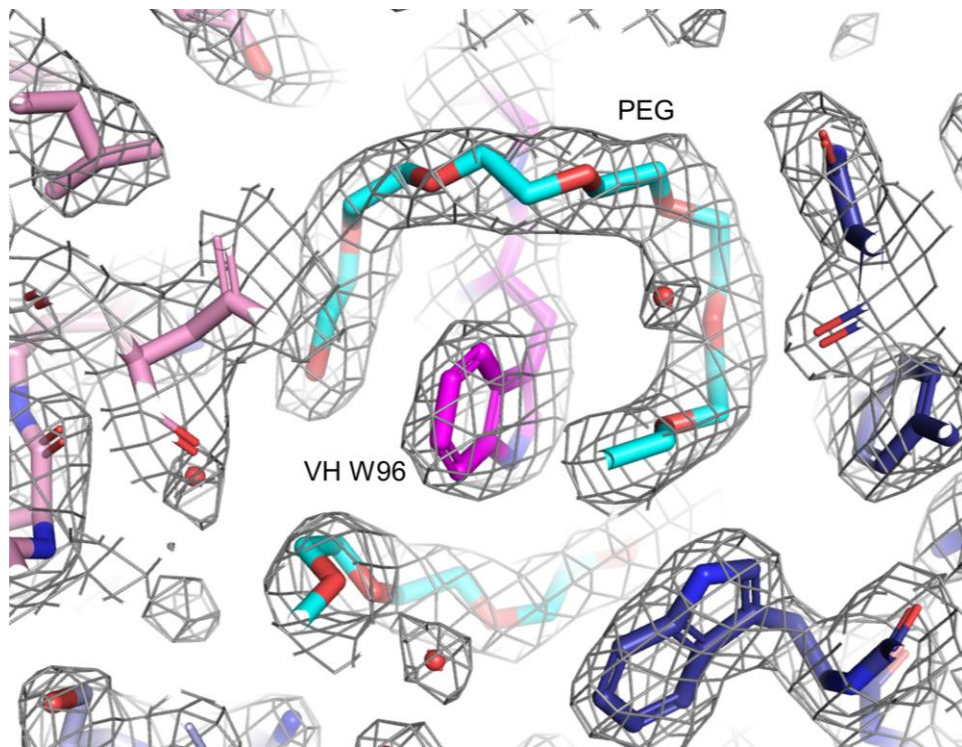


1 **Structure of an anti-PEG antibody reveals an open ring that captures highly**
2 **flexible PEG polymers**

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4 Huckaby et al.

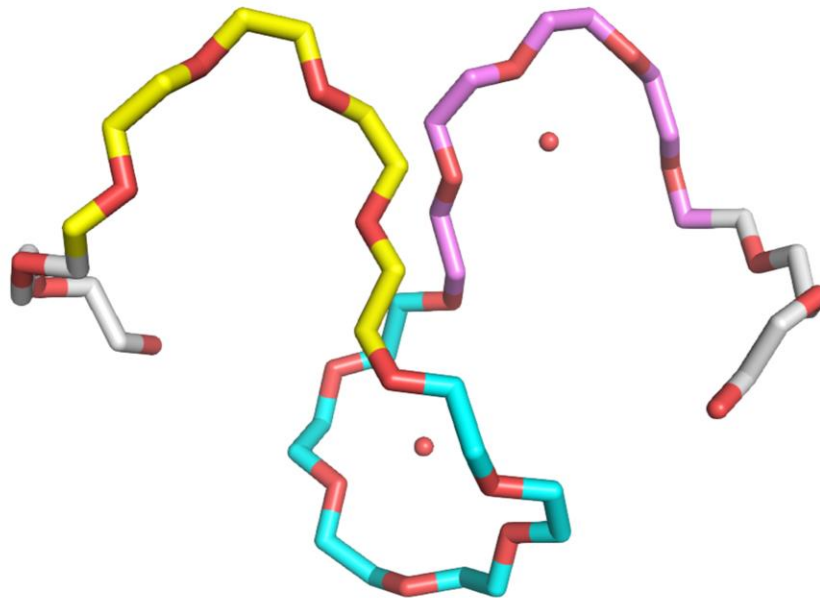
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Supplementary Figure 1: Representative electron density for the Fab-PEG interface in the crystal structure

