

**Characterization of three *Stenotrophomonas* strains and proposal of
Stenotrophomonas mori sp. nov., and *Stenotrophomonas lacuserhaii* sp.
nov.**

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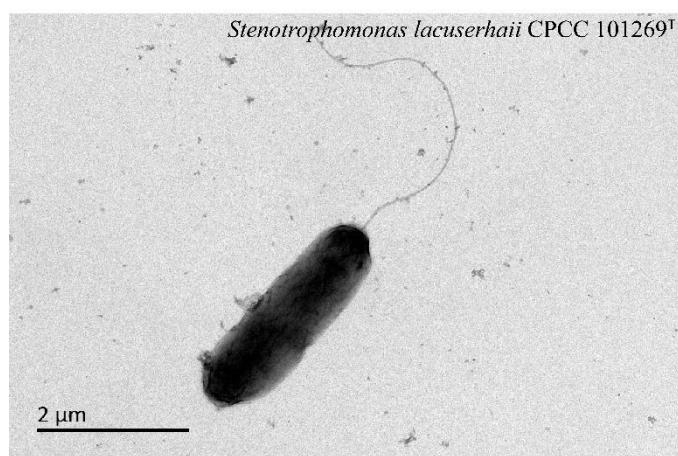
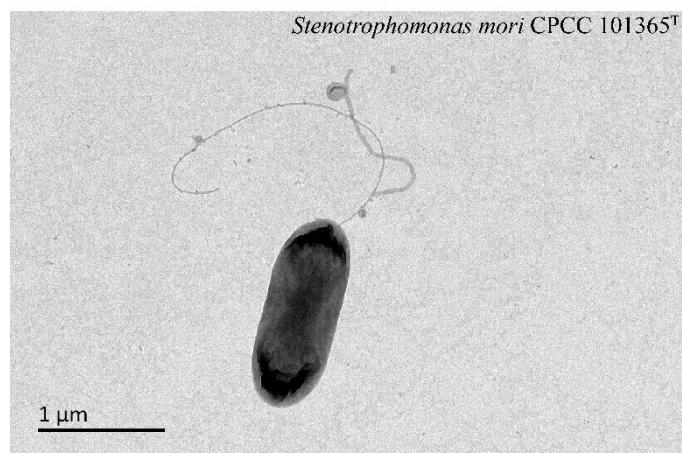
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Keywords: *Stenotrophomonas mori*, *Stenotrophomonas lacuserhaii*, genome, ANI, dDDH, IAA

Abbreviations: DPG, diphasphatidylglycerol; PG, phosphatidylglycerol; PE, phosphatidylethanolamine; ANI, average nucleotide identity; dDDH, digital DNA-DNA hybridization; IAA, indole-3-acetic acid; MALDI-TOF MS, matrix-assisted laser desorption/ionization time-of-flight mass spectrometry.

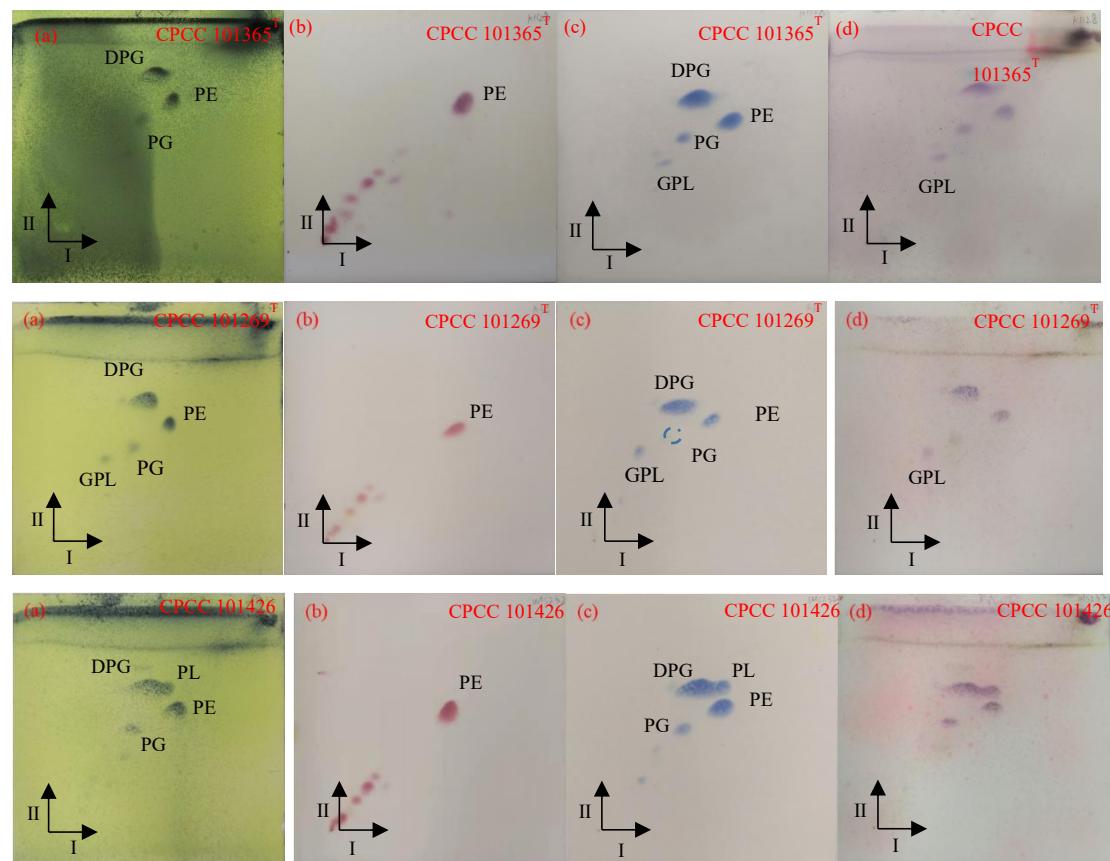
Supplementary Figure S1. Transmission electron micrographs of motile cells of strains CPCC 101365^T and CPCC 101269^T.



