

Supplementary Information

Biosensor-assisted titratable CRISPRi high-throughput (BATCH) screening for over-production phenotypes

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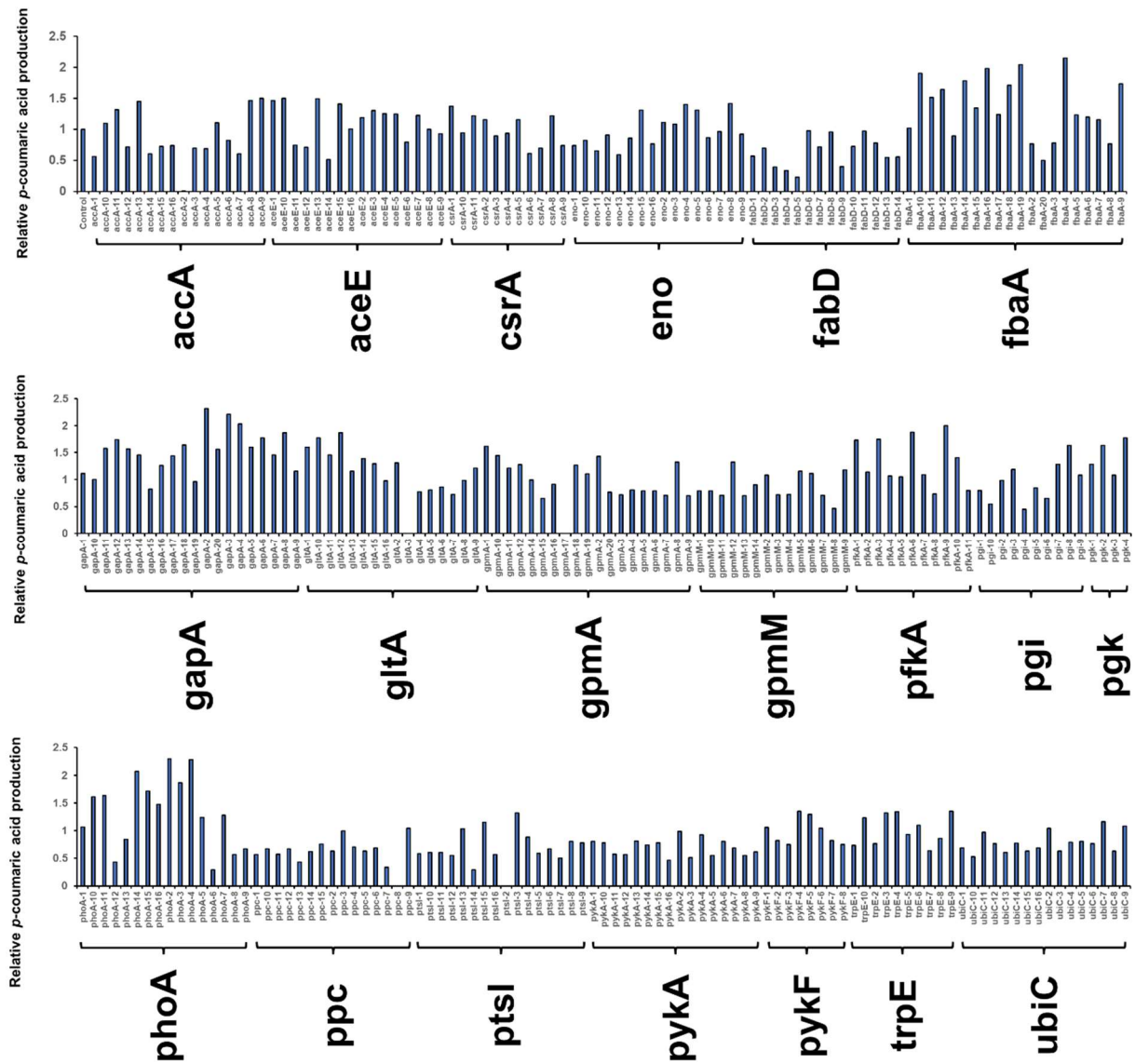


Figure S1. Relative *p*-coumaric acid titer of selected sgRNA variants in test tubes. The medium used for screening was M9Y with 20 g/L glycerol.