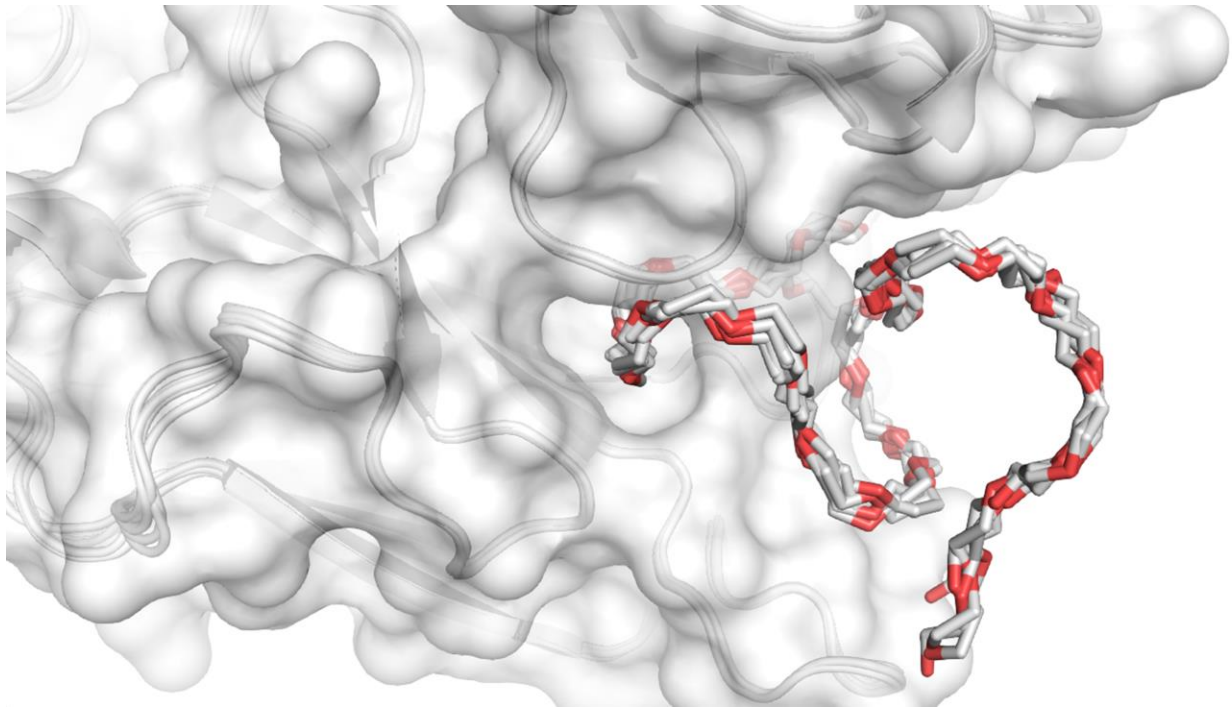
Image showing the 2Fo-Fc electron density map for the crystal structure contoured at a 1sigma level to show the quality of the data in the vicinity of the PEG molecule and the correspondingly bound Fab.



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Supplementary Figure 2: PEG polymer antigen colored by semi-circular domain with relevant solvent molecules added

Water solvent molecules make alternating hydrogen bonds to ether oxygen atoms of the PEG polymer chain, contributing to the various semi-circular domains (colored yellow, cyan, and magenta) and overall spiral shape of the PEG molecule while in complex with the anti-PEG Fab.

