Supporting Information for

Genetically Encoded Aryl Alkyne for Raman Spectral Imaging of Intracellular a-

Synuclein Fibrils

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- Table S1. Raman F_{CC} peak positions for Figure 4
- Figure S1. Raman spectra for F_{CC} in water/DMSO mixture shown in Figure 2
- Figure S2. Fingerprint region of the Raman spectra for proteins shown in Figure 4
- Figure S3. Corresponding wide-field fluorescence image of Figure 5.
- Figure S4. *z*-Step scans showing cellular internalization of F_{CC} fibrils.
- Figure S5. Corresponding Raman spectra for Figure 7.

Figure S6. Additional Raman maps collected for internalized Y39F_{CC} fibrils.

Protein	Soluble v_{max} (cm ⁻¹)	Fibrillar v_{max} (cm ⁻¹)	$\Delta v_{max} (\mathrm{cm}^{-1})$
F4F _{CC}	2109.6	2108.9	-0.7
Y39F _{CC}	2109.3	2107.8	-1.5
F94F _{CC}	2108.9	2105.9	-3.0
Y125F _{CC}	2108.7	2107.7	-1.0

Table S1. Raman shifts[‡] of the F_{CC} alkyne stretching band in α -syn constructs in the soluble and aggregated states shown in **Fig. 4**.

^{\pm} Raman shifts were determined by fitting the alkyne stretching band to a Lorentzian function, which was found to have an accuracy of ± 1 cm⁻¹.



Figure S1. C=C stretching bands of 10 mM (**A**) F_{CC} and (**B**) HPG in solvent mixtures from 100% DMSO (red) to 100% phosphate buffer (20 mM sodium phosphate, 100 mM NaCl, pH 7.4, blue).



Figure S2. Fingerprint region from the Raman spectra of fibrillar $F4F_{CC}$ (blue), Y39F_{CC} (green), F94F_{CC} (orange), and Y125F_{CC} (red). Solid lines represent averages across multiple aggregates with shaded areas indicating the standard deviation (n = 8).



Figure S3. Combined bright-field and wide-field fluorescence image of the cell shown in **Figure 5**. The fluorescence signal represents ThT emission from internalized F_{CC} fibrils.



Figure S4. Fibril clusters are fully internalized within SH-SY5Y cells. (**A**, **C**) Raman maps of lipids (magenta, methylene C–H stretching band 2825–2860 cm⁻¹), nucleotides (blue, nucleotide ring breathing mode 1557–1587 cm⁻¹), and F94F_{CC} fibrils (yellow, C=C stretching band 2090–2130 cm⁻¹). Scale bars are 5 μ m. (**B**, **D**) *z*-Position scans recorded at the fibril clusters marked in (**A**) and (**C**) at a single location (blue squares indicated by white arrows) showing lipids (magenta, 2825–2860 cm⁻¹), proteins (cyan, 2900–2950 cm⁻¹), and F94F_{CC} fibrils (yellow, 2090–2130 cm⁻¹). Data were collected with 1-s accumulation at each *z*-position with 0.2 μ m steps. The cell in (**A**) is the same cell shown in **Figure 5**.



Figure S5. Raman spectra from the individual pixels (blue boxes) as indicated in the corresponding panels in **Figure 7 A**, **D**, **G**, and **K**. Amide-I (**A**, **D**, **G**, **K**), C=C stretching (**B**, **E**, **H**, **L**), and C–H stretching (**C**, **F**, **J**, **M**) regions are shown as red curves. The *in vitro* fibril spectrum is also shown as a reference (black curves) in each panel. Dashed lines serve as guides for comparison.



Figure S6. Cellular treatments with Y39F_{CC} fibrils. (**A**, **E**, **J**) Raman maps of lipids (magenta, methylene C–H stretching band 2825–2860 cm⁻¹), nucleotides (blue, nucleotide ring breathing mode 1557–1587 cm⁻¹), and Y39F_{CC} fibrils (yellow, C=C stretching band 2090–2130 cm⁻¹). Scale bars are 5 μ m. Raman spectra (red) of the amide-I (**B**, **F**, **K**), C=C stretching (**C**, **G**, **L**), and C–H stretching regions (**D**, **H**, **M**) collected from the individual pixels in (**A**, **E**, and **J**, blue boxes indicated by white arrows) are shown. Raman spectra collected for *in vitro* Y39F_{CC} fibrils are also shown for comparison (black) with the difference spectra (cell–*in vitro* fibril, blue) shown below. Data are normalized to the intensity of the C=C stretching band. Dashed lines are shown as guides for comparison.