

A Gain-of-Function TPC2 Variant R210C Increases Affinity to PI(3,5)P₂ and Causes Lysosome Acidification and Hypopigmentation

Wang Q, et al.

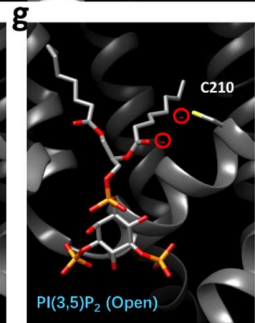
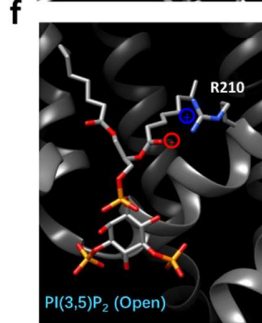
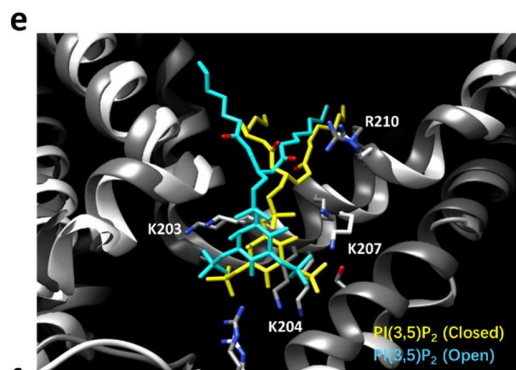
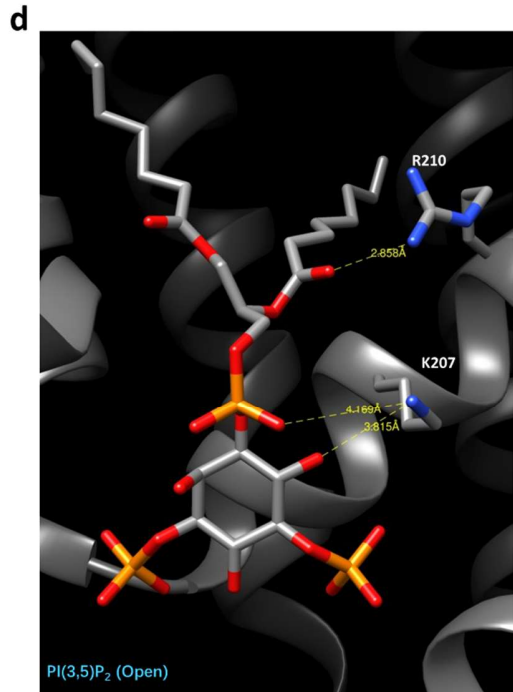
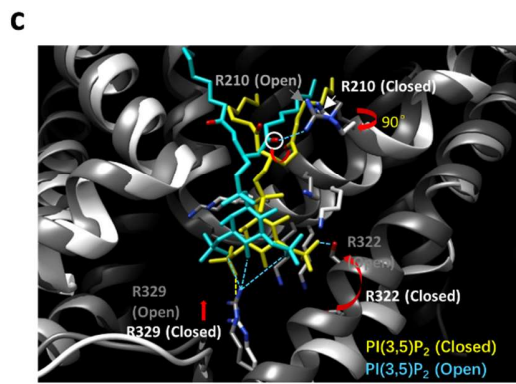
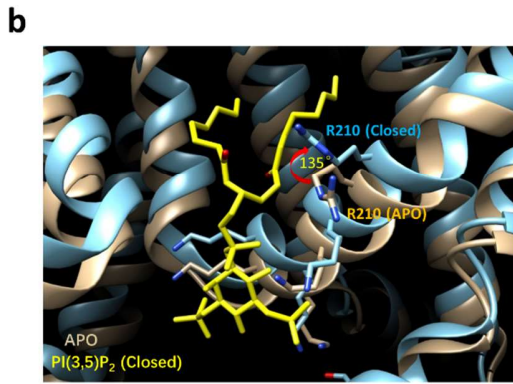
Supplementary Figures and Figure Legends