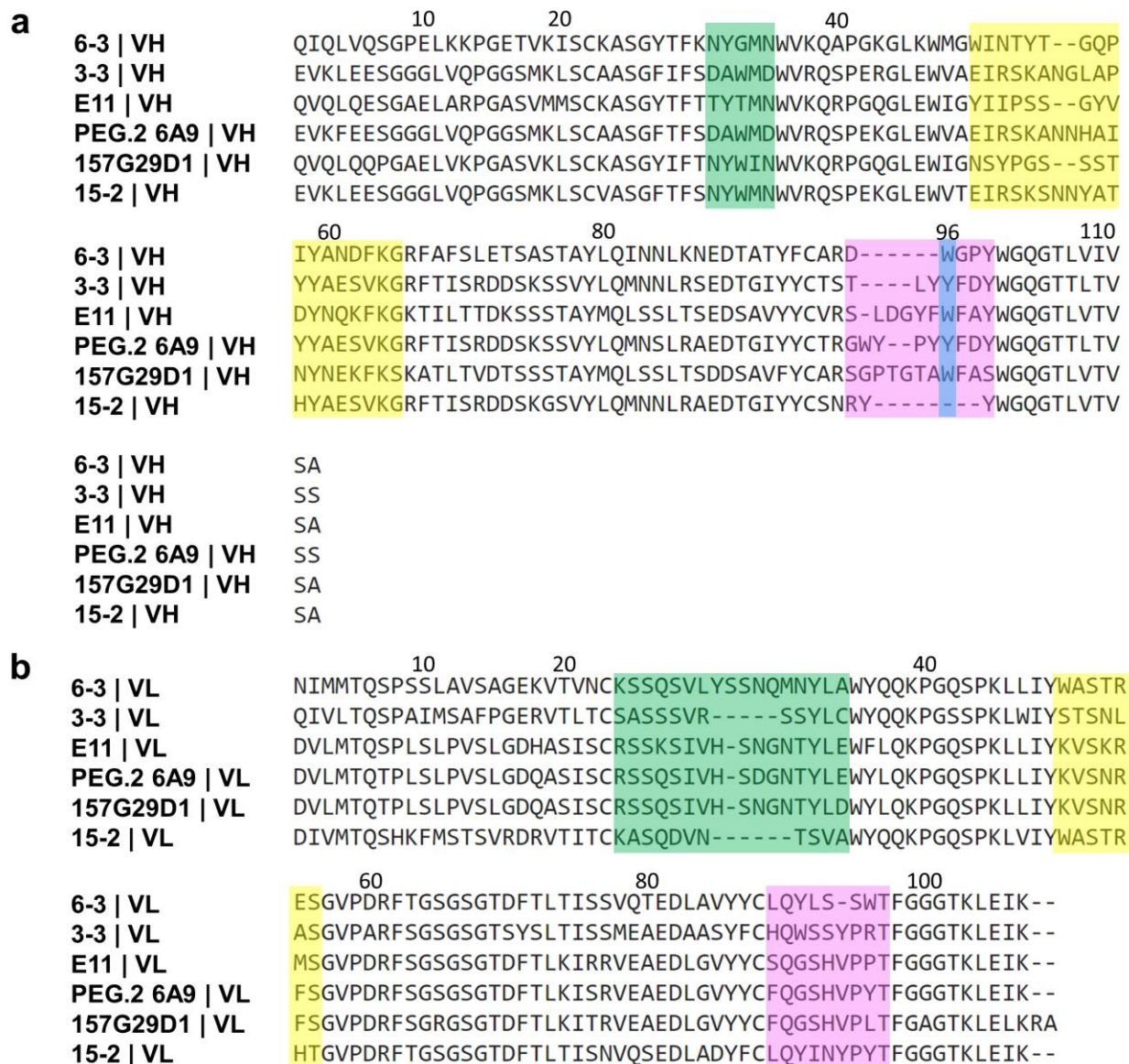


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25 **Supplementary Figure 3: Superposition of all antibody-antigen structure**
26 **complexes**

27 Superimposing all complexes from both of the crystal structures reveals strong
28 alignment for protein atoms of the Fab molecules as well as atoms along the PEG
29 antigens, but alignment begins to diverge near the terminal ends of the polymer chains.
30 This divergent behavior is likely a result of inherent PEG polymer chain flexibility distal
31 to the antigen-binding interface region of the complex.