Supplementary Figure S2. Polar lipid profiles of strains CPCC 101365^T, CPCC 101269^T, and CPCC 101426 after separation by two-dimensional TLC.

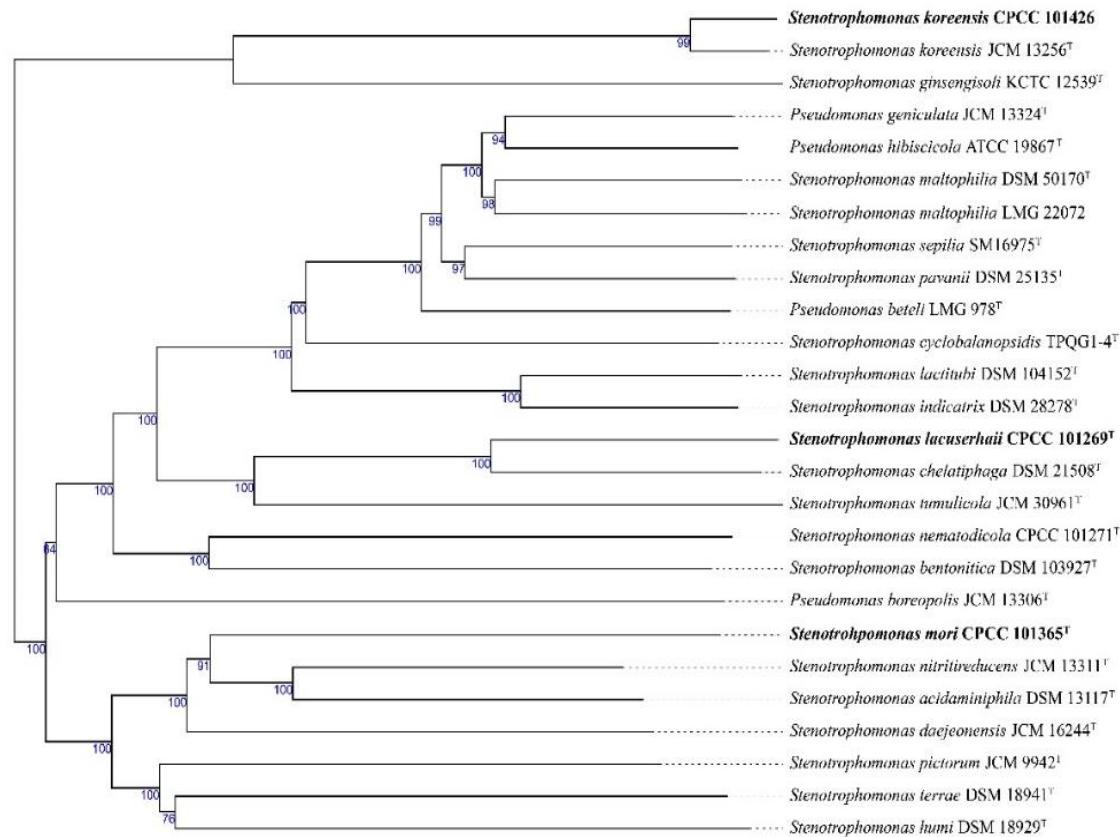
(a) Detection by spraying with molybdatophosphoric acid reagent. (b) Detection by spraying with ninhydrin stain reagent. (c) Detection by spraying with molybdenum blue stain reagent. (d) Detection by spraying with p-anisaldehyde stain reagent.

DPG, diphosphatidylglycerol; PG, phosphatidylglycerol; PE, phosphatidylethanolamine; GPL, unidentified glycoprophospholipid; PL, unidentified phospholipid.



