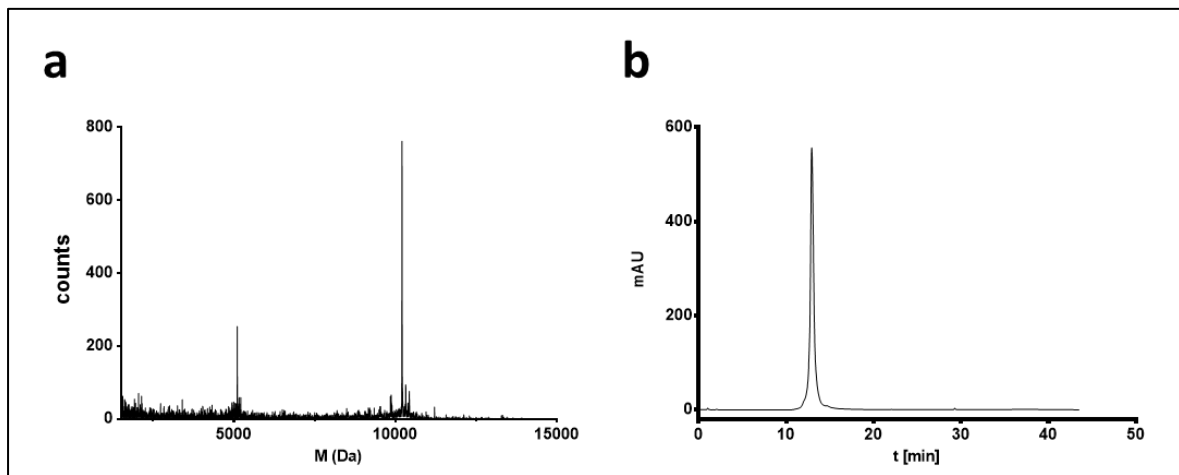


## Supplementary Materials for

### Antibody-oligonucleotide conjugate achieves central nervous system delivery in animal models for spinal muscular atrophy

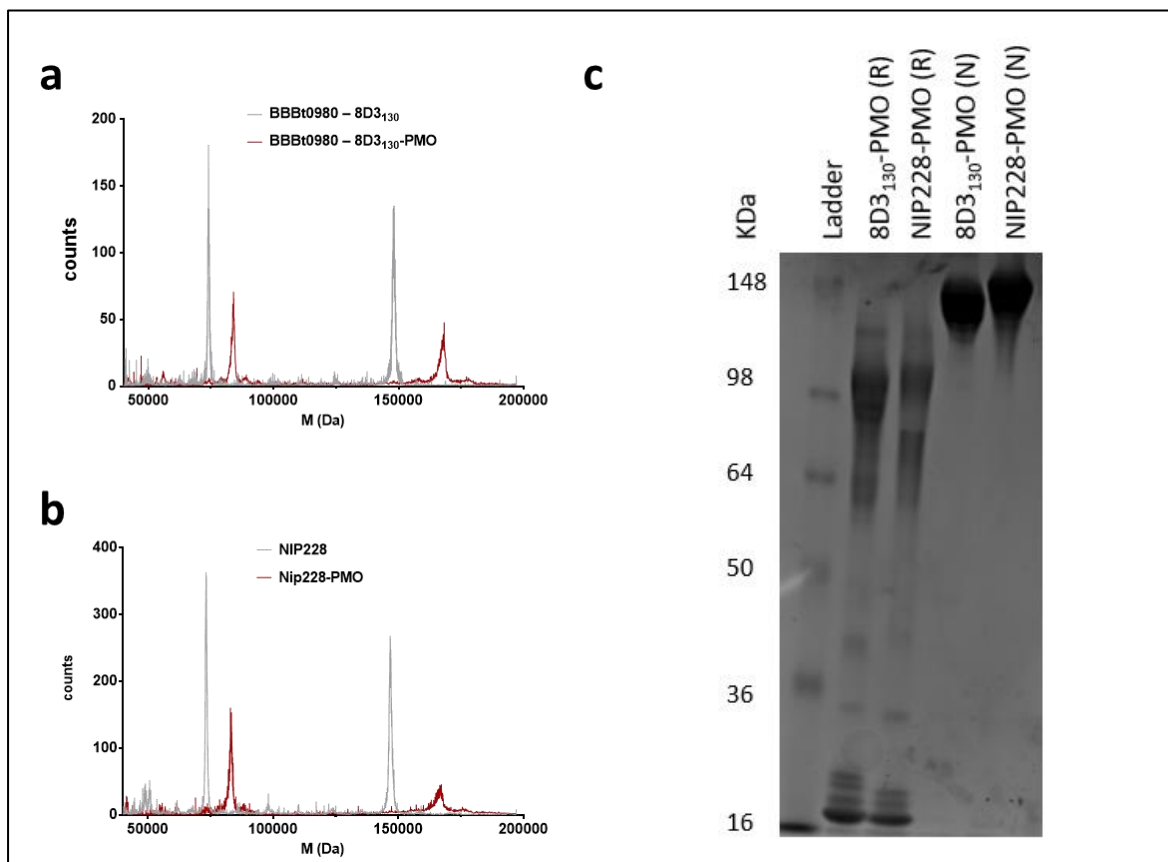
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[carl.webster@astrazeneca.com](mailto:carl.webster@astrazeneca.com)



