# Chemoselective Decarboxylative Protonation Enabled by Cooperative Earth-Abundant Element Catalysis

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#### **General Information**

All reagents were purchased from the commercially available sources and used without further purification. All reactions were carried out in a vial with magnetic stirring. All reactions were monitored by either <sup>1</sup>H NMR or thin layer chromatography (TLC) carried out on 0.25 mm precoated silia plates (F-254) purchased from Silicycle, Quebec, Canada, using shortwave UV light as visualizing agent and KMnO<sub>4</sub> or phosphomolybdic acid (PMA) as developing agents. Flash column chromatography was performed using SiliaFlash-P60 silica gel (40 – 63  $\mu$ m) purchased from Silicycle, Quebec, Canada. <sup>1</sup>H and <sup>13</sup>C spectra were recorded on a Bruker DRX-600 spectrometers operating at 600 MHz for proton nuclei and 151 MHz for carbon nuclei. Peaks were calibrated using residual undeuterated solvent as an internal reference (CDCl<sub>3</sub>: 7.26 ppm <sup>1</sup>H NMR and 77.20 ppm <sup>13</sup>C NMR). For reporting NMR peak multiplicities, the following abbreviations were used: s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, hept = heptet, m = multiplet. High-resolution mass spectra (HRMS) were recorded on an Agilent UHPLC TOF mass spectrometer using electrospray ionization time-of-flight (ESI-TOF) or chemical ionization time-of-flight (CI-TOF) reflectron experiments. Kessil lamp (390 nm and 427 nm) used in this work was purchased from the following website: <u>https://kessil.com/science/PR160L.php</u>

#### **Experimental Section**

#### General procedure for the iron-catalyzed decarboxylative protonation



4-Oxo-4-phenylbutanoic acid **1** (71.3 mg, 0.4 mmol, 1.0 equiv), Fe(NO<sub>3</sub>)<sub>3</sub>·9H<sub>2</sub>O (4.04 mg, 0.01 mmol, 2.5 mol%), di(2-picolyl)amine (1.99 mg, 0.01 mmol, 2.5 mol%), Na<sub>2</sub>CO<sub>3</sub> (4.24 mg, 0.04 mmol, 10 mol%) and TRIP disulfide (9.41 mg, 0.02 mmol, 5 mol%) were loaded to a vial; then evacuated and charged with N<sub>2</sub> twice. DCE (2 mL) and H<sub>2</sub>O (2 mL) were subsequently added to the mixture, sparged with nitrogen balloon for 5 min and sealed the vial with parafilm. The reaction was stirred vigorously under the irradiation of 390 nm LED light (5 to 10 cm to the reaction vial) for 20 h. Upon completion, the reaction was extracted with CH<sub>2</sub>Cl<sub>2</sub>, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo for column purification to afford decarboxylation product **2** (46.7 g, 0.348 mmol, 87%).

\* For volatile compounds **5**, **6**, **14**, **21**, **22** and readily decomposed compound **17**, product formations were characterized directly by crude NMR spectra without further purification (These compounds were reported previously).

# Procedure for the iron-catalyzed decarboxylative protonation on gram scale



Loxoprofen (1.5 g, 6.09 mmol, 1.0 equiv),  $Fe(NO_3)_3 \cdot 9H_2O$  (122.8 mg, 0.304 mmol, 5 mol%), di(2picolyl)amine (60.57 mg, 0.304 mmol, 5 mol%),  $Na_2CO_3$  (64.4 mg, 0.608 mmol, 10 mol%) and TRIP disulfide (143.1 mg, 0.304 mmol, 5 mol%) were loaded to a 200 mL round bottle flask; then evacuated and charged with  $N_2$  twice. DCE (30 mL) and  $H_2O$  (30 mL) were subsequently added to the mixture and sparged with nitrogen balloon for 20 min. The reaction was stirred vigorously under the irradiation of 390 nm LED light (5 cm to the reaction vial) for 20 h. Upon completion, the reaction was extracted with  $CH_2Cl_2$ , dried over  $Na_2SO_4$ , filtered and concentrated in vacuo for column purification to afford decarboxylation product **30** (1.01 g, 4.99 mmol, 82%).



Figure S1: Gram-scale reaction setup

# General procedure for the iron-catalyzed decarboxylative coupling with styrene



4-Oxo-4-phenylbutanoic acid **1** (71.3 mg, 0.4 mmol, 1.0 equiv), styrene (124.98 mg, 1.2 mmol, 3.0 equiv),  $Fe(NO_3)_3 \cdot 9H_2O$  (8.08 mg, 0.02 mmol, 5 mol%), di(2-picolyl)amine (3.98 mg, 0.02 mmol, 5 mol%),  $Na_2CO_3$  (4.24 mg, 0.04 mmol, 10 mol%) and TRIP disulfide (9.41 mg, 0.02 mmol, 5 mol%) were loaded to a vial; then evacuated and charged with  $N_2$  twice. DCE (2 mL) and  $H_2O$  (2 mL) were subsequently added to the mixture, sparged with nitrogen balloon for 5 min and sealed the vial with parafilm. The reaction was stirred vigorously under the irradiation of 390 nm LED light (5 to 10 cm to the reaction vial) for 20 h. Upon completion, the reaction was extracted with  $CH_2Cl_2$ , dried over  $Na_2SO_4$ , filtered and concentrated in vacuo for preparative TLC purification to afford decarboxylation coupling product **C1**. The reaction yield (27%) was determined by <sup>1</sup>H NMR spectroscopy using trimethoxybenzene as an internal standard.

\*C1 and decarboxylative protonation product of 1 possess similar polarity on TLC.

\*No product was observed in the absence of TRIP disulfide.

#### Synthesis of (E)-5-phenylpent-4-enoic acid substrates<sup>51</sup>



To a solution of (3-carboxypropyl)triphenylphosphonium bromide (8.81 mmol, 1.2 equiv) in THF (10 mL) was added LiHMDS (17.6 mmol, 2.4 equiv, 1M in THF) at 0 °C and stirred for 30 min while keeping the solution at 0 °C. Anisaldehyde solution (7.34 mmol, 1.0 equiv) in THF (3 mL) was then slowly added and allowed to react at RT for overnight. Until completion, the reaction mixture was quenched by NaOH solution then extracted with ethyl acetate. The separated aqueous layer was collected, acidified by HCl until the indication of acidic solution (or the appearance of precipitate) and extracted with ethyl acetate. The resulting organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo for column purification to afford the product.