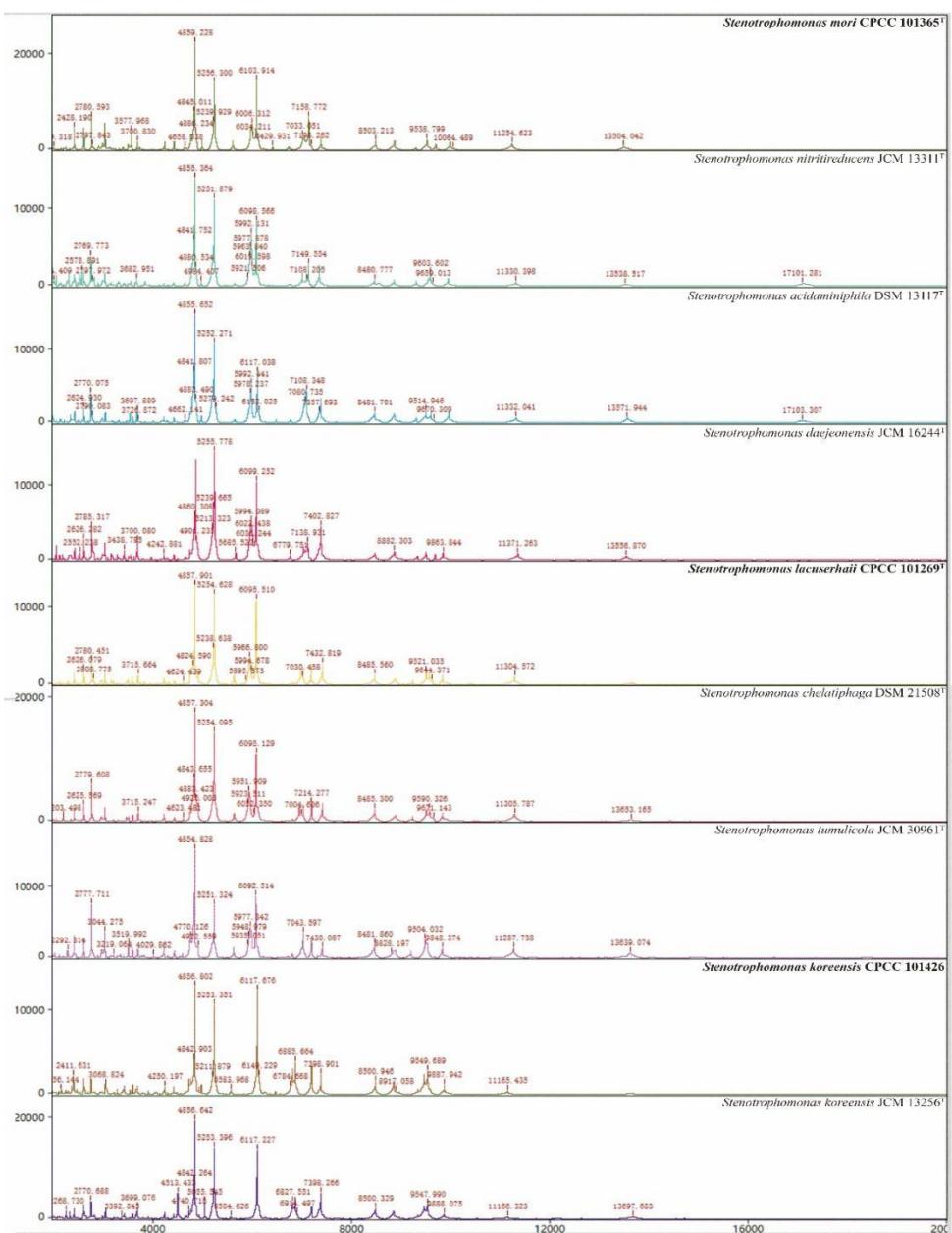
Supplementary Figure S3. Whole-genome phylogenetic tree generated for strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and other representatives of the family *Lysobacteraceae*.

The phylogenetic tree was inferred with the FastME algorithm using GBDP distances calculated from genome sequences. Branch lengths are scaled based on the GBDP distance formula d5. Numbers above branches show GBDP pseudo-bootstrap support values from 100 replicates.