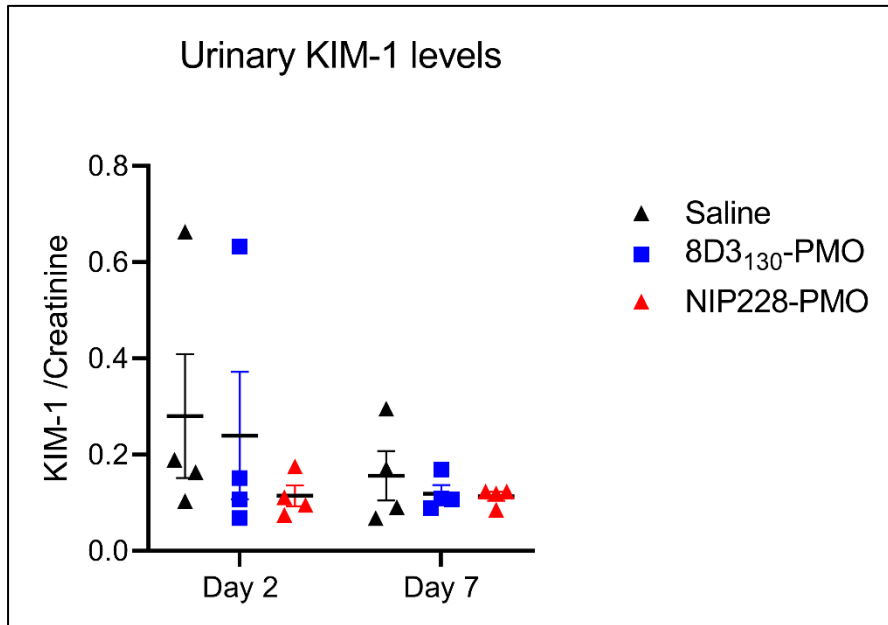
#### Supplemental Figure S1.

(A) MALDI-TOF spectra of 25-mer PMO targeting ISS-N1 and directly conjugated it to a short maleimide functionalized peptide linker, Mal-C3-FB[RB]<sub>6</sub>-PMO. (B) LC-trace (260nm) of Mal-C3-FB[RB]<sub>6</sub>-PMO.



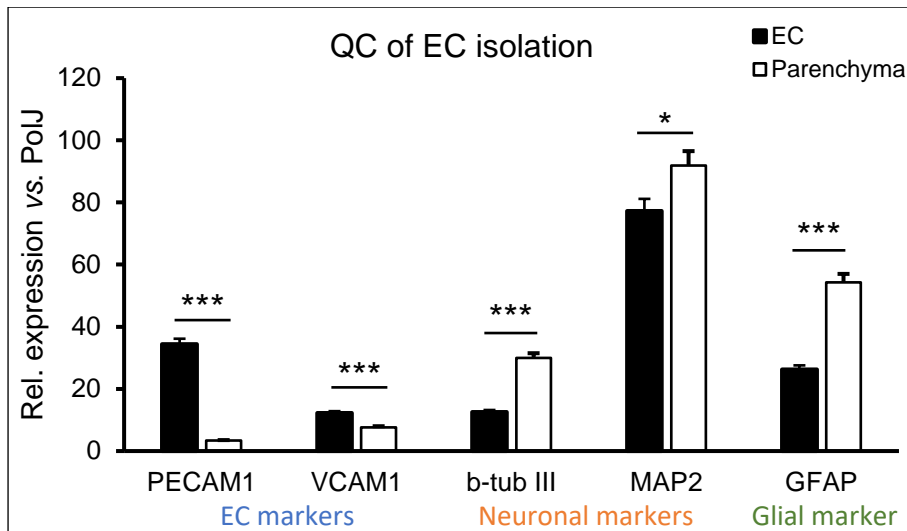
### Supplemental Figure S2.