#### Kinetic studies to determine the reaction rate law



4-Oxo-4-phenylbutanoic acid **1**, Fe(NO<sub>3</sub>)<sub>3</sub>·9H<sub>2</sub>O, di(2-picolyl)amine, Na<sub>2</sub>CO<sub>3</sub> and TRIP disulfide were loaded to a 20 mL vial according to Table S1; then evacuated and charged with N<sub>2</sub> twice. DCE (5 mL) and H<sub>2</sub>O (5 mL) were subsequently added to the mixture, sparged with nitrogen balloon for 5 min and sealed the vial with parafilm. The reaction was stirred vigorously for extra 5 min before shining the light to make sure all solid has dissolved. The sample was then irradiated with a 390 nm LED Kessil lamp (5 to 10 cm to the reaction vial). 0.05 mL aliquots (DCE phase) were transferred from the solution via syringe at specific time points directly to NMR tubes.



Figure S2: Reaction setup for kinetic studies (one lamp)



Figure S3: Reaction setup for kinetic studies (two lamps)

Entry	mmol acid	mmol iron/ligand	mmol disulfide	Initial rate (M/s)
1	1	0.025	0.05	4.33 x 10 <sup>-6</sup>
2	1	0.0125	0.05	4.05 x 10 <sup>-6</sup>
3	1	0.0375	0.05	4.19 x 10 <sup>-6</sup>
4	1	0.025	0.025	4.22 x 10 <sup>-6</sup>
5	1	0.025	0.075	4.33 x 10 <sup>-6</sup>
6	0.5	0.025	0.05	4.20 x 10 <sup>-6</sup>
7	1.5	0.025	0.05	4.34 x 10 <sup>-6</sup>
8 (2 lamps)	1	0.025	0.05	7.1 x 10 <sup>-6</sup>

#### **Table S1:** Initial rate data with different initial concentrations of acid and catalysts

(a)



(c)



(d)



(e)



#### Kinetic studies to determine the rate law of TRIPSH



\*The experimental protocol was identical to the kinetic studies using TRIP disulfide.



	5 mol% TRIPSH	10 mol% TRIPSH
initial rate(M/s)	2.5 x 10 <sup>-6</sup>	2.71 x 10 <sup>-6</sup>

#### **KIE studies**



1-(4-chlorophenyl)cyclobutane-1-carboxylic acid (210.66 mg, 1.0 mmol, 1 equiv),  $Fe(NO_3)_3 \cdot 9H_2O$ (20.2 mg, 0.05 mmol, 5 mol%), di(2-picolyl)amine (9.96 mg, 0.05 mmol, 5 mol%), TRIP disulfide (23.54 mg, 0.05 mmol, 5 mol%) and Na<sub>2</sub>CO<sub>3</sub> (10.59 mg, 0.1, 10 mol%) were loaded to a 20 mL vial; then evacuated and charged with N<sub>2</sub> twice. DCE (5 mL) and D<sub>2</sub>O (or H<sub>2</sub>O) (5 mL) were subsequently added to the mixture, sparged with nitrogen balloon for 5 min and sealed the vial with parafilm. The reaction was stirred vigorously for extra 5 min before shining the light to make sure all solid has dissolved. The sample was then irradiated with a 390 nm LED Kessil lamp (5 to 10 cm to the reaction vial). 0.05 mL aliquots (DCE phase) were transferred from the solution via syringe at specific time points directly to NMR tubes.

\*Note: Pure product was isolated using preparative TLC due to its similar polarity with TRIP disulfide/thiol.





time (s)

• H2O • D2O



Table S2: Scope exploration of ligand-free conditions

\* Same  $Fe(NO_3)_3 \cdot 9H_2O$  loadings were subjected in accordance with the general conditions shown in manuscript, Table 2.

### Characterization of decarboxylative protonation compounds

propiophenone (2)<sup>S2</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.96 – 7.91 (m, 2H), 7.55 – 7.49 (m, 1H), 7.45 – 7.39 (m, 2H), 2.97 (q, *J* = 7.26 Hz, 2H), 1.20 (t, *J* = 7.2 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 200.9, 137.0, 133.0, 128.6, 128.1, 31.9, 8.3. HRMS (APCl): calc'd for C<sub>9</sub>H<sub>11</sub>O [M+H]<sup>+</sup> 135.0804; Found 135.0806.

#### 1-ethyl-4-methoxybenzene (3)<sup>S3</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.15 (d, J = 8.5 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 3.81 (s, 3H), 2.62 (q, J = 7.6 Hz, 2H), 1.24 (t, J = 7.6 Hz, 3H).
 <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 157.8, 136.5, 128.8, 113.9, 55.4, 28.1, 16.0. HRMS (APCl): calc'd for C<sub>9</sub>H<sub>13</sub>O [M+H]<sup>+</sup> 137.0961; Found 137.0956.

#### 1-(benzyloxy)-4-ethylbenzene (4)<sup>S4</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.53 (d, J = 8.7 Hz, 2H), 7.47 (t, J = 7.7 Hz, 2H), 7.41 (t, J = 7.4 Hz, 1H), 7.21 (d, J = 8.3 Hz, 2H), 7.01 (dt, J = 8.5, 3.1 Hz, 2H), 5.12 (s, 2H), 2.69 (q, J = 7.6 Hz, 2H), 1.31 (t, J= 7.6 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 157.0, 137.4, 136.8, 128.9, 128.7, 128.0, 127.6, 114.8, 70.1, 28.1, 16.0. HRMS (APCl): calc'd for C<sub>15</sub>H<sub>15</sub>O [M-H]<sup>+</sup> 211.1117; Found 211.1114.

#### ethylbenzene (5)<sup>S3</sup>



<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>):** δ 7.27 (d, *J* = 15.6 Hz, 2H), 7.21 – 7.12 (m, 3H), 2.63 (q, *J* = 7.6 Hz, 2H), 1.22 (t, *J* = 7.6 Hz, 3H).

1-chloro-4-ethylbenzene (6)<sup>S5</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.22 (d, *J* = 6.8 Hz, 2H), 7.11 (d, *J* = 7.9 Hz, 2H), 2.60 (q, *J* = 7.3 Hz, 2H), 1.22 – 1.17 (m, 3H).

