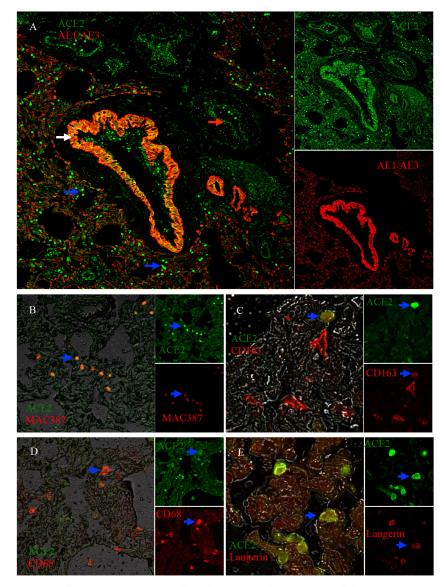
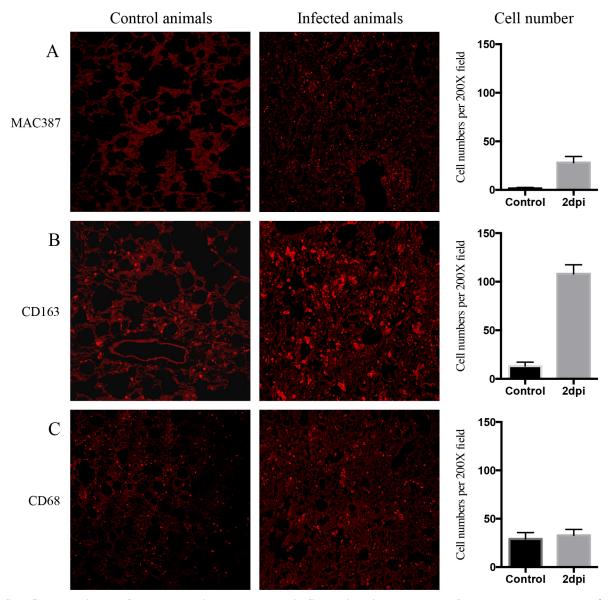


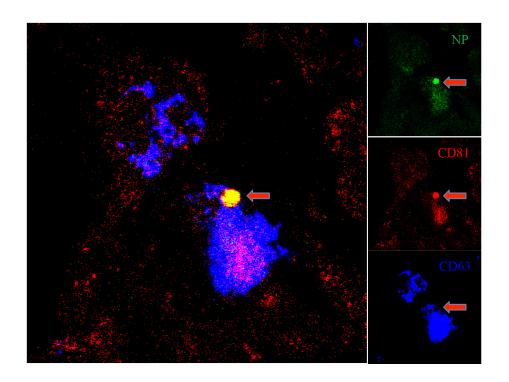
Supplementary Figure S1. Distribution of monocytes/macrophages and DCs in the upper respiratory mucosa. These sections are representative samples of pharyngeal mucosa tissue sections from infected animals at 2 dpi (right panel) or control animals (left panel) stained for SARS-CoV nucleoprotein, CD68, CD163, MAC387 and Langerin (400x). A. Viral NP and Langerin. B. Viral NP and CD68. C. Viral NP and CD163. D. Viral NP and MAC387. Figures A and B show that there are no significant changes in the numbers of Langerin⁺ cells (A, red) and CD68+ (B, red) cells in the pharyngeal mucosa at 2 dpi in infected animals compared with uninfected animals. C and D showed increased numbers of CD163⁺ (B, red) and MAC387⁺ (C, red) cells in the pharyngeal mucosa at 2 dpi in infected animals compared with uninfected animals.



