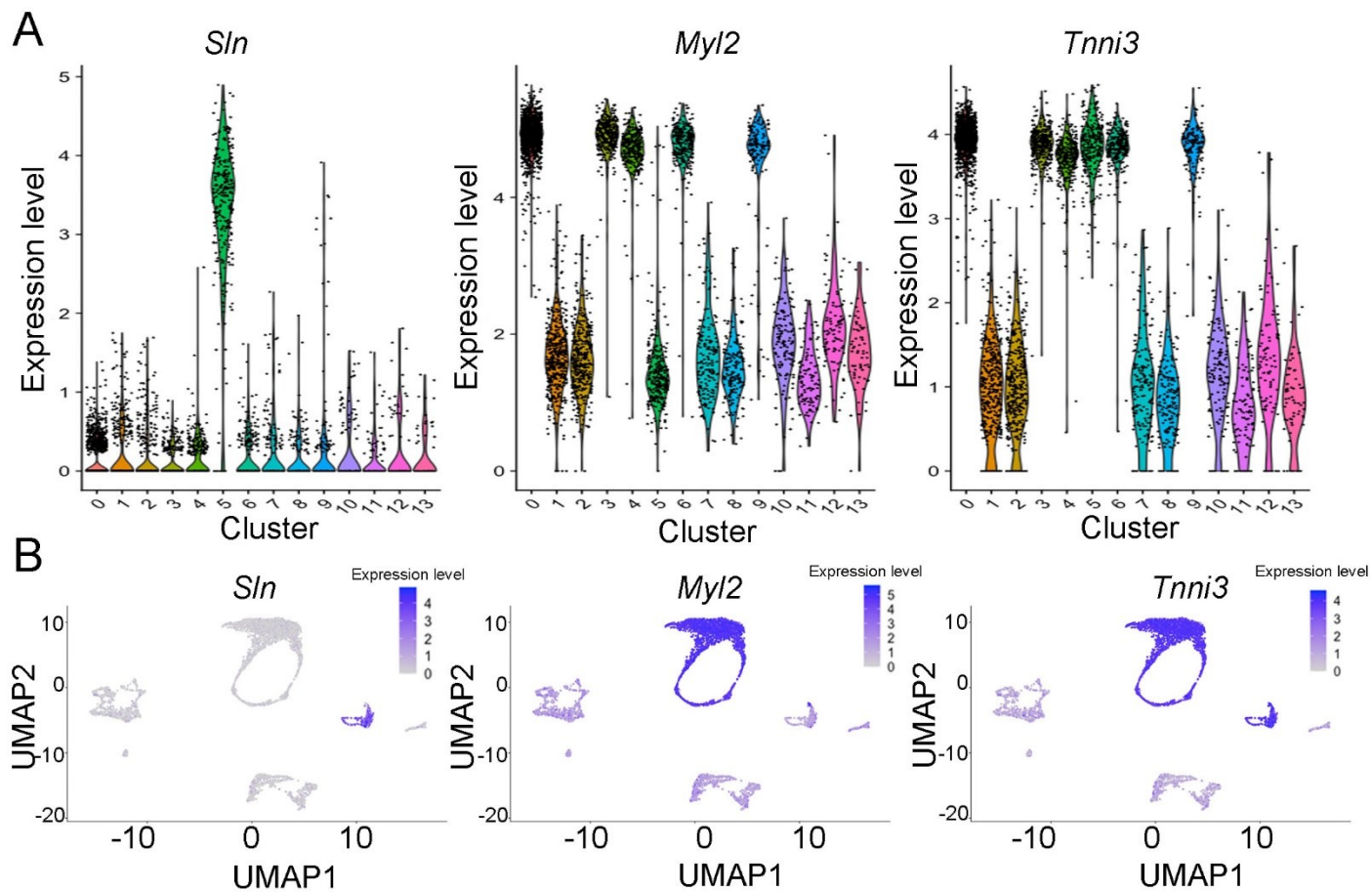


Fig S3. The expression pattern of *Sln* and *Myl2* in E18\_P1 single cells. (A, B) Violin plots and Feature plots show that the clusters with atrial and ventricular CMs (*Tnni3* positive) highly express *Sln* and *Myl2*, respectively.

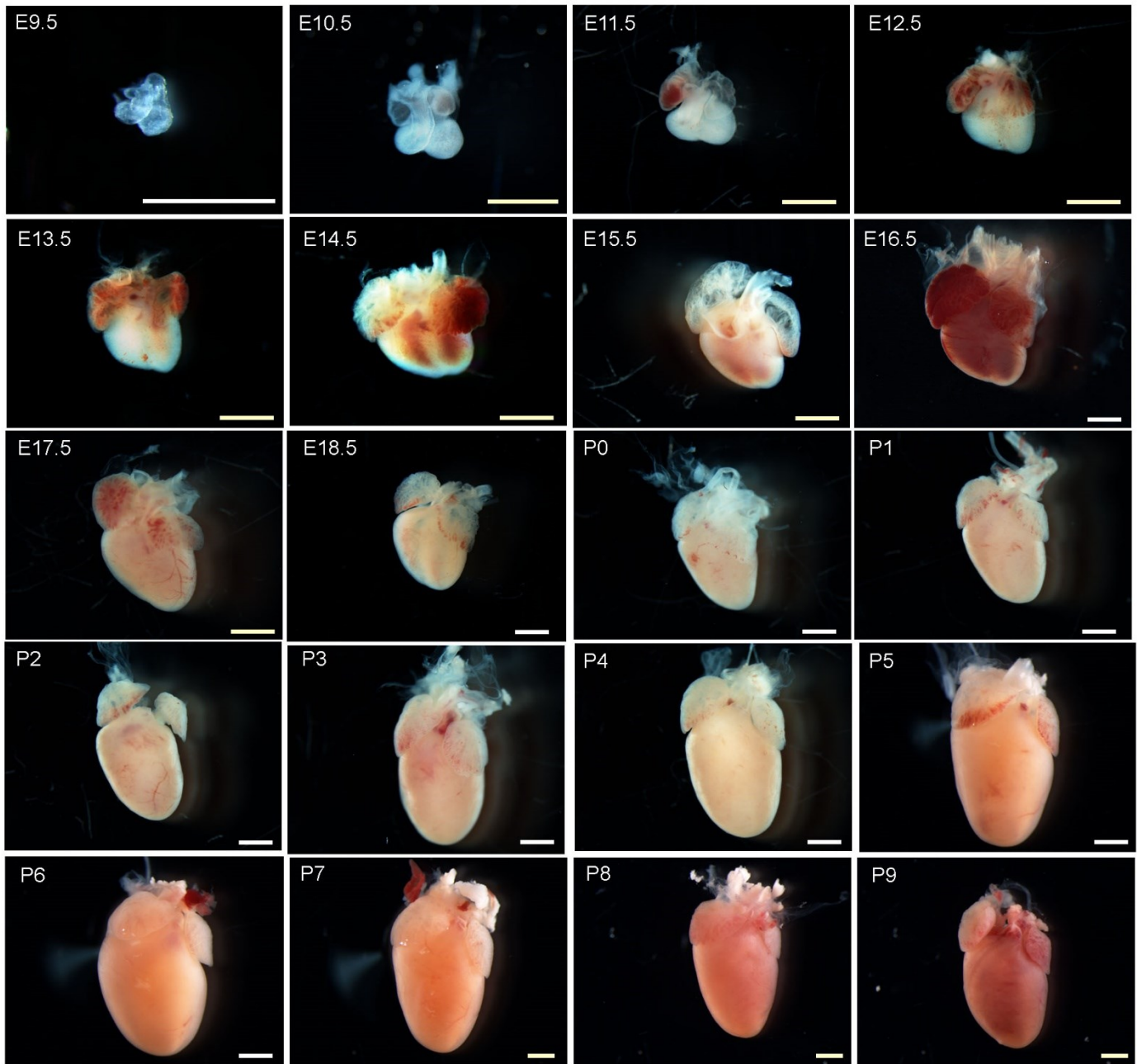


Fig S4. Representative images of staged hearts. Scale bar=1mm.



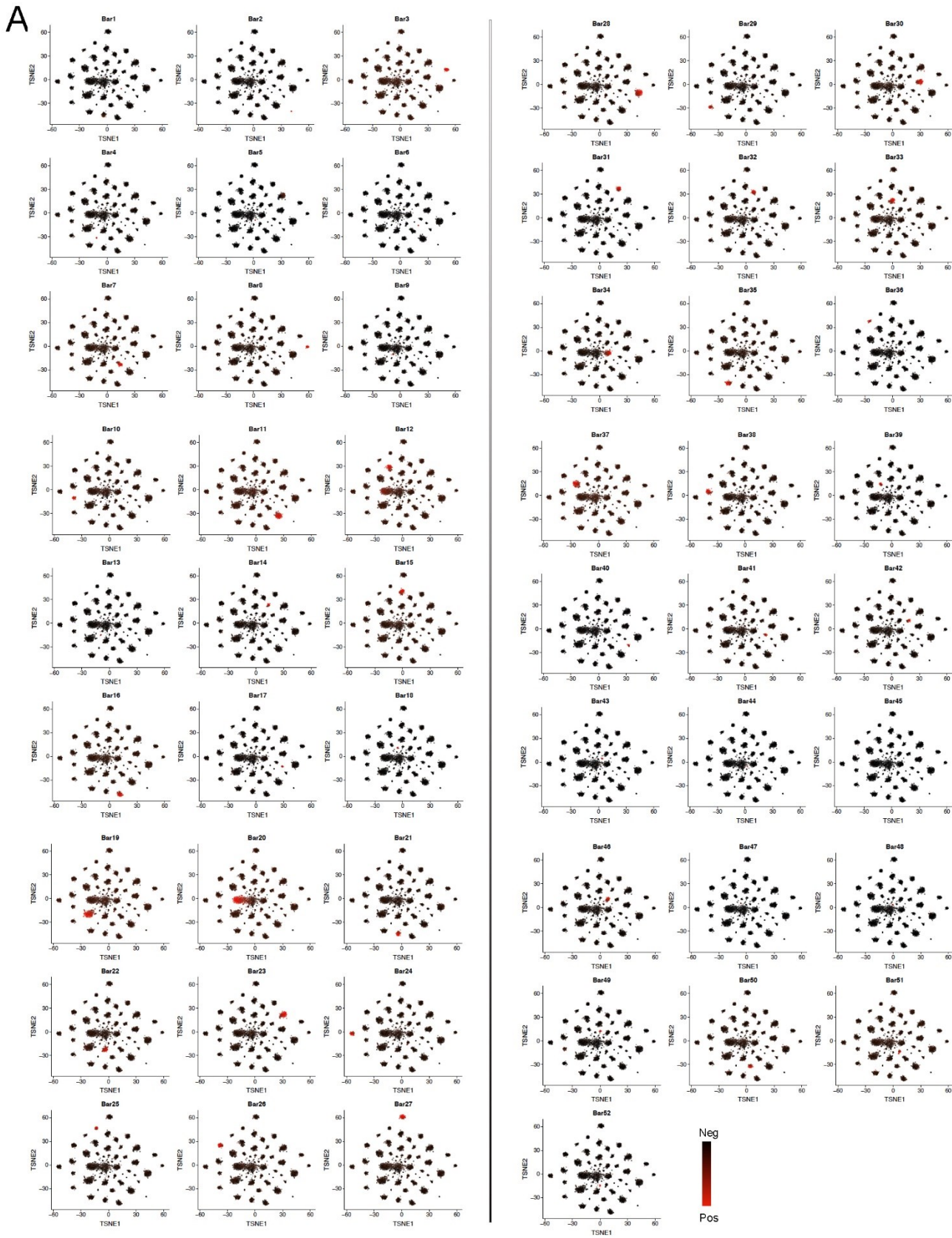


Fig S5. The expression pattern of MULTI-seq barcodes in the 5k\_1 dataset. Red color represents a positive signal, and black represents the background. Each barcode was mainly enriched in one cluster of cells.

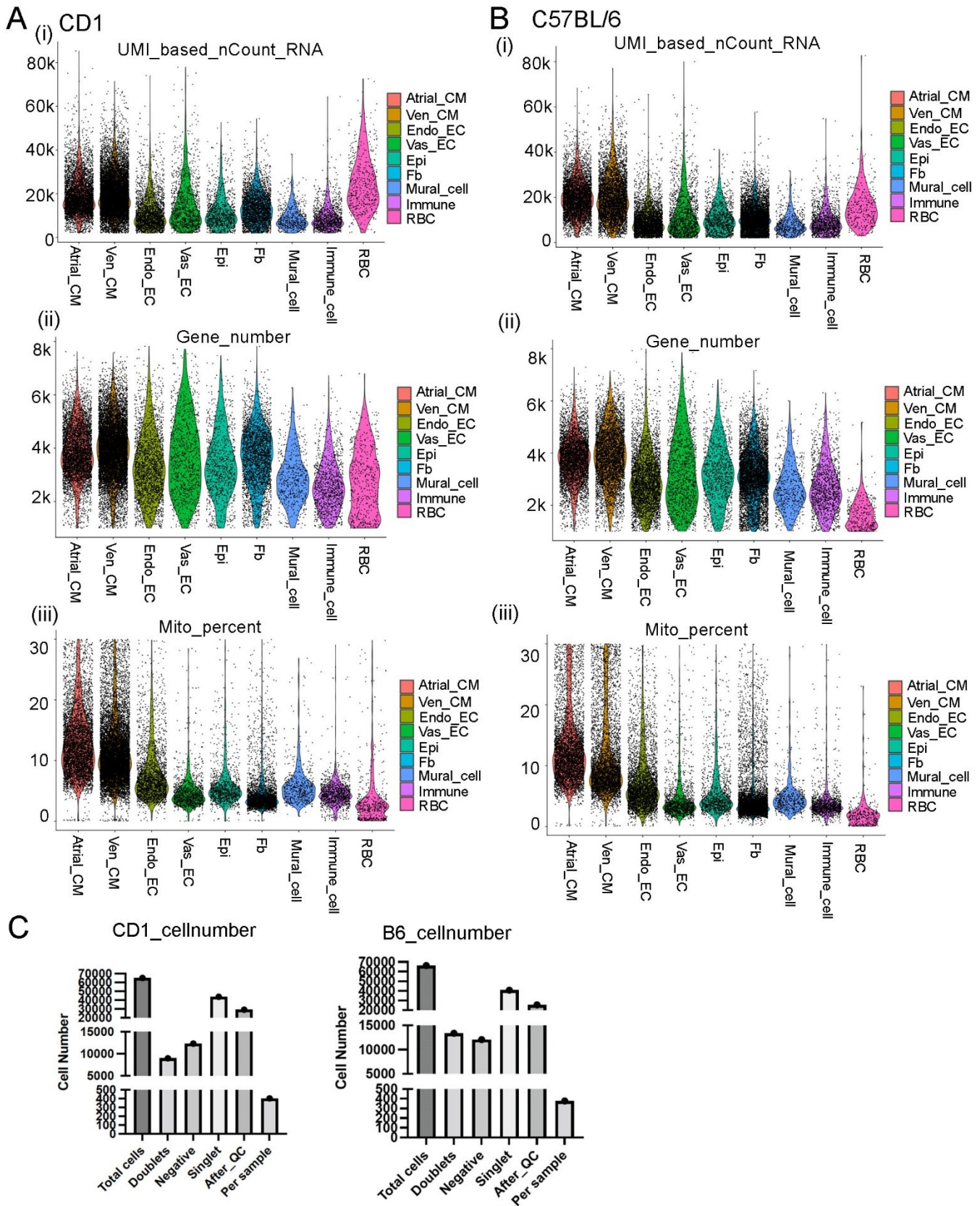


Fig S6. Quantitative analysis of the cells in CD1 and C57BL/6 datasets. (A, B) The number of sequencing reads, gene numbers, and percentage of mitochondria genes at each cell type in CD1 and C57BL/6. (C, D) The number of cells at different QC steps in CD1 and C57BL/6 datasets.



