Figure S1

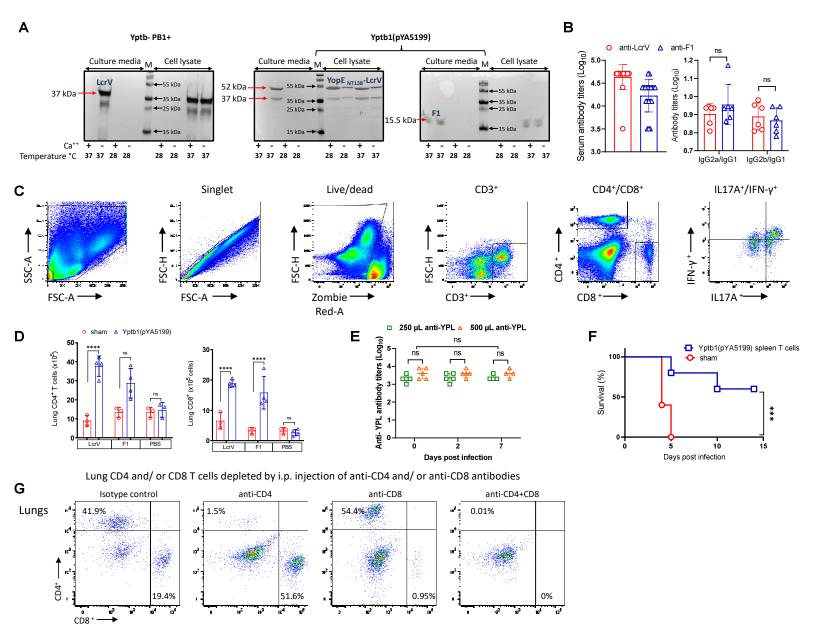


Figure S1. Characterization and immunogenicity of Yptb1(pYA5199) in Swiss webster mice. (A) Western blot analysis of YopE_{NT138}-LcrV, LcrV, and F1 synthesis in Yptb1(pYA5199) that was cultured in a medium replete with or deprived of Ca²⁺. (B) Antibody responses to *Y. pestis* LcrV and F1 antigens in Yptb1(pYA5199)-immunized mice (n=15, mixed males and females) at 28 dpv. Serum total IgG titers to LcrV and F1 and ratios of IgG2a/IgG1 and IgG2b/IgG1 to a respective antigen. (C) Gating strategy for CD4+/CD8+ T cells with IFN-γ, TNF-α, and/or IL-17A production. (D) Quantitative analysis of LcrV- or F1-specific CD4+ and CD8+ T cells in the lungs of Yptb1(pYA5199)-immunized mice (n=6 females) at 42 dpv. (E) Serum anti-YPL antibody titers in naïve mice (n=5 females) that adoptively received 250 μl and 500 μl of serum from Yptb1(pYA5199)-immunized mice were measured at days 0, 2, and 7 post-pulmonary *Y. pestis* challenge (10 LD₅₀). (F) The protective role of spleen T cells against pneumonic plague. Naïve irradiated (5 Gy) mice were i.v. injected with purified spleen (CD4+ CD8+) T cells isolated from Yptb1(pYA5199)-immunized and sham mice at 42 dpv. At 24 h post-transfer, mice (n=5 females) were intranasally infected with 10 LD₅₀ of *Y. pestis*, and survival was recorded for 14 days. (G) A representative flow plot showing depletion of CD4+ and CD8+ T cells in the lungs by i.p. injection of 500 μg of mouse anti-CD4 and anti-CD8 monoclonal antibodies (mAbs). Each symbol in the bar graph represents a data point obtained from an individual mouse. Data obtained from experiments are presented as the mean ± SD. The statistical analysis is described in the Materials and Methods.