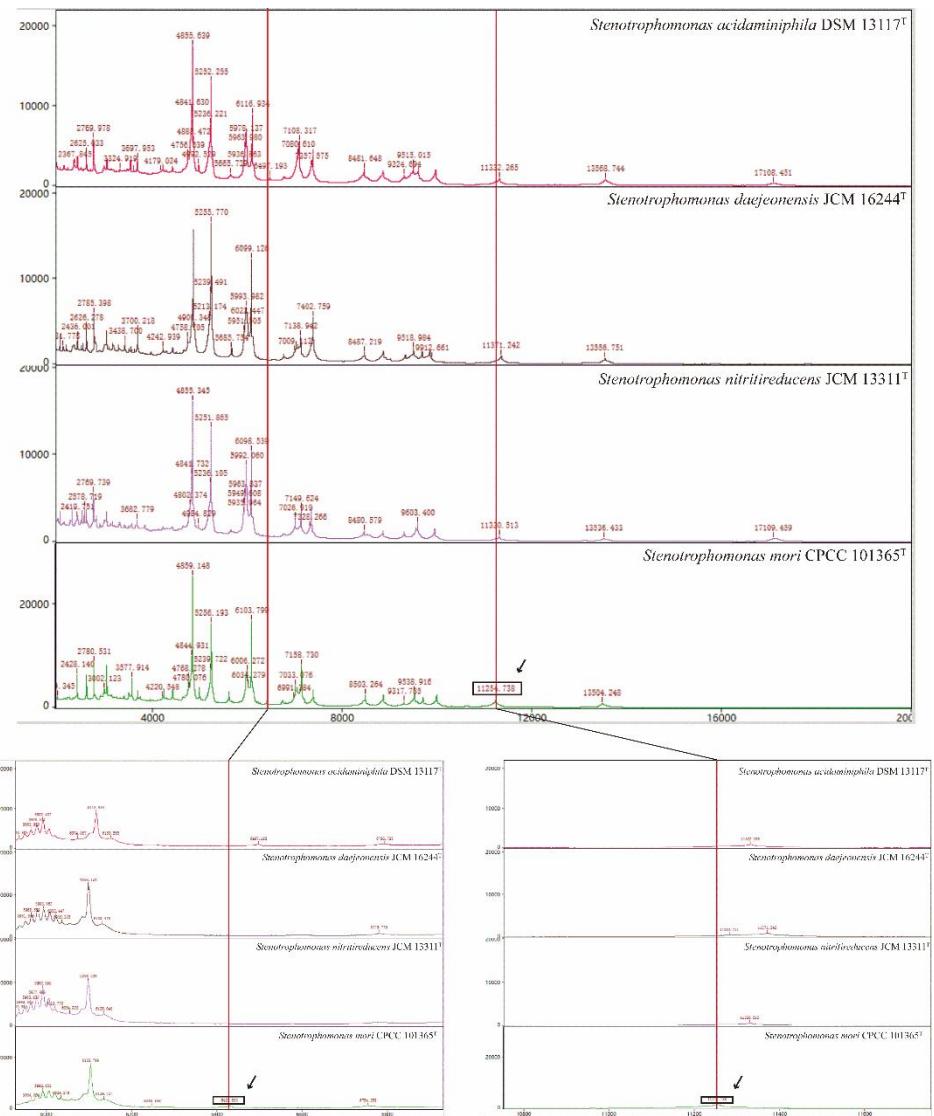


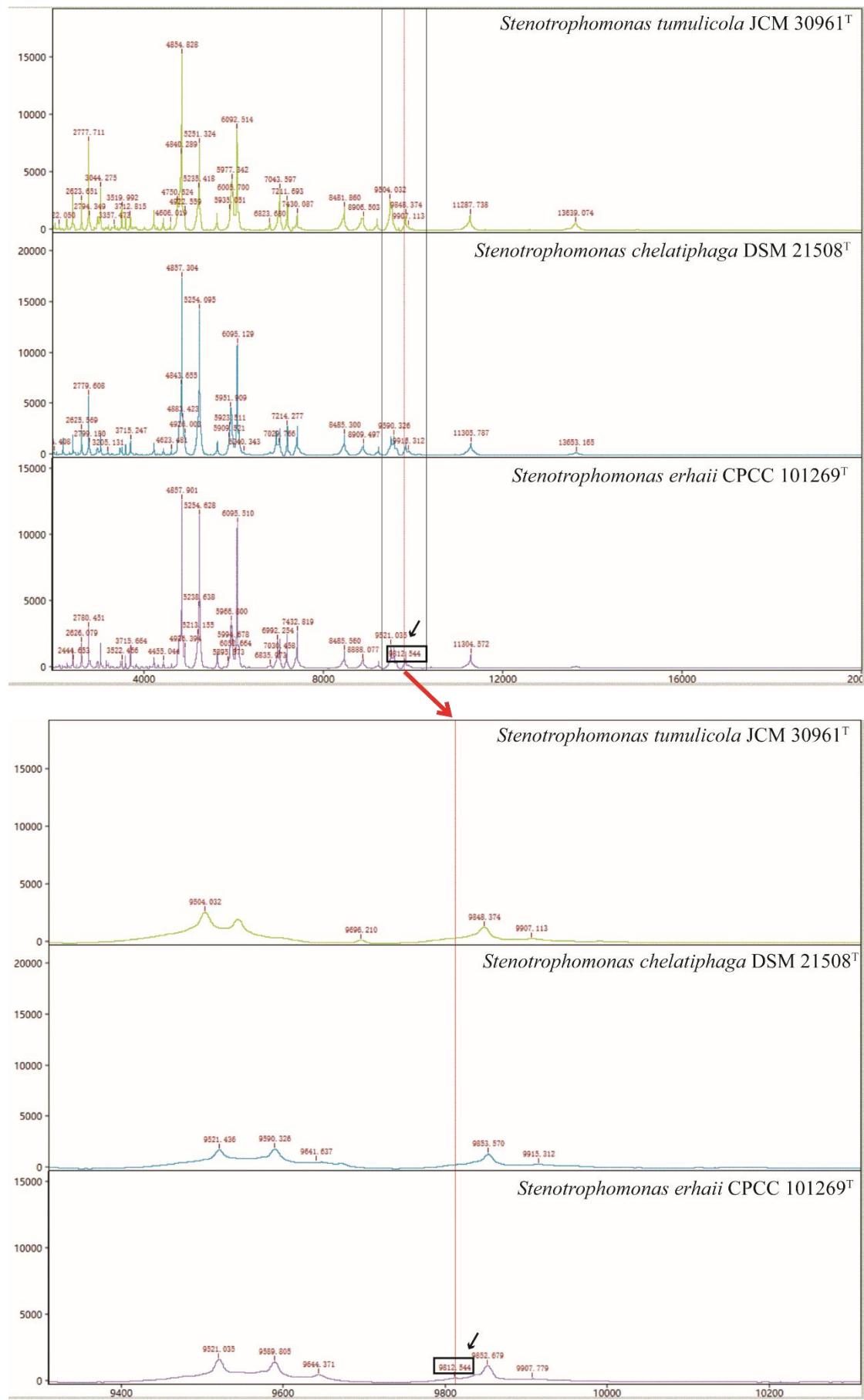
Supplementary Figure S4. MALDI-TOF MS spectra profiles of strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and closely related *Stenotrophomonas* strains.