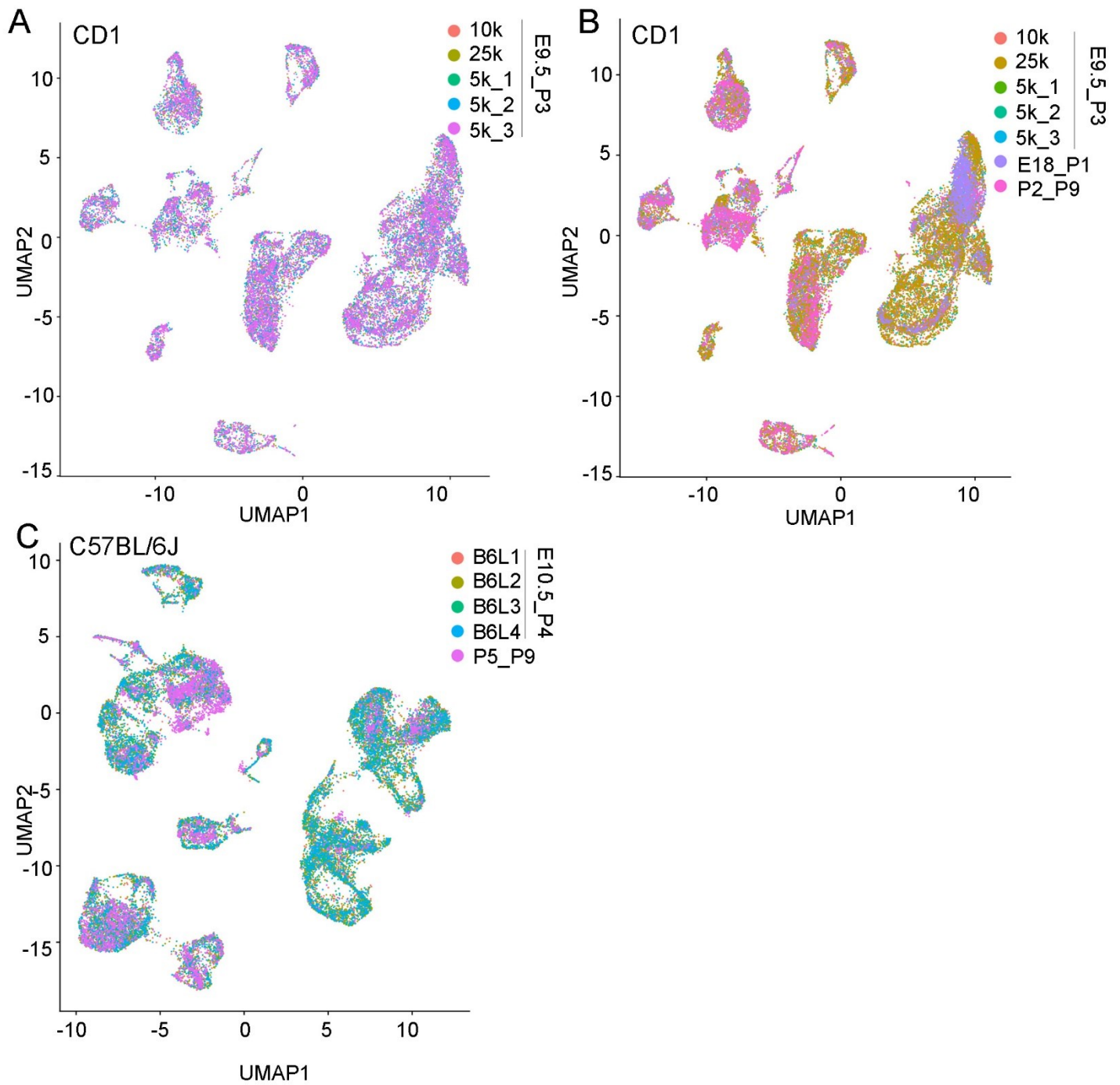


Fig S7. Integrative analysis of the single cells from different experiments. (A) The CD1 E9.5\_P3 single cells were from five different loading wells with different targeting cell numbers. (B) UMAP plot of the CD1 data from different experiments (E9.5\_P3, E18\_P1, P2\_P9). (C) The C57BL/6 single cell data profiled at different experiments. The UMAP plots were labeled by loading wells or experiments.

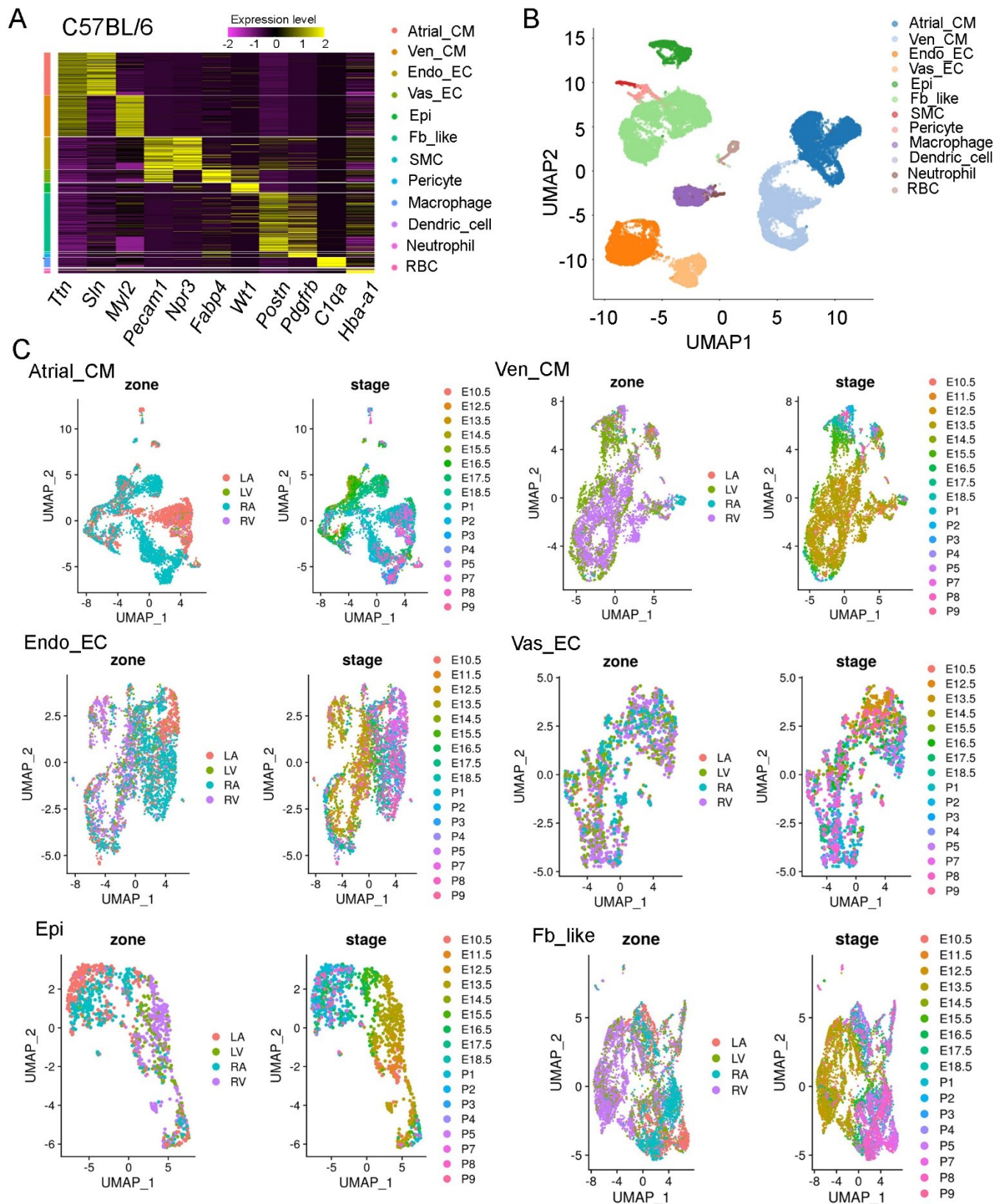


Fig S8. Analysis of the C57BL/6 single cell data. (A) Heatmap showing the expression pattern of cardiac lineage genes. (B) UMAP plot of the single cells labeled by cell types. (C) Zone and stage analysis of the cells in each cell type.





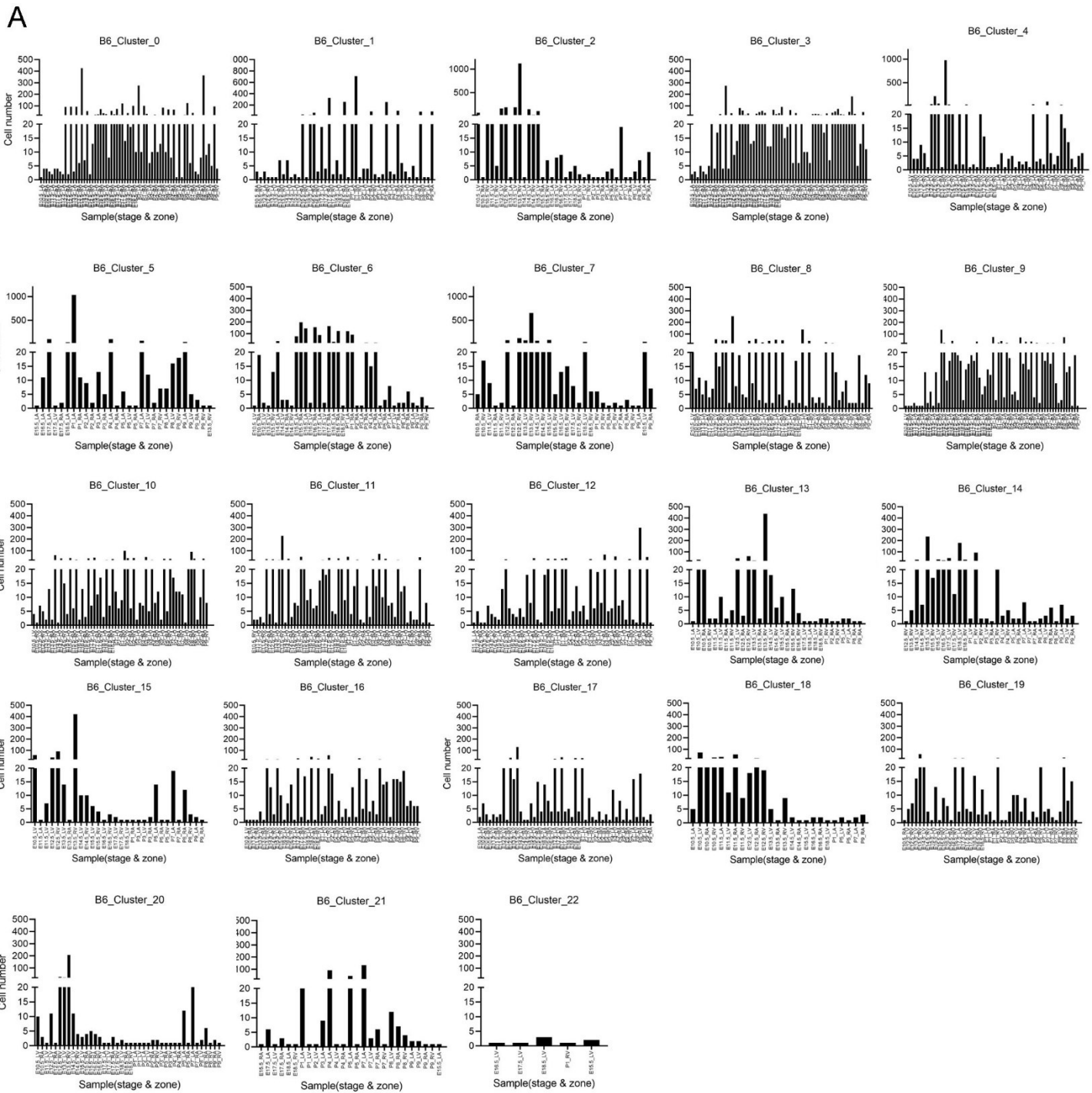


Fig S10. The number of cells in each sample at each C57BL/6 cluster.

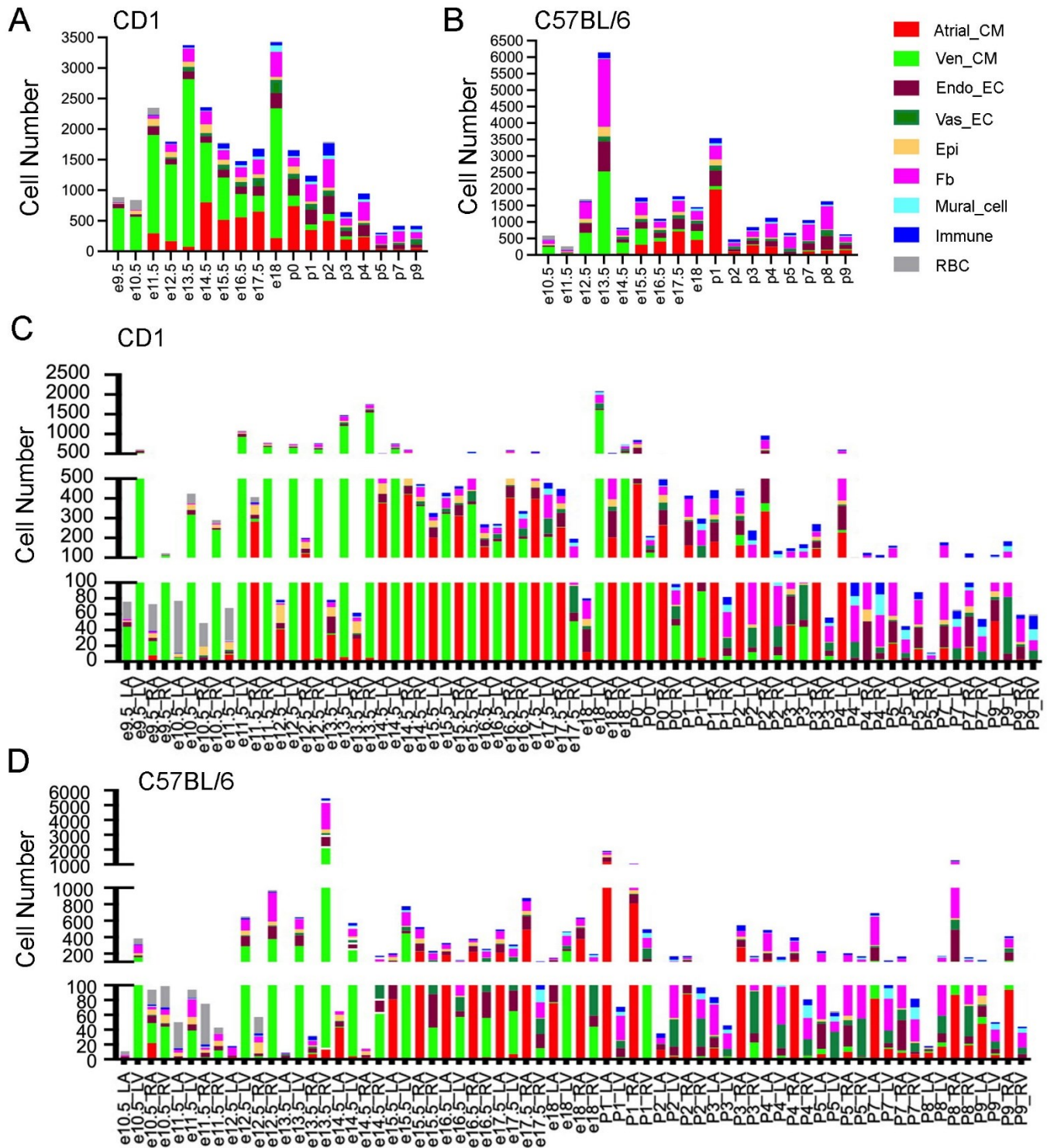


Fig S11. The cell type compositions at each stage and sample. (A, B) The cell type compositions at each stage in CD1 and C57BL/6 datasets. (C, D) The cell type compositions in each sample in CD1 and C57BL/6 datasets.