a

Species	Accession	Sequence	Residue
[Danio rerio TPC1]	XP_695923.4	SHLRVTRALRPIFLVDCRYCGAVRRNLRQIFQSLPPFIDILLLLFFMVI	242
[Mus musculus TPC1]	NP_665852.1	SHVRVTRALRCIFLVDCRYCGVRRNLRQIFQSLPPFMDILLLLFFMI	246
[Homo sapiens TPC1]	NP_001137291.1	SHVRVTRALRCIFLVDCRYCGVRRNLRQIFQSLPPFMDILLLLFFMI	317
[Oryctolagus cuniculus TPC1]	ON502176,	SHVRVTRALRCIFLVDCRYCGVRRNLRQIFQSLPPFMDILLLLFFMI	246
[Arabidopsis thaliana TPC1]	NP_567258.1	LPFRIAPYVRVIFIL--SIRELRDTLVLLSGMLGTYLNILALWMLFLF	229
[Danio rerio TPC2]	NP_001071190.1	ENLRVRRILRPFFLLQ--NSSLMKKTLCIKRRLPEIASVILLALHICL	243
[Gallus gallus TPC2]	XP_040529179.1	ETVRRRILRPFFLLQ--NSSMKKTLKINSTLPEMASVLLAVHLSL	216
[Mus musculus TPC2]	NP_666318.2	EPLRMRLLRPFFLLQ--NSSMKKTLKTRWSLPEMASVGLLATHLCL	213
[Homo sapiens TPC2]	NP_620714.2	EPLRIRLLRPFFLLQ--NSSMKKTLKTRWSLPEMASVGLLATHLCL	229
[Oryctolagus cuniculus TPC2]	ON502177	EPLRIRLLRPFFLLQ--NSSMKKTLKTRWSLPEMASVGLLAVHLSL	231
[Danio rerio TPC3]	NP_001170916.1	YGIKRSRVLPLLLVNVTEGRQIRRAFSTIRNALPQISYVFFLFMFSVLV	212
[Oryctolagus cuniculus TPC3]	EU344155.1	RGIRWSRAFQPVFLINFPESRQIRRALRSIRNTLPDILYVFLFMFVLI	195
[Gallus gallus TPC3]	NP_001137403.2	KSVRWSRIVRPIFLINFAESRQIRRAFSTIRNTLPEITYVFLFMFSLLM	183

poly-basic motif

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Supplementary Fig. 1 Partial sequence alignment of the TPC protein family and structural insights of R210 in TPC2 gating.

(a) Sequence alignment of TPC proteins. The blue box represents a poly-basic motif, and the black arrow indicates the position of R210 in human TPC2 and residues at the equivalent position in TPC2 of other species and TPC paralogs.