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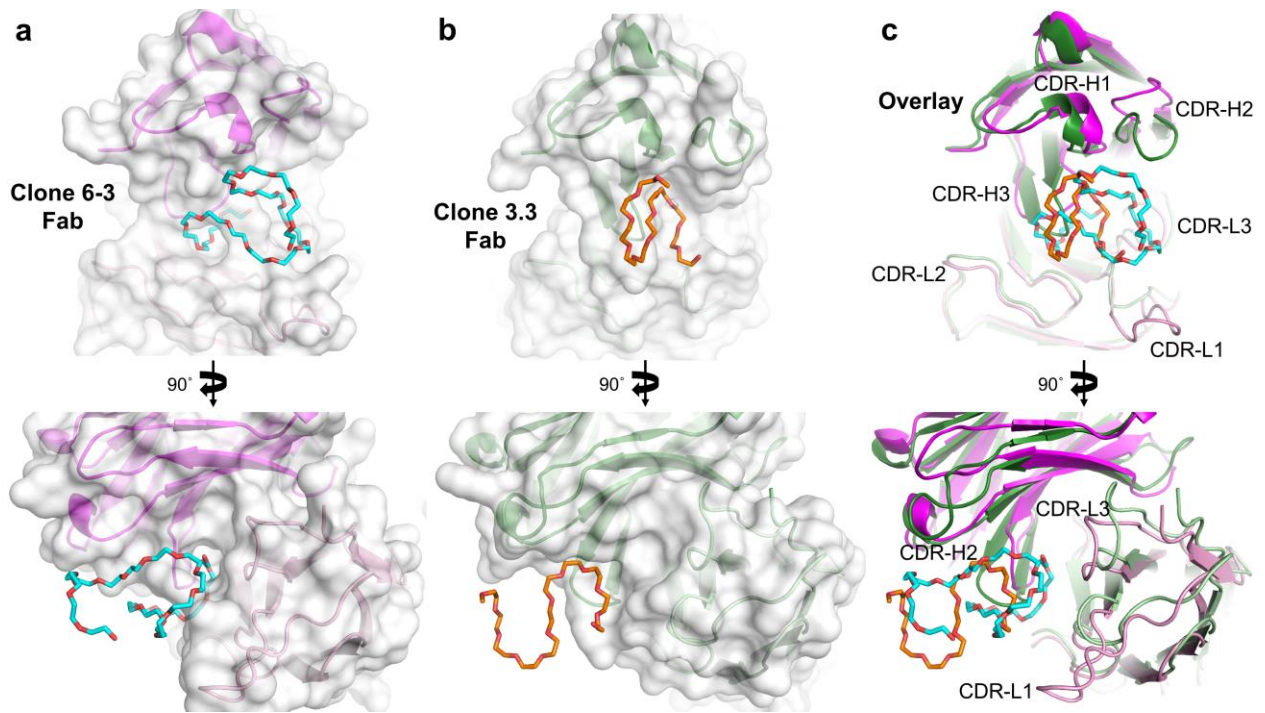
35 **Supplementary Figure 4: Variable domain amino acid sequence alignments for**
36 **various APA clones**

37 **a** Variable heavy (VH) domain amino acid sequence alignments for six different APA
38 clones (6-3, 3-3, E11, PEG.2 6A9, 157G29D1, and 15-2) with Kabat numbering and
39 CDRs highlighted (CDR1, green; CDR2, yellow; CDR3, pink). The 96th residue is
40 labeled and highlighted blue to show conservation of the ring-forming tryptophan (W) or

41 similarly bulky and hydrophobic tyrosine (Y) residue across other APA clones in the VH
42 domain.

43 **b** Variable light (VL) domain amino acid sequence alignments for six different APA
44 clones (6-3, 3-3, E11, PEG.2 6A9, 157G29D1, and 15-2) with Kabat numbering and
45 CDRs highlighted (CDR1, green; CDR2, yellow; CDR3, pink).

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49 **Supplementary Figure 5: Overlay of two different APA Fab clone structures**