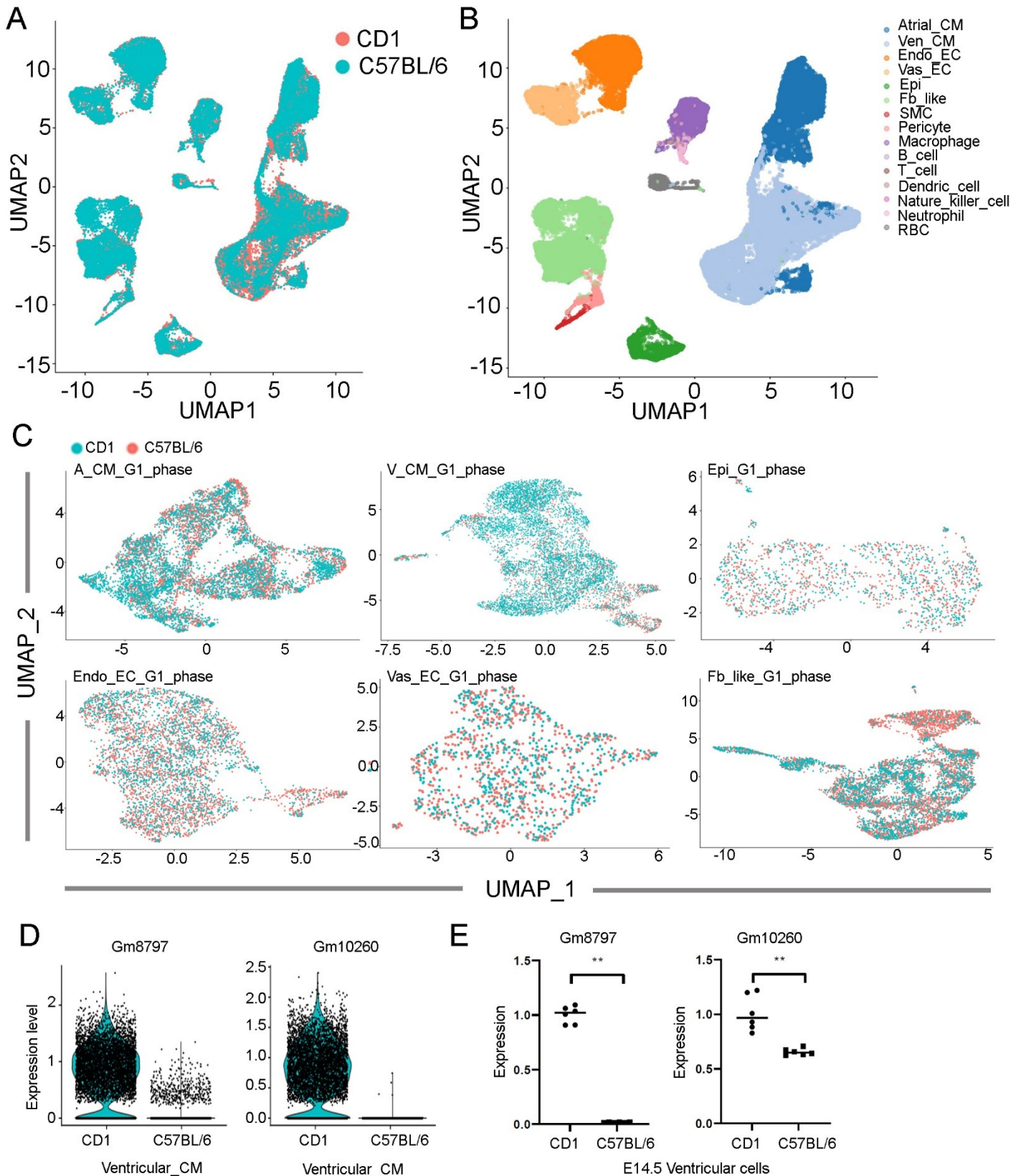


Fig S12. Comparative analysis of the CD1 and C57BL/6 single cell datasets. (A, B) UMAP plot of the integrated CD1 and C57BL/6 datasets labeled by mouse strain or cell type. (C) UMAP plot of the integrated G1 phased cells in each cell type. (D) The two representative genes differentially expressing in CD1 and C57BL/6 ventricular\_CMs. (E) qPCR analysis confirmed the differential expression of the two genes between strains. **N=2 biologically independent experiments with 3 replicates in each experiment. Student's t-test with two-tailed distribution was used for the statistical analysis.** \*\* indicates the significance with p value <0.01.



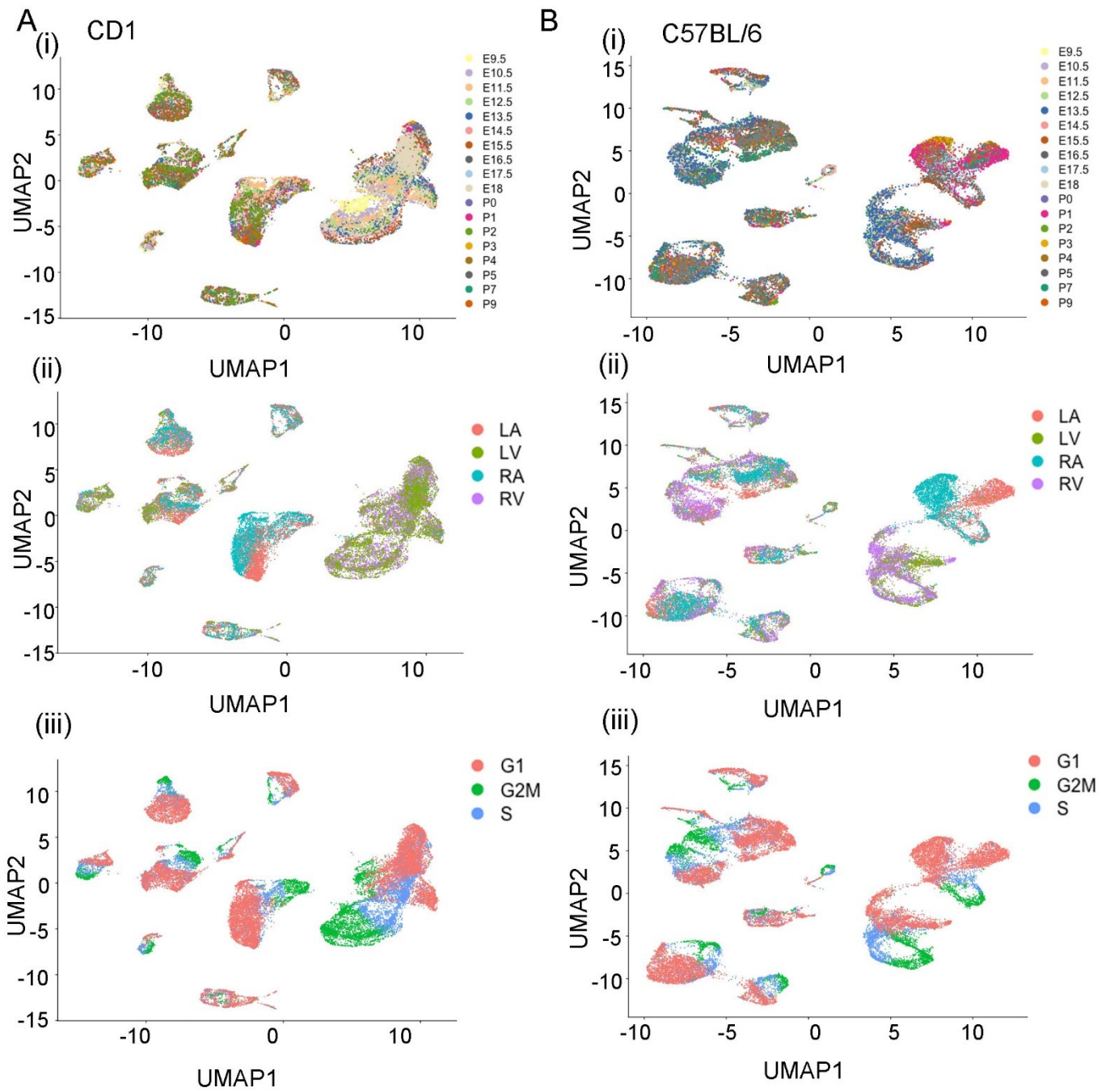


Fig S13. UMAP plots of the CD1 and C57BL/6 single cells with different labels. (A, B) The single cells from CD1 and C57BL/6 strains were labeled by stage, zone, or cell cycle phase.

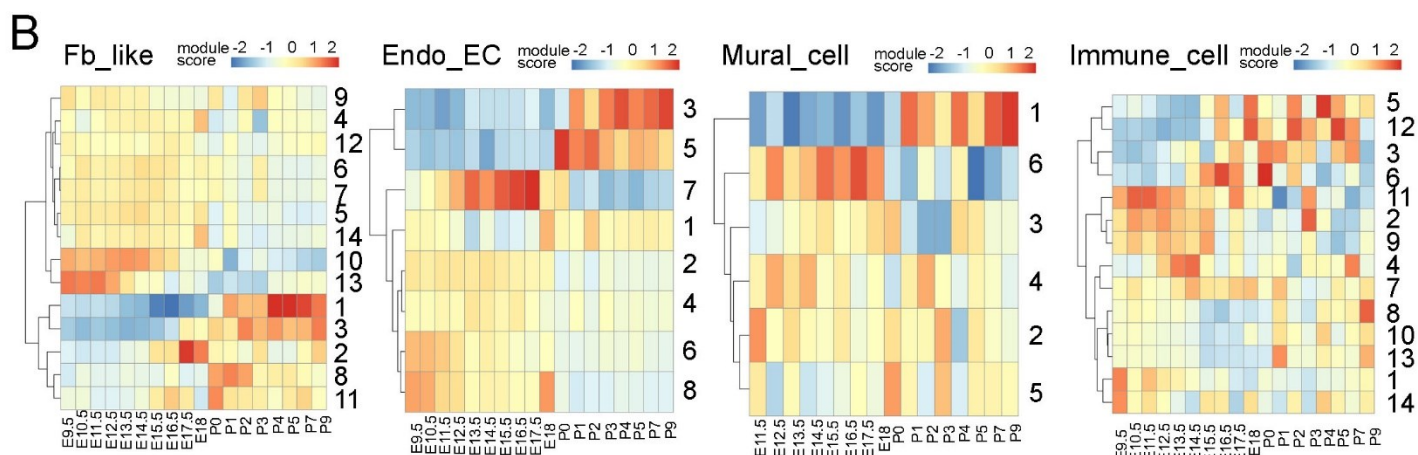
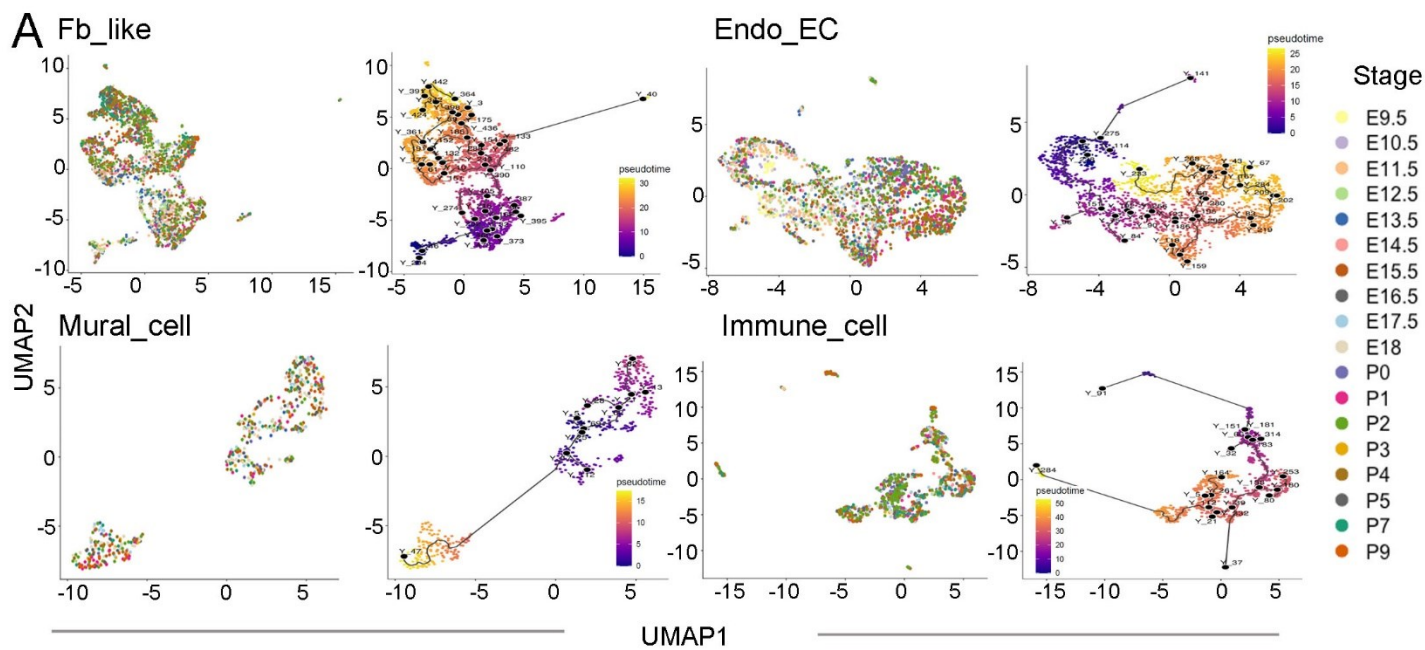


Fig S14. Stage analysis of the single cells in Fb\_like, Endo\_EC, Mural cell, and Immune cell. (A) UMAP plot of the single cells labeled by stage or pseudotime. (B) Identification of gene modules in each cell type. The color represents module score.

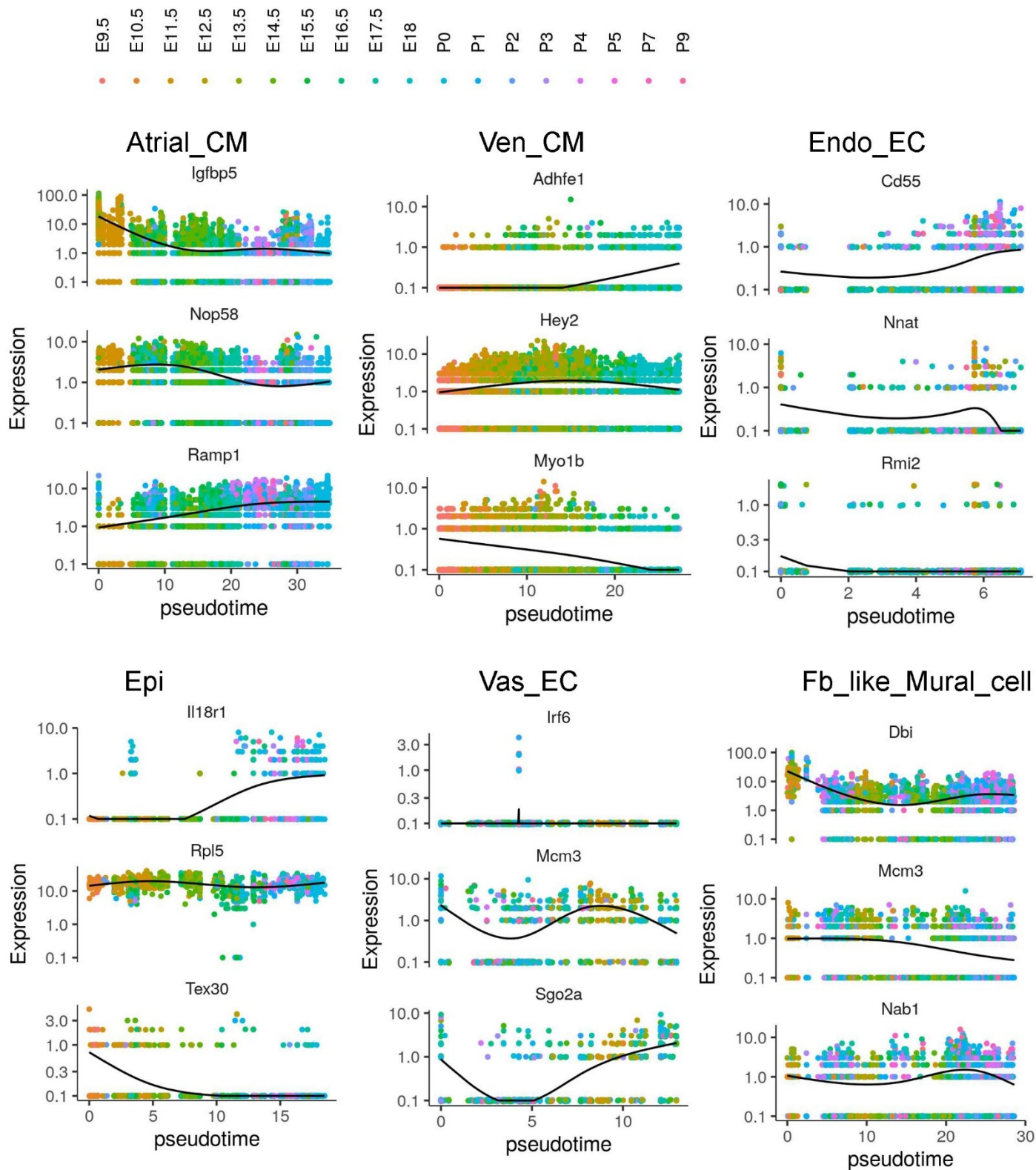


Fig S15. Representative genes that display pseudotime stage-specific expression pattern in each cell type. The color represents real stages. The cells were ordered along the x-axis by pseudotime, and the Y-axis represents gene expression level.