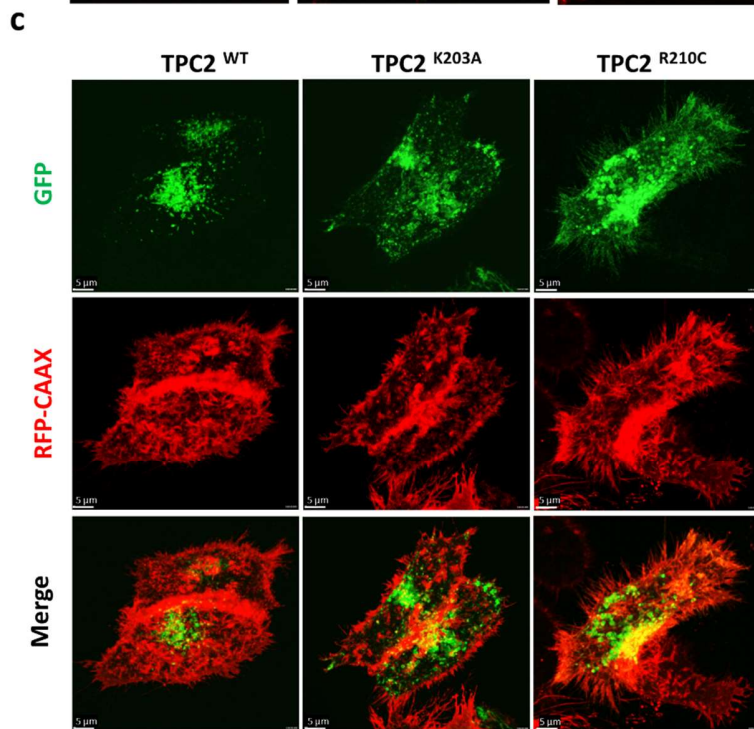
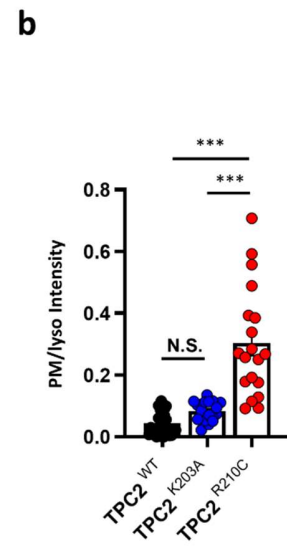
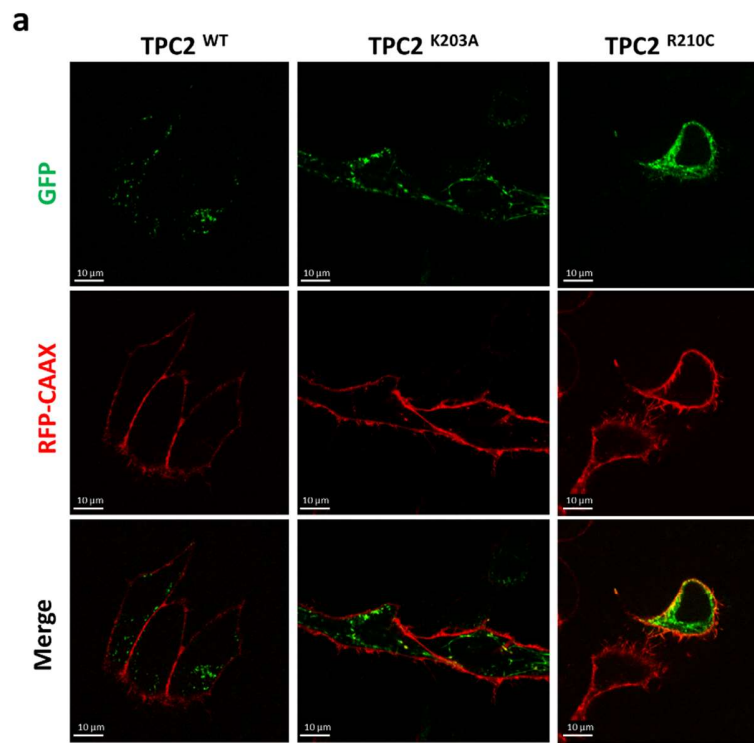
(b) Mobilization of R210 side chain when PI(3,5)P₂ (*highlighted in yellow*) binds to TPC2 during the conformational transition from the Apo state (*golden helix and golden-blue guanidinium*) to the ligand-bound closed state (*light blue helix and light-blue and blue guanidinium*), based on the human TPC2 cryo-EM structures. Note that there is a 135° twist of guanidine during the transition, indicating that R210 plays a role in regulating PI(3,5)P₂ binding. Also note that R210 is juxtaposed to the sn-1 fatty acyl chain of PI(3,5)P₂ in the ligand-bound closed state.

(c) Mobilization of R210 side chain and the flip of the PI(3,5)P₂ fatty acyl chains during the conformational transition from the ligand-bound closed state (*light grey*) to ligand-bound open state (*dark grey*). PI(3,5)P₂ is highlighted in yellow in the closed state, but in cyan in the open state. Note that R210 side chain rotated 90° and PI(3,5)P₂ fatty acyl chains flipped to bring the sn-2 carbonyl group (*marked by a white circle*) close to the guanidine group of R210, sufficient to form an electrostatic interaction. R329 and S322 on IS6 helix move toward PI(3,5)P₂ and establish salt bridges and hydrogen bond with the phosphates on the head of PI(3,5)P₂ (*shown in blue dot line*) to open the channel.

(d) Measured distance between the R210 side chain and the carbonyl groups on the fatty acyl chain of PI(3,5)P₂. In the ligand-bound closed state, the distance is 4.668 Å from sn-1 carbonyl group. In the ligand-bound open state, this distance is shortened to 2.858 Å from sn-2 carbonyl group. The closer distance increases stronger electrostatic interaction by ~2.66 folds according to the Coulomb's law.

(e) PI(3,5)P₂ interaction with TPC2 at the open state, highlighting the electrostatic interaction between R210 guanidine group and the sn-2 carbonyl group of PI(3,5)P₂, as well as the hydrophobic interaction between the fatty acyl chains and the S3 and S4 TM segments of TPC2.

(f, g) Electrostatic property change from R210 to C210, which represents that a positive-to-negative attractive force is changed into a negative-to-negative repulsive force, supporting our **Attraction-Repulsion (AR) Model** to explain the profound influence of PI(3,5)P₂-regulated TPC2 gating.