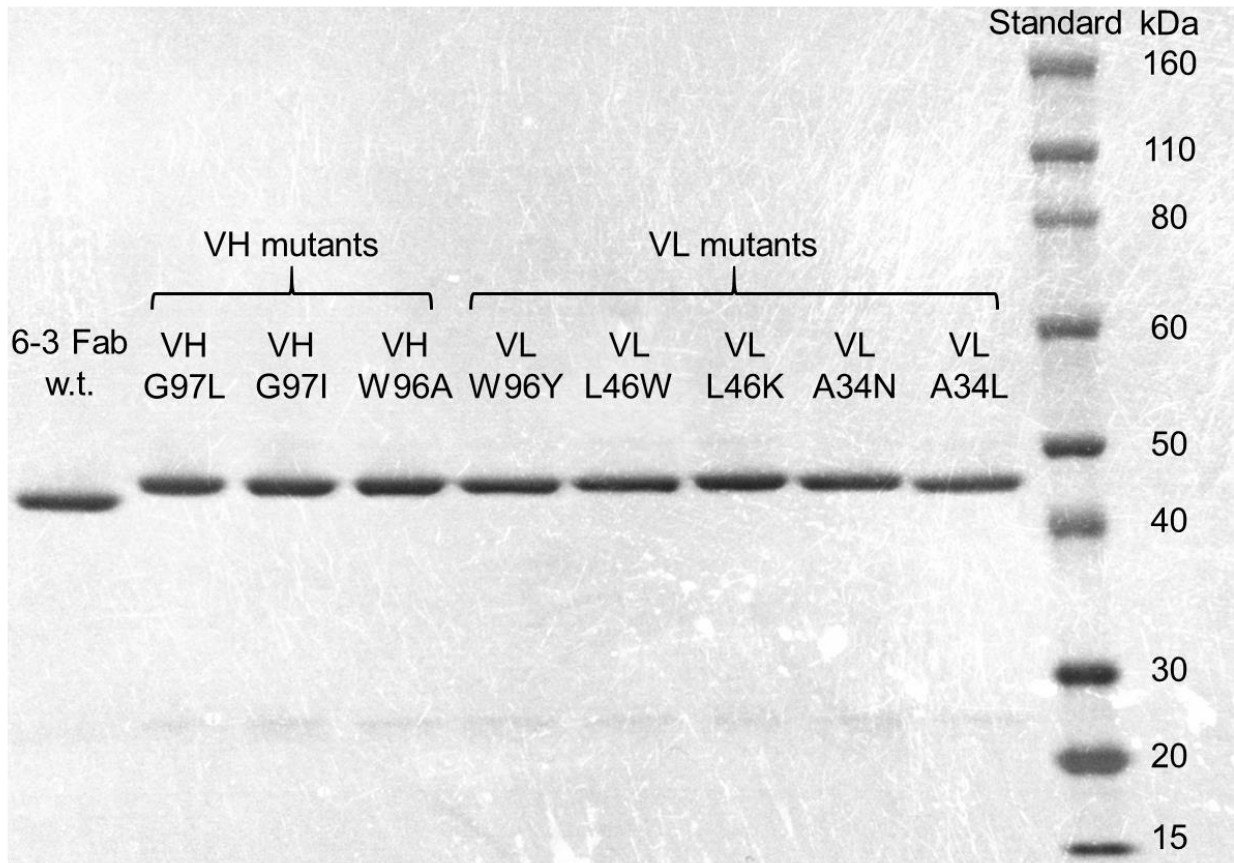
50 **reveals different binding mechanisms to PEG backbone**

51 **a** Ribbon (magenta) and surface model representations of clone 6-3 Fab structure in
 52 complexation with PEG backbone antigen (carbon, cyan; oxygen, red).

53 **b** Ribbon (green) and surface model representations of clone 3.3 Fab structure in
 54 complexation with PEG backbone antigen (carbon, orange; oxygen, red).

55 **c** Overlay of clone 6-3 Fab (magenta) and clone 3.3 Fab (green) structures represented
 56 as ribbon models in complex with PEG backbone antigens. Complementarity-
 57 determining regions (CDRs) in the variable domains at the antigen-binding interface are
 58 labeled to show structural differences in the hypervariable regions leading to different
 59 PEG binding mechanisms.