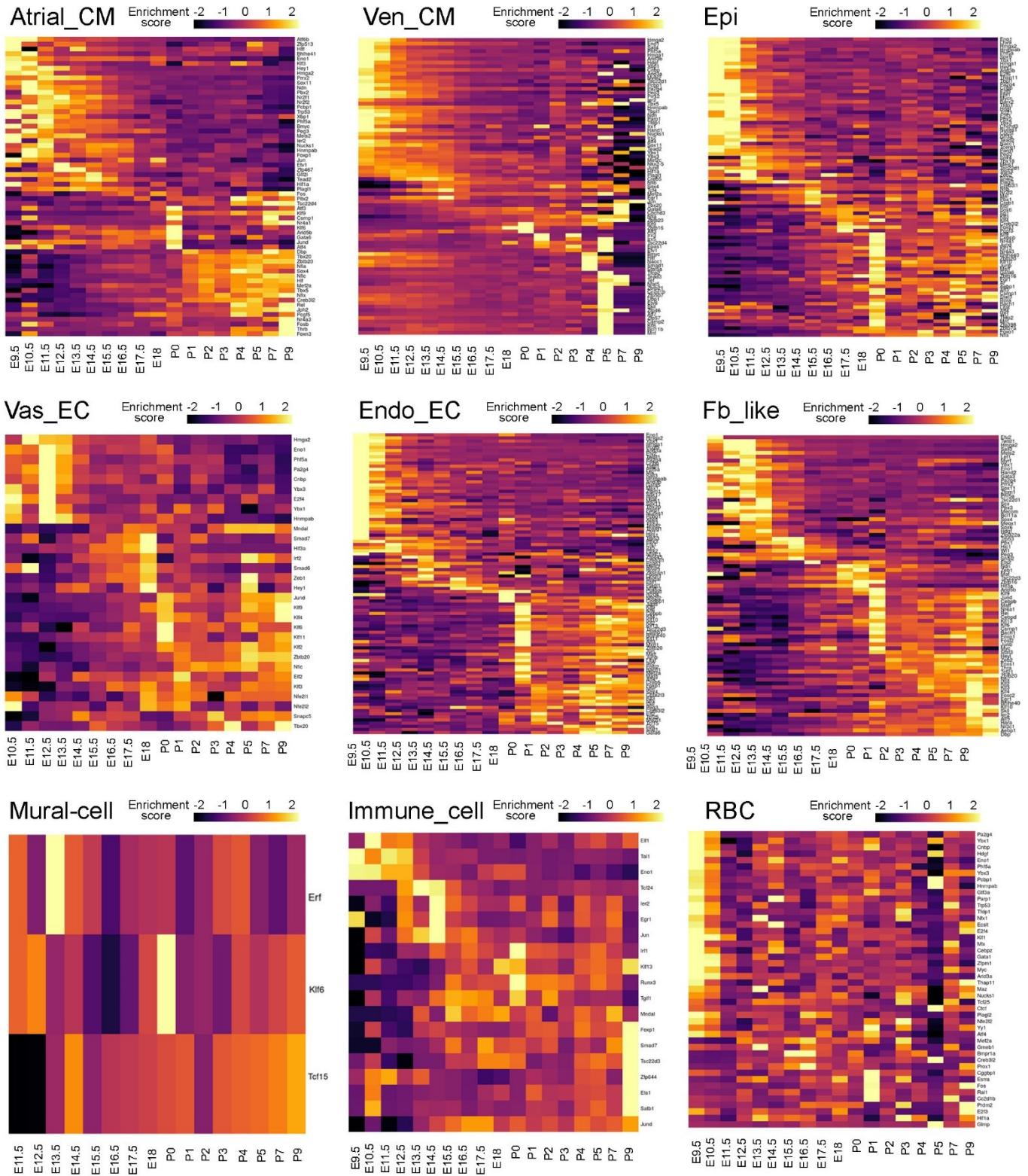


Fig S16. The stage specifically expressed transcription factors in each cell type. The gene expression level reduced from yellow to blue to black.



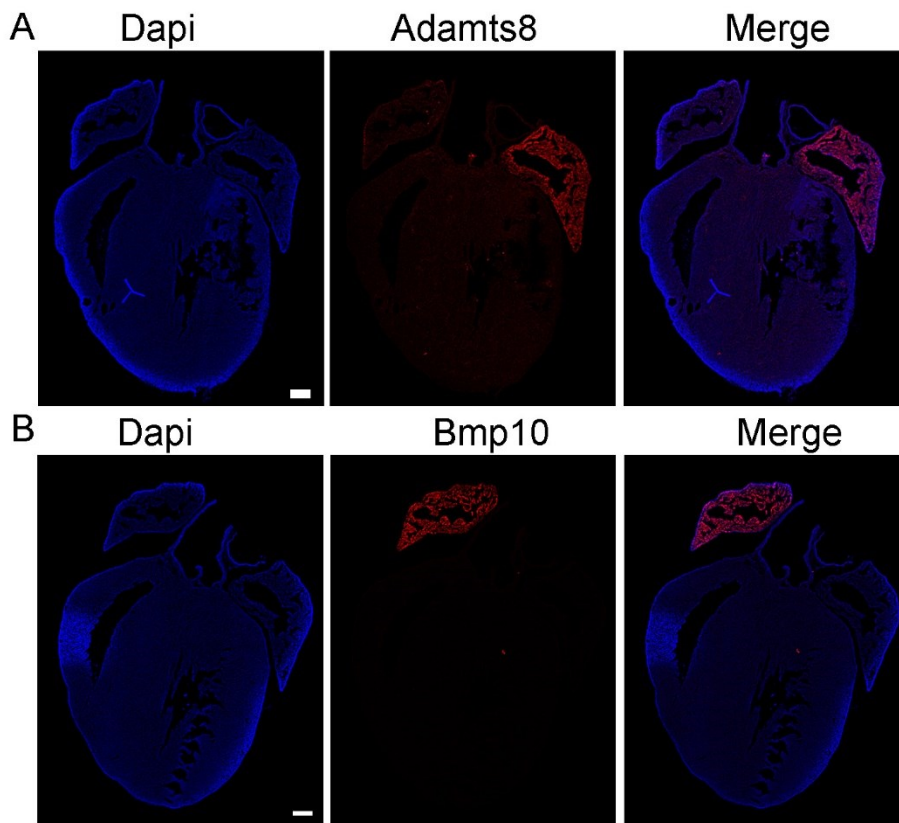


Fig S17. Staining analysis of *Adamts8* and *Bmp10*. (A, B) mRNA staining showed that *Adamts8* and *Bmp10* were specifically expressed in LA and RA at P3 hearts, respectively. The staining experiments were repeated twice with similar results. Scale bar=500  $\mu$ m.

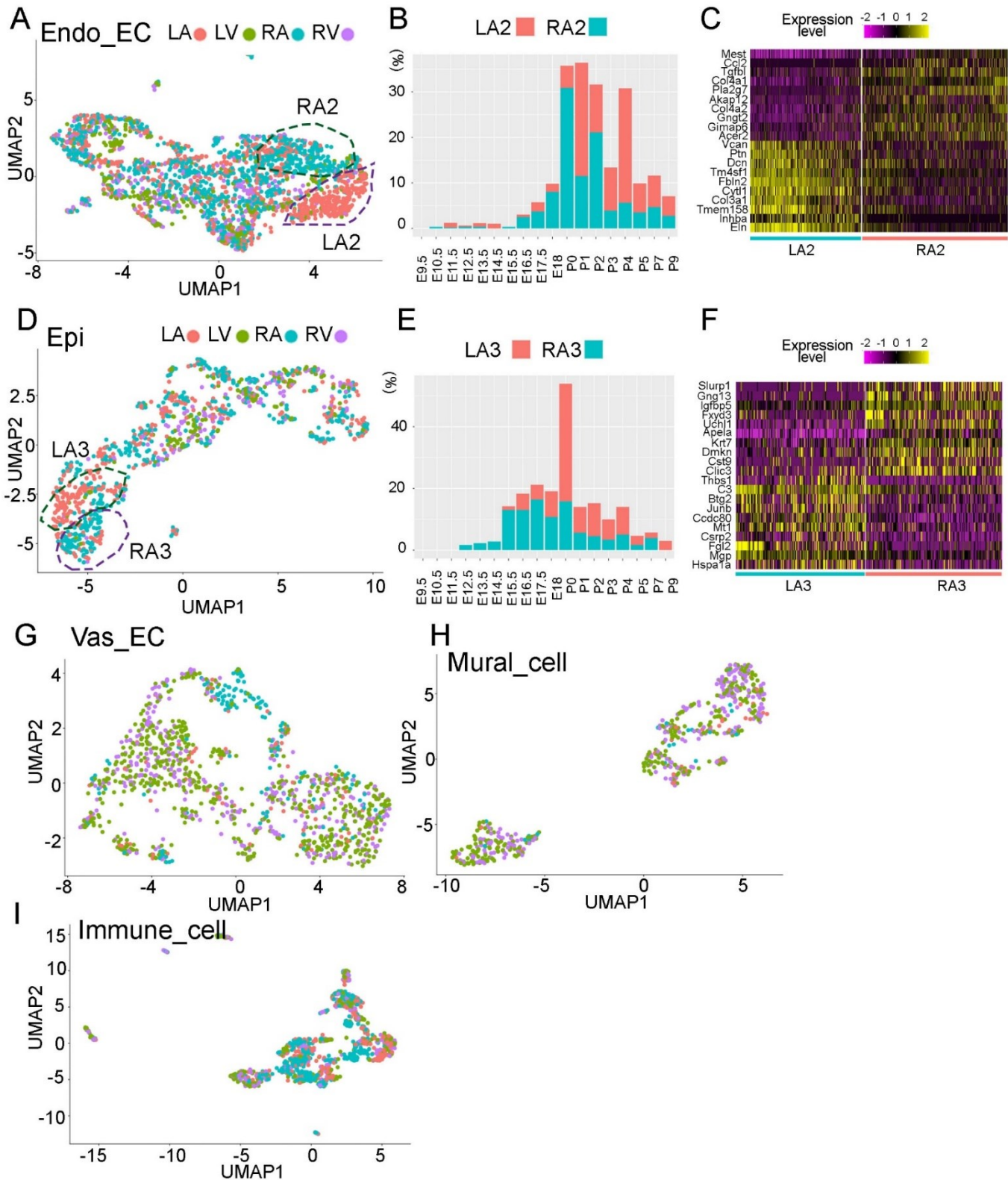


Fig S18. The zone-specific molecular signatures in five cell types. (A) UMAP plot of Endo\_EC cells labeled by zones. One LA and one RA-specific cell populations (LA2, RA2) were identified. (B) Stage analysis of the cells in LA2 and RA2. (C) Expression heatmap of the top 20 genes that were differentially expressed in LA2 and RA2 cells. (D) UMAP plot of epicardial cells labeled by zones. One LA and one RA-specific cell populations (LA3, RA3) were identified. (E) Stage analysis of the cells in LA3 and RA3. (F) Expression heatmap of the top 20 genes that were differentially expressed in LA3 and RA3 cells. (G-I) UMAP plots of Vas\_EC, Mural\_cells, and Immune cells labeled by zones.

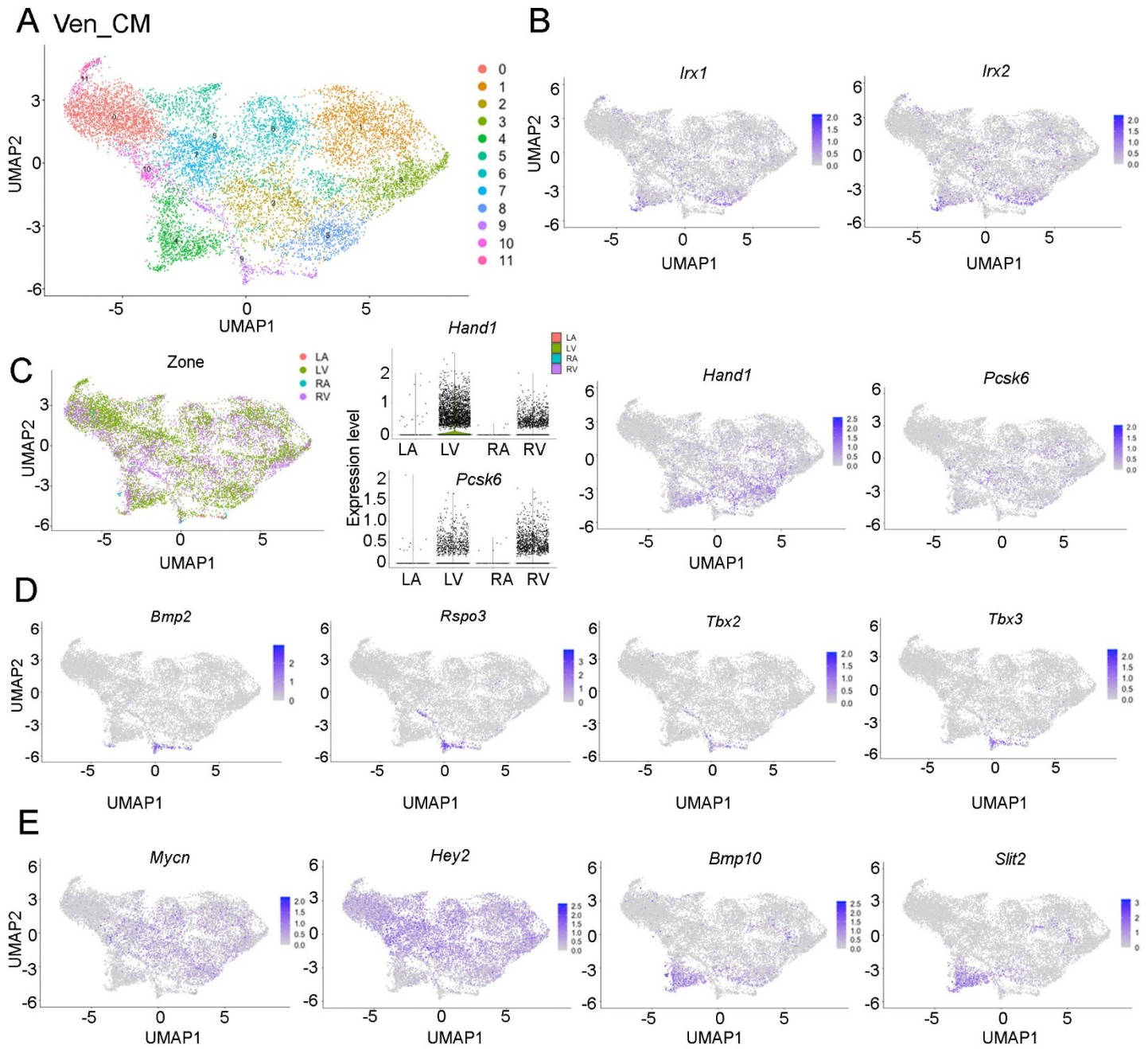
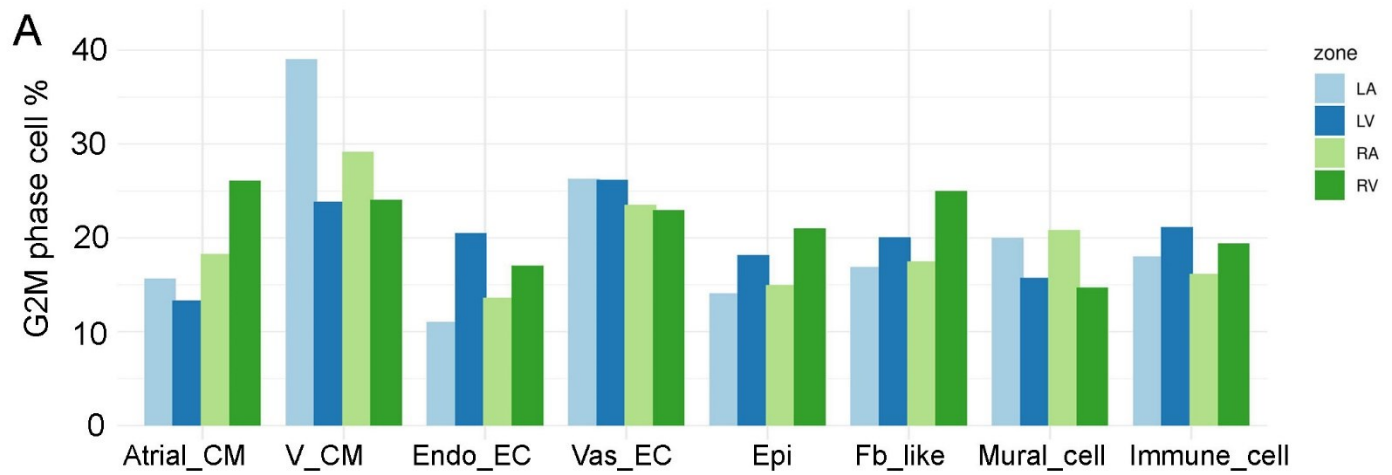


Fig S19. The cellular heterogeneity in Ven\_CM. (A) UMAP plot of Ven\_CM labeled by clusters. (B) *Irx1* and *Irx2* positive CMs were distributed in multiple cell clusters. (C) UMAP plot of Ven\_CMs labeled by zone. No LV and RV-specific clusters were identified. Violin plots showed a preferential expression of *Hand1* and *Pcsk6* in LV and RV CMs, respectively. The feature plots did not identify cluster-specific expressions of *Hand1* and *Pcsk6*. (D) The cluster 9 CMs highly expressed AVC marker genes *Bmp2*, *Rspo3*, *Tbx2*, and *Tbx3*. (E) The cluster 4 CMs highly specifically expressed trabecular myocardium genes *Bmp10* and *Slit2*, and the cells in other clusters expressed compact myocardium genes *Mycn* and *Hey2*.