#### 2-ethyl-1,4-dimethoxybenzene (7)<sup>56</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 6.81 – 6.77 (m, 2H), 6.74 – 6.69 (m, 1H), 3.81 (s, 3H), 3.80 (s, 3H), 2.66 (q, *J* = 7.5 Hz, 2H), 1.26 – 1.19 (m, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 153.6, 151.7, 134.0, 115.6, 111.1, 110.5, 56.0, 55.7, 23.5, 14.3. HRMS (APCl): calc'd for C<sub>10</sub>H<sub>15</sub>O<sub>2</sub> [M+H]<sup>+</sup> 167.1067; Found 167.1062.

#### 5-ethyl-1,2,3-trimethoxybenzene (8)<sup>57</sup>

MeO. MeO OMe

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 6.40 (s, 2H), 3.84 (s, 6H), 3.81 (s, 3H), 2.58 (q, J = 7.6 Hz, 2H), 1.22 (t, J = 7.6 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 153.1, 140.1, 136.0, 104.7, 60.9, 56.0, 29.3, 15.7. HRMS (APCl): calc'd for C<sub>11</sub>H<sub>17</sub>O<sub>3</sub> [M+H]<sup>+</sup> 197.1172; Found 197.1167.

(E)-but-1-en-1-ylbenzene (9a)<sup>S8</sup>

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.36 (d, J = 7.4 Hz, 2H), 7.31 (t, J = 7.5 Hz, 2H), 7.21 (t, J = 7.3 Hz, 1H), 6.39 (d, J = 15.8 Hz, 1H), 6.33 – 6.26 (m, 1H), 2.25 (quint, J = 7.2 Hz, 2H), 1.11 (t, J = 7.5 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 138.1, 132.8, 128.9, 128.6, 126.9, 126.0, 26.2, 13.8. HRMS (APCl): calc'd for C<sub>10</sub>H<sub>13</sub> [M+H]<sup>+</sup> 133.1012; Found 133.1009.

(E)-1-(but-1-en-1-yl)-4-methoxybenzene (10a)<sup>S8</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.30 (d, J = 8.3 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 6.35 (d, J = 15.2 Hz, 1H), 6.19 – 6.12 (m, 1H), 3.81 (s, 3H), 2.23 (quint, J = 7.2 Hz, 2H), 1.11 (t, J = 7.8 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 158.7, 130.9, 130.7, 128.3, 127.1, 114.1, 55.4, 26.2, 14.0. HRMS (APCl): calc'd for C<sub>11</sub>H<sub>15</sub>O [M+H]<sup>+</sup> 163.1117; Found 163.1114.

#### 5-(4-methoxybenzyl)dihydrofuran-2(3H)-one (10b)<sup>59</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.14 (d, J = 8.5 Hz, 2H), 6.85 (d, J = 8.6 Hz, 2H), 4.69 (quint, J = 6.8 Hz, 1H), 3.79 (s, 3H), 3.00 (dd, J = 5.9, 14.1 Hz, 1H), 2.88 (dd, J = 6.2, 14.1 Hz, 1H), 2.50 – 2.40 (m, 1H), 2.39 – 2.30 (m, 1H), 2.28 – 2.19 (m, 1H), 1.99 – 1.89 (m, 1H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ

177.3, 158.8, 130.7, 128.0, 114.2, 81.1, 55.4, 40.6, 28.8, 27.1. **HRMS (APCI):** calc'd for C<sub>12</sub>H<sub>15</sub>O<sub>3</sub> [M+H]<sup>+</sup> 207.1016; Found 207.1010.

#### (E)-1-(but-1-en-1-yl)-4-fluorobenzene (11a)<sup>S10</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.35 – 7.28 (m, 2H), 7.02 – 6.97 (m, 2H), 6.36 (d, J = 15.8 Hz, 1H), 6.23 – 6.16 (m, 1H), 2.24 (quint, J = 10.8 Hz, 2H), 1.11 (t, J = 7.5 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 162.1 (d,  $J_{C-F} = 243.9$  Hz), 134.2 (d,  $J_{C-F} = 3.2$  Hz), 132.5 (d,  $J_{C-F} = 2.0$  Hz), 127.8, 127.4 (d,  $J_{C-F} = 7.7$ Hz), 115.4 (d,  $J_{C-F} = 20.9$  Hz), 26.2, 13.8. <sup>19</sup>F NMR (565 Hz, CDCl<sub>3</sub>): δ -115.87 – -116.04 (m, 1F). HRMS (APCl): calc'd for C<sub>10</sub>H<sub>12</sub>F [M+H]<sup>+</sup> 151.0918; Found 151.0914.

#### (ethoxymethyl)benzene (12)<sup>S11</sup>

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<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.38 – 7.24 (m, 5H), 4.50 (s, 2H), 3.54 (q, J = 7.0 Hz, 2H), 1.25 (t, J = 7.0 Hz, 3H).
 <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 138.7, 128.5, 127.8, 127.6, 72.9, 65.9.
 15.4. HRMS (APCl): calc'd for C<sub>9</sub>H<sub>11</sub>O [M-H]<sup>+</sup> 135.0804; Found 135.0805.

dodecane (13)<sup>S12</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 1.34 − 1.22 (m, 20H), 0.89 (t, *J* = 6.9 Hz, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 32.2, 30.0, 29.9, 29.6, 22.9, 14.3.

*tert*-butylcyclohexane (14)<sup>S13</sup>

tBu

<sup>1</sup>H NMR (Crude spectrum, 600 MHz, CDCl<sub>3</sub>): δ 1.79 – 1.71 (m, 4H), 1.66 – 1.61 (m, 1H), 1.23 – 1.04 (m, 3H), 1.00 – 0.87 (m, 3H), 0.83 (s, 9H).

#### adamantane (15)<sup>S14</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 1.87 (s, 4H), 1.77 − 1.72 (m, 12H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 37.9, 28.5. HRMS (APCl): calc'd for C<sub>10</sub>H<sub>7</sub> [M+H]<sup>+</sup> 137.1325; Found 137.1327.