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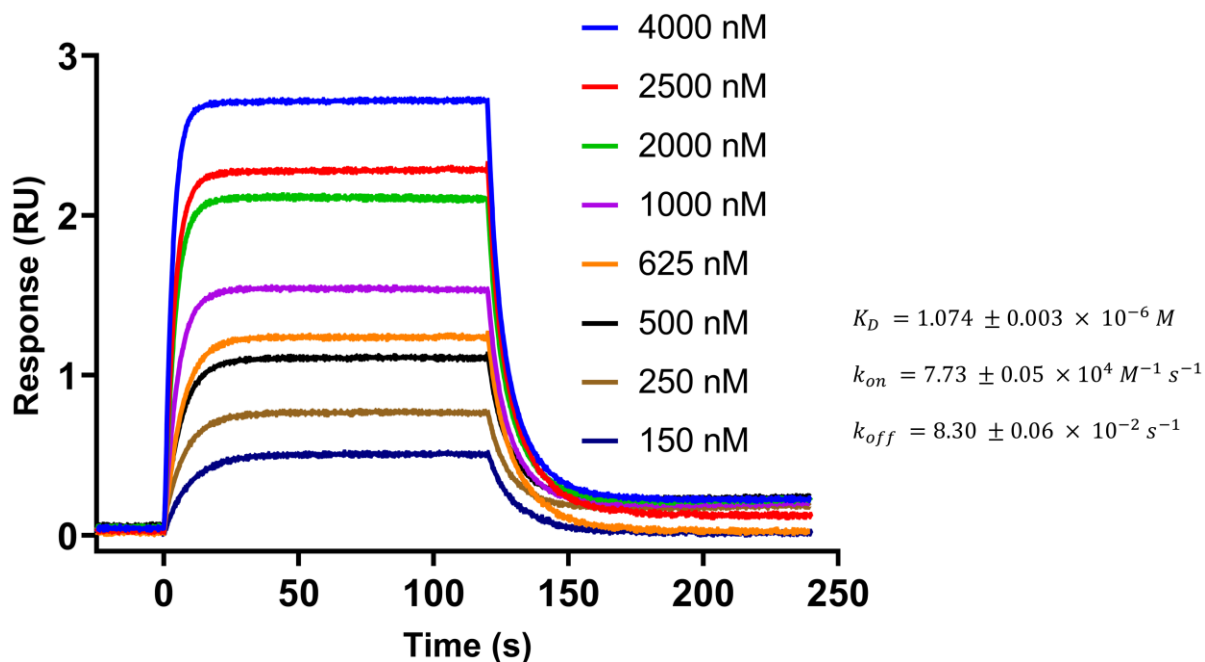
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63 **Supplementary Figure 6: Size and purity of wild type (w.t.) and mutant anti-PEG**

64 **Fabs**

65 SDS-PAGE stained with Coomassie revealed high purity (single stained protein band
66 per sample well) and similar molecular sizes (~ 50 kDa) across all APA Fabs expressed,
67 including wild type (w.t.) and mutant samples.

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71 **Supplementary Figure 7: Anti-PEG Fab binding kinetics and affinity to PEG**
72 **polymer backbone antigen**

73 Bio-layer interferometry (BLI) was used to measure the binding affinity constant (K_D),
74 association rate constant (k_{on}), and dissociation rate constant (k_{off}) between APA Fab
75 and PEG antigen. Binding to immobilized PEG was measured at various APA Fab
76 concentrations above and below the calculated binding affinity constant.

77 **Supplementary Table 1: Crystallographic data collection and model refinement**
 78 **statistics**
 79

	6VL9	6VL8
Space group	P 21 21 2	P 1 21 1
Unit cell parameters <i>a</i> , <i>b</i> , <i>c</i> (Å)	90.711, 169.788, 69.394	112.108, 87.791, 116.074
Unit cell parameters α , β , γ (°)	90.00, 90.00, 90.00	90.00, 113.66, 90.00
Resolution (Å)	43.819-2.634 (2.700-2.634) ¹	41.925-2.423 (2.484-2.423)
No. of reflections collected	189233	477285
No. of unique reflections	31825	78203
CC1/2	0.973 (0.863)	0.981 (0.916)
CC*	0.993 (0.963)	0.995 (0.978)
Mean I/ σ I	11.9 (3.08)	17.07 (2.42)
R _{pim}	0.86 (0.238)	0.075 (0.262)
Completeness (%)	98.0 (100.0)	99.8 (98.6)
Redundancy	5.9 (6.4)	6.1 (4.5)
<i>R</i> _{work} / <i>R</i> _{free} (%)	20.95/26.28 (27.33/33.31)	20.76/25.67 (28.57/31.00)
Number of protein atoms (non-hydrogen)	6545 (2 Fabs in the AU)	13106 (4 Fabs in the AU)
Number of ligand atoms	59	144 unique (288 total)
Number of solvent atoms (non-hydrogen)	78	168
Average B value for protein atoms (Å ²)	38.10	44.78
Average B value for ligand atoms (Å ²)	36.98	41.34
Average B value for solvent atoms (Å ²)	31.81	37.63
RMSD bond lengths (Å)	0.002	0.003
RMSD bond angles (°)	0.542	0.583
Clashscore	3.77	3.85
Ramachandran favored (%)	96.20	96.33
Ramachandran outliers (%)	0.36	0.36
¹ Values in parentheses correspond to the highest resolution shell.		

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