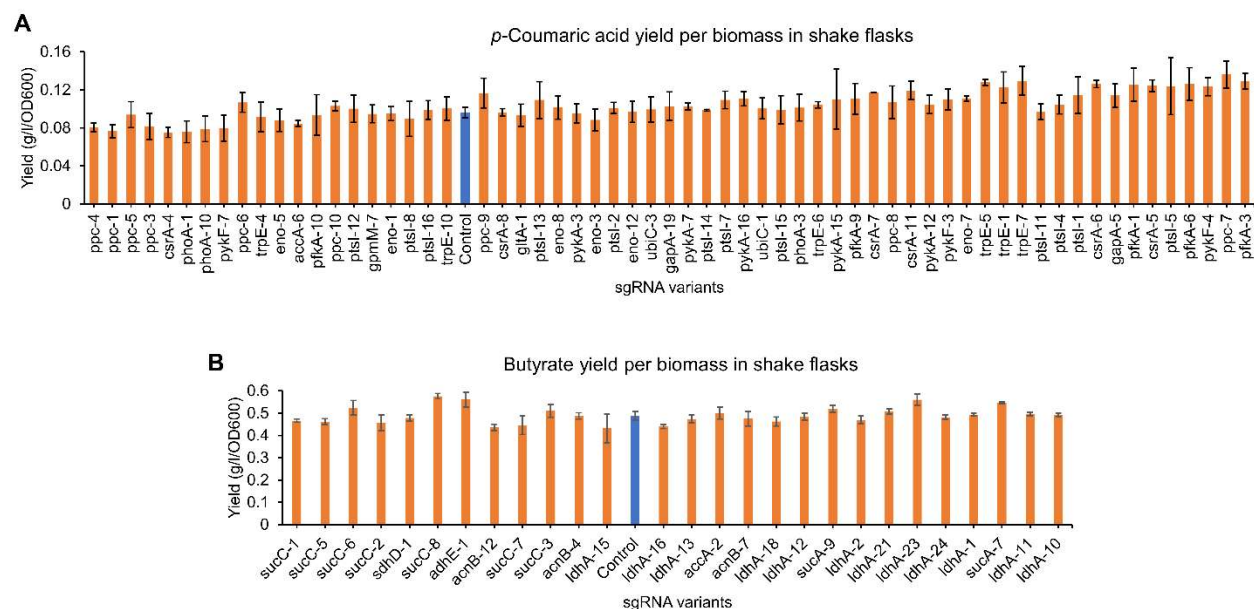


Figure S2. The overall yield (product titer/biomass (OD600)) profiles of *p*-coumaric acid (A) and butyrate (B) in shake flask experiments. The data used here corresponded to those from Figure 4C and 5E, respectively. All data represent the mean of three biologically independent samples and error bars show standard deviation.