#### 1-bromoheptane (16)<sup>S15</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 3.40 (t, *J* = 6.9 Hz, 2H), 1.85 (quint, *J* = 6.9 Hz, 2H), 1.45 – 1.37 (m, 2H), 1.34 – 1.23 (m, 6H), 0.87 (t, *J* = 7.0 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 34.2, 33.0, 31.8, 28.6, 28.3, 22.7, 14.2. HRMS (APCl): calc'd for C<sub>7</sub>H<sub>16</sub>Br [M+H]<sup>+</sup> 179.0430; Found 179.0283.



<sup>1</sup>H NMR (Crude spectrum, 600 MHz, CDCl<sub>3</sub>): δ 2.18 (td, *J* = 2.6, 7.1 Hz, 2H), 1.94 (t, *J* = 2.64 Hz, 1H), 1.54 − 1.49 (m, 2H), 1.43 − 1.35 (m, 2H), 1.33 − 1.25 (m, 8H), 0.88 (t, *J* = 7.0 Hz, 3H).

phenyl(piperidin-1-yl)methanone (18)<sup>S17</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.38 (s, 5H), 3.71 (br, 2H), 3.33 (br, 2H), 1.67 (br, 4H), 1.50 (br, 2H).
 <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 170.3, 136.5, 129.3, 128.4, 126.8, 48.7, 43.1, 26.5, 25.6, 24.6. HRMS (APCl): calc'd for C<sub>12</sub>H<sub>16</sub>NO [M+H]<sup>+</sup> 190.1226; Found 190.1220.

#### methyl bicyclo[2.2.2]octane-1-carboxylate (19)<sup>S18</sup>

MeOOC

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  3.60 (s, 3H), 1.73 – 1.66 (m, 6H), 1.61 – 1.51 (m, 7H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  179.0, 51.6, 38.3, 28.1, 25.4, 23.8. HRMS (APCI): calc'd for C<sub>10</sub>H<sub>17</sub>O<sub>2</sub> [M+H]<sup>+</sup> 169.1223; Found 169.1223.

#### benzyl 4-methyl-5-oxooxazolidine-3-carboxylate (20)<sup>S19</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.39 – 7.32 (m, 5H), 5.40 (br, 1H), 5.32 – 5.12 (m, 3H), 4.28 (br, 1H), 1.54 (br, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 172.9, 152.8, 135.5, 128.8, 128.6, 128.3, 77.4, 67.8, 50.7, 16.7. HRMS (APCl): calc'd for C<sub>12</sub>H<sub>14</sub>NO<sub>4</sub> [M+H]<sup>+</sup> 236.0917; Found 236.0914.

#### propylbenzene (21)<sup>S20</sup>

<sup>1</sup>H NMR (Crude spectrum, 600 MHz, CDCl<sub>3</sub>): δ 7.30 – 7.24 (m, 2H), 7.19 – 7.15 (m, 3H), 2.59 (t, J = 7.6 Hz, 2H), 1.64 (sext, J = 7.4 Hz, 2H), 0.94 (t, J = 7.4 Hz, 3H).

sec-butylbenzene (22)<sup>S21</sup>

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.29 (t, *J* = 5.8 Hz, 2H), 7.18 (d, *J* = 6.9 Hz, 3H), 2.63 – 2.54 (m, 1H), 1.63 – 1.54 (m, 2H), 1.29 – 1.21 (m, 3H), 0.82 (t, *J* = 7.8 Hz, 3H).

#### 1-chloro-4-cyclopropylbenzene (23)<sup>S22</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.25 – 7.19 (m, 2H), 7.03 – 6.97 (m, 2H), 1.90 – 1.84 (m, 1H), 1.00 – 0.94 (m, 2H), 0.70 – 0.63 (m, 2H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 142.6, 131.0, 128.4, 127.1, 15.0, 9.4. HRMS (APCl): calc'd for C<sub>9</sub>H<sub>10</sub>Cl [M+H]<sup>+</sup> 153.0466; Found 153.0461.

#### 1-chloro-4-cyclobutylbenzene (24)<sup>S23</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.25 (d, J = 8.5 Hz, 2H), 7.14 (d, J = 8.3 Hz, 2H), 3.51 (quint, J = 8.5 Hz, 1H), 2.40 – 2.29 (m, 2H), 2.17 – 2.06 (m, 2H), 2.06 – 1.96 (m, 1H), 1.90 – 1.80 (m, 1H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  144.8, 131.4, 128.4, 127.8, 39.9, 29.9, 18.3. HRMS (APCl): calc'd for C<sub>10</sub>H<sub>12</sub>Cl [M+H]<sup>+</sup> 167.0622; Found 167.0619.

#### tert-butyl pyrrolidine-1-carboxylate (25)<sup>S24</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 3.28 (s, 2H), 3.22 (s, 2H), 1.78 (s, 4H), 1.41 (s, 9H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 154.8, 78.9, 46.0, 45.7, 28.6, 25.8, 25.1. HRMS (APCI): calc'd for C<sub>9</sub>H<sub>18</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 172.1332; Found 172.1332.

(E)-7-hydroxy-5-methoxy-4-methyl-6-(3-methylpent-2-en-1-yl)isobenzofuran-

1(3H)-one (26)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.61 (br, 1H), 5.15 (s, 3H), 3.74 (s, 3H), 3.35 (d, J = 6.8 Hz, 2H), 2.12 (s, 3H), 1.95 (q, J = 7.3 Hz, 2H), 1.75 (s, 3H), 0.93 (t, J = 7.4 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 173.0, 163.8, 153.7, 144.0, 137.8, 122.7, 120.6, 116.8, 106.4, 70.1, 61.1, 32.4, 22.7, 16.2, 12.7, 11.6. HRMS (APCl): calc'd for C<sub>16</sub>H<sub>21</sub>O<sub>4</sub> [M+H]<sup>+</sup> 277.1434; Found 277.1433.

#### 1-ethyl-4-isobutylbenzene (27)<sup>S25</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.13 (d, J = 7.6 Hz, 2H), 7.09 (d, J = 7.4 Hz, 2H), 2.65 (q, J = 7.6 Hz, 2H), 2.46 (d, J = 7.1 Hz, 2H), 1.87 (sept, J = 6.7 Hz, 1H), 1.25 (t, J = 7.6 Hz, 3H), 0.92 (d, J = 6.5 Hz, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  141.6, 139.0, 129.2, 127.7, 45.2, 30.4, 28.6, 22.6, 15.8. HRMS (APCl): calc'd for C<sub>12</sub>H<sub>19</sub> [M+H]<sup>+</sup> 163.1481; Found 163.1482.