**B** pHH3 staining

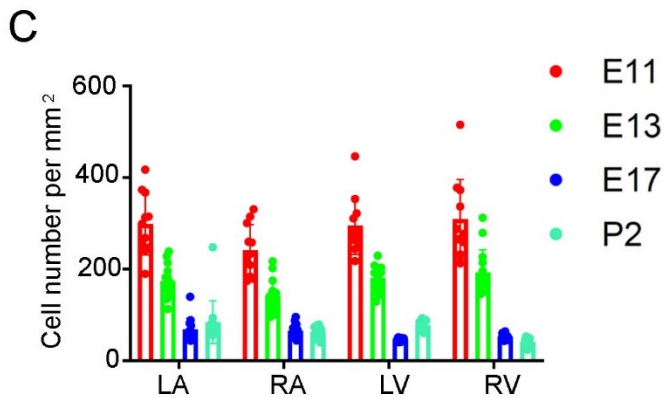
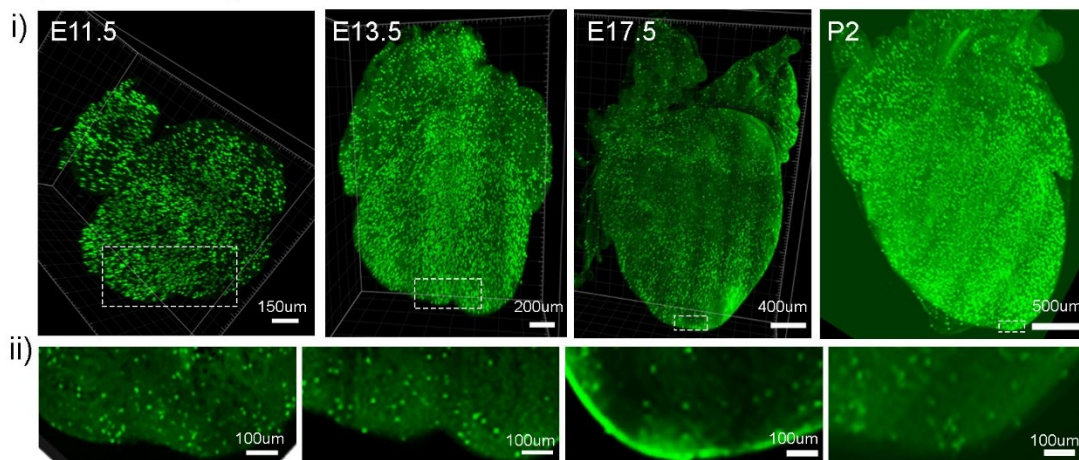


Fig S20. The proportion of cells in proliferation at different stages and zones. (A) The percentages of G2M phased cells in each cell type at each chamber. (B) (i) Overview and (ii) enlarged portion of cleared hearts with pHH3 staining at four stages. (C) The pHH3 positive cells declined along developmental progression in all four chambers. iDISCO images scale bar: E11.5 = 150  $\mu$ m; E13.5 = 200  $\mu$ m; E17.5 = 400  $\mu$ m; P2 = 500  $\mu$ m. **N= around 10 section images from one cleared heart at each stage were analyzed. The error bars represent SD. ANOVA with Tukey's multiple comparisons were used for the statistical analysis.** All comparisons except the E17 and P2 pair are statistical significant with p value < 0.01.



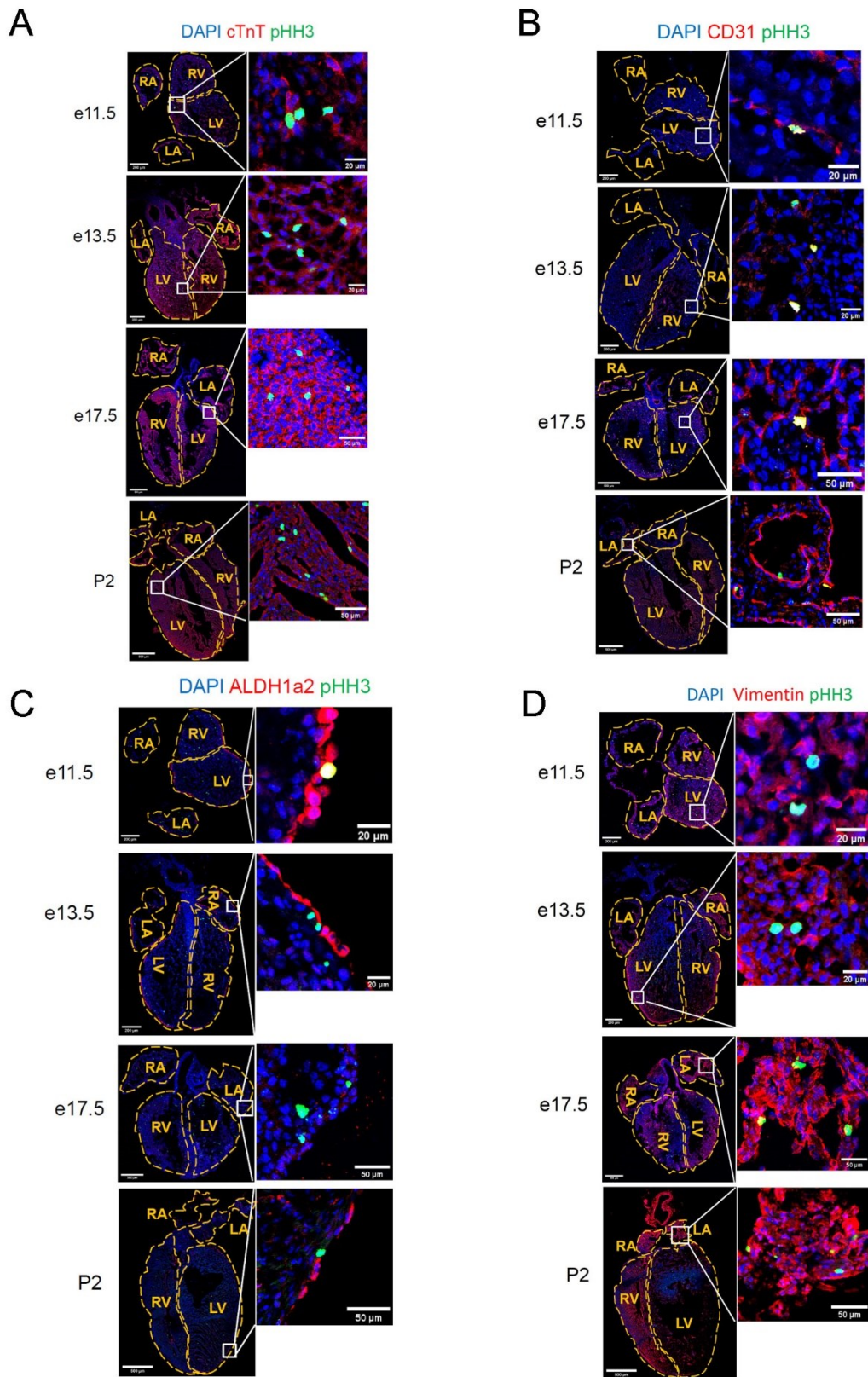


Fig S21. Representative images staining for pHH3 together with (A) CM lineage gene cTNT, (B) endothelial cell gene CD31, (C) epicardial cell gene ALDH1A2, and (D) fibroblast gene VIM. Scale bar=200 $\mu$ m in the images with whole heart sections at E11.5 and E13.5. Scale bar=500 $\mu$ m in the images with whole heart sections at E17.5 and P2. Scale bar=20 $\mu$ m in the enlarged images at E11.5 and E13.5. Scale bar=50 $\mu$ m in the E17.5 and P2 enlarged images. **The staining experiments were repeated on three heart sections with similar results.**

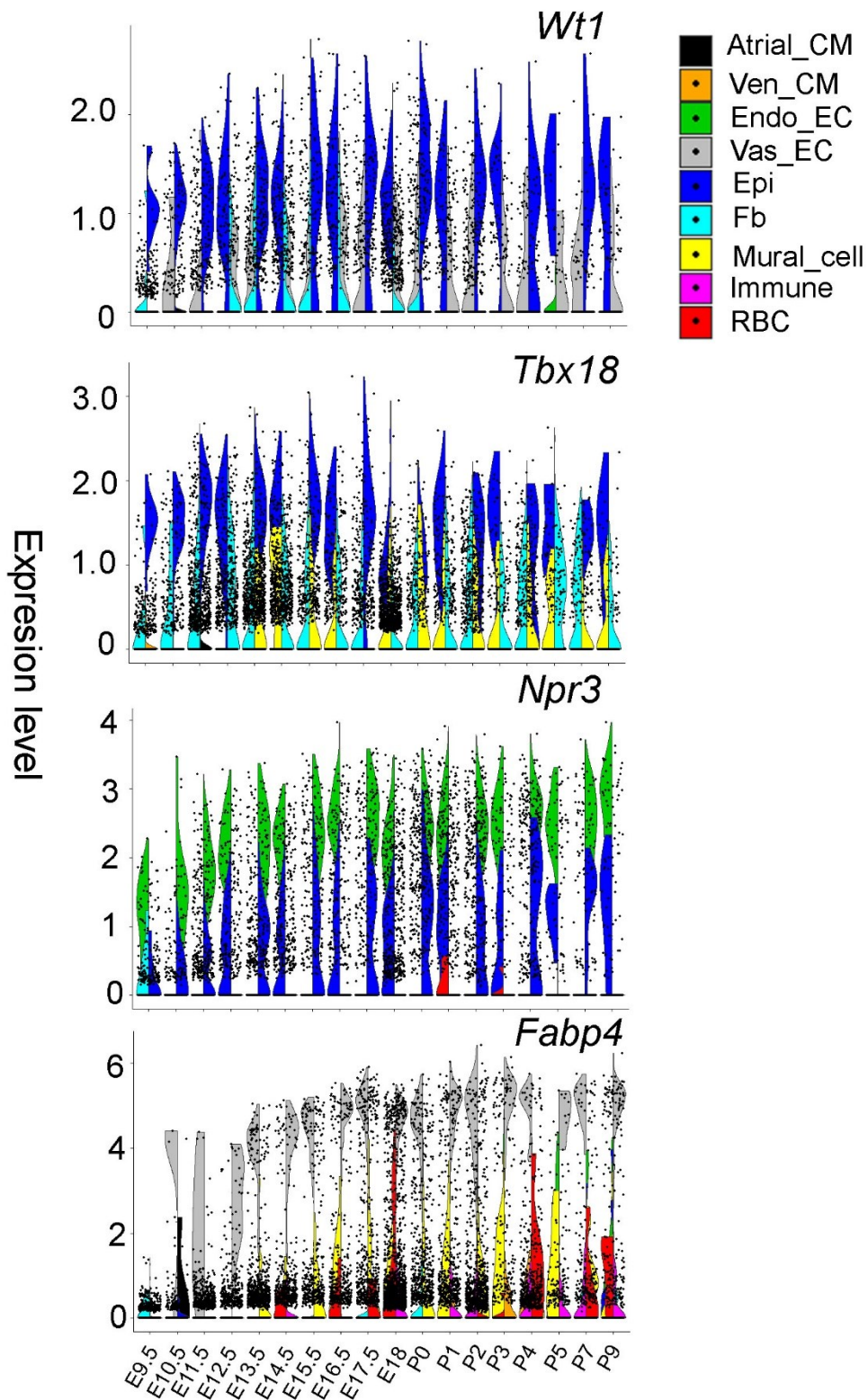


Fig S22. Violin plots showing the expression of genes *Wt1*, *Tbx18*, *Npr3*, and *Fabp4* in each cell type across the stages.



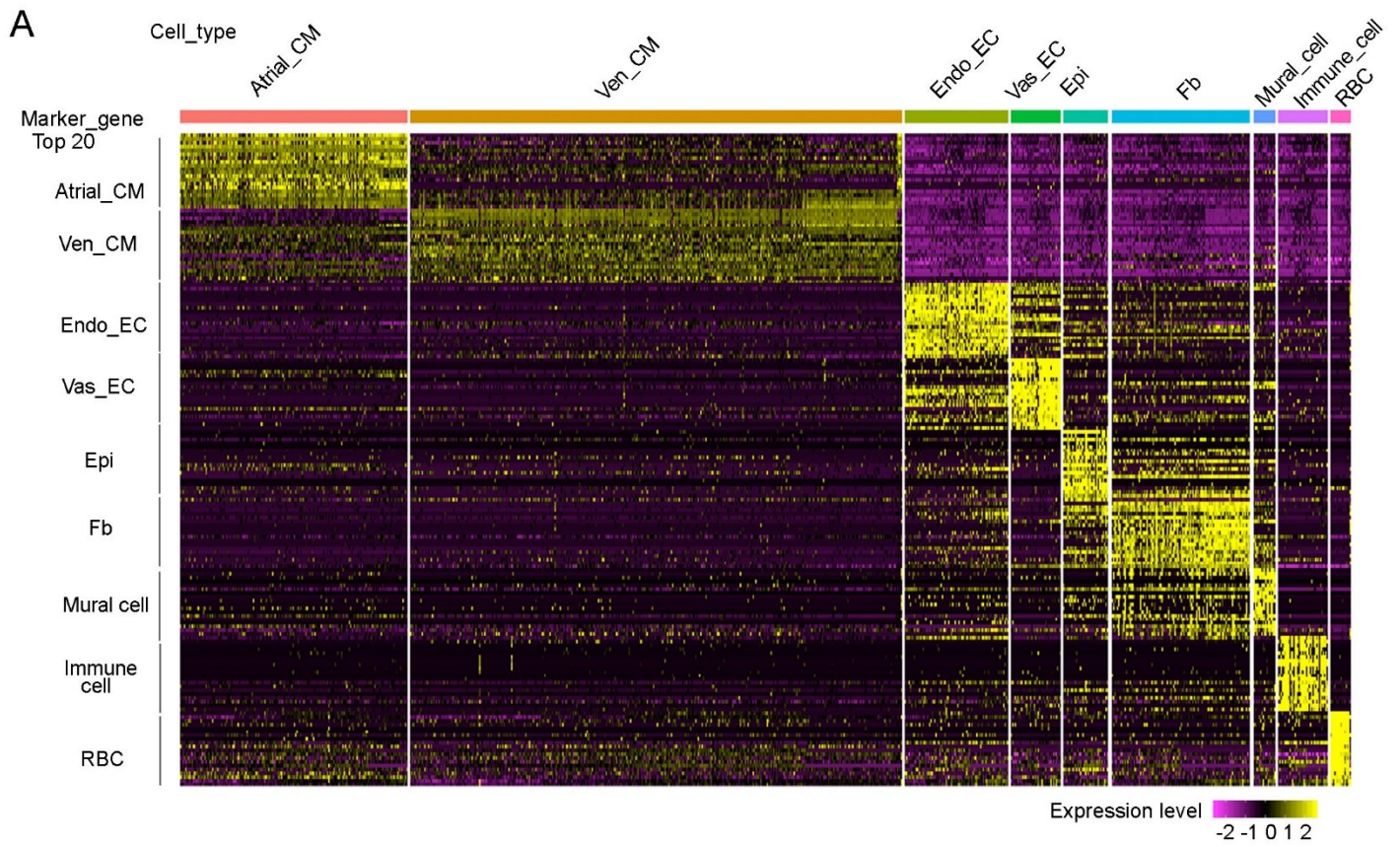


Fig S23. Expression heatmap of the top 20 genes that were uniquely expressed in each cell type.