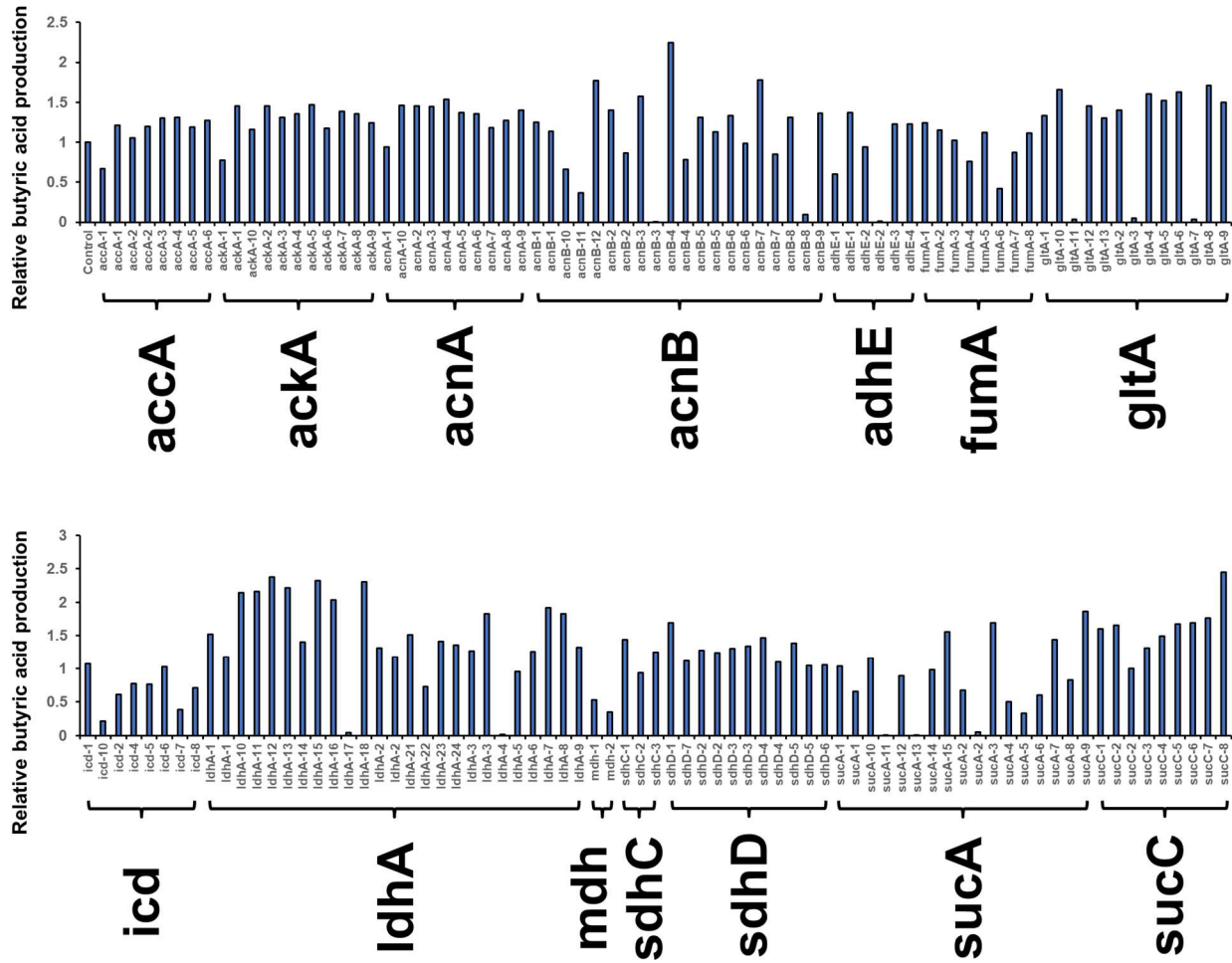


Figure S3. Relative butyric acid titer of selected sgRNA variants in test tubes. The medium used for screening was M9Y with 20 g/L glucose.

