Supplemental Methods

Antibodies used to determine surface expression of various markers on cell lines by flow cytometry

PerCP-Cy5.5-conjugated antibodies used were anti-CD38 (BD Biosciences, cat # 561106), anti-CD46 (cat# 352413), anti-CD55 (cat# 311315), anti-CD20 (cat# 302326), and IgG1 and IgG2b isotype controls (cat #s 400150 and 400338, respectively) from Biolegend. FITC-conjugated antibodies used were anti-CD59 (Biolegend, cat#304706) and its isotype control (Sigma, cat # F6522).

Blocking antibodies used in CDC

Blocking antibodies used were IgG1 anti-CD55 antibody (BRIC-216) (ThermoFisher, cat# MA1-91161) and IgG2 anti-CD59 antibody (MEM-43) (Novus Biologicals, Littleton, CO, cat# NB500-330). Isotype matched antibodies used were IgG1 control antibody (ThermoFisher, cat# 16-4714-85) and IgG2 control antibody (Novus, cat# MAB003R).

Supplemental Figure Legends

Supplemental Figure 1. Flow cytometry showing gating strategy and phenotypic profiles of PEL cells, B cells, and plasma cells from patients. Representative flow cytometric analyses are shown for cells in the pleural fluid of one PEL patient (A) Flow cytometry analysis showing gating strategy for normal B cells (blue) and plasma cells (green) using antibodies against CD19, CD20, and CD38. (B) Representative flow cytometry showing combinational alternative non-CD38 gating strategy used to identify PEL cells (red circles) for the purpose of CD38 measurement. (C) Representative flow cytometric analysis showing the phenotypic profiles of PEL cells (red), B cells (blue) and plasma cells (green). The PEL cells express CD38 but lacks CD19 and CD20. The normal plasma cells express both CD19 and bright CD38 but are negative for CD20. Normal B cells express CD19, CD20, and variable CD38.

Supplemental Figure 2. Dara does not lead to CDC of PEL cell lines in part due to high expression of complement-inhibitory proteins. (A) CDC in the presence of 20% pooled normal human serum was performed on BCBL-1, BC-1, BC-3, BC-2, and JSC-1 PEL cell lines, treated with 0, 1, 10, or 100 μg/mL Dara. Cells were stained with propidium iodide (PI) and % lysis was determined based on PI-positive cells using flow cytometry after 2 hours. (B) Levels of complement-inhibitory proteins CD46, CD55, and CD59 on the surface of Daudi or PEL cell lines were measured by flow cytometry after staining the cells with PerCP-Cy5.5-conjugated anti-CD46 or anti-CD55 antibodies or FITC-conjugated anti-CD59 antibody. (C) Levels of CD46, CD55, and CD59 on PEL cell lines expressed as fold changes over Daudi from histograms in (B).

Supplemental Figure 3. CD38 expression and growth of ATRA-treated PEL cell lines. (A) BC-2 and JSC-1 cell lines were treated with 0 to 40 nM ATRA for 96 hours and levels of surface CD38 were measured by flow cytometry using PerCP-Cy5.5 conjugated anti-CD38 antibody. Data shows CD38 levels expressed as median fluorescent intensity (MFI). (B) Levels of surface CD38 on BC-2, JSC-1, BCBL-1, BC-1, and BC-3 cell lines treated with DMSO (0 nM ATRA) or indicated concentrations of ATRA for 72 hours as measured by flow cytometry using PerCP-Cy5.5 conjugated anti-CD38 antibody. (C) PEL cell lines were treated for 72 hours with DMSO control or 10 nM ATRA and then live cell number was counted after trypan blue-staining. Live cell number in ATRA-treated cells is presented as a percentage of DMSO-treated cells.