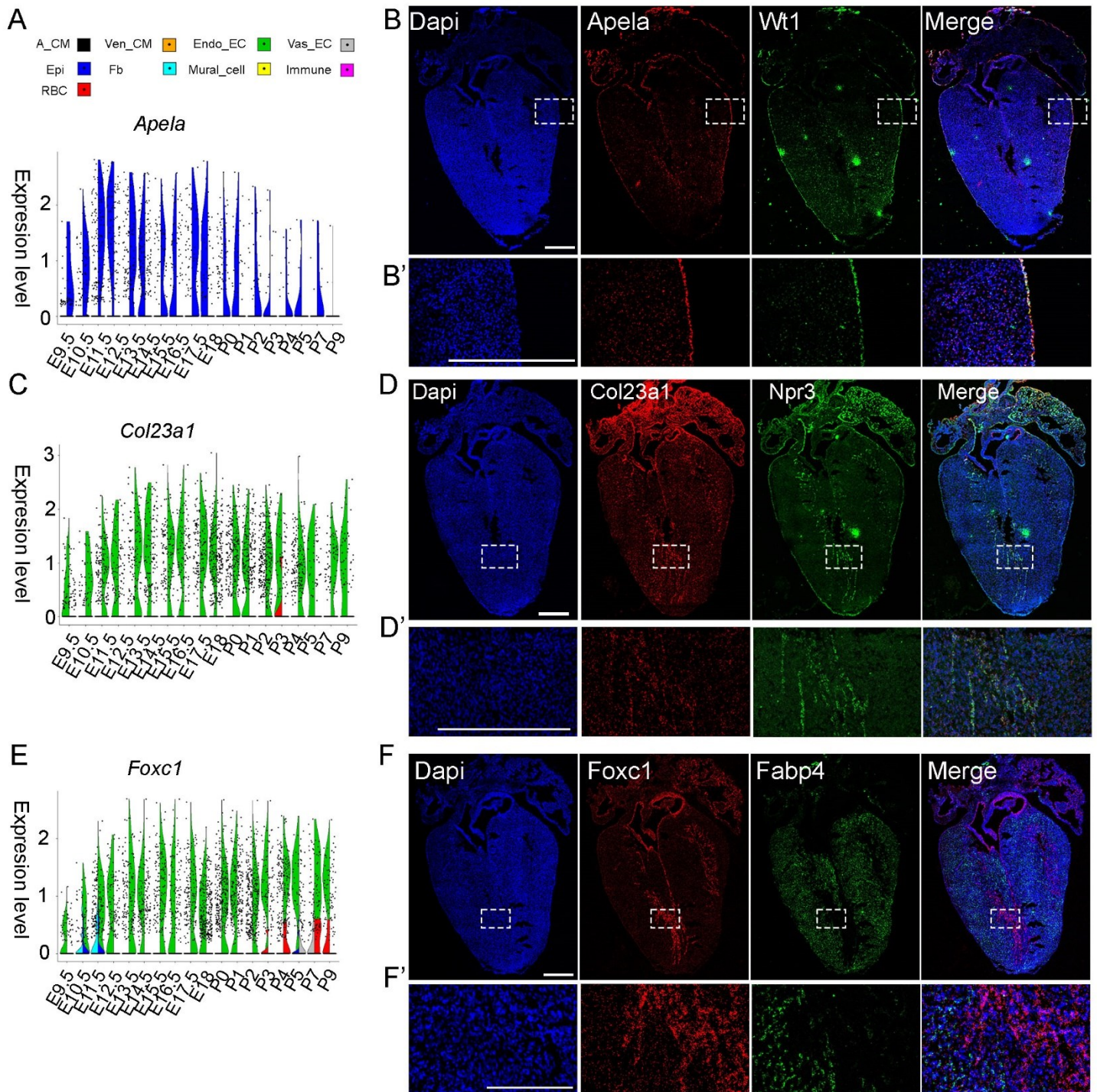


Fig S24. Expression analysis of the newly identified lineage genes. (A) Violin plot showing that *Apela* specifically expressed in epicardial cells at all stages. (B, B') *In situ* RNA staining of *Apela* confirmed its epicardium-specific expression in P2 hearts. (C, E) *Col23a1* and *Foxc1* were found to be specifically expressed in Endo\_EC at most stages. (D, D') *In situ* RNA staining of *Col23a1* with *Npr3* at P2 hearts confirmed its Endo\_EC-specific expression. (F, F') *In situ* RNA staining of *Foxc1* and *Fabp4* confirmed their complement expression pattern, indicating *Foxc1* expresses in Endo\_EC. **The staining experiments were repeated twice with similar results.** Scale bar=500  $\mu$ m.

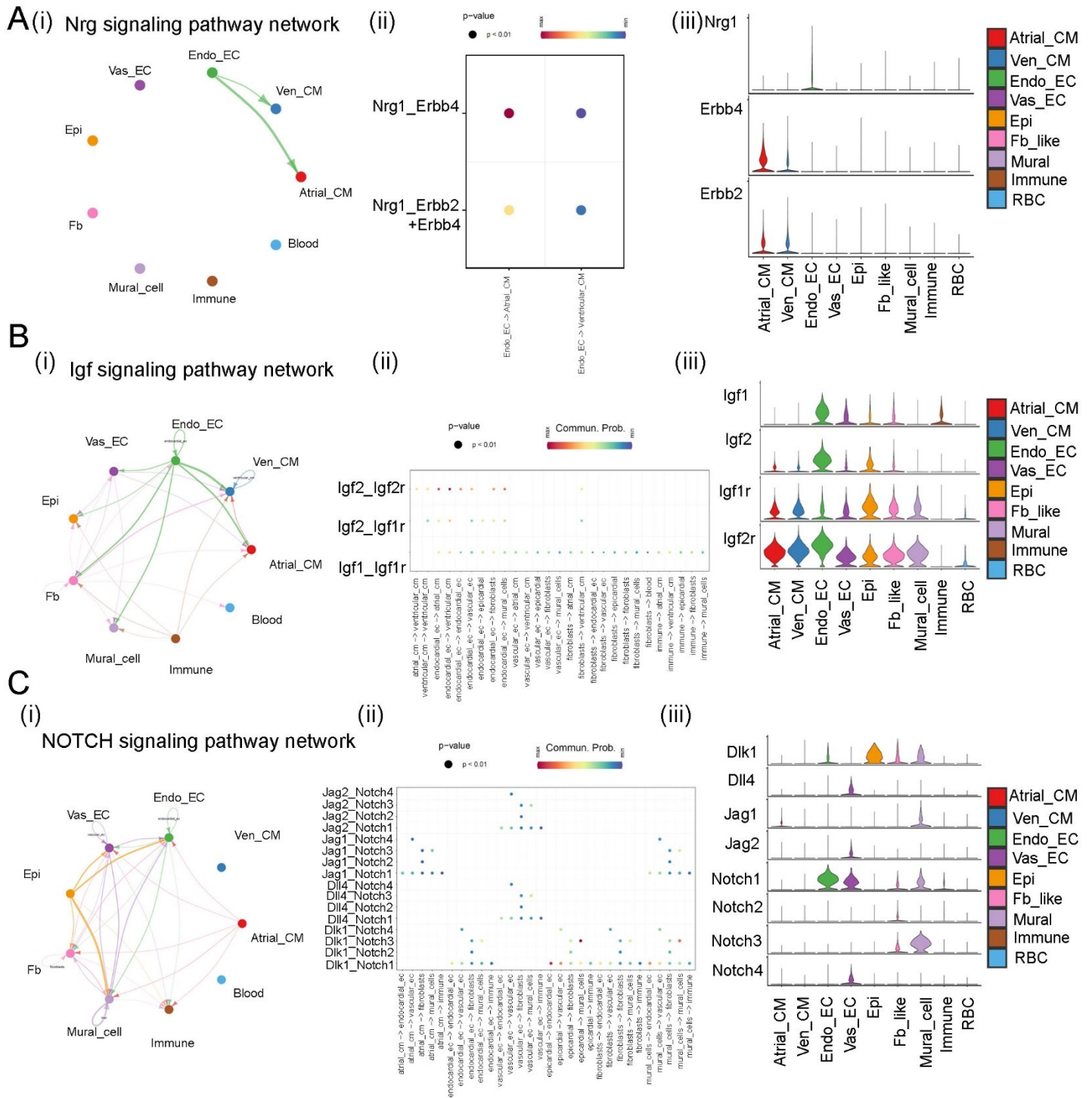


Fig S25. The ligand-receptor interactions in three representative signaling pathways. (A)(i) The network of Nrg signaling pathway. The Loops outside represent autocrine signals for each cell type; the lines connecting cell types represent paracrine signals. The lines with the same color as the cell type dots indicate the cell type secreted ligand in the interactions; the line thickness correlates with the amount of interactions. (ii) The specific ligand-receptor interactions. Each dot represents interactions between the pair of cell types. The color represents interaction probabilities. (iii) Violin plots showing the expression of ligands and receptors at each cell type. (B-C) The signaling pathway networks, ligand-receptor interactions, and ligand and receptor expression patterns in Igf and Notch signaling pathways.

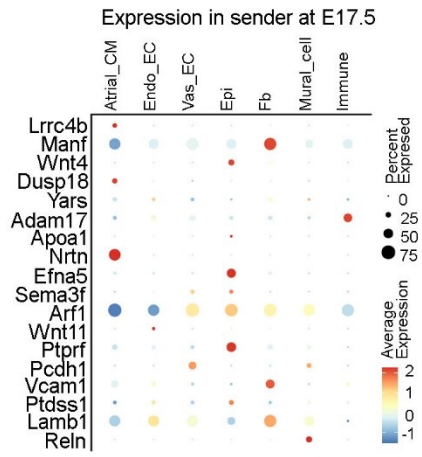


Fig S26. The expression pattern of epicardial cell derived ligands that potentially regulated genes expression in Ven\_CMs at E17.5.



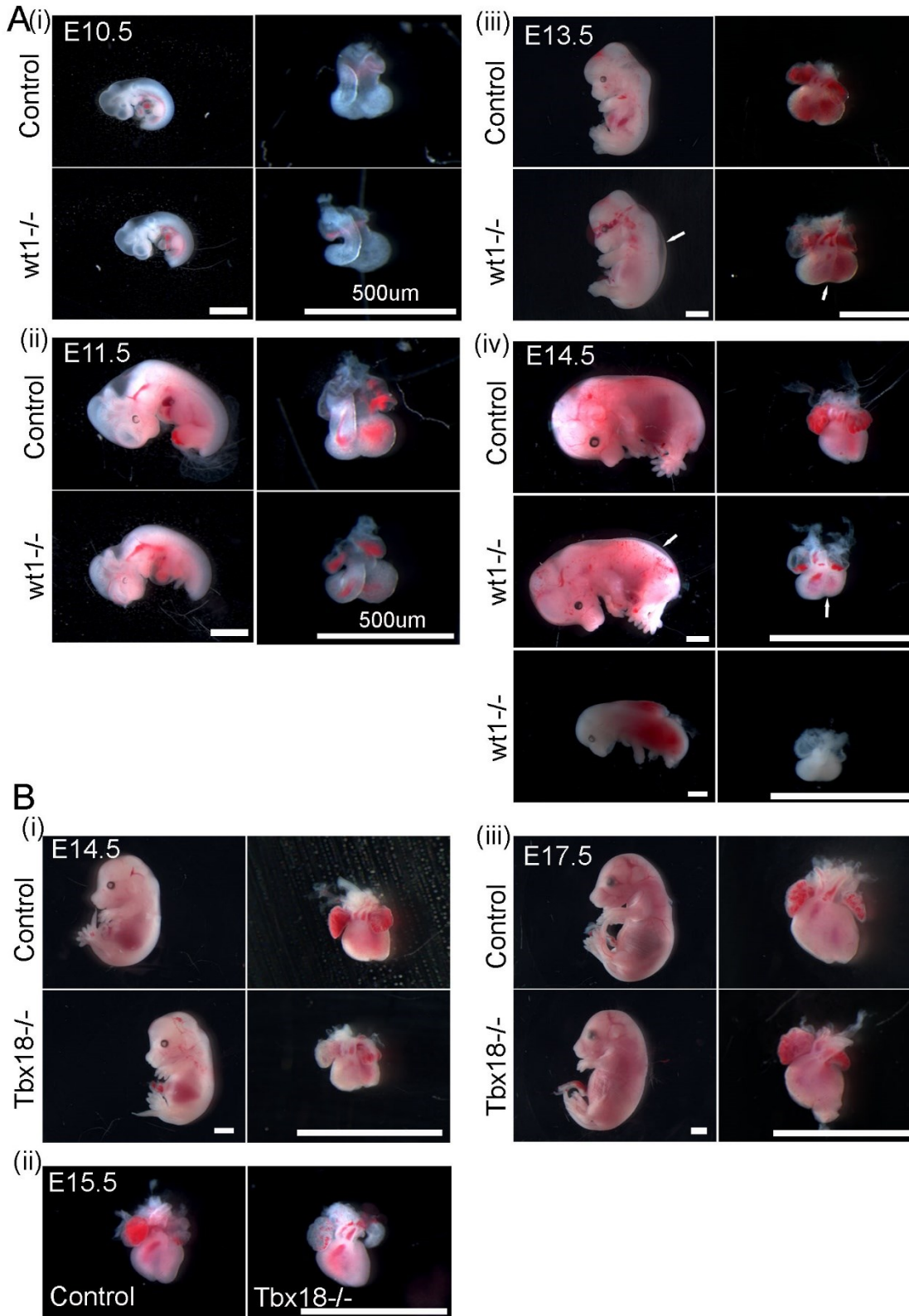


Fig S27. Representative images of control and *Wt1* or *Tbx18* mutant embryos and hearts. (A)(i-iv) Representative embryos and hearts of *Wt1* mutants and wildtype controls from the same litter at different stages. At E13.5 and E14.5, the *Wt1* mutant embryos have obvious body wall edema, and their hearts have more rounded and bifid apices, as pointed by the arrows. (B) (i-iii) Representative *Tbx18* mutant and control embryos and hearts at E14.5, E15.5, and E17.5. Except for the two labeled scale bars that represent 500um, all the other scale bars=1mm.