Table S1. Primers for constructing mismatched sgRNA arrays

Name	Sequence (5'-3') ^a
sgegfp-rbs-F (ApaLI)	ggaaaGTGCACA TTTCTCCTCTTTNNTGAATT GTTTTAGAG CTAGAAATAGCAAGTT
sgegfp-cds2-F (ApaLI)	ggaaaGTGCACA CTGTACATAACCNNCGGGCA GTTTTAGA GCTAGAAATAGCAAGTT
sgpgi-F(ApaLI)	ggaaaGTGCACA TGCCAGGCAGCGNNCTGCGT GTTTTAGA GCTAGAAATAGCAAGTT
sgpfkA-F(ApaLI)	ggaaaGTGCACA CGAATTGCGGCGNNCATGCC GTTTTAGA GCTAGAAATAGCAAGTT
sgfbaA-F(ApaLI)	ggaaaGTGCACA TCATCACCAGTGNNTACGCC GTTTTAGA GCTAGAAATAGCAAGTT
sggapA-F(ApaLI)	ggaaaGTGCACA GGAAAACAATGCNNCCGATA GTTTTAG AGCTAGAAATAGCAAGTT
sgpgk-F(ApaLI)	ggaaaGTGCACA TTTCCCAGCAAGNNCCAGAT GTTTTAGA GCTAGAAATAGCAAGTT
sggpmA-F(ApaLI)	ggaaaGTGCACA CGTCGTACCAACNNGTGAA GTTTTAGA GCTAGAAATAGCAAGTT
sggpmM-F(ApaLI)	ggaaaGTGCACA TCCAGAATCACCNNTACCAT GTTTTAGA GCTAGAAATAGCAAGTT
sgeno-F(ApaLI)	ggaaaGTGCACA ACCGATGATTTTNNCGATTT GTTTTAGA GCTAGAAATAGCAAGTT

sgpykA-F(ApaLI)	ggaaaGTGCACA TTTTGTTCTGCGNNGCCTTC GTTTTAGAG CTAGAAATAGCAAGTT
sgpykF-F(ApaLI)	ggaaaGTGCACA TCCGATGGTGCANNCAATTT GTTTTAGA GCTAGAAATAGCAAGTT
sgaceE-F(ApaLI)	ggaaaGTGCACA CGATCGGATCCANNTCATT GTTTTAGA GCTAGAAATAGCAAGTT
sggltA-F(ApaLI)	ggaaaGTGCACA AACAGCTGTATCNCGTTG AGTTTAGA GCTAGAAATAGCAAGTT
sgppc-F(ApaLI)	ggaaaGTGCACA ACTGACATTACTNNGCAATG GTTTTAGA GCTAGAAATAGCAAGTT
sgaccA-F(ApaLI)	ggaaaGTGCACA CAATCGGCTGTTNNAATCA GTTTTAGA GCTAGAAATAGCAAGTT
sgfabD-F(ApaLI)	ggaaaGTGCACA CGGTTTGAGAACNNTGTCC AGTTTAGA GCTAGAAATAGCAAGTT
sgcsrA-F(ApaLI)	ggaaaGTGCACA GACCTCATCCCNNTCATG AGTTTAGA GCTAGAAATAGCAAGTT
sgmurA-F(ApaLI)	ggaaaGTGCACA ACTTCGCCCTGGNNCTTCGT GTTTTAGA GCTAGAAATAGCAAGTT
sgphoA-F(ApaLI)	ggaaaGTGCACA GTCACAGGGGTANNCAGTAA GTTTTAGA GCTAGAAATAGCAAGTT
sgptsI-F(ApaLI)	ggaaaGTGCACA TTTACCGAAAGCNNTACCCG GTTTTAGA GCTAGAAATAGCAAGTT