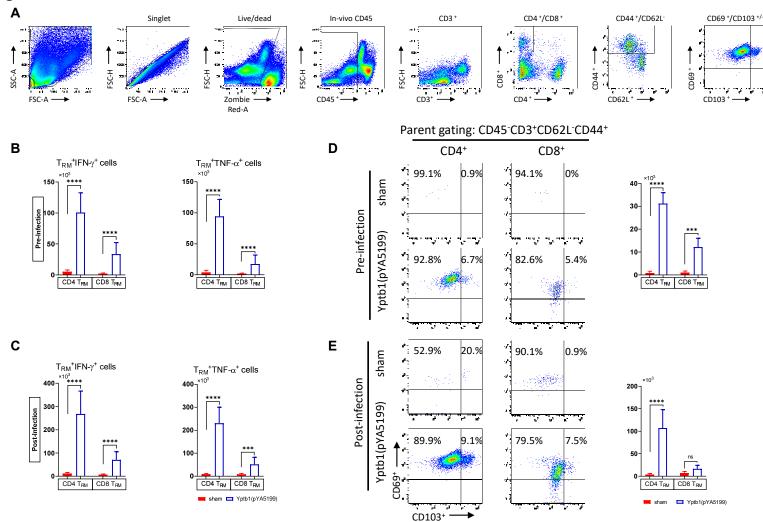


Figure S2: Activation of lung CD4⁺ and CD8⁺ T_{RM} cells in Yptb1(pYA5199)-immunized mice. (A) A gating strategy for T_{RM} cells in the lung. The number of CD4⁺ and CD8⁺ T_{RM} cells expressing IFN-γ⁺ or TNF-α⁺ before infection (B) and after infection (C). Lung single cells isolated from mice (n=5 females) at 42 dpv (pre-infection) or at 2 dpi with *Y. pestis* (post-infection) were stained for T_{RM} surface markers (CD45⁻, CD4⁺/CD8⁺, CD44⁺, and CD69⁺) and intracellular cytokines (IFN-γ or TNF-α) for flow cytometry. (D) Representative flow plots showing lung CD4⁺ and CD8⁺ T_{RM} cells with CD103⁺ expression in pre-infected mice (n=6 females). (E) Lung CD4⁺ and CD8⁺ T_{RM} cells with CD103⁺ expression at 2 dpi. Bar graphs showing quantitative analysis of CD4⁺ or CD8⁺ T_{RM} cells expressing CD103⁺. Data obtained from experiments were pooled and analyzed and are presented as the mean \pm SD. The statistical analysis is described in the Materials and Methods.

