

## **Supplemental information**

### **Molecular basis for antiviral activity of two pediatric neutralizing antibodies targeting SARS-CoV-2 Spike RBD**

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**Table S1. Summary of biolayer interferometry kinetic constants. Related to Table 1 and Figure S2C.**

Analyte	Immobilized ligand	Fitting mode	$k_a$ ( $M^{-1}s^{-1}$ )	$k_d$ ( $s^{-1}$ )	$K_D$ ( $\mu M$ )	$K_D$ reduction fold compared to $RBD_{WT}$
EH3 IgG	$RBD_{WT}$	1:1	$1.01 \times 10^5$	$6.09 \times 10^{-7}$	6.06	1
	$RBD_{E484K}$	1:1	$1.11 \times 10^5$	$4.46 \times 10^{-7}$	4.04	0.67
EH8 IgG	$RBD_{WT}$	1:1	$2.11 \times 10^5$	$3.55 \times 10^{-7}$	1.68	1
	$RBD_{E484K}$	1:1	$1.83 \times 10^5$	$6.01 \times 10^{-7}$	3.29	1.96

**Table S2. Data collection and model refinement statistics. Related to Figure 3.**

	<b>EH3-RBD</b>	<b>EH8-RBD</b>
<b>Data collection</b>		
Wavelength, Å	0.979	0.979
Resolution range, Å	34.38 - 2.65 (2.745 - 2.65)	46.91 - 2.49 (2.58 - 2.49)
Space group	P 2 <sub>1</sub> 2 <sub>2</sub> 1	P 2 <sub>1</sub> 2 <sub>2</sub> 1
Unit cell parameter		
a, b, c, Å	86.7, 103.1, 112.3	81.8, 100.7, 106.0
α, β, γ, °	90.0, 90.0, 90.0	90.0, 90.0, 90.0
Redundancy	6.6	18.8
Completeness, %	99.21 (98.85)	99.20 (92.82)
Mean I/sigma(I)	10.37 (2.28)	30.40 (2.67)
R <sub>merge</sub> <sup>a</sup>	0.190 (0.720)	0.144 (1.037)
R <sub>pim</sub> <sup>b</sup>	0.079 (0.301)	0.034 (0.259)
CC <sub>1/2</sub> <sup>c</sup>	0.933 (0.772)	0.996 (0.877)
Wilson B <sub>factor</sub> <sup>d</sup> (1/Å <sup>2</sup> ) <sup>d</sup>	39.62	57.01
<b>Refinement</b>		
R <sub>work</sub> <sup>e</sup>	0.179 (0.245)	0.244 (0.425)
R <sub>free</sub> <sup>f</sup>	0.221 (0.305)	0.270 (0.431)
Resolution, Å	34.38 - 2.65	46.91 - 2.49
# of non-hydrogen atoms		
proteins	4968	4829
ligands		
water	144	102
Overall B <sub>factor</sub> <sup>d</sup> (Å <sup>2</sup> )		
proteins	45.13	62.54
ligands	99.79	96.86
water	42.48	49.96
RMS (bond lengths), Å	0.004	0.005
RMS (bond angles), °	0.74	0.89
Ramachandran <sup>g</sup>		
Favored, %	97.11	96.96
Allowed, %	2.73	3.04
Outliers, %	0.16	0.00
PDB ID	7UL1	7UL0

Statistics for the highest-resolution shell are shown in parentheses.

<sup>a</sup>R<sub>merge</sub> =  $\sum |I - \langle I \rangle| / \sum I$ , where  $I$  is the observed intensity and  $\langle I \rangle$  is the average intensity obtained from multiple observations of symmetry-related reflections after rejections

<sup>b</sup>R<sub>pim</sub> = as defined in [45].