sgubiC-F(ApaLI) ggaaaGTGCACA**CACGCAGTTGCGNNAACGCG**GTTTTAGA
GCTAGAAATAGCAAGTT

sgtrpE-F(ApaLI) ggaaaGTGCACA**CAGGTTAGCAGTNNGAGAGT**GTTTTAGA
GCTAGAAATAGCAAGTT

sgldhA-F(ApaLI) ggaaaGTGCACA**GTACTGTTTTGTNNTATAAA**GTTTTAGA
GCTAGAAATAGCAAGTT

sgadhE-F(ApaLI) ggaaaGTGCACA**GAAACTGGCATANNCACGCT**GTTTTAGA
GCTAGAAATAGCAAGTT

sgackA-F(ApaLI) ggaaaGTGCACA**ACCATTTACTGCNNCGATGA**GTTTTAGA
GCTAGAAATAGCAAGTT

sgacnA-F(ApaLI) ggaaaGTGCACA**GTCCTTACTGGCNNCTCGTA**GTTTTAGA
GCTAGAAATAGCAAGTT

sgacnB-F(ApaLI) ggaaaGTGCACA**CACGCTCAGCTANNTGCTTA**GTTTTAGA
GCTAGAAATAGCAAGTT

sgicd-F(ApaLI) ggaaaGTGCACA**GTGATCTTCTTGNNTTGTGC**GTTTTAGA
GCTAGAAATAGCAAGTT

sgsucA-F(ApaLI) ggaaaGTGCACA**GAGGTAAGAAGANNCCAACC**GTTTTAG
AGCTAGAAATAGCAAGTT

sgsucC-F(ApaLI) ggaaaGTGCACA**CGGTGCTGGTAANNCATAGC**GTTTTAGA
GCTAGAAATAGCAAGTT

sgsdhC-F(ApaLI) ggaaaGTGCACA**GTCTGTAGGTCCNNATTAAC**GTTTTAGA
GCTAGAAATAGCAAGTT

sgsdhD-F(ApaLI)	ggaaaGTGCACA GCCATTGCGTCCNNATGCGGG GTTTTAGA GCTAGAAATAGCAAGTT
sgfumA-F(ApaLI)	ggaaaGTGCACA AAAGGAGCCTGANNATGAAAG GTTTTAG AGCTAGAAATAGCAAGTT
sgmdh-F(ApaLI)	ggaaaGTGCACA CAATACCGCCAGNNGCGCCG GTTTTAGA GCTAGAAATAGCAAGTT
sgRNA-R(ApaLI)	ggaaagtgcAcagtatcttgttatccgctcacaat

^a The sgRNA spacers were shown in red.

Table S2. DNA sequences of the screened sgRNA variants.

sgRNA variants	Spacer DNA sequence (fully matched base pair)
For <i>p</i>-coumaric acid screening	
csrA-5	GACCTCATCCCC T TCATGA (AA)
csrA-6	GACCTCATCCCC T TCATGA (AA)
csrA-7	GACCTCATCCCC A TTCATGA (AA)
csrA-8	GACCTCATCCCC T GTCATGA (AA)
csrA-11	GACCTCATCCCC C GTCATGA (AA)
eno-1	ACCGATGATTTT T CCGATTT (TA)
eno-3	ACCGATGATTTT G GCGATTT (TA)
eno-5	ACCGATGATTTT T CCGATTT (TA)
eno-7	ACCGATGATTTT T CCGATTT (TA)
eno-8	ACCGATGATTTT T TCGATTT (TA)
eno-12	ACCGATGATTTT G GCGATTT (TA)
gapA-5	GGAAAACAATGC A GCCGATA (GA)
gapA-19	GGAAAACAATGC C GCCGATA (GA)
gltA-1	AACAGCTGTATC A ACGTTGA (CC)
gpmM-7	TCCAGAATCACCT A TACCAT (AG)
pfkA-3	CGAATTGCGGCG T TTCATGCC (TT)
pfkA-6	CGAATTGCGGCG T GTCATGCC (TT)
phoA-3	GTCACAGGGGTA A ACAGTAA (AA)
ppc-7	ACTGACATTACT T GGCAATG (AC)
ppc-8	ACTGACATTACT T GGCAATG (AC)