The name of each strain was labeled on the upper right corner of each spectrum. (a) MALDI-TOF MS spectra profiles of strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and their closely related *Stenotrophomonas* strains; (b) MALDI-TOF MS profiles of strain CPCC 101365^T compared with *Stenotrophomonas acidaminiphila* DSM 13117^T and *Stenotrophomonas nitritireducens* JCM 13311^T, with the detail of peptide peaks of 6,429.9 Da and 11,254.6 Da in the spectra of strain CPCC 101365^T, *S. nitritireducens* JCM 13311^T and *S. acidaminiphila* DSM 13117^T; (c) MALDI-TOF MS profiles of strain CPCC 101269^T compared with *Stenotrophomonas chelatiphaga* DSM 21508^T and *Stenotrophomonas tumulicola* JCM 30961^T, with the detail of the peptide peak of 9,812.5 Da in the spectra of strain CPCC 101269^T, *Stenotrophomonas chelatiphaga* DSM 21508^T and *Stenotrophomonas tumulicola* JCM 30961^T.



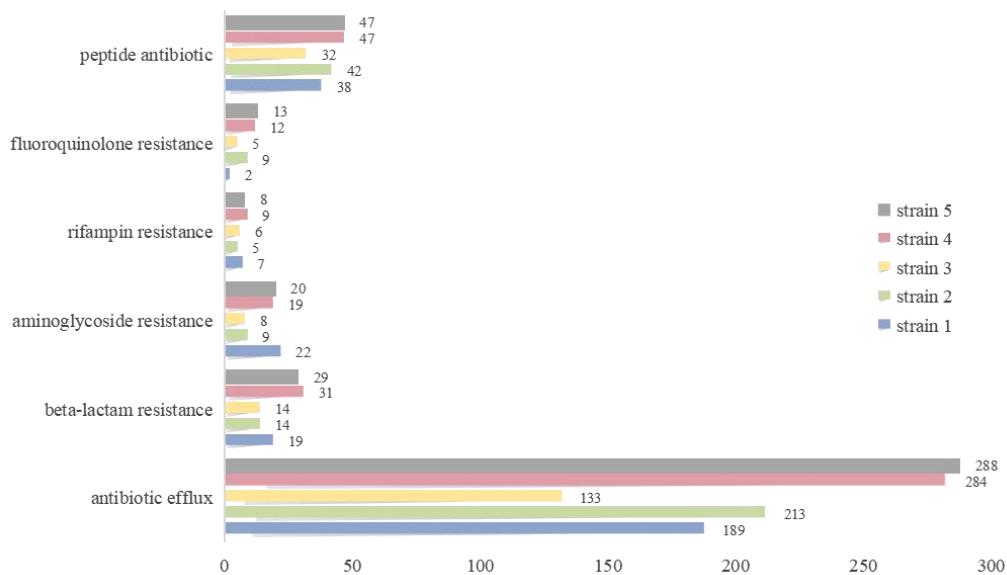
(a)





Supplementary Figure S5. Antibiotic resistance genes predicted from the genomes of CPCC 101365^T, CPCC 101269^T, CPCC 101426, *S. maltophilia* DSM 50170^T, and *S. maltophilia* K279a.

Strain 1, CPCC 101365^T; Strain 2, CPCC 101269^T; Strain 3, CPCC 101426; Strain 4, *S. maltophilia* DSM 50170^T; Strain 5, *S. maltophilia* K279a.



Supplementary Table S1. Cellular fatty acid profiles of strains CPCC 101365^T, CPCC 101269^T, CPCC 101426 and other closely related strains.

Strains: 1, CPCC 101365^T; 2, *Stenotrophomonas nitritireducens* JCM 13311^T; 3, *Stenotrophomonas acidaminiphila* DSM 13117^T. 4, *Stenotrophomonas daejeonensis* JCM 16244^T; 5, CPCC 101269^T; 6, *Stenotrophomonas chelatiphaga* DSM 21508^T; 7, *Stenotrophomonas tumulicola* JCM 30961^T; 8, CPCC 101426; 9, *Stenotrophomonas koreensis* JCM 13256^T. All data were generated in this study. -, not detected.

Antesio-C ₁₅ :0	9.5	5.9	4.6	3.9	16.6	14.8	8.8	4.7	6.1
Antesio-C ₁₇ :0							-	-	0.5
Hydroxy fatty acids									
C ₁₁ :0 3OH	0.5	0.5	-	-	-	-	-	0.9	0.7
C ₁₂ :0 3OH	1.2	1.7	1.1	0.8	1.9	2.5	1.5	0.6	-
C ₁₃ :0 2OH	1.1	1.1		0.5	0.9	1.5	0.5	-	-
Iso-C ₁₁ :0 3OH	2.8	3.9	0.5	3.2	1.7	2.7	1.7	2.7	3.1
Iso-C ₁₂ :0 3OH	1.7	1.8	5.8	0.5	-	1.0	-	0.7	-
Iso-C ₁₃ :0 3OH	2.3	2.9	2.9	2.2	1.5	2.5	1.9	1.5	1.6
Others									
Sum In Feature 3 (C ₁₆ :1 ω 7c/C ₁₆ :1 ω 6c)	5.1	-	2.0	3.4	5.1	6.7	1.7	2.5	1.3
Sum In Feature 8 (C ₁₈ :1 ω 7c/C ₁₈ :1 ω 6c)	-	-	-	-	0.8	-	1.0	-	-
Sum In Feature 9 (iso- C ₁₇ :1 ω 9c/C ₁₆ :0 10-methyl)	5.3	2.5	10.2	4.4	3.3	1.9	1.8	4.2	5.4

Supplementary Table S2. Pairwise comparison of the 16S rRNA gene sequences of strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and other closely related strains.