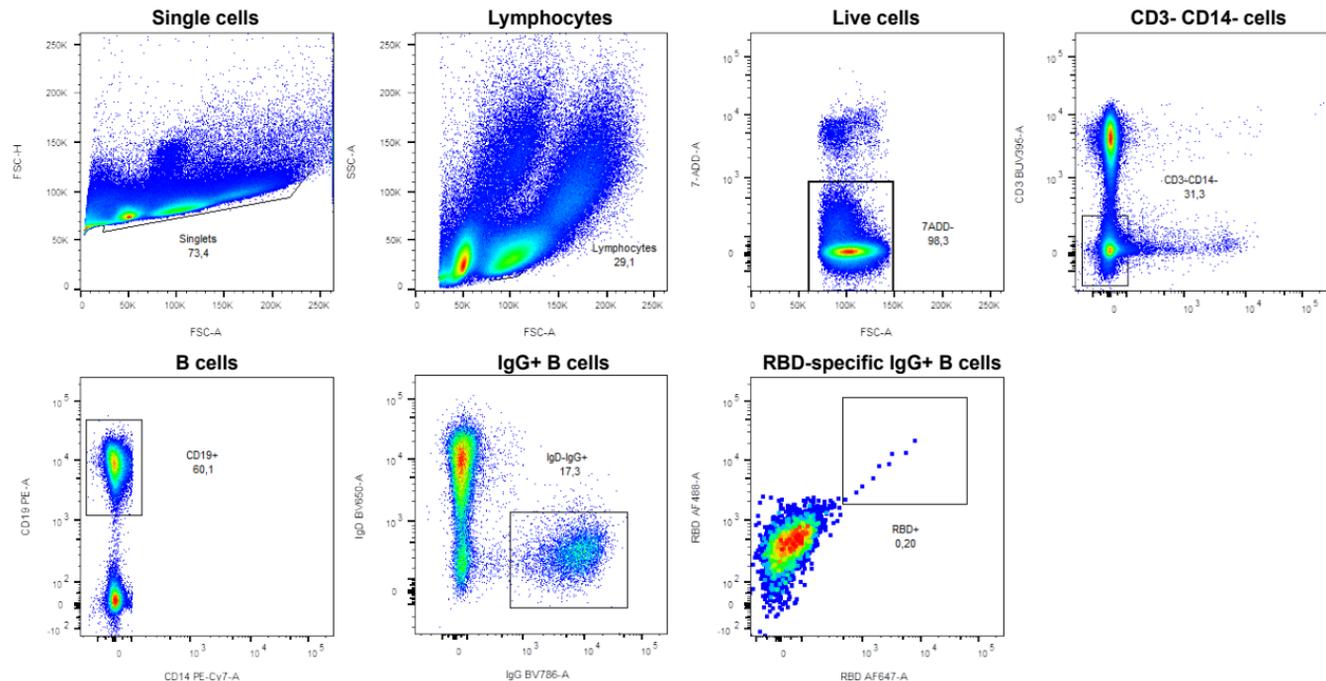
<sup>c</sup>CC<sub>1/2</sub> = as defined by Karplus and Diederichs [46]

<sup>d</sup>Wilson B<sub>factor</sub> as calculated in [47]

<sup>e</sup>R =  $\sum \|F_o| - |F_c\| / \sum |F_o|$ , where  $F_o$  and  $F_c$  are the observed and calculated structure factors, respectively.