ppc-9	ACTGACATTACTATGCAATG (AC)
ptsI-1	TTTACCGAAAGCATTACCCG (GA)
ptsI-4	TTTACCGAAAGCCTTACCCG (GA)
ptsI-5	TTTACCGAAAGCCGTACCCG (GA)
ptsI-7	TTTACCGAAAGCTTTACCCG (GA)
ptsI-11	TTTACCGAAAGCTGTACCCG (GA)
ptsI-14	TTTACCGAAAGCAGTACCCG (GA)
ptsI-15	TTTACCGAAAGCTTTACCCG (GA)
pykA-3	TTTTGTTCTGCGTGGCCTTC (AA)
pykA-7	TTTTGTTCTGCGGAGCCTTC (AA)
pykA-12	TTTTGTTCTGCGGAGCCTTC (AA)
pykA-15	TTTTGTTCTGCGGGCCTTC (AA)
pykA-16	TTTTGTTCTGCGGCGCCTTC (AA)
pykF-3	TCCGATGGTGCAGACAATTT (AA)
pykF-4	TCCGATGGTGCAGGCAATTT (AA)
pykF-7	TCCGATGGTGCATCAATTT (AA)
trpE-1	CAGGTTAGCAGTTCGAGAGT (TC)
trpE-5	CAGGTTAGCAGTGGGAGAGT (TC)
trpE-6	CAGGTTAGCAGTCGGAGAGT (TC)
trpE-7	CAGGTTAGCAGTCGGAGAGT (TC)
ubiC-1	CACGCAGTTGCGGCAACGCG (TT)
ubiC-3	CACGCAGTTGCGGCAACGCG (TT)

For butyrate screening

accA-2	CAATCGGCTGTTATAAATCA (CA)
acnB-4	CACGCTCAGCTAAATGCTTA (CG)
acnB-7	CACGCTCAGCTAGATGCTTA (CG)
acnB-12	CACGCTCAGCTATTTGCTTA (CG)
ldhA-2	GTACTGTTTTGTTATATAAA (GC)
ldhA-10	GTACTGTTTTGTCTATAAA (GC)
ldhA-11	GTACTGTTTTGTCTATAAA (GC)
ldhA-12	GTACTGTTTTGTGCTATAAA (GC)
ldhA-16	GTACTGTTTTGTAAATAAA (GC)
ldhA-18	GTACTGTTTTGTGTATAAA (GC)
ldhA-21	GTACTGTTTTGTATTATAAA (GC)
ldhA-23	GTACTGTTTTGTTTATAAA (GC)
ldhA-24	GTACTGTTTTGTGTATAAA (GC)
sdhD-1	GCCATTGCGTCCGCATGCGG (TA)
sucA-7	GAGGTAAGAAGAATCCAACC (GT)
sucA-9	GAGGTAAGAAGATCCAACC (GT)
sucC-1	CGGTGCTGGTAATGCATAGC (GC)
sucC-2	CGGTGCTGGTAACGCATAGC (GC)
sucC-3	CGGTGCTGGTAAATCATAGC (GC)
sucC-5	CGGTGCTGGTAAAGCATAGC (GC)
sucC-6	CGGTGCTGGTAAGGCATAGC (GC)
sucC-7	CGGTGCTGGTAAACATAGC (GC)
sucC-8	CGGTGCTGGTAAGGCATAGC (GC)

Table S3. Sequences of hybrid promoters used in *p*-coumaric acid responsive biosensor system

promoter	DNA sequences ^a
P1	AACATGTAAATAGTTACATGATTTGACATAAATACCAC TGGCGGTGATACTGAGCACATCAGCAGGACGCACTGAC CGAATTCATTAAAGAGGAGAAA
P2	TAAATTATCTCTGGCGGTGTTGACA AACATGTAAATAGT TACATGATGATACTGAGCACATCAGCAGGACGCACTGA CCGAATTCATTAAAGAGGAGAAA
P7	AACATGTAAATAGTTACATGATTTGACAATGGTGTTAA AGTGAACATGTA GATACTGAGCACATCAGCAGGACGCA CTGACCGAATTCATTAAAGAGGAGAAA
P9	TAAATTATCTCTGGCGGTGTTGACATAAATACCACTGGC GGTGATACTG AACATGTAAATAGTTACATGATGAATTC ATTAAAGAGGAGAAA

^a The PadR binding box 1 was labeled in blue while the PadR binding box 2 was labeled in brown. The RBS sequence was labeled in green.