Supplemental Figure 4. ATRA does not change the levels of complement-inhibitory proteins in PEL cell lines. Levels of complement-inhibitory proteins CD46, CD55, and CD59 on the surface of BC-2 and JSC-1 cell lines were measured 72 hours after treatment with DMSO (0 nM ATRA) or indicated concentrations of ATRA by flow cytometry after staining the cells with PerCP-Cy5.5-conjugated anti-CD46 or anti-CD55 antibodies or FITC-conjugated anti-CD59 antibody.

Supplemental Figure 5. PBMC-mediated ADCC of BC-2 and JSC-1 cell lines. BC-2 and JSC-1 cells were treated with DMSO control or ATRA (10 nM) or Pom (1μM for BC-2 and 0.5μM for JSC-1) for 3 days prior to measuring PBMC-mediated ADCC using calcein-AM release assay. Data is presented as lysis induced by 0.1 and 1 μg/mL Dara as a percent of that by 0 Dara. (A and B) Lysis of DMSO control-treated BC-2 (A) and JSC-1 (B) cells induced by 0, 0.1, or 1 μg/mL Dara. (C and D) Lysis of DMSO, ATRA, or Pom-treated BC-2 (C) or JSC-1 (D)

cells induced by 0.1 or 1 μg/mL Dara. Data represents averages and standard deviations from 4 separate experiments. Statistically significant difference (*P≤0.05) between 0 and 1 μg/mL ATRA as calculated using 2-tailed t-test is indicated.

Supplemental Figure 6. Levels of CD59 in pomalidomide-treated PEL cell lines. BC-2 and JSC-1 cell lines were treated with DMSO (0 Pom) or indicated concentrations of Pom for 72 hours and levels of surface CD59 were measured by flow cytometry using FITC-conjugated anti-CD59 antibody. (B) shows histograms from one representative experiment. (C) shows median fluorescent intensity (MFI) for CD59 on DMSO and Pom-treated cells expressed as average from 3 separate experiments. Error bars represent standard deviations.

Supplemental Figure 7. Level of CD20 in ATRA and pomalidomide-treated PEL cell lines. PEL cell lines BCBL-1 and BC-3 were treated for 72 hours with indicated concentrations of ATRA, pomalidomide, or DMSO control, and surface level of CD20 was measured by flow cytometry using PerCP-Cy5.5 conjugated anti-CD20 antibody. (A) Level of CD20 in BCBL-1 and BC-3 cells after 10 nM and 100 nM ATRA treatment. (B) Level of CD20 in BCBL-1 and BC-3 cells after 1 μM or 10 μM pomalidomide treatment.

Supplemental Figure 8. CD38 levels on PEL cells from patients treated with Dara. Flow cytometric analysis showing the phenotypic profiles of PEL cells (red) isolated from patients 1 and 2 prior to starting Dara-treatment. PEL cells were identified using a panel of antibodies as described in materials and methods. CD38, CD45, and CD19 levels on PEL cells from cerebrospinal fluid (CSF) (A) and peritoneal fluid (B) of patient 1, and from CSF of patient 2 (C)

are shown. The median MFIs for CD38 in the patients were 10,647 (CSF) and 28,078 (peritoneal fluid) for patient 1, and 33,265 (CSF) for patient 2.

Supplemental Tables

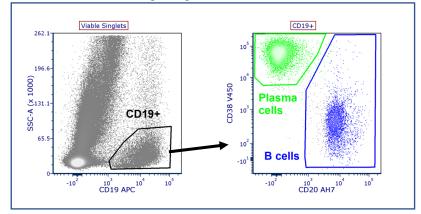
Supplemental Table 1. Characteristics and history of patient 1 prior to Dara treatment