1, CPCC 101365^T; 2, *Stenotrophomonas nitritireducens* JCM 13311^T; 3, *Stenotrophomonas acidaminiphila* DSM 13117^T; 4, *Stenotrophomonas daejeonensis* JCM 16244^T; 5, CPCC 101269^T; 6, *Stenotrophomonas chelatiphaga* DSM 21508^T; 7, *Stenotrophomonas tumulicola* JCM 30961^T; 8, CPCC 101426; 9, *Stenotrophomonas koreensis* JCM 13256^T; 10, *Stenotrophomonas ginsengisoli* KCTC 12539^T.

16S rRNA gene sequence similarities (%) > 98.65 % were highlighted in red.

Species	1	2	3	4	5	6	7	8	9	10
1	100.0									
2	99.1	100.0								
3	98.8	98.7	100.0							
4	98.0	97.0	98.4	100.0						
5	97.3	97.3	97.3	97.1	100.0					
6	97.4	97.3	97.1	96.6	99.9	100.0				
7	97.8	97.4	97.3	96.6	99.4	99.4	100.0			
8	97.1	97.1	97.2	96.3	97.3	97.4	97.6	100.0		
9	97.2	97.2	97.3	96.4	97.6	97.5	97.6	99.8	100.0	
10	96.7	97.1	96.7	96.7	97.6	97.5	97.6	98.3	98.4	100.0

Supplementary Table S3. Genome features of strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and other closely related strains.

Strains: 1, CPCC 101365^T; 2, *Stenotrophomonas nitritireducens* JCM 13311^T; 3, *Stenotrophomonas acidaminiphila* DSM 13117^T; 4, *Stenotrophomonas daejeonensis* JCM 16244^T; 5, CPCC 101269^T; 6, *Stenotrophomonas chelatiphaga* DSM 21508^T; 7, *Stenotrophomonas tumulicola* JCM 30961^T; 8, CPCC 101426; 9, *Stenotrophomonas koreensis* JCM 13256^T.

*Data were derived from this study except: ^a, Finkmann et al., 2000; ^b, Assih et al., 2002; ^c, Lee et al., 2011; ^d, Kaparullina et al., 2009; ^e, Handa et al., 2016; ^f, Yang et al., 2006.

Characteristic	1	2	3	4	5	6	7	8	9
Isolation source	Mulberry root	Ammonia-supplied	Upflow anaerobic	Sewage water sample ^c	Fresh water reservoir	Sewage sludge ^d	Major contaminant of	Desert sandy soil	Compost ^f

	biofilters ^a			sludge blanket ^b			stone chamber		
							interior ^c		
No. of contigs	11	95	126	124	27	28	26	16	58
N50 Length (bp)	468,441	167,790	71,728	46,611	381,161	227,000	313,801	521,188	188,329
Genome size (Mbp)	3.4	4.0	3.9	3.3	4.0	4.0	4.3	3.1	3.0
G+C (%)	70.2	68.3	68.8	68.6	66.4	66.9	65.6	66.2	66.1
No. of CDS	2,860	3,490	3,554	2,959	3,439	3,378	3,774	2,706	2,598
DDBJ/EMBL/GenBank accession number of draft genome	JAIKTS000000 000	LDJG00000000	LDJO00000000	LDJP00000000	JACHQY000000 000	LDJK00000000	JACGXS000000 000	JACIUV000000 000	LDJH00000000
GenBank/RefSeq assembly accession number	GCA_0235563 35.1	GCA_0014314 25.1	GCA_0014315 95.1	GCA_0014315 05.1	GCA_01459629 5.1	GCA_0014315 35.1	GCA_01411721 5.1	GCA_01414581 5.1	GCA_0014315 25.1

Supplementary Table S4. Average nucleotide identity (ANI) (%) values between strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and other *Stenotrophomonas* type strains.

Strains: 1, CPCC 101365^T; 2, CPCC 101269^T; 3, CPCC 101426; 4, *S. acidaminiphila* DSM 13117^T; 5, *S. bentonitica* DSM 103927^T; 6, *S. chelatiphaga* DSM 21508^T; 7, *S. daejeonensis* JCM 16244^T; 8, *S. ginsengisoli* KCTC 12539^T; 9, *S. humi* DSM 18929^T; 10, *S. indicatrix* DSM 28278^T; 11, *S. koreensis* JCM 13256^T; 12, *S. lactitubi* DSM 104152^T; 13, *S. maltophilia* DSM 50170^T; 14, *S. nitritireducens* JCM 13311^T; 15, *S. pavanii* DSM 25135^T; 16, *S. pictorum* JCM 9942^T; 17, *S. rhizophila* JCM 13333^T; 18, *S. terrae* DSM 18941^T; 19, *S. tumulicola* JCM 30961^T.