(A) MALDI-TOF spectra of 8D3<sub>130</sub> before (grey line) and after (red line) conjugation reaction with Mal-PPMO. (B) MALDI-TOF spectra of NIP228 before (grey line) and after (red line) conjugation reaction with Mal-PPMO. (C) SDS-Page of reduced antibody-PMO conjugates (R) and non-reduced antibody-PMO conjugates (N).



### Supplemental Figure S3.

Preliminary *in vivo* toxicity data for antibody-PMOs in male adult transgenic mice bearing the human *SMN2* gene. Tail vein administration of 8D3<sub>130</sub>-PMO and NIP228-PMO were given at 8 weeks of age and urine collected on 2- and 7-days post-administration. Urinary KIM-1 levels were measured via an ELISA and normalised to urinary creatinine levels measured using a clinical chemistry analyser (MRC Harwell Institute, UK). Data shown as mean  $\pm$  S.E.M., n=4 for each group. Data were analysed via 2-way ANOVA corrected for multiple comparisons using Tukey Test. P values adjusted to account for each comparison, confidence level 0.95%. \*p, <0.05; \*\*p, <0.005; \*\*\*p, <0.0005; \*\*\*\*p, <0.0001; #p, <0.05; ##p, <0.005; ###p, <0.0005; ####p, <0.0001; ns, not significant.



#### Supplemental Figure S4.

Endothelial cell isolation quality control data. The relative expression of endothelial cell (EC) markers, Pcam1 and Vcam1, were significantly higher in the EC fraction than the parenchyma fraction. On the other hand, the expression of neuronal markers  $\beta$ -tubulin III (Tubb3) and microtubule-associated protein 2 (Map2), was significantly higher in parenchyma fractions than in the EC. In addition, the expression of Glial fibrillary acidic protein (Gfap) was significantly higher in parenchyma fractions than in the EC ( $54.3 \pm 3.6$  vs.  $26.3 \pm 1.2$ ) as expected. These all indicate that the EC isolation resulted in enriched endothelial cells.

**Supplemental Table S1.**

QPCR primers for SMN2 expression

<b>Primer/Probe</b>	<b>FLSMN2 (5'-3')</b>	<b>Exon/Exon junction</b>
Reverse Primer	TCGTTTCTTTAGTGGTGTCAATTTAG	Ex8
Forward Primer	TATCATACTGGCTATTATATGGGTTTT	Ex6-Ex7
Probe	AAGGAGAAATGCTGGCATAGAGCAGC	Ex7-Ex8
	<b>TotalSMN2 (5'-3')</b>	
Reverse Primer	TCAGTGCTGTATCATCCCAAATG	Ex2a
Forward Primer	CAGGAGGATTCCGTGCTGTT	Ex1
Probe	CGGCACAGGCCAGAGCGATG	Ex1-Ex2a

**Supplemental Table S2.**

QPCR primers for endothelial cell isolation quality control data

<b>Primer/Probe</b>	<b><i>Pecam1</i> (NM_008816)</b>
Forward	TGGTTGTCATTGGAGTGGTC
Probe	CACGGGTTTCTGTTTGGCCTTGG
Reverse	TTCTCGCTGTTGGAGTTCAG
	<b><i>Vcam1</i> (NM_011693)</b>
Forward	GCAAAGGACACTGGAAAAGAG
Probe	CACTTGTGCATGGGAGACCTGTCA
Reverse	TCAAAGGGATACACATTAGGGAC
	<b><i>Tubb3</i> (NM_023279)</b>
Forward	CGCCTTTGGACACCTATTCAG
Probe	CGCCCTCCGTATAGTGCCCTTTG
Reverse	TTCTCACACTCTTCCGCAC
	<b><i>Map2</i> (NM_008632.2)</b>
Forward	CAGGGCACCTATTCAGATACC
Probe	CAGCTCTCCGTTGATCCCGTTCT
Reverse	TCCTTCTTTGTTACCTTTCAG
	<b><i>Gfap</i> (NM_010277)</b>
Forward	GAAAACCGCATCACCATTC
Probe	AGACTTTCTCCAACCTCCAGATCCGA
Reverse	CTTAATGACCTCACCATCCCG