Age at PEL diagnosis	34		
Sex/Gender	Male/Cis		
HIV status	+		
EBV status	+		
KSHV disease history			
Disease	MCD (2016)		
Treatment	Anthracycline-containing combination chemotherapy plus rituximab		
Outcome	Remission		
Initial PEL diagnosis (2017			
PEL location	Systemic PEL involving abdominal cavity		
Treatments	Treatment	Outcome	
	8 cycles of anthracycline-containing combination chemotherapy and splenectomy	Refractory PEL	
	One cycle of platinum-containing combination chemotherapy	Progressive PEL with ECOG status 4 and patient transferred to NCI	
PEL management at NCI (2019)			
PEL location	Extensive abdominal cavity PEL with leptomeningeal involvement		
Prior treatments	Treatment	Outcome	
	200 mg pembrolizumab every 3 weeks, 4 mg pomalidomide daily 21 out of 28 days, and intrathecal methotrexate	Partial response of systemic PEL but persistent leptomeningeal disease	

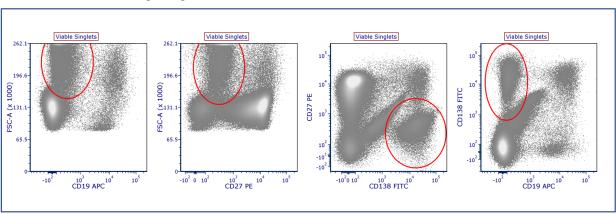
Supplemental Table 2. Characteristics and history of patient 2 prior to Dara treatment

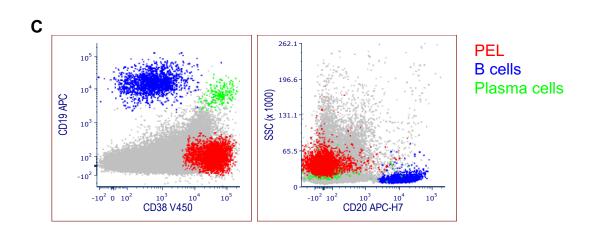
Age at PEL diagnosis	50	
Sex/Gender	Male/Cis	
HIV status	+	
EBV status	+	
KSHV disease history		
Disease	KS and MCD (2004)	
Treatment	Liposomal doxorubicin and rituximab	
Outcome	Complete remission	
PEL diagnosis (2017)	•	
PEL location	Extracavitary PEL with asymptomatic leptomeningeal involvement	
Prior treatments		
Prior treatments	Treatment	Outcome
Prior treatments	Treatment Combination anthracycline-containing chemotherapy and intrathecal methotrexate, cytarabine, and hydrocortisone	Outcome Remission of systemic PEL with refractory leptomeningeal PEL
Prior treatments	Combination anthracycline-containing chemotherapy and intrathecal methotrexate,	Remission of systemic PEL with refractory
Prior treatments	Combination anthracycline-containing chemotherapy and intrathecal methotrexate, cytarabine, and hydrocortisone Temozolomide, etoposide, liposomal	Remission of systemic PEL with refractory leptomeningeal PEL
Prior treatments	Combination anthracycline-containing chemotherapy and intrathecal methotrexate, cytarabine, and hydrocortisone Temozolomide, etoposide, liposomal doxorubicin, and dexamethasone	Remission of systemic PEL with refractory leptomeningeal PEL Refractory leptomeningeal PEL