1,4-dimethyl-2-((4-methylpentyl)oxy)benzene (28)<sup>526</sup>



<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>):** δ 7.07 (d, *J* = 7.5, 1H), 6.72 (d, *J* = 7.5, 1H), 6.70 (s, 1H), 3.99 (t, *J* = 6.5 Hz, 2H), 2.38 (s, 3H), 2.26 (s, 3H), 1.90 – 1.83 (m, 2H), 1.70 (hept, *J* = 6.7 Hz, 1H), 1.47 – 1.40 (m, 2H), 1.01 (s, 3H), 1.00 (s, 3H). <sup>13</sup>**C NMR (150 MHz, CDCl<sub>3</sub>):** δ 157.3, 136.6, 130.4, 123.8, 120.7, 112.1, 68.3, 35.5, 28.0, 27.5, 22.8, 21.6, 16.0. **HRMS (APCl):** calc'd for C<sub>14</sub>H<sub>23</sub>O [M+H]<sup>+</sup> 207.1743; Found 207.1743.

heptadec-8-ene (29)<sup>S27</sup>



<sup>1</sup>H NMR (major isomer, 600 MHz, CDCl<sub>3</sub>): δ 5.37 – 5.32 (m, 2H), 2.07 – 1.98 (m, 4H), 1.38 – 1.21 (m, 22H), 0.88 (t, *J* = 6.8 Hz, 6H). <sup>13</sup>C NMR (major isomer, 150 MHz, CDCl<sub>3</sub>): δ 130.1, 32.1, 32.1, 30.0, 29.9, 29.8, 29.6, 29.5, 29.46, 29.44, 27.4, 22.9, 14.3. HRMS (APCl): calc'd for C<sub>17</sub>H<sub>35</sub> [M+H]<sup>+</sup> 239.2733; Found 239.2734.

#### 2-(4-ethylbenzyl)cyclopentan-1-one (30)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.12 (d, *J* = 8.0 Hz, 2H), 7.09 (d, *J* = 8.1 Hz, 2H), 3.12 (dd, *J* = 4.1, 13.9 Hz, 1H), 2.62 (q, *J* = 7.6 Hz, 2H), 2.52 (dd, *J* = 9.5, 13.9 Hz, 1H), 2.38 – 2.29 (m, 2H), 2.15 – 2.05 (m, 2H), 2.00 – 1.91 (m, 1H), 1.78 – 1.68 (m, 1H), 1.61 – 1.52 (m, 1H), 1.24 (t, *J* = 7.6 Hz, 3H). <sup>13</sup>C NMR

(**150** MHz, CDCl<sub>3</sub>): δ 142.0, 137.2, 128.9, 127.9, 51.1, 38.3, 35.2, 29.2, 28.5, 20.6, 15.6. HRMS (APCl): calc'd for C<sub>14</sub>H<sub>19</sub>O [M+H]<sup>+</sup> 203.1430; Found 203.1430.

(1*R*,2*S*,4b*R*,7*S*,9a*R*)-1-methyl-8-methylene-13-oxo-1,2,5,6,8,9,10,10a-octahydro-4a,1-(epoxymethano)-7,9a-methanobenzo[*a*]azulene-2,7(4b*H*)-diyl diacetate (31)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 6.34 (d, J = 9.4 Hz, 1H), 5.78 (dd, J = 3.8, 9.3 Hz, 1H), 5.27 (d, J = 3.8 Hz, 1H), 5.07 (s, 1H), 4.90 (s, 1H), 2.79 (dd, J = 8.0, 11.2 Hz, 1H), 2.50 – 2.44 (m, 1H), 2.32 (dd, J = 8.7, 12.6 Hz, 1H), 2.25 (d, J = 15.5 Hz, 1H), 2.09 – 2.02 (m, 4H), 1.97 (s, 3H), 1.94 – 1.87 (m, 2H), 1.79 – 1.70 (m, 2H), 1.67 – 1.59 (m, 2H), 1.15 (s, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 177.6, 169.93, 169.9, 154.9, 134.8, 128.9, 107.0, 91.7, 84.9, 70.4, 52.3, 51.1, 50.4, 48.2, 46.7, 39.9, 36.7, 35.0, 22.2, 20.9, 16.6, 14.6. HRMS (APCl): calc'd for C<sub>22</sub>H<sub>27</sub>O<sub>6</sub> [M+H]<sup>+</sup> 387.1802; Found 387.1802.

(4aR,6aS,6bR,10S,12aS)-10-hydroxy-2,4a,6a,6b,9,9,12a-heptamethyl-

1,3,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,12b,14b-octadecahydropicen-13(2*H*)one (32)



<sup>1</sup>H NMR (major isomer (1.8:1 dr), 600 MHz, CDCl<sub>3</sub>): δ 5.54 (s, 1H), 4.05 (q, *J* = 7.1 Hz, 1H), 3.18 – 3.12 (m, 1H), 2.76 – 2.68 (m, 1H), 2.28 (s, 1H), 2.13 – 0.98 (m, 27H), 0.98 – 0.71 (m, 13H), 0.64 (d,

*J* = 11.7 Hz, 1H). <sup>13</sup>C NMR (isomeric mixture (1.8:1 dr)150 MHz, CDCl<sub>3</sub>): δ 200.4, 200.3, 171.1, 170.9, 170.4, 127.95, 127.93, 78.6, 61.8, 60.4, 54.9, 51.7, 45.4, 45.39, 45.36, 43.4, 43.3, 41.4, 40.8, 39.16, 39.1, 37.7, 37.1, 34.3, 33.3, 32.79, 32.77, 32.38, 30.6, 28.9, 28.7, 28.1, 27.6, 27.3, 26.8, 26.63, 26.6, 26.5, 26.4, 23.34, 23.3, 22.4, 21.0, 18.7, 18.67, 17.5, 16.9, 16.3, 15.6, 14.2. HRMS (APCl): calc'd for C<sub>29</sub>H<sub>47</sub>O<sub>2</sub> [M+H]<sup>+</sup> 427.3571; Found 427.3570.