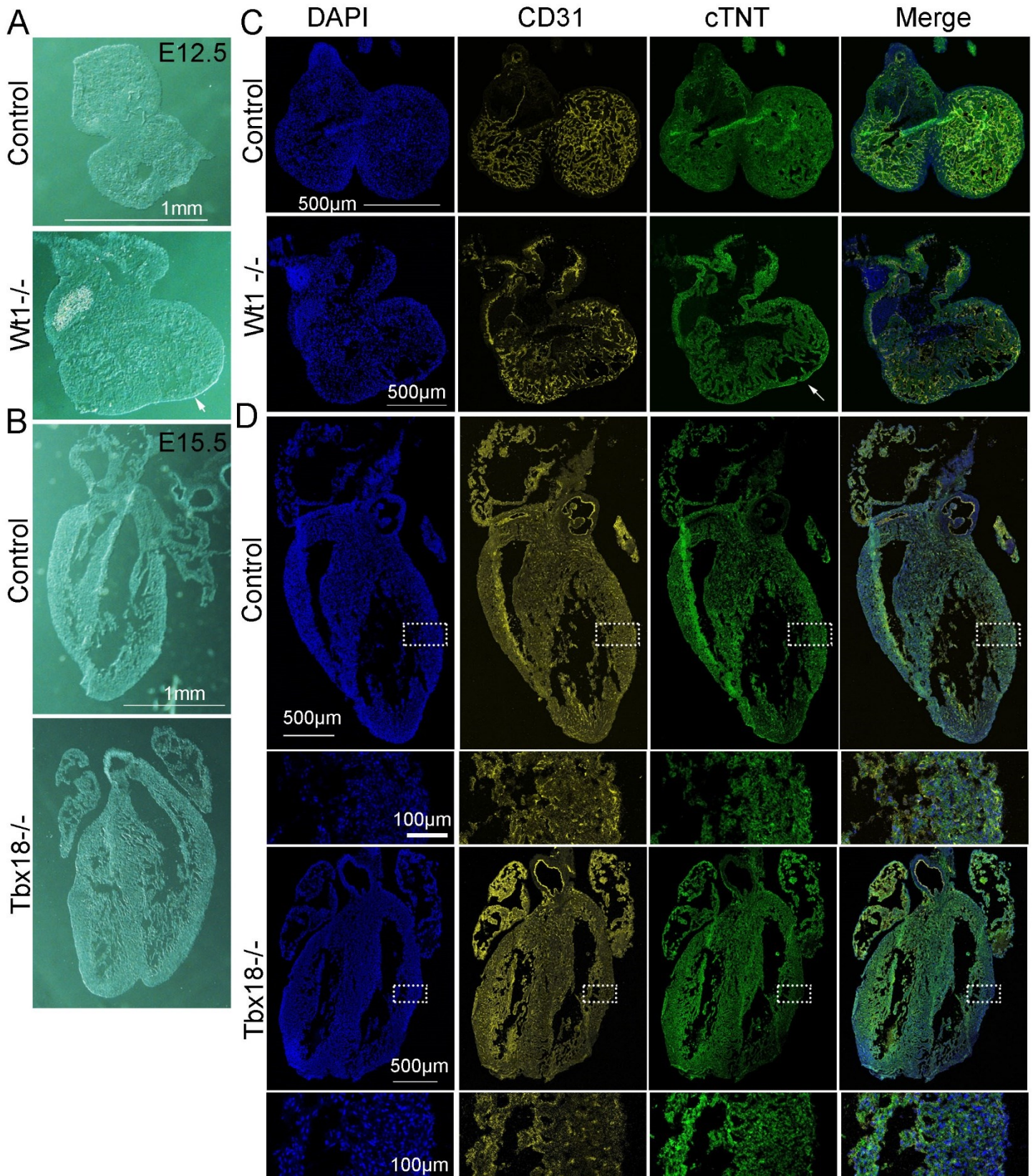


Fig S28. Histological and staining analysis of *Wt1* and *Tbx18* control and mutant hearts. (A, B) Histological images (cryo-sectioning) of *Wt1* control and mutant hearts at E12.5 and *Tbx18* control and mutant hearts at E15.5. (C) Staining analysis of CD31 (endothelial cell marker) and cTNT (cardiomyocyte marker) on control and *Wt1* mutant heart sections at E12.5. Note that thinner myocardium was observed in *Wt1* mutant hearts than the control hearts (point by arrow). (D) Staining analysis of CD31 and cTNT on *Tbx18* control and mutant heart sections at E15.5. No obvious changes in vessel density were observed between controls and mutants. **The staining experiments were repeated on three heart sections with similar results.**

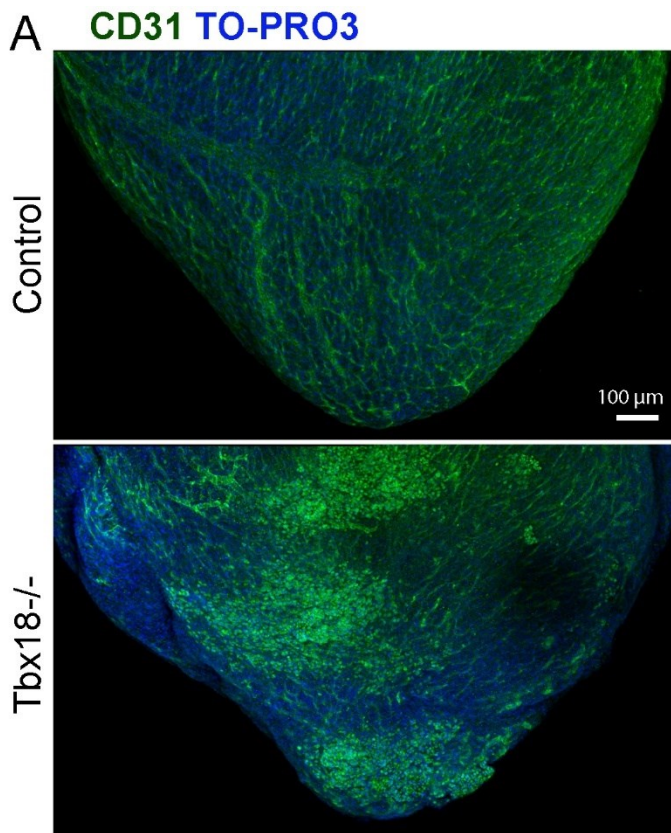


Fig S29. Whole mount staining analysis of CD31 in *Tbx18* control (*Tbx18*<sup>+/-</sup>) and mutant (*Tbx18*<sup>-/-</sup>) hearts at E17.5. Ectopic nodules with CD31-positive cells were observed in *Tbx18* mutant hearts. Only one mutant heart at this stage was analyzed, and its defects were observed. Scale bar=100  $\mu$ m.



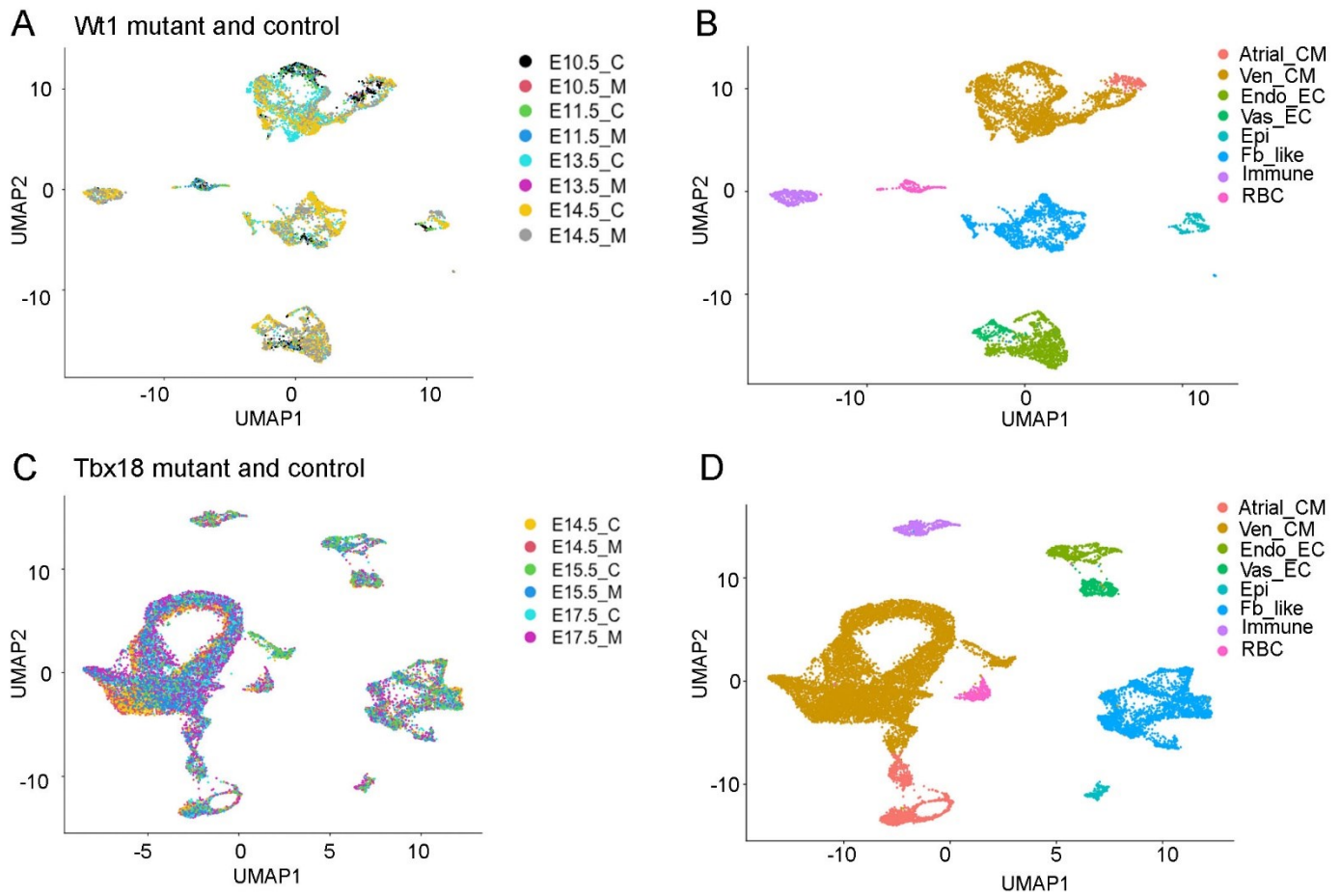


Fig S30. Unsupervised clustering analysis of the scRNA-seq data from *Wt1* and *Tbx18* mutant and control samples. (A, B) UMAP plots of *Wt1* mutant and control cells labeled by sample or cell type. (C, D) UMAP plots of *Tbx18* mutant and control cells labeled by sample or cell type.



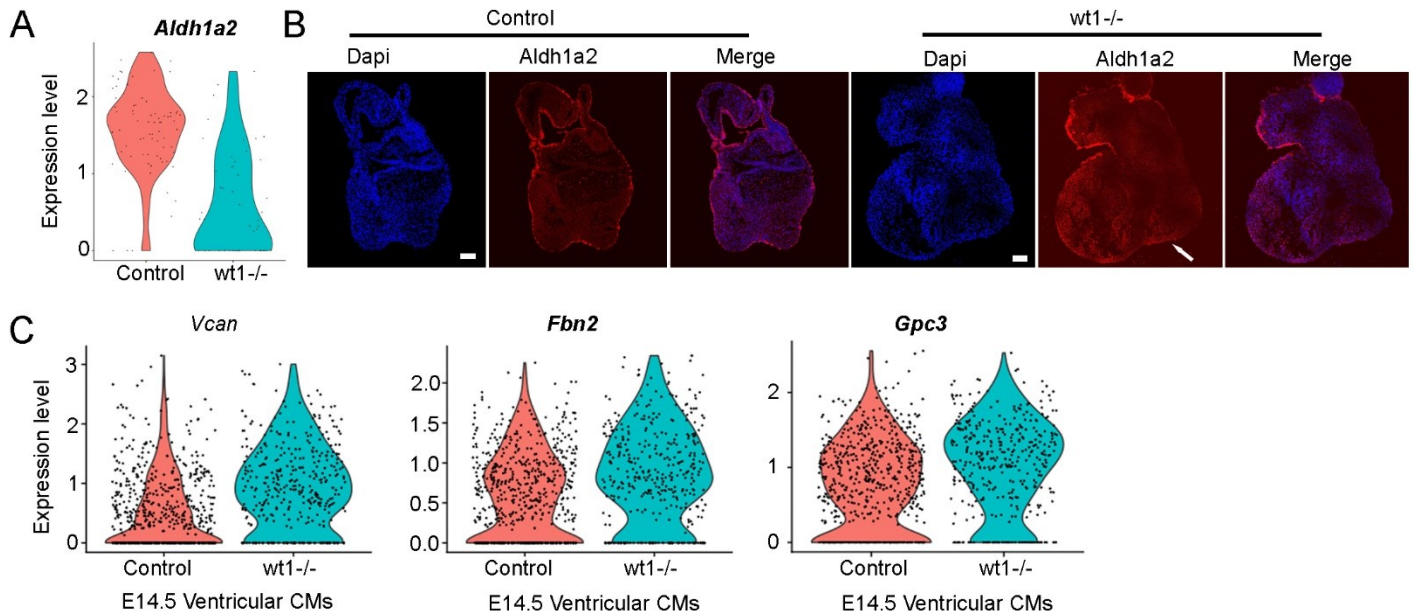


Fig S32. Gene expression analysis of the *Wt1* mutant hearts. (A) ScRNA-seq data revealed a reduction of *Aldh1a2* expression in *Wt1* mutant epicardial cells. (B) Antibody staining confirmed the reduction of *Aldh1a2* in *Wt1* mutant ventricular cells (arrow). However, the reduction was not observed in atrial epicardial cells. **The staining experiments were repeated on three heart sections with similar results.** (C) ScRNA-seq data revealed an upregulation of *Tgfb3* target genes *Vcan*, *Fbn2*, and *Gpc3* in *Wt1* mutant ventricular CMs compared to control CMs. Scale bar=100 μm.





BC22	CCTTGGCACCCGAGAATTCCACTACGACAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC23	CCTTGGCACCCGAGAATTCCAAGAAGAGGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC24	CCTTGGCACCCGAGAATTCCAGTACGCATAAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC25	CCTTGGCACCCGAGAATTCCACGAAGCCAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC26	CCTTGGCACCCGAGAATTCCAACATGCGTAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC27	CCTTGGCACCCGAGAATTCCATAAGGCTCAAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC28	CCTTGGCACCCGAGAATTCCAGGAAGGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC29	CCTTGGCACCCGAGAATTCCACCATGGCGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC30	CCTTGGCACCCGAGAATTCCAAAAGGGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC31	CCTTGGCACCCGAGAATTCCATTACGGTGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC32	CCTTGGCACCCGAGAATTCCAGCATGTACAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
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BC34	CCTTGGCACCCGAGAATTCCAATACGTGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC35	CCTTGGCACCCGAGAATTCCAGCAGTATCAAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC36	CCTTGGCACCCGAGAATTCCAAGATTCCGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
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BC38	CCTTGGCACCCGAGAATTCCAGAACTCTGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC39	CCTTGGCACCCGAGAATTCCACGATTGACAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC40	CCTTGGCACCCGAGAATTCCAACAGTGCTAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC41	CCTTGGCACCCGAGAATTCCATAACTGGCAAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC42	CCTTGGCACCCGAGAATTCCACCAGTTAGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC43	CCTTGGCACCCGAGAATTCCAGGCTAACTAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC44	CCTTGGCACCCGAGAATTCCACCCGAAGCAAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
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BC47	CCTTGGCACCCGAGAATTCCACACCACGAAAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA

BC48	CCTTGGCACCCGAGAATTCCATCCGAGATAAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC49	CCTTGGCACCCGAGAATTCCAGACCAGCCAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
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BC68	CCTTGGCACCCGAGAATTCCAAGCGCTGAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
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BC70	CCTTGGCACCCGAGAATTCCAAGCGCTGAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC71	CCTTGGCACCCGAGAATTCCAAGCGCTGAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA
BC72	CCTTGGCACCCGAGAATTCCAAGCGCTGAAAAAAAAAAAAAAAAA AAAAAAAAAAAAA



Table S2. The sequence of H probes that were used for single molecular in situ hybridization (PLISH) analysis.