A B-cell/Plasma-cell gating

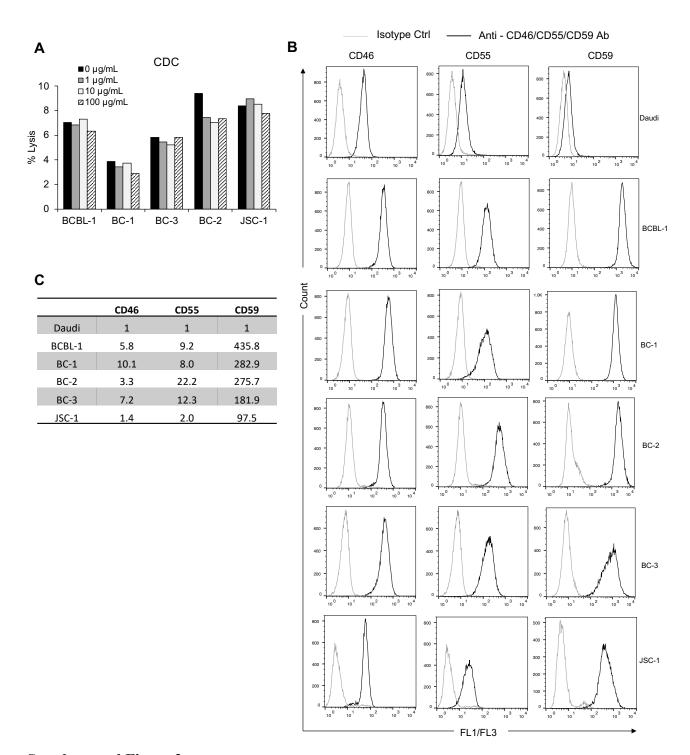


B Combinational PEL gating

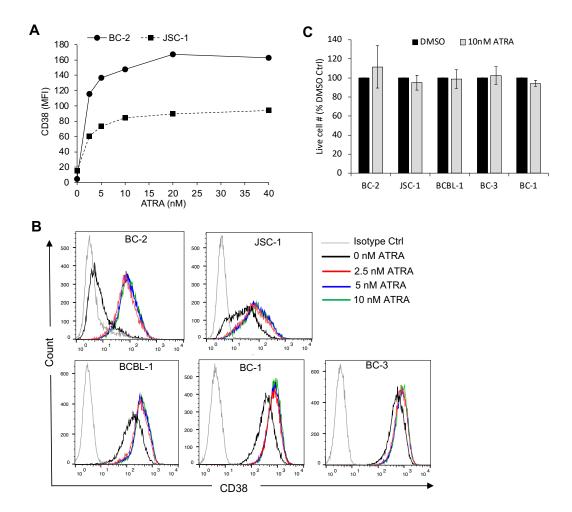




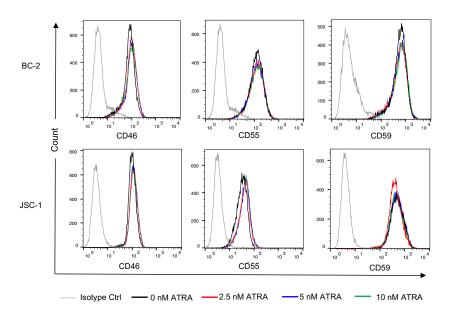
Supplemental Figure 1



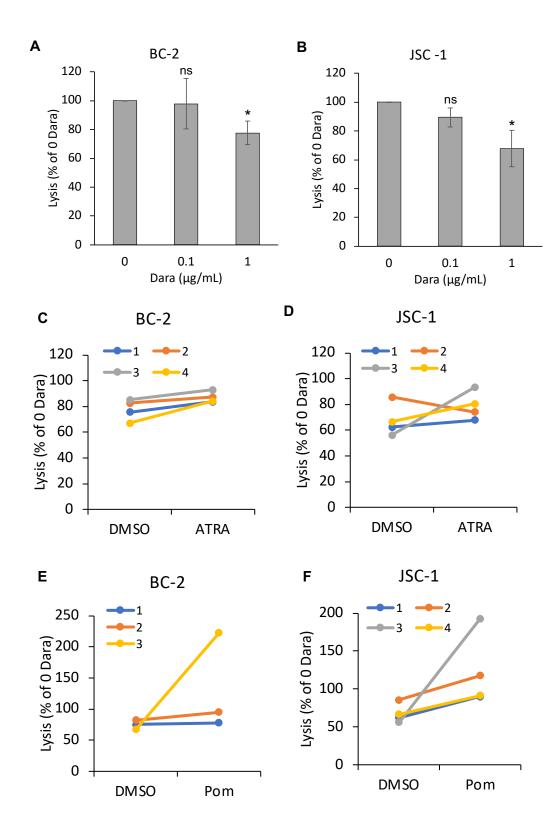
Supplemental Figure 2