(10S,13R)-17-((R)-sec-butyl)-10,13-dimethyldodecahydro-3H-

cyclopenta[a]phenanthrene-3,7,12(2H,4H)-trione (33)<sup>S18</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 2.93 – 2.79 (m, 3H), 2.37 – 2.15 (m, 6H), 2.14 – 1.90 (m, 6H), 1.86 – 1.78 (m, 1H), 1.59 (td, *J* = 4.4, 14.5 Hz, 1H), 1.51 – 1.42 (m, 1H), 1.38 (s, 3H), 1.32 – 1.06 (m, 5H), 1.05 (s, 3H), 0.85 (t, *J* = 7.4 Hz, 3H), 0.81 (d, *J* = 6.5 Hz, 2H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 212.2, 209.2, 208.9, 57.0, 51.9, 49.1, 46.9, 45.6, 45.5, 45.1, 42.9, 38.7, 37.5, 36.6, 36.1, 35.3, 27.9, 27.8, 25.3, 22.0, 18.5, 11.9, 10.9. HRMS (APCl): calc'd for C<sub>23</sub>H<sub>35</sub>O<sub>3</sub> [M+H]<sup>+</sup> 359.2581; Found 359.2578.

(2R,3S)-3-methyl-2-((Z)-pent-2-en-1-yl)cyclopentan-1-one (34)



<sup>1</sup>H NMR (major isomer (10:1), 600 MHz, CDCl<sub>3</sub>): δ 5.44 – 5.37 (m, 1H), 5.28 – 5.22 (m, 1H), 2.39
– 2.24 (m, 3H), 2.11 – 2.00 (m, 4H), 1.93 – 1.85 (m, 1H), 1.72 – 1.65 (m, 1H), 1.41 – 1.31 (m, 1H),
1.13 (d, *J* = 6.4 Hz, 3H), 0.94 (t, *J* = 7.5 Hz, 3H). <sup>13</sup>C NMR (major isomer(10:1), 150 MHz, CDCl<sub>3</sub>): δ

220.9, 133.6, 125.7, 56.7, 38.4, 36.5, 29.7, 25.2, 20.7, 19.8, 14.3. **HRMS (APCI):** calc'd for C<sub>11</sub>H<sub>19</sub>O [M+H]<sup>+</sup> 167.1430; Found 167.1429.

(4-chlorophenyl)(5-methoxy-2,3-dimethyl-1H-indol-1-yl)methanone (35)<sup>S18</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.67 – 7.61 (m, 2H), 7.48 – 7.42 (m, 2H), 6.92 (d, J = 8.9 Hz, 1H), 6.90 (d, J = 2.5 Hz, 1H), 6.67 (dd, J = 2.6, 8.9 Hz, 1H), 3.85 (s, 3H), 2.31 (s, 3H), 2.19 (s, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 168.3, 156.0, 138.9, 134.5, 133.8, 132.0, 131.1, 130.9, 129.1, 115.5, 115.0, 111.2, 101.3, 55.8, 13.4, 8.86. HRMS (APCl): calc'd for C<sub>18</sub>H<sub>17</sub>ClNO<sub>2</sub> [M+H]<sup>+</sup> 314.0942; Found 314.0942.

#### 4-chloro-N-(4-isopropoxyphenethyl)benzamide (36)<sup>528</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.62 (d, *J* = 8.5 Hz, 2H), 7.35 (d, *J* = 6.7 Hz, 2H), 7.11 (d, *J* = 7.4, 2H), 6.84 (d, *J* = 8.2 Hz, 2H), 6.23 (br, 1H), 4.52 (hept, *J* = 6.1 Hz, 1H), 3.66 (d, *J* = 5.8 Hz, 2H), 2.85 (t, *J* = 6.8 Hz, 2H), 1.32 (d, *J* = 6.1 Hz, 6H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  166.6, 166.5, 156.8, 137.7, 133.2, 130.7, 129.9, 128.9, 128.4, 116.3, 70.1, 41.5, 34.8, 22.2. HRMS (APCI): calc'd for C<sub>18</sub>H<sub>21</sub>ClNO<sub>2</sub> [M+H]<sup>+</sup> 318.1255; Found 318.1255.

#### 2-ethyldibenzo[b,f]thiepin-10(11H)-one (37)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 8.19 (d, *J* = 6.7 Hz, 1H), 7.58 (d, *J* = 7.9 Hz, 1H), 7.53 (d, *J* = 7.9 Hz), 7.43 – 7.36 (m, 1H), 7.32 – 7.26 (m, 2H), 7.01 (d, *J* = 7.9 Hz, 1H), 4.33 (s, 2H), 2.63 (q, *J* = 7.6 Hz, 2H), 1.24 – 1.18 (m, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 191.8, 146.8, 140.8, 137.7, 136.3, 132.5, 131.6, 131.5, 131.3, 130.9, 129.1, 126.9, 126.8, 51.2, 28.7, 15.5. HRMS (APCl): calc'd for C<sub>16</sub>H<sub>15</sub>OS [M+H]<sup>+</sup> 255.0838; Found 255.0829.

#### N,N-bis(2-chloroethyl)-4-propylaniline (38)<sup>526</sup>



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.11 (d, J = 8.6 Hz, 2H), 6.66 (d, J = 8.7 Hz, 2H), 3.72 (t, J = 7.3 Hz, 4H), 3.65 (t, J = 7.3 Hz, 4H), 2.53 (t, J = 7.5 Hz, 2H), 1.63 (sex, J = 7.4 Hz, 2H), 0.97 (t, J = 7.3 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  144.2, 132.1, 129.7, 112.2, 53.8, 40.7, 37.0, 24.9, 14.0. HRMS (APCl): calc'd for C<sub>13</sub>H<sub>20</sub>Cl<sub>2</sub>N [M+H]<sup>+</sup> 260.0960; Found 260.0967.

#### 2,6-dichloro-N-(o-tolyl)aniline (39)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.38 (d, J = 8.1 Hz, 2H), 7.20 (d, J = 7.4 Hz, 1H), 7.10 – 7.01 (m, 2H), 6.90 (t, J = 7.3 Hz, 1H), 6.45 (d, J = 8.0 Hz, 1H), 5.54 (br, 1H), 3.12 (s, 3H). <sup>13</sup>C NMR (150 MHz, **CDCl<sub>3</sub>):** δ 141.7, 137.5, 130.7, 130.6, 129.0, 126.5, 126.1, 124.9, 121.3, 115.6, 18.0. **HRMS (APCl):** calc'd for C<sub>13</sub>H<sub>12</sub>Cl<sub>2</sub>N [M+H]<sup>+</sup> 252.0341; Found 252.0334.