Name	Sequence
cy5-mApela-Right-1	atgtgtggaagacggccatTTATACGTCGAGTTGAACGTCGTAACA
cy5-mApela-left-1	TAGCGCTAACAACTTACGTCGTTATGaggacgtgatgtactggtat
cy5-mApela-Right-2	attcagacaaacgcatgtgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mApela-left-2	TAGCGCTAACAACTTACGTCGTTATGtttaacctcgtctgtttcc
cy5-mApela-Right-3	tgaaaagccatccacggtacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mApela-left-3	TAGCGCTAACAACTTACGTCGTTATGtctgaaacagcctctgttgc
cy5-mApela-Right-4	cgtctgtaaatcgcagtatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mApela-left-4	TAGCGCTAACAACTTACGTCGTTATGgtccggctcaccacacatc
cy5-mApela-Right-5	ccattcaggcagcagtaacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mApela-left-5	TAGCGCTAACAACTTACGTCGTTATGttaagcgggaagtctctgca
cy5-mLrrn4-Right-1	gttaccagaaggccagttttTATACGTCGAGTTGAACGTCGTAACA
cy5-mLrrn4-left-1	TAGCGCTAACAACTTACGTCGTTATGaactgtaactctggaatac
cy5-mLrrn4-Right-2	tactccgtatatacctcagtTTATACGTCGAGTTGAACGTCGTAACA
cy5-mLrrn4-left-2	TAGCGCTAACAACTTACGTCGTTATGaccaagtgagatactacact
cy5-mLrrn4-Right-3	tactgagggatatacggagtTTATACGTCGAGTTGAACGTCGTAACA
cy5-mLrrn4-left-3	TAGCGCTAACAACTTACGTCGTTATGactgaccaatgtaaaggatg
cy5-mLrrn4-Right-4	ggagctagtaagacagggctTTATACGTCGAGTTGAACGTCGTAACA
cy5-mLrrn4-left-4	TAGCGCTAACAACTTACGTCGTTATGagtgagaacaactataggag
cy5-mLrrn4-Right-5	cagctcaaggccaacagaacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mLrrn4-left-5	TAGCGCTAACAACTTACGTCGTTATGtcattgtgctaaatcgggc
cy5-mAdamts8-Right-1	AGGTCAGCACATAAGAGGGCTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdamts8-left-1	TAGCGCTAACAACTTACGTCGTTATGTGATAGTGCAGCTTGCTGAG
cy5-mAdamts8-Right-2	AGTGGTAATAGGTAGGGACTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdamts8-left-2	TAGCGCTAACAACTTACGTCGTTATGTGTTCCAGGTTAATAGCACC
cy5-mAdamts8-Right-3	CTAGACAATTTTCATTGGTGTTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdamts8-left-3	TAGCGCTAACAACTTACGTCGTTATGAAGATCACACTTTAGTTCCGG
cy5-mAdamts8-Right-4	ATTTTTCCCCGCTGTGAGGTTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdamts8-left-4	TAGCGCTAACAACTTACGTCGTTATGAGATTGATTATTGGGGAAAC
cy5-mAdamts8-Right-5	CGGACGCAGATGGACAGAGTTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdamts8-left-5	TAGCGCTAACAACTTACGTCGTTATGATCCGGTCCACACAGAGTGC
cy5-mDgit4I-Right-1	ccagagaactgctaacgggtTTATACGTCGAGTTGAACGTCGTAACA
cy5-mDgit4I-left-1	TAGCGCTAACAACTTACGTCGTTATGaaaagatctatgtccatgcc
cy5-mDgit4I-Right-2	ggcccagctctgtgaagactTTATACGTCGAGTTGAACGTCGTAACA
cy5-mDgit4I-left-2	TAGCGCTAACAACTTACGTCGTTATGagctccgtgtacctcccca
cy5-mDgit4I-Right-3	ttgctgaatagaacagagttTTATACGTCGAGTTGAACGTCGTAACA
cy5-mDgit4I-left-3	TAGCGCTAACAACTTACGTCGTTATGacgtggaaggtaaattgtct
cy5-mDgit4I-Right-4	tttcatatctagtctgtggtTTATACGTCGAGTTGAACGTCGTAACA
cy5-mDgit4I-left-4	TAGCGCTAACAACTTACGTCGTTATGagcaacaacatctcagagtc
cy5-mDgit4I-Right-5	acaagcagcaatgtgacctTTATACGTCGAGTTGAACGTCGTAACA
cy5-mDgit4I-left-5	TAGCGCTAACAACTTACGTCGTTATGagtggatccgaaagggacgg
cy5-mAdm-Right-1	tagttccctctcccacgacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdm-left-1	TAGCGCTAACAACTTACGTCGTTATGtagcggccacttattccac

cy5-mAdm-Right-2	tctgggtaggaactgtcgtcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdm-left-2	TAGCGCTAACAACTTACGTCGTTATGtcatcagcagtgcccgtagg
cy5-mAdm-Right-3	gcttcgctctgattgtggcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdm-left-3	TAGCGCTAACAACTTACGTCGTTATGttgtagggggccagttgtgt
cy5-mAdm--Right-4	tcaatgctgtcaccgcaccTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdm--left-4	TAGCGCTAACAACTTACGTCGTTATGtatatcctaaagagtctgga
cy5-mAdm--Right-5	cgcaggcgccaacgggatacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mAdm--left-5	TAGCGCTAACAACTTACGTCGTTATGtcgcccactgttcaatgct
cy5-mBmp10-Right-1	cttctccagggggcactgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-1	TAGCGCTAACAACTTACGTCGTTATGtcaaggccataatggggct
cy5-mBmp10-Right-2	tgaagcaataccatcttgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-2	TAGCGCTAACAACTTACGTCGTTATGtccgtgaagatatcatcaa
cy5-mBmp10-Right-3	cctcgctaccgtctgcactcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-3	TAGCGCTAACAACTTACGTCGTTATGtctagtacctcaaaaatggt
cy5-mBmp10-Right-4	ctcctcgctaccgtctgcacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-4	TAGCGCTAACAACTTACGTCGTTATGtctctagtacctcaaaaatg
cy5-mBmp10-Right-5	ctgttggtccgtagatctcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-5	TAGCGCTAACAACTTACGTCGTTATGgttgatactaaagaccagca
cy5-mBmp10-Right-6	cactgttggtccgtagatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mBmp10-left-6	TAGCGCTAACAACTTACGTCGTTATGtctgttgatactaaagaccag
cy5-mMest-Right-1	aagaaatcaaggcgatcacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mMest-left-1	TAGCGCTAACAACTTACGTCGTTATGtcatggaacctcagggtca
cy5-mMest-Right-2	ccgaccacaccgacagaatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mMest-left-2	TAGCGCTAACAACTTACGTCGTTATGttggtagaagatgcgtaggc
cy5-mMest-Right-3	gccgaggcgcccgcagcgtTTATACGTCGAGTTGAACGTCGTAACA
cy5-mMest-left-3	TAGCGCTAACAACTTACGTCGTTATGacaggatcggagggtggcgtc
cy5-mMest-Right-4	attaatgtactgtaaaagacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mMest-left-4	TAGCGCTAACAACTTACGTCGTTATGtgcgatgaccagggtgccc
cy5-mMest-Right-5	ctgcatttgggctatggaacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mMest-left-5	TAGCGCTAACAACTTACGTCGTTATGtgcgcaagcgtatcggcgc
cy5-mSfrp2-Right-1	atttcgctgacatggcaaccTTATACGTCGAGTTGAACGTCGTAACA
cy5-mSfrp2-left-1	TAGCGCTAACAACTTACGTCGTTATGtaagtgcacggtataagcca
cy5-mSfrp2-Right-2	ggccggcaggagggtggtcgc TTATACGTCGAGTTGAACGTCGTAACA
cy5-mSfrp2-left-2	TAGCGCTAACAACTTACGTCGTTATGtactagcaggggggatgcag
cy5-mSfrp2-Right-3	cacggctggatggtctcatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mSfrp2-left-3	TAGCGCTAACAACTTACGTCGTTATGtaggtcgtcgagacagacag
cy5-mSfrp2-Right-4	acgatgcaatggtcagagcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mSfrp2-left-4	TAGCGCTAACAACTTACGTCGTTATGtcatgaggccataaaacgag
cy5-mSfrp2-Right-5	gcagccgcatgttctgttacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mSfrp2-left-5	TAGCGCTAACAACTTACGTCGTTATGtcatgcccgtggcacagctg
cy5-mCldn5-Right-1	cctacttcaccgatgaaatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mCldn5-left-1	TAGCGCTAACAACTTACGTCGTTATGtgagcgttccaccacgtcg
cy5-mCldn5-Right-2	agcgtctctcccgtgagtgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mCldn5-left-2	TAGCGCTAACAACTTACGTCGTTATGtaccctgccttaactgggc
cy5-mCldn5-Right-3	ctggacattaaggcagcatcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mCldn5-left-3	TAGCGCTAACAACTTACGTCGTTATGtagtgccccaggatctcag

cy5-mCldn5-Right-4	cggtcttcccacgcgggctTATACGTCGAGTTGAACGTCGTAACA
cy5-mCldn5-left-4	TAGCGCTAACAACTTACGTCGTTATGtagctgcgggaagagcccc
cy5-mCldn5-Right-5	gcaccgtcggatcatagaacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mCldn5-left-5	TAGCGCTAACAACTTACGTCGTTATGtcgaggacaacgatgttggc
cy5-mCol23a1-Right-1	ctgtgacaactcaaagctttTATACGTCGAGTTGAACGTCGTAACA
cy5-mCol23a1-left-1	TAGCGCTAACAACTTACGTCGTTATGaaggacagggaaatctagtaa
cy5-mCol23a1-Right-2	cagctagtaaccatcggaatTTATACGTCGAGTTGAACGTCGTAACA
cy5-mCol23a1-left-2	TAGCGCTAACAACTTACGTCGTTATGaaaactcaccagaggtgac
cy5-mCol23a1-Right-3	acagatgagtggtgttatttTATACGTCGAGTTGAACGTCGTAACA
cy5-mCol23a1-left-3	TAGCGCTAACAACTTACGTCGTTATGaagatgccgtaacaagggct
cy5-mCol23a1-Right-4	gccctaacgatgttgttcttTATACGTCGAGTTGAACGTCGTAACA
cy5-mCol23a1-left-4	TAGCGCTAACAACTTACGTCGTTATGaggatgaaggaggagtgctt
cy5-mCol23a1-Right-5	ttcaggtcatggtgtccgtTATACGTCGAGTTGAACGTCGTAACA
cy5-mCol23a1-left-5	TAGCGCTAACAACTTACGTCGTTATGaccatttactaaggaggcct
cy5-mFoxc1-Right-1	agttgtcaagccgatccgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mFoxc1-left-1	TAGCGCTAACAACTTACGTCGTTATGtgagactcgaacatttccc
cy5-mFoxc1-Right-2	ggaggaacgttcggatcaacTTATACGTCGAGTTGAACGTCGTAACA
cy5-mFoxc1-left-2	TAGCGCTAACAACTTACGTCGTTATGttccaggcgcagtcggggca
cy5-mFoxc1-Right-3	cagtctctgcgcggccgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mFoxc1-left-3	TAGCGCTAACAACTTACGTCGTTATGttaaggaagcattgggaaaa
cy5-mFoxc1-Right-4	gggctcggctgcggcgatTTATACGTCGAGTTGAACGTCGTAACA
cy5-mFoxc1-left-4	TAGCGCTAACAACTTACGTCGTTATGaaggcccgtaggcgcggcc
cy5-mFoxc1-Right-5	gttcattccatttgctctTATACGTCGAGTTGAACGTCGTAACA
cy5-mFoxc1-left-5	TAGCGCTAACAACTTACGTCGTTATGaagagtgcgggaatagggt
cy5-mPlvap-Right-1	ggaagctggcgtgatgcgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mPlvap-left-1	TAGCGCTAACAACTTACGTCGTTATGtccatctcacgtcgcgtagt
cy5-mPlvap-Right-2	ctaagatccaccgggaacgcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mPlvap-left-2	TAGCGCTAACAACTTACGTCGTTATGtcatctacaattgaggccct
cy5-mPlvap-Right-3	cgcgctctagctccaactc TTATACGTCGAGTTGAACGTCGTAACA
cy5-mPlvap-left-3	TAGCGCTAACAACTTACGTCGTTATGtggcggcgcagttctgcatt
cy5-mPlvap-Right-4	atctgaaaaccctagtgggcTTATACGTCGAGTTGAACGTCGTAACA
cy5-mPlvap-left-4	TAGCGCTAACAACTTACGTCGTTATGtttaagagggaggtggaaa
cy5-mPlvap-Right-5	cggcatcagctggtagtggtTATACGTCGAGTTGAACGTCGTAACA
cy5-mPlvap-left-5	TAGCGCTAACAACTTACGTCGTTATGagggcagggttccaaggt
cy3-mWt1-Right-1	cggaaccatgaggtgcggcTTATACGTCGAGTTGACCGACGTATTG
cy3-mWt1-left-1	TATTCGTTTGAACCTTACGTCGTTATGtcggatgcggacggttctcc
cy3-mWt1-Right-2	aaagtgaccgtgctgatccTTATACGTCGAGTTGACCGACGTATTG
cy3-mWt1-left-2	TATTCGTTTGAACCTTACGTCGTTATGttggtgcggatggtaggct
cy3-mWt1-Right-3	gttactcaaacctatgagcTTATACGTCGAGTTGACCGACGTATTG
cy3-mWt1-left-3	TATTCGTTTGAACCTTACGTCGTTATGttaacaatcgctgtagaa
cy3-mWt1-Right-4	cacacagtgatgcagctaatTTATACGTCGAGTTGACCGACGTATTG
cy3-mWt1-left-4	TATTCGTTTGAACCTTACGTCGTTATGaccatacacctcctaattt
cy3-mNpr3-Right-1	ttgcaaggagagctgttggTATACGTCGAGTTGACCGACGTATTG
cy3-mNpr3-left-1	TATTCGTTTGAACCTTACGTCGTTATGatgctccacgattctggtct
cy3-mNpr3-Right-2	taagctgaacagactctgtTATACGTCGAGTTGACCGACGTATTG
cy3-mNpr3-left-2	TATTCGTTTGAACCTTACGTCGTTATGacatcttcagccagactagc



cy3-mNpr3-Right-3	cctagctcactgtccaagctTTATACGTCGAGTTGACCGACGTATTG
cy3-mNpr3-left-3	TATTCGTTTGAACCTTACGTCGTTATGatgcatcatttgggtggct
cy3-mNpr3-Right-4	tagaataggaagctatatgtTTATACGTCGAGTTGACCGACGTATTG
cy3-mNpr3-left-4	TATTCGTTTGAACCTTACGTCGTTATGagctgggtgcagctttgactc
cy3-mFabp4-Right-1	ctgcagcacaggagggtgctTTATACGTCGAGTTGACCGACGTATTG
cy3-mFabp4-left-1	TATTCGTTTGAACCTTACGTCGTTATGatgagcctctgaagtccaga
cy3-mFabp4-Right-2	gtggcaaagcccactcccacTTATACGTCGAGTTGACCGACGTATTG
cy3-mFabp4-left-2	TATTCGTTTGAACCTTACGTCGTTATGttctttcatgtaatcatcga
cy3-mFabp4-Right-3	tggtgaccaaatacccatttTTATACGTCGAGTTGACCGACGTATTG
cy3-mFabp4-left-3	TATTCGTTTGAACCTTACGTCGTTATGacgctgatgatcatgttggg
cy3-mFabp4-Right-4	gtggaagtcacgcctttcatTTATACGTCGAGTTGACCGACGTATTG
cy3-mFabp4-left-4	TATTCGTTTGAACCTTACGTCGTTATGaacacattccaccaccagct

Table S3. The sequence of qPCR primers and mouse genotyping primers.

Atf3 Forward Primer	GAGGATTTTGCTAACCTGACACC
Atf3 Reverse Primer	TTGACGGTAACTGACTCCAGC
Eno3 Forward Primer	CACAGCCAAGGGTCGATTCC
Eno3 Reverse Primer	CCCAGGTATCGTGCTTTGTCT
Klf9 Forward Primer	TTATTGCACGCTGGTCACTATC
Klf9 Reverse Primer	CTCATCGGGACTCTCCAGAC
Fhl2 Forward Primer	ATGACTGAACGCTTTGACTGC
Fhl2 Reverse Primer	CGATGGGTGTTCCACACTCC
Rps2 Forward Primer	GGGGCTCGTGGAGGTAAAG
Rps2 Reverse Primer	TCTCAGACTCCTTAATGGGCAG
Ranbp1 Forward Primer	CGAGGACCATGATACTTCCACA
Ranbp1 Reverse Primer	CCTCCAGCGTTTTAATTTCTTGC
Per1 Forward Primer	GAATTGGAGCATATCACATCCGA
Per1 Reverse Primer	CCCGAAACACATCCCCTTTG
Gapdh Forward Primer	GGATTTGGTCGATTGGG
Gapdh Reverse Primer	GGAAGATGGTGATGGGATT
GM8797 Forward Primer	GGCAAGACCATCACCTAGA
GM8797 Reverse Primer	TAATAGCCACCCCTCAGACG
GM10260 Forward Primer	GATCCCAGACTGGTTCCTGA
GM10260 Reverse Primer	TAAGAGCAAAGGCCAGAGA
Fbn2 Forward Primer	CAACTCCGAAGGAAGCTACG
Fbn2 Reverse Primer	AAGAGCCCTTCGTGTTCTCA
Gpc3 Forward Primer	CCAACATGCTGCTCAAGAAA
Gpc3 Reverse Primer	CTGGAAAGAGGCTGTGCAAC
Vcan Forward Primer	GGCCAACAGTGGTTTCAAGT
Vcan Reverse Primer	TTCTGTGAAGGCTGTGATG

#### Mouse genotyping primers

Tbx18-Cre wt Forward Primer	AACGCCAGAGAAAGAGGAAAC
Tbx18-Cre wt Reverse Primer	TCAGTGCCTCCACAGAGAAG
Tbx18-Cre mutant Forward Primer	AACGCCAGAGAAAGAGGAAAC
Tbx18-Cre mutant Reverse Primer	AGGCAAATTTTGGTGTACGG
Wt1-Cre wt Forward Primer	CCTACCATCCGCAACCAAG

Wt1-Cre wt Reverse Primer	CCCTGTCCGCTACTTTCAGA
Wt1-Cre mutant Forward Primer	ATCGCAGGAGCGGAGAAC
Wt1-Cre mutant Reverse Primer	GAACTTCAGGGTCAGCTTGC