ANI values >95 % were highlighted in red.

Supplementary Table S5. Digital DNA–DNA hybridization (dDDH) (%) values between strains CPCC 101365^T, CPCC 101269^T, CPCC 101426, and other *Stenotrophomonas* type strains.

Strains: 1, CPCC 101365^T; 2, CPCC 101269^T; 3, CPCC 101426; 4, *S. acidaminiphila* DSM 13117^T; 5, *S. bentonitica* DSM 103927^T; 6, *S. chelatiphaga* DSM 21508^T; 7, *S. daejeonensis* JCM 16244^T; 8, *S. ginsengisoli* KCTC 12539^T; 9, *S. humi* DSM 18929^T; 10, *S. indicatrix* DSM 28278^T; 11, *S. koreensis* JCM 13256^T; 12, *S. lactitubi* DSM 104152^T; 13, *S. maltophilia* DSM 50170^T; 14, *S. nitritireducens* JCM 13311^T; 15, *S. pavanii* DSM 25135^T; 16, *S. pictorum* JCM 9942^T; 17, *S. rhizophila* JCM 13333^T; 18, *S. terrae* DSM 18941^T; 19, *S. tumulicola* JCM 30961^T.

dDDH values > 70 % were highlighted in red.

JCM 16244 ^T														
S.	LDJM00000000	21.1	20.6	27.2	21.2	20.7	20.7	21.7	100.					
<i>ginsengisoli</i>									0					
KCTC														
12539 ^T														
<i>S. humi</i> DSM	LDJI00000000	24.8	22.2	21.0	25.8	22.6	22.1	26.0	21.1	100.				
18929 ^T										0				
<i>S. indicatrix</i>	PEJS00000000	22.9	25.2	20.9	23.3	23.3	25.6	23.5	21.0	22.5	100.			
DSM 28278 ^T										0				
<i>S. koreensis</i>	LDJH00000000	21.6	20.5	78.3	21.7	20.6	20.6	22.1	27.4	21.2	20.9	100.		
JCM 13256 ^T										0				
<i>S. lactitubi</i>	PHQX00000000	22.8	25.1	21.0	23.1	24.9	25.5	23.6	20.7	22.3	52.5	21.0	100.	
DSM											0			
104152 ^T														
<i>S. maltophilia</i>	BCUI00000000	23.3	25.2	21.4	23.7	25.0	25.5	24.1	21.3	21.3	32.0	21.3	32.3	100.
DSM 50170 ^T											0			
<i>S. nitritireducens</i>	LDJG00000000	31.3	23.0	22.0	39.9	24.5	23.4	32.3	21.8	26.3	23.5	22.0	23.6	24.1
<i>s</i>	JCM										0			
13311 ^T														
<i>S. pavonii</i>	LDJN00000000	23.2	25.3	20.8	23.7	25.1	25.6	23.8	21.2	22.8	32.4	21.0	32.4	43.0
DSM 25135 ^T												0		
<i>S. pictorum</i>	LLXS00000000	25.2	21.8	21.0	26.3	22.9	22.0	26.5	20.9	26.1	22.4	21.0	22.2	23.0
JCM 9942 ^T												0		
<i>S. rhizophila</i>	CP007597	23.3	24.5	21.0	23.7	29.9	24.5	24.2	21.1	22.8	24.8	21.1	24.5	24.7
JCM 13333 ^T												0		

Supplementary Table S6. Genetic features predicted from the genomes of strains CPCC 101365^T, CPCC 101269^T, and CPCC 101426 that may be potentially involved in plant growth-promotion and environmental stress tolerance. -, not detected.

Gene	Enzyme code	Product	CPCC 101365 ^T	CPCC 101269 ^T	CPCC 101426
Biosynthesis of indole-3-acetic acid					
<i>trpE</i>	EC 4.1.3.27	Anthranilate synthase component II	K5L01_RS12200	H4O12_RS06550	H4O09_RS01845
<i>trpG</i>	EC 4.1.3.27	Anthranilate synthase component I	K5L01_RS12210	H4O12_RS06555	H4O09_RS01850
<i>trpD</i>	EC 2.4.2.18	Anthranilate phosphoribosyltransferase	K5L01_RS12185	H4O12_RS06545	H4O09_RS01835
<i>trpF</i>	EC 5.3.1.24	Phosphoribosylanthranilate isomerase	K5L01_RS05645	H4O12_RS13755	H4O09_RS07325
<i>trpC</i>	EC 4.1.1.48	Indole-3-glycerol phosphate synthase	K5L01_RS12180	H4O12_RS06540	H4O09_RS01830
<i>trpA</i>	EC 4.2.1.20	Tryptophan synthase subunit alpha	K5L01_RS05625	H4O12_RS13770	H4O09_RS07280
<i>trpB</i>	EC 4.2.1.20	Tryptophan synthase subunit beta	K5L01_RS05635	H4O12_RS13765	H4O09_RS07290
	EC 1.2.7.8	Indolepyruvate ferredoxin oxidoreductase	K5L01_RS14095	H4O12_RS08805	-
Production and uptake of siderophores					
<i>aroA</i>	EC 2.5.1.19	3-phosphoshikimate 1-carboxyvinyltransferase	K5L01_RS04530	H4O12_RS02960	H4O09_RS05170
<i>aroC</i>	EC 4.2.3.5	Chorismate synthase	K5L01_RS05675	H4O12_RS13725	H4O09_RS10235
<i>aroQ</i>	EC 5.4.99.5	Gamma subclass chorismate mutase	-	H4O12_RS15100	-
<i>pheA</i>	EC 4.2.1.51	Prephenate dehydratase	K5L01_RS04535	H4O12_RS02955	H4O09_RS05165
<i>tonB</i>		TonB-dependent siderophore receptor	K5L01_RS06265, K5L01_RS09170, K5L01_RS09675, K5L01_RS04030	H4O12_RS03035, H4O12_RS03085, H4O12_RS03135, H4O12_RS04010, H4O12_RS04930, H4O12_RS08610, H4O12_RS00795	-
		TonB family protein	K5L01_RS04520, K5L01_RS08370	-	H4O09_RS12115, H4O09_RS14025
<i>exbB</i>		TonB-system energizer	K5L01_RS09450	H4O12_RS09260	H4O09_RS03300
<i>exbD</i>		Biopolymer transporter	K5L01_RS09455, K5L01_RS09460, K5L01_RS09455	H4O12_RS01135, H4O12_RS09250, H4O12_RS09255	H4O09_RS07115, H4O09_RS03290, H4O09_RS03295
<i>tolA</i>		Cell envelope integrity protein	K5L01_RS01620	H4O12_RS11265	H4O09_RS11665
<i>tolQ</i>		MotA/TolQ/ExbB proton channel family protein	K5L01_RS05475	H4O12_RS17900	H4O09_RS07120
<i>feoB</i>		Ferrous iron transporter	K5L01_RS03155	H4O12_RS16055	H4O09_RS06280

<i>ahpC</i>	Peroxiredoxin	-	H4O12_RS12965	H4O09_RS00175
<i>fiu</i>	Catecholate siderophore receptor	-	H4O12_RS03715	-
<i>bfr</i>	Bacterioferritin	-	H4O12_RS06475, H4O12_RS00590	H4O09_RS01770
<i>fecR</i>	periplasmic ferric-dicitrate binding protein	K5L01_RS01055, K5L01_RS02360, K5L01_RS05375, K5L01_RS12910	-	H4O09_RS10730

Supplementary Table S7. Genes related to heat shock and suicide mechanisms predicted from the genomes of strains CPCC 101365^T, CPCC 101269^T, CPCC 101442, *S. maltophilia* K279a, and *S. maltophilia* DSM 50170^T.

Gene	Product	Biological role	CPCC 101365 ^T	CPCC 101269 ^T	CPCC 101426	<i>S. maltophilia</i> K279a	<i>S. maltophilia</i> DSM 50170 ^T
<i>rpoH</i>	Sigma 32	Heat shock sigma factor	K5L01_RS12545	H4O12_RS06290	H4O09_RS09420	SMLT_RS20270	CKW06_RS21085
<i>dnaJ</i>	Chaperone DnaJ	Stress response	K5L01_RS02800	H4O12_RS02310	H4O09_RS04855	SMLT_RS09635	CKW06_RS10350
<i>dnaK</i>	Chaperone DnaK	Stress response	K5L01_RS02795	H4O12_RS02305	H4O09_RS04850	SMLT_RS09630	CKW06_RS10345
<i>grpE</i>	Chaperone complex	Prevents the aggregation of stress-denatured proteins	K5L01_RS02790	H4O12_RS02300	H4O09_RS04845	SMLT_RS09625	CKW06_RS10340
<i>groL</i>	Chaperonin GroEL	Heat shock chaperone	K5L01_RS09590	H4O12_RS06075	H4O09_RS01565	SMLT_RS20060	CKW06_RS20875
<i>htpG</i>	Heat shock chaperone	Heat shock response	K5L01_RS03590	H4O12_RS01760	H4O09_RS05765	SMLT_RS08735	CKW06_RS09760
<i>clpA</i>	ATPase with chaperone activity	Stress response	K5L01_RS04170	H4O12_RS04830	H4O09_RS13075	SMLT_RS11235	CKW06_RS11950
<i>hslV</i>	Protease	Degradation of abnormal/stress-denatured proteins	K5L01_RS10195	H4O12_RS14930	H4O09_RS09155	SMLT_RS19390	CKW06_RS20190
<i>htpX</i>	Protease	Degradation of abnormal/stress-denatured proteins due to heat shock	K5L01_RS05205	H4O12_RS14930	H4O09_RS13225	SMLT_RS15810	CKW06_RS16180

<i>lon</i>	Lon Protease	Degradation of abnormal/stress- denatured proteins	K5L01_RS07160	H4O12_RS11945	H4O09_RS14160	SMLT_RS04765	CKW06_RS04660
	Killer protein	Suicide mechanism	K5L01_RS05285	H4O12_RS11850	-	SMLT_RS06410, SMLT_RS17165, SMLT_RS20265, SMLT_RS00305, SMLT_RS01585	CKW06_RS17925, CKW06_RS21080, CKW06_RS00205, CKW06_RS01455
	Entericidin EcnAB	Suicide mechanism	K5L01_RS08140, K5L01_RS08145	H4O12_RS08170, H4O12_RS08175	H4O09_RS10945, H4O09_RS10950	SMLT_RS01620	CKW06_RS17925