#### tert-butyl but-1-en-1-ylcarbamate (40)<sup>S29</sup>



*E* isomer: <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 6.40 (t, *J* = 12.2 Hz, 1H), 6.18 (br, 1H), 5.01 – 4.92 (m, 1H), 2.01 – 1.91 (m, 2H), 1.43 (s, 9H), 0.95 (t, *J* = 7.4 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 153.1, 123.0, 111.7, 80.2, 28.4, 23.0, 14.5. HRMS (APCl): calc'd for C<sub>9</sub>H<sub>18</sub>NO<sub>2</sub> [M+H]<sup>+</sup> 172.1332; Found 172.1332. *Z* isomer: <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 6.35 (t, *J* = 9.6 Hz, 1H), 6.21 (br, 1H), 4.54 (q, *J* = 7.6 Hz, 1H), 2.01 – 1.91 (m, 2H), 1.45 (s, 9H), 0.98 (t, *J* = 7.5 Hz, 3H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>): δ 153.0, 121.6, 110.1, 80.4, 28.4, 18.9, 14.2.

1,5-diphenylpentan-1-one (C1)



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.97 – 7.92 (m, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.45 (t, *J* = 7.9 Hz, 2H), 7.30 – 7.25 (m, 2H), 7.21 – 7.15 (m, 3H), 2.99 (t, *J* = 7.1 Hz, 2H), 2.67 (t, *J* = 7.7 Hz, 2H), 1.83 – 1.68 (m, 4H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  200.4, 142.4, 137.2, 133.1, 128.7, 128.5, 128.4, 128.2, 125.9, 38.5, 35.9, 31.2, 24.1. HRMS (APCl): calc'd for C<sub>17</sub>H<sub>19</sub>O [M+H]<sup>+</sup> 239.1436; Found 239.1437.

#### References

- (S1) Y. Yamamoto, Y. Nakanishi, K. -i. Yamada, K. Tomioka, Aminolithiation–arylation consecutive cyclization of N-(2-fluorophenyl)methylaminoalkylstyryls giving arylsubstituted pyrido[1,2-b]isoquinolines. *Tetrahedron* 2018, 74, 5309 – 5318.
- (S2) Wu, J. -C.; Gong, L. B.; Xia, Y.; Song, R.- J.; Xie, Y. -X.; and Li, J. -H. Nickel-Catalyzed Kumada Reaction of Tosylalkanes with Grignard Reagents to Produce Alkenes and Modified Arylketones. *Angew. Chem. Int. Ed.* **2012**, *51*, 9909 – 9913.
- (S3) Kang, Q. -K.; Li, Y.; Chen, K.; Zhu, H.; Wu, W. -Q.; Lin, Y.; Shi, H. Rhodium-Catalyzed Stereoselective Deuteration of Benzylic C–H Bonds via Reversible η6-Coordination. *Angew. Chem. Int. Ed.* **2022**, *61*, e202117381.
- (S4) Epifanov, M.; Mo, J. Y.; Dubois, R.; Yu, H.; Sammis, G. M. One-Pot Deoxygenation and Substitution of Alcohols Mediated by Sulfuryl Fluoride. *J. Org. Chem.* 2021, *86*, 3768 – 3777.
- (S5) Gao, J.; Ma, R.; Feng, L.; Liu, Y.; Jackstell, R.; Jagadeesh, R. V.; Beller, M. Ambient
   Hydrogenation and Deuteration of Alkenes Using a Nanostructured Ni-Core–Shell
   Catalyst. Angew. Chem. Int. Ed. 2021, 60, 18591 18598.
- (S6) Roberts, W. P.; Ebner, C. L. Synthesis of monoalkyl derivatives of 7,7,8,8-tetracyano-pquinodimethane from 2,5-dimethoxybenzoic acid. *J. Org. Chem.* **1987**, *52*, 2297 – 2299.
- (S7) Pincock, J. A.; Wedge, P. J. The Photochemistry of Methoxy-Substituted Benzyl Acetates and Benzyl Pivalates: Homolytic vs Heterolytic Cleavage. J. Org. Chem. 1994, 59, 5587 – 5595.
- (S8) Yu, X.; Zhao, H.; Li, P.; Koh, M. J. Iron-Catalyzed Tunable and Site-Selective Olefin Transposition. J. Am. Chem. Soc. 2020, 142, 18223 – 18230.
- (S9) Ariyarathna, J. P.; Wu, F.; Colombo, S. K.; Hillary, C. M.; Li, W. Aerobic Catalytic Features in Photoredox- and Copper-Catalyzed Iodolactonization Reactions. *Org. Lett.* **2018**, *20*, 6462–6466.
- (S10) Jia, Z. -J.; Gao, S.; Arnold, F. H. Enzymatic Primary Amination of Benzylic and Allylic
   C(sp<sup>3</sup>)–H Bonds. J. Am. Chem. Soc. 2020, 142, 10279 10283.