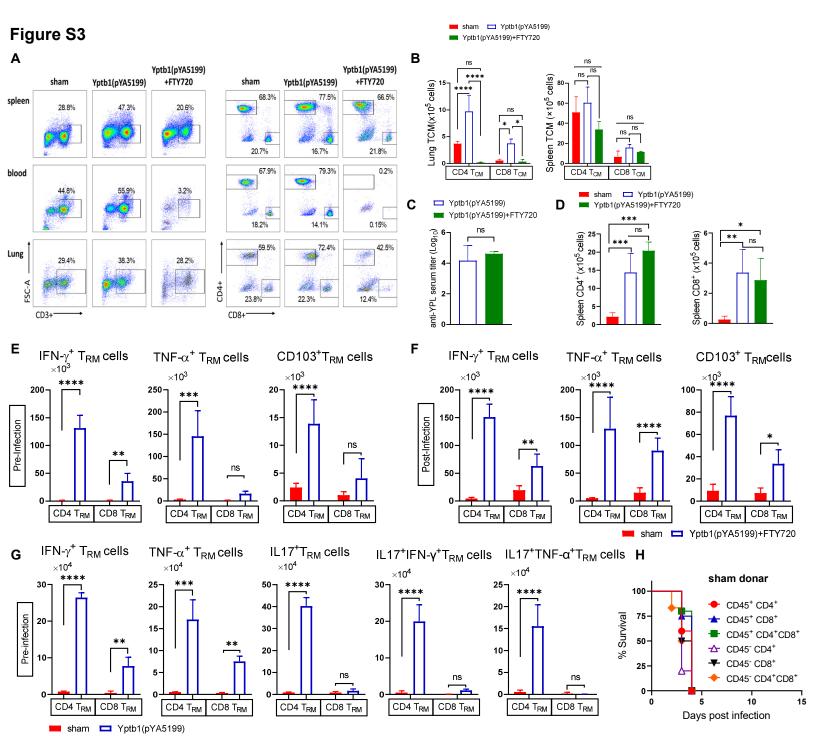


Figure S3. Resident and central memory T cells in Yptb1(pYA5199)-immunized mice treated with or without FTY720. (A) Flow cytometric analysis of the ability of FTY720 treatment to block circulating T cells. Flow plot showing CD3⁺, CD4⁺, and CD8⁺ T cells in the blood, lung, and spleen obtained from sham, Yptb1(pYA5199)immunized, and FTY720-treated Yptb1(pYA5199)-immunized mice (n=6 females). (B) Quantitative analysis of T_{CM} cells (CD62L⁺) in the lung and spleen of sham mice and immunized mice with and without FTY720 treatment. (C) Total anti-YPL IgG titers in Yptb1(pYA5199)-immunized mice (n=10, mixed males and females) with and without FTY720 treatment. (D) Spleen CD4⁺ and CD8⁺ T-cell responses in Yptb1(pYA5199)-immunized mice (n=6 females) with and without FTY720 treatment at 42 dpv. (E) Lung CD4⁺ and CD8⁺ T_{RM} cells (CD45⁻ CD44⁺ CD69⁺ CD4⁺/CD8⁺) expressing IFN-γ, TNF-α, or CD103 in sham and Yptb1(pYA5199)+FTY720 mice (n=5 females) at 42 dpv (preinfection). (F) Lung CD4⁺ and CD8⁺ T_{RM} cells expressing IFN-γ, TNF-α, or CD103 at 2 dpi (postinfection). (G) Lung T cells FACS sorted at 42 dpv were induced in vitro with 20 µg of LcrV for 48 h. Cells were surface stained with CD4+ and CD8+ T cells expressing IL17A, IFN-γ, and TNF-α markers. (H) Adoptive transfer of FACS-sorted lung circulating (CD45⁺) T cells or T_{RM} cells (CD45⁻) from sham mice. At 24 h post intratracheal administration, recipient mice (n=5 females) were i.n. challenged with 10 LD₅₀ of Y. pestis and survival was recorded for 14 days. Data obtained from experiments were pooled and analyzed and are presented as the mean \pm SD. The statistical analysis is described in the Materials and Methods.

Figure S4

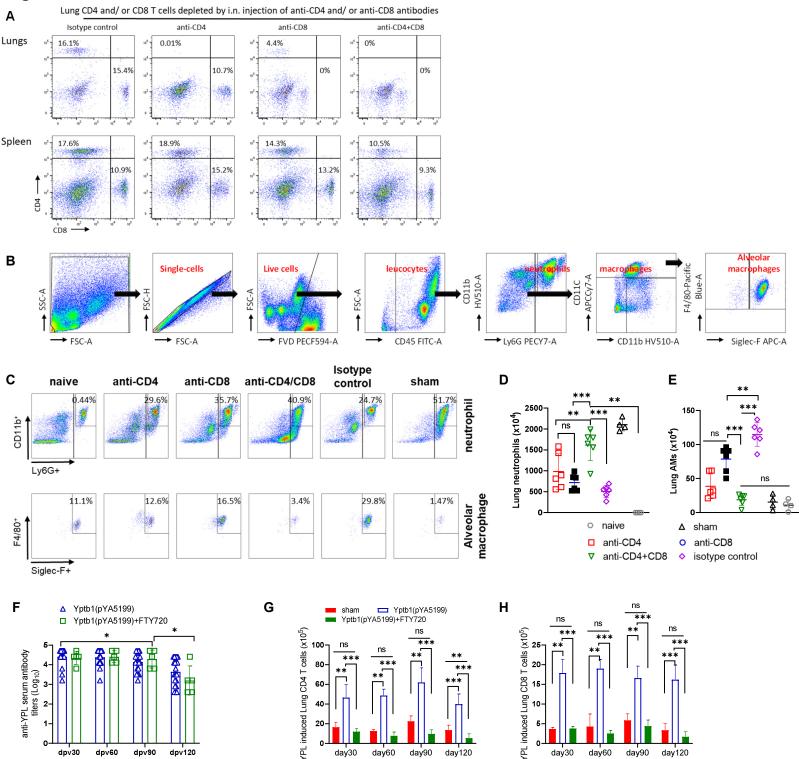


Figure S4. Alterations in lung neutrophil and alveolar macrophage populations in T cells depleted mice and the assessment of long-term immune responses in Yptb1(pYA5199) and Yptb1(pYA5199)+FTY720 mice. (A) A representative flow plot showing depletion of CD4⁺ and CD8⁺ T cells in the lungs by i.n. injection of 200 μg of mouse anti-CD4 and anti-CD8 monoclonal antibodies (mAbs). (B) Gating strategy to analyze neutrophil and macrophage populations in the lung and BAL fluid. (C) Representative flow plots showing the percentages of lung neutrophil and alveolar macrophage populations in naïve, sham, anti-CD4 mAb-treated, anti-CD8 mAb-treated, both anti-CD4 and anti-CD8 mAb-treated, and isotype IgG-treated mice (n=6 females.) at 2 dpi. (D) The number of lung neutrophils and (E) alveolar macrophages (AMs) in Yptb1(pYA5199)+FTY720 mice after different treatments. Sham and naïve mice were considered controls. (F) Kinetics of anti-YPL serum antibodies and (G) YPL-specific CD4⁺ and (H) CD8⁺ T cells in the lung of Yptb1(pYA5199) and Yptb1(pYA5199)+FTY720 mice (n=10, equal number of males and females) at 30, 60, 90, and 120 dpv. The statistical analysis is described in the Materials and Methods.