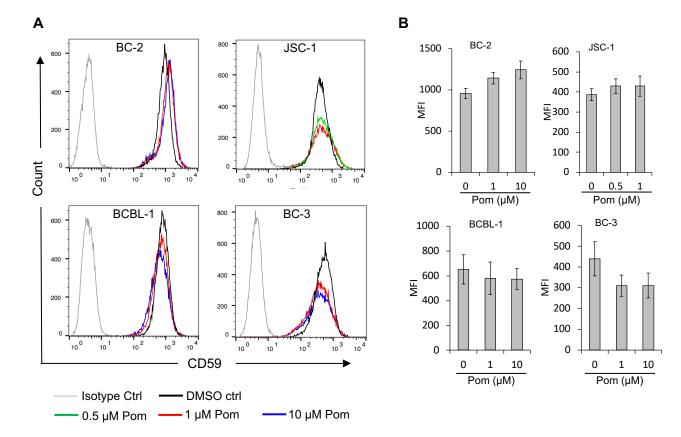
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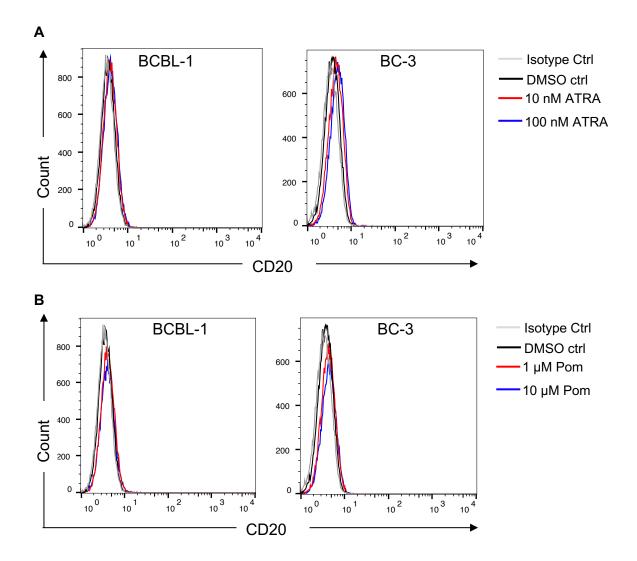
Supplemental Figure 4



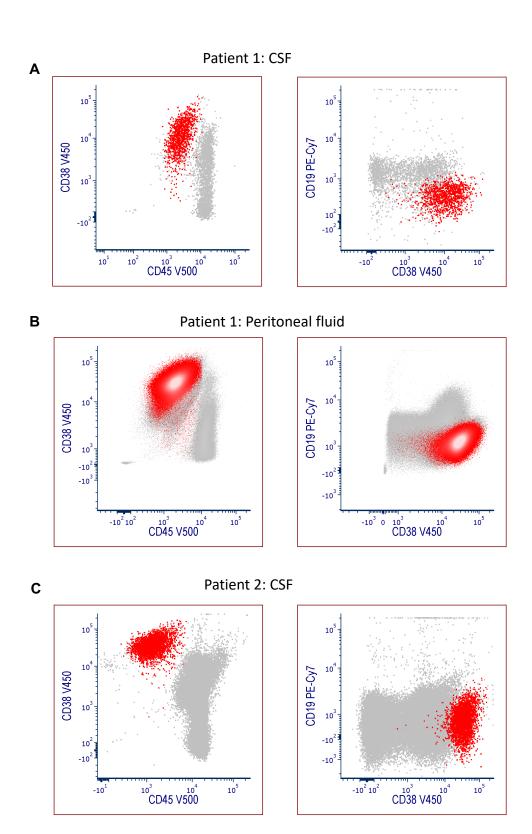
Supplemental Figure 5



Supplementary Figure 6



Supplemental Figure 7



Supplemental Figure 8