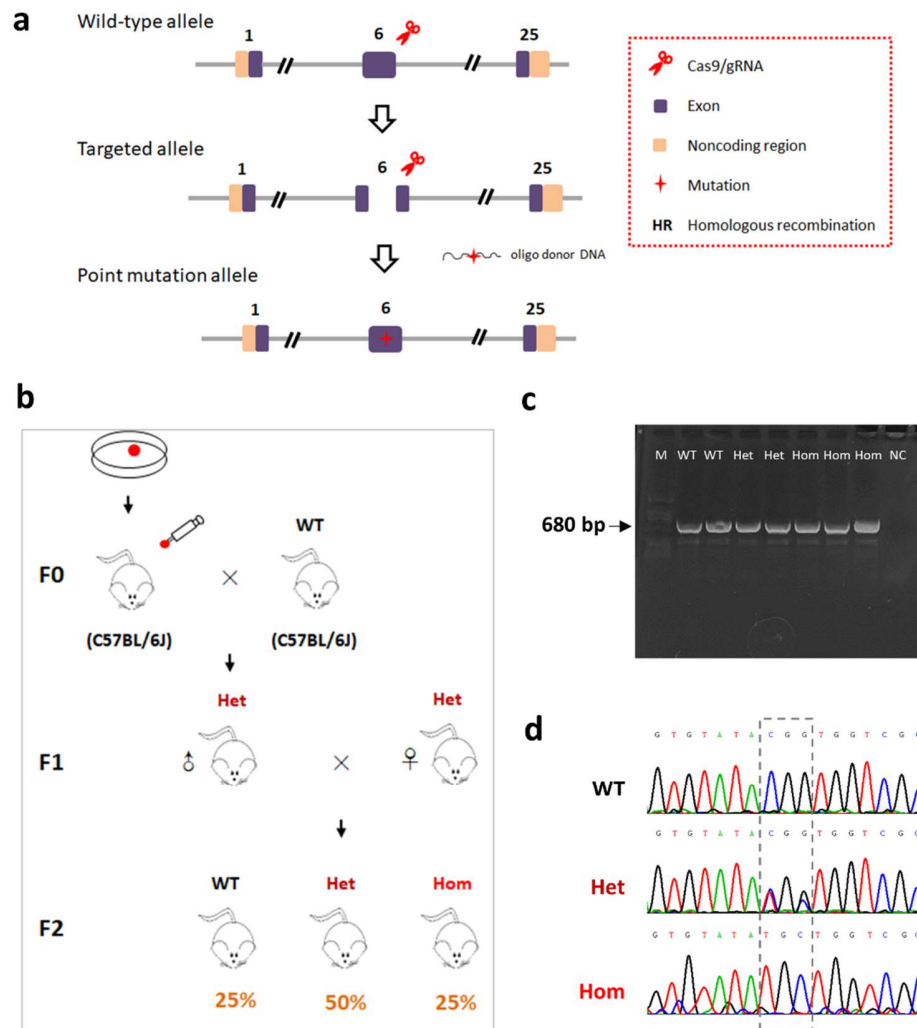


Supplementary Fig. 2 Altered localization and enlarged vacuoles in TPC2^{R210C} overexpressed cells.

(a) Representative confocal images of HeLa cells co-transfected with EGFP-TPC2 and RFP-CAAX. Overexpression of TPC2^{R210C} made cells round up and the EGFP signal was partially localized at the PM compared to TPC2^{WT} and TPC2^{K203A}. Scale bar, 10 μ m.

(b) Statistics ratio of EGFP fluorescent intensity on the PM to that associated with lysosomes. The ratios (Means \pm SEM) are: TPC2^{WT}, 0.04 \pm 0.01 (n = 27); TPC2^{K203A}, 0.08 \pm 0.01 (n = 19), and TPC2^{R210C}, 0.30 \pm 0.04 (n = 19). *** p < 0.001, N.S., not significant, by one-way ANOVA. Source data are provided as a Source Data file.

(c) 3D scanning images of HeLa cells co-transfected with EGFP-TPC2 and RFP-CAAX. Cells overexpressing TPC2^{R210C} exhibited a considerable amount of green fluorescence that co-localized with RFP-CAAX in the pseudopods. Scale bar, 5 μ m. This assay was repeated 3 times.



Supplementary Fig 3 Generation of *Tpcn2* R194C knock-in mouse strain with CRISPR-Cas9 editing strategy.

(a) A schematic flowchart for the generation of the mouse knock-in R194C variant with a gRNA targeting exon 6 of *Tpcn2* on the C57BL/6J background.

(b) Mouse mating processes. The mouse obtained from microinjection was mated with C57BL/6J to produce F1 offspring carrying heterozygous knock-in variant. The F1 mice were mated to give rise to F2 offspring.

(c) PCR amplicon product of 680 bp was confirmed on a 1% agarose gel and used for sequencing to conduct genotyping. This assay was repeated twice.

(d) The genotypes of offspring were verified by Sanger sequencing. The dashed grey box indicates the codon site of R194C variant.