- (S11) Bakos, M.; Gyömöre, Á.; Domján, A.; Soós, T. Auto-Tandem Catalysis with Frustrated Lewis Pairs for Reductive Etherification of Aldehydes and Ketones. *Angew. Chem. Int. Ed.* 2017, 56, 5217 – 5221.
- (S<u>12</u>) Huang, J. -L.; Dai, X. -J.; Li, C. -J. Iridium-Catalyzed Direct Dehydroxylation of Alcohols. *Eur. J. Org. Chem.* **2013**, 6496 – 6500.
- (S13) Miyamura, H.; Suzuki, A.; Yasukawa, T.; Kobayashi, S. Polysilane-Immobilized Rh–Pt
   Bimetallic Nanoparticles as Powerful Arene Hydrogenation Catalysts: Synthesis,
   Reactions under Batch and Flow Conditions and Reaction Mechanism. J. Am. Chem. Soc.
   2018, 140, 11325 11334.
- (S14) Fukuyama, T.; Fujita, Y.; Miyoshi, H.; Ryu, I.; Kao, S. -C.; Wu, Y. -K. Electron transferinduced reduction of organic halides with amines. *Chem. Commun.* 2018, *54*, 5582 – 5585.
- (S15) Galli, M.; Fletcher, C. J.; Pozo, M. d.; Goldup, S. M. Scalable anti-Markovnikov
   hydrobromination of aliphatic and aromatic olefins. *Org. Biomol. Chem.* 2016, 14, 5622 –
   5626.
- (S16) Miao, C.; Zhi, J.; Sun, S.; Yang, X.; Hu, A. Formation of conjugated polynaphthalene via Bergman cyclization. J. Polym. Sci. A Polym. Chem. 2010, 48, 2187 – 2193.
- (S17) Zhao, Q.; Li, H.; Wang, L. The direct amidation of α-diketones with aminesvia TBHPpromoted oxidative cleavage of C(sp<sup>2</sup>)–C(sp<sup>2</sup>) bonds. *Org. Biomol. Chem.* **2013**, *11*, 6772 – 6779.
- Qin, T.; Malins, L. R.; Edwards, J. T.; Merchant, R. R.; Novak, A. J. E.; Zhong, J. Z.; Mills, R. B.; Yan, M.; Yuan, C.; Eastgate, M. D.; Baran, P. S. Nickel-Catalyzed Barton
  Decarboxylationand Giese Reactions: A Practical Take on Classic Transforms. *Angew. Chem. Int. Ed.* 2017, *56*, 260 265.
- (S19) Walter, M. W.; Adlington, R. M.; Baldwin, J. E.; Schofield, C. J. Reaction of (Trifluoromethyl)trimethylsilane with Oxazolidin-5-ones: Synthesis of Peptidic and Nonpeptidic Trifluoromethyl Ketones. J. Org. Chem. 1998, 63, 5179 – 5192.
- (S20) Carter, T. S.; Guiet, L.; Frank, D. J.; West, J.; Thomas, S. P. Iron-Catalysed Reduction of Olefins using a Borohydride Reagent. *Adv. Synth. Catal.* **2013**, *355*, 880 – 885.

- (S21) Yang, C. -T.; Zhang, Z. -Q.; Liang, J.; Liu, J. -H.; Lu, X. -Y.; Chen, H. -H.; Liu, L. Copper-Catalyzed Cross-Coupling of Nonactivated Secondary Alkyl Halides and Tosylates with Secondary Alkyl Grignard Reagents. J. Am. Chem. Soc. 2012, 134, 11124 – 11127.
- (S22) Gieuw, M. H.; Ke, Z.; Yeung, Y. -Y. Lewis Base-Promoted Ring-Opening 1,3-Dioxygenation of Unactivated Cyclopropanes Using a Hypervalent Iodine Reagent. *Angew. Chem. Int. Ed.* 2018, *57*, 3782 – 3786.
- (S23) Greb, A.; Poh, J. -S.; Greed, S.; Battilocchio, C.; Pasau, P.; Blakemore, D. C.; Ley, S. V. A Versatile Route to Unstable Diazo Compounds via Oxadiazolines and their Use in Aryl– Alkyl Cross-Coupling Reactions. *Angew. Chem. Int. Ed.* **2017**, *56*, 16602 – 16605.
- (S24) Wang, Y.; Cao, X.; Zhao, L.; Pi, C.; Ji, J.; Cui, X.; Wu, Y. Generalized Chemoselective Transfer Hydrogenation/Hydrodeuteration. *Adv. Synth. Catal.* **2020**, *362*, 4119 – 4129.
- (S25) Brunard, E.; Boquet, V.; Elslande, E. V.; Saget, T.; Dauban, P. Catalytic Intermolecular C(sp<sup>3</sup>)–H Amination: Selective Functionalization of Tertiary C–H Bonds vs Activated Benzylic C–H Bonds. J. Am. Chem. Soc. 2021, 143, 6407 6412.
- (S26) Zheng, C.; Wang, G. -Z.; Shang, R. Catalyst-free Decarboxylation and Decarboxylative Giese Additions of Alkyl Carboxylates through Photoactivation of Electron Donor-Acceptor Complex. Adv. Synth. Catal. 2019, 361, 4500 – 4505.
- (S27) Li, D.; Han, T.; Xue, J.; Xu, W.; Xu, J.; Wu, Q. Engineering Fatty Acid Photodecarboxylase to Enable Highly Selective Decarboxylation of trans Fatty Acids. *Angew. Chem. Int. Ed.* 2021, 60, 20695 20699.
- (S28) Beato, E. d. P.; Spinnato, D.; Zhou, W.; Melchiorre, P. A General Organocatalytic System for Electron Donor–Acceptor Complex Photoactivation and Its Use in Radical Processes. J. Am. Chem. Soc. 2021, 143, 12304 – 12314.
- (S29) Hashimoto, T.; Nakatsu, H.; Takiguchi, Y.; Maruoka, K. Axially Chiral Dicarboxylic Acid Catalyzed Activation of Quinone Imine Ketals: Enantioselective Arylation of Enecarbamates. J. Am. Chem. Soc. 2013, 135, 16010 – 16013.

# NMR spectra of decarboxylation compounds





















## <sup>1</sup>H NMR spectrum of **10b** (600 MHz, CDCl<sub>3</sub>)



# <sup>19</sup>F NMR spectrum of **11a** (565 MHz, CDCl<sub>3</sub>)















<sup>1</sup>H NMR spectrum of **16** (600 MHz, CDCl<sub>3</sub>)

Crude <sup>1</sup>H NMR spectrum of **17** (600 MHz, CDCl<sub>3</sub>)

























# <sup>13</sup>C NMR spectrum of **30** (150 MHz, CDCl<sub>3</sub>)















## <sup>13</sup>C NMR spectrum of **32** (150 MHz, CDCl<sub>3</sub>)









# $^{13}\text{C}$ NMR spectrum of 35 (150 MHz, CDCl\_3)







# <sup>13</sup>C NMR spectrum of **37** (150 MHz, CDCl<sub>3</sub>)





# <sup>13</sup>C NMR spectrum of **39** (150 MHz, CDCl<sub>3</sub>)









# <sup>1</sup>H NMR spectrum of deuterated **2**' (600 MHz, Acetone-d6)



1.18 1.17 1.16 1.15 1.14 1.13 1.12 1.11 1.10 1.09 1.08