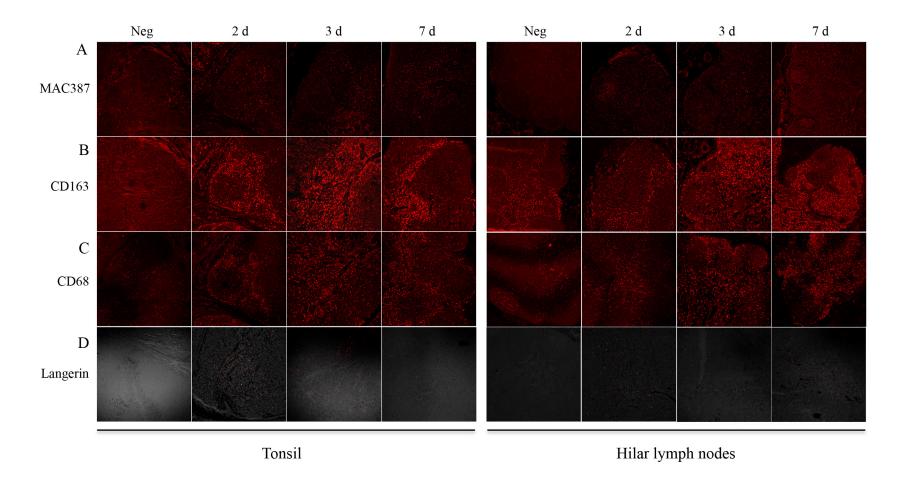
Supplementary Figure S2. ACE2 expression on monocytes/macrophages and Langerhans cells. These sections are representative samples of SARS-CoV receptor (ACE2)⁺ cells in the lungs that are associated with the respiratory epithelium (A, white arrows), endothelium (A, red arrow), and MAC387⁺ (B, blue arrow), CD163⁺ (C, blue arrow), CD68⁺ (D, blue arrow) and Langerin⁺ cells (E, blue arrow). Lung tissue samples are stained for ACE2 (FITC), AE1/AE3 (TRITC) (A, 100x), MAC387 (TRITC) (B, 400x), CD163 (TRITC) (C, 630x), CD68 (TRITC) (D, 400x) and Langerin (TRITC) (E, 630x). In each figure, the left panel shows a low magnification overview. The right panel shows the single colors from the left panel.



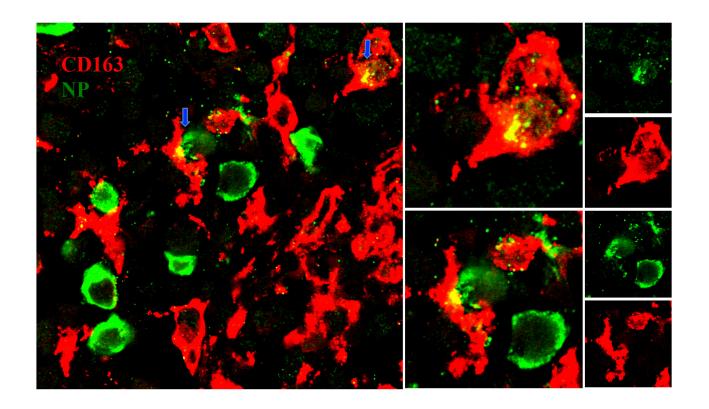
Supplementary Figure S3. Comparison of monocyte/macrophage infiltration in the lungs from each monkey. These sections are representative 2-dpi lung tissues from infected animals (middle panel) or control animals (left panel) that were stained for MAC387 (A), CD163 (B) and CD68 (C). Figure A, B and C shows increased number of CD68+ (A, red), CD163+ (B, red) and MAC387+ (C, red) cells in the lungs at 2 dpi in infected animals compared with uninfected animals. The right panel shows the comparison of numbers of monocytes/macrophages between the infection group and the control group. Horizontal bars represent numbers of positive cells in a 200x field.



Supplementary Figure S4. SARS-CoV virions accumulate in a CD81+/CD63- intracellular compartment in macrophages. Lung tissue samples were stained for SARS-CoV nucleoprotein (FITC), CD81 (TRITC) and CD63 (blue). The left panel shows a merged image. The right panel shows the single colors from the middle panel. This section is a representative sample of the viral antigen (NP)-positive signals that are in the CD81-rich and CD63-negative intracellular monocyte/macrophages compartment in the lungs (Red arrow, 600x).



Supplementary Figure S5. Comparison of the accumulation of monocytes/macrophages and Langerhans cells in the tonsils and hilar lymph nodes of each macaque. These sections are representative tissue samples of tonsils (left panel) and hilar lymph nodes (right panel) from infected (2, 3 and 7 dpi) or control animals that were stained for MAC387 (A), CD163 (B), CD68 (C) and Langerin (D). Figure A shows no significantly increased number of MAC387⁺ cells in the lymphoid tissue section of infected animals in comparison with uninfected animals. Figures B, C and D show increased numbers of CD163⁺ (B), CD68⁺ (C) and Langerin⁺ (D) cells in the tonsils at 2 dpi and hilar lymph nodes at 3 dpi in infected animals, respectively, compared with uninfected animals (200x).



Supplementary Figure S6. Infectious synapse formed between CD163⁺ cells and NP⁺ cells. Hilar lymph nodes are stained for SARS-CoV NP (FITC) and CD163 (TRITC). Left panel shows a low magnification overview (600x). Right panel shows a higher magnification of the boxed area in left panel. These sections are representative samples of infectious synapses formed between CD163⁺ monocytes and infected cells (blue arrows) and trans-infection, which is accomplished via the recruitment of virus to the sites of cell-cell contact and transfer of virions at the infectious synapse into the cytoplasm.