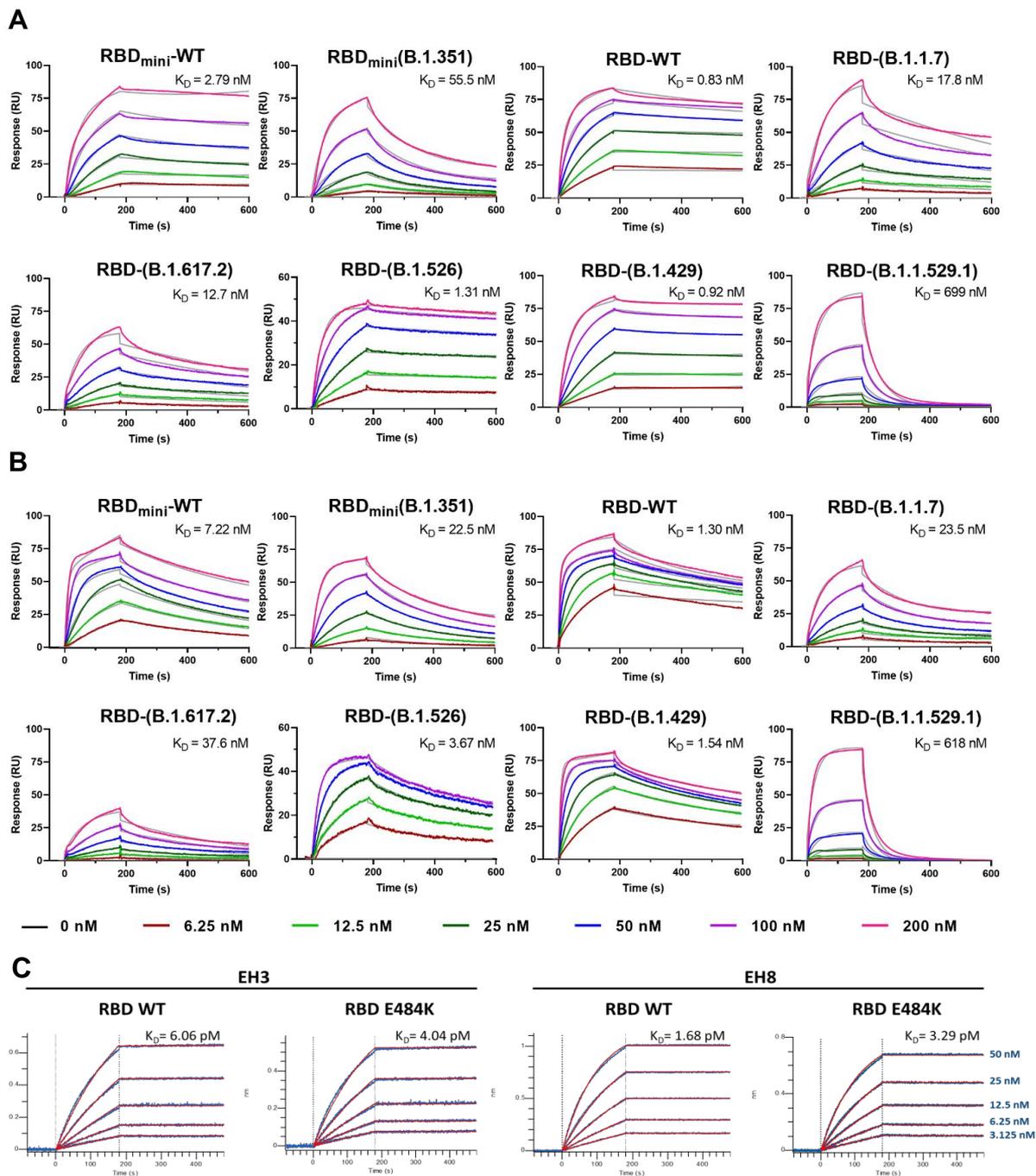
<sup>f</sup>R<sub>free</sub> = as defined by Brünger [48]

<sup>g</sup>Calculated with MolProbity [49].

