

*Sci Rep.* [Original Research]

# Substrate stiffness controls proinflammatory responses in human gingival fibroblasts

Watcharaphol Tiskratok<sup>1,2</sup>, Masahiro Yamada<sup>1\*</sup>, Jun Watanabe<sup>1</sup>, Nadia Kartikasari<sup>1</sup>, Tsuyoshi Kimura<sup>3</sup>, and Hiroshi Egusa<sup>1,4\*</sup>

<sup>1</sup> *Division of Molecular and Regenerative Prosthodontics, Tohoku University Graduate School of Dentistry, Sendai, Miyagi, Japan.*

<sup>2</sup> *Institute of Dentistry, Suranaree University of Technology, Mueang Nakhon Ratchasima District, Nakhon Ratchasima 30000, Thailand.*

<sup>3</sup> *Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University, Chiyoda-ku, Tokyo, Japan*

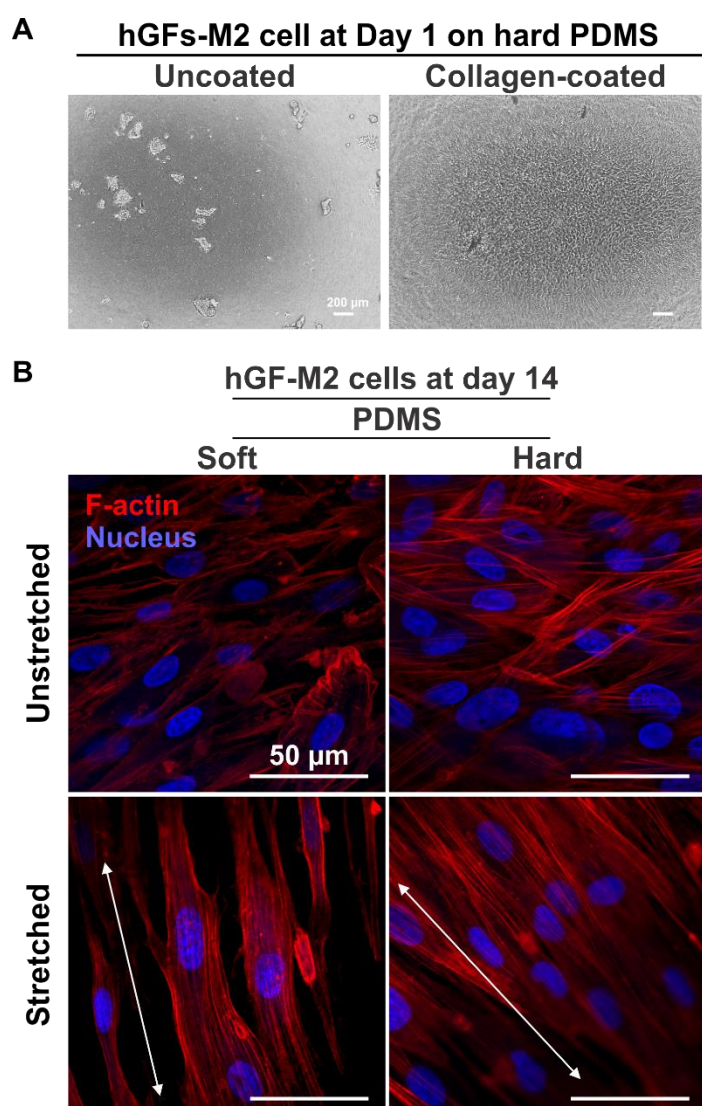
<sup>4</sup> *Center for Advanced Stem Cell and Regenerative Research, Tohoku University Graduate School of Dentistry, Sendai, Miyagi, Japan.*

**Keywords:** Cytokines, extracellular matrix, gingivitis, inflammation, mechanotransduction, mucosal immunity

**\* Address corresponding to:**

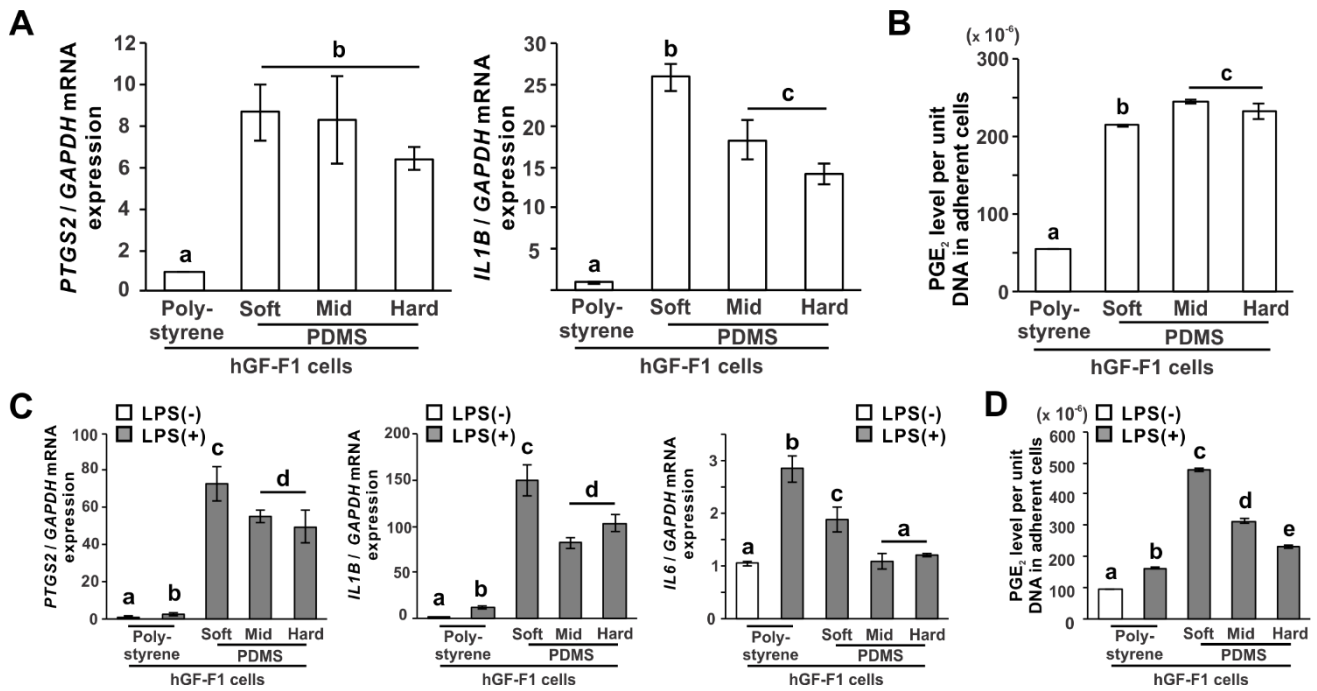
Masahiro Yamada, DDS, PhD  
Division of Molecular and Regenerative Prosthodontics,  
Tohoku University Graduate School of Dentistry  
4-1 Seiryomachi, Aoba-ku, Sendai, 980-8575, Japan  
tel: +81-22-717-8363; fax: +81-22-717-8367  
E-mail: [masahiro.yamada.a2@tohoku.ac.jp](mailto:masahiro.yamada.a2@tohoku.ac.jp)

Hiroshi Egusa, DDS, PhD  
Division of Molecular and Regenerative Prosthodontics,  
Tohoku University Graduate School of Dentistry  
4-1 Seiryomachi, Aoba-ku, Sendai, 980-8575, Japan  
tel: +81-22-717-8363; fax: +81-22-717-8367  
E-mail: [egu@tohoku.ac.jp](mailto:egu@tohoku.ac.jp)