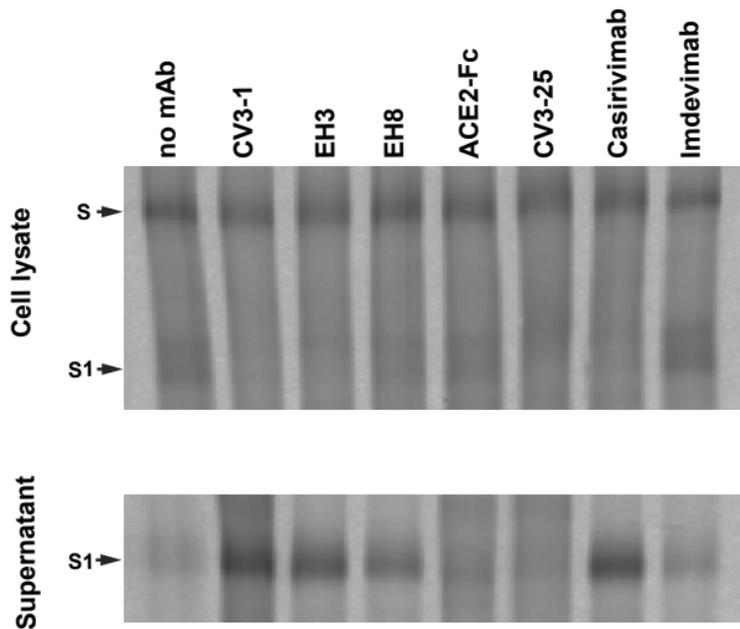


**Figure S1. Gating strategy for the isolation of RBD-specific mAbs. Related to Figure 1.**

Flow cytometry gates to identify RBD-specific B cells from PBMCs of a pediatric patient (Patient 12). RBD-specific B cells were identified according to cell morphology by light-scatter parameters and excluding doublets cells while gating on live cells (7-AAD<sup>-</sup>). Cells were then gated based on lineage markers: CD3<sup>-</sup> (T cell marker), CD14<sup>-</sup> (monocyte marker) and CD19<sup>+</sup> (B cell marker). Cells were further gated for isotypic expression: IgD<sup>-</sup> (naïve) and IgG<sup>+</sup> (class-switched). Finally, RBD-specific IgG<sup>+</sup> B cells were identified using a dual staining with fluorescent RBD probes (RBD-AF647 and RBD-AF488). The nine RBD-specific B cells were sorted as single cells to clone their BCR sequence.



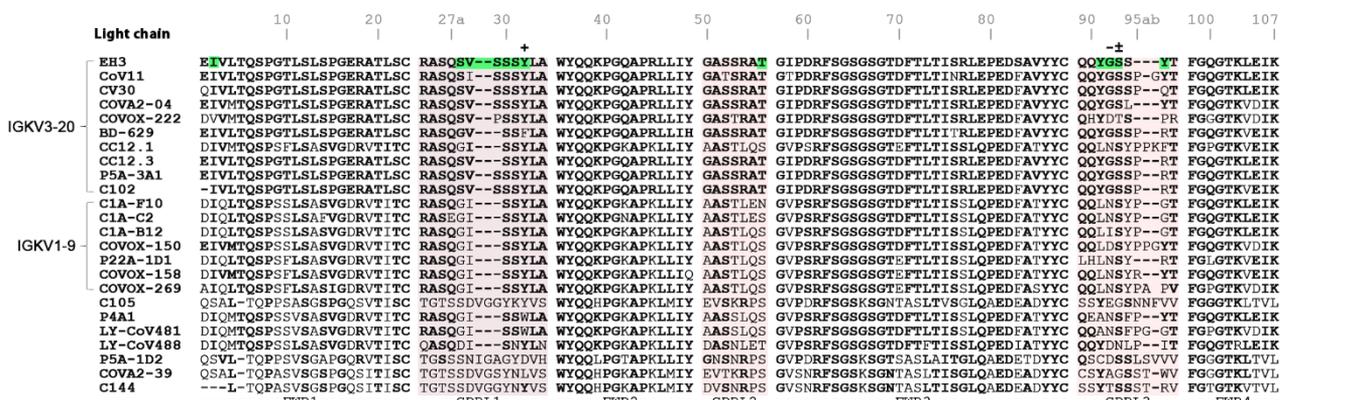
**Figure S2. Binding affinity of RBDs of SARS-CoV-2 VOCs to EH3 and EH8. Related to Table 1.**  
**(A-B)** SPR kinetic measurement of SARS-CoV-2 RBDs binding to immobilized **(A)** EH3 IgG and **(B)** EH8 IgG. Either IgGs (~250-350 RU) were immobilized on a Protein A chip and 2-fold dilutions of SARS-CoV-2 RBDs of WT and 6 VOCs (6.25-200nM) were injected as flow analytes. The experimental sensorgrams are colored in indicated colors and the 1:1 Langmuir fitting are in grey. RBD<sub>mini</sub> is the segment of residue 329-527 and otherwise are residue 319-537. The detailed SPR kinetic constants are listed in **Table 1**. **(C)** Binding kinetics between SARS-CoV-2 RBD (WT or E484K) and EH3 or EH8 mAbs assessed by bi-layer interferometry. Biosensors loaded with RBD proteins were soaked in two-fold dilution series of indicated mAbs (50 nM–3.125 nM) at 25°C. Raw data are shown in blue and 1:1 binding model is shown in red. The detailed bi-layer interferometry kinetic constants are listed in **Table S1**.



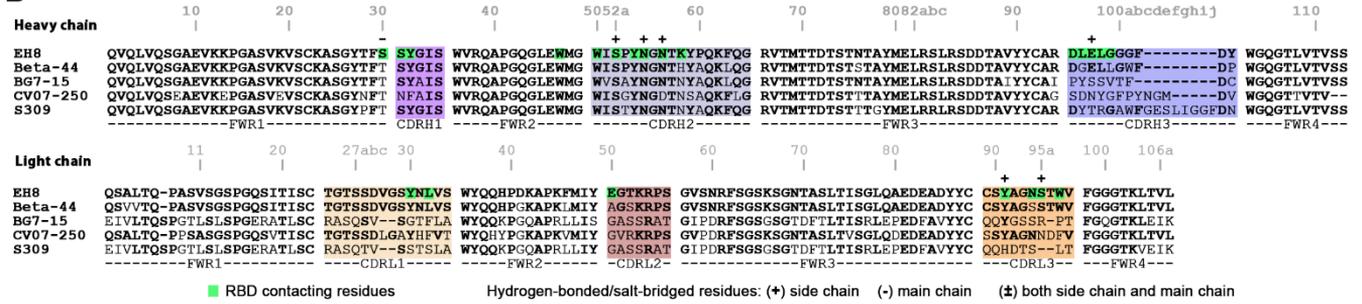
**Figure S3. The ability of RBD-specific mAbs to induce S1 shedding is epitope-dependent. Related to Figure 4.**

S1 shedding was evaluated by transfection of 293T cells to express SARS-CoV-2 S D614G followed by radiolabeling using  $^{35}\text{S}$ -methionine/cysteine mix in presence of RBD-specific mAbs (CV3-1, EH3, EH8), ACE2-Fc, S2-specific CV3-25, or two FDA-approved RBD-specific mAbs (Casirivimab and Imdevimab). This was followed by immunoprecipitation of cell lysates and supernatant with CV3-25 and a rabbit antiserum raised against SARS-CoV-2 RBD produced in-house. Samples were analyzed by SDS-PAGE and autoradiography.

**A**



**B**



**Figure S4. V<sub>H</sub> and V<sub>L</sub> sequence alignments of EH3 and EH8 to other structurally available RBD-specific mAbs. Related to Figure 4.**

(A) Alignments of EH3 and other 23 IGHV3-53 encoded Abs. (B) Alignments of EH8 and other 4 IGHV1-18 Abs. The RBD-contacting residues (BSA>0) in EH3-RBD and EH8-RBD crystal structures are shaded in green. Residues involved in salt-bridges or H-bonds to the RBD are marked above the sequence with (+) for the side chain, (-) for the main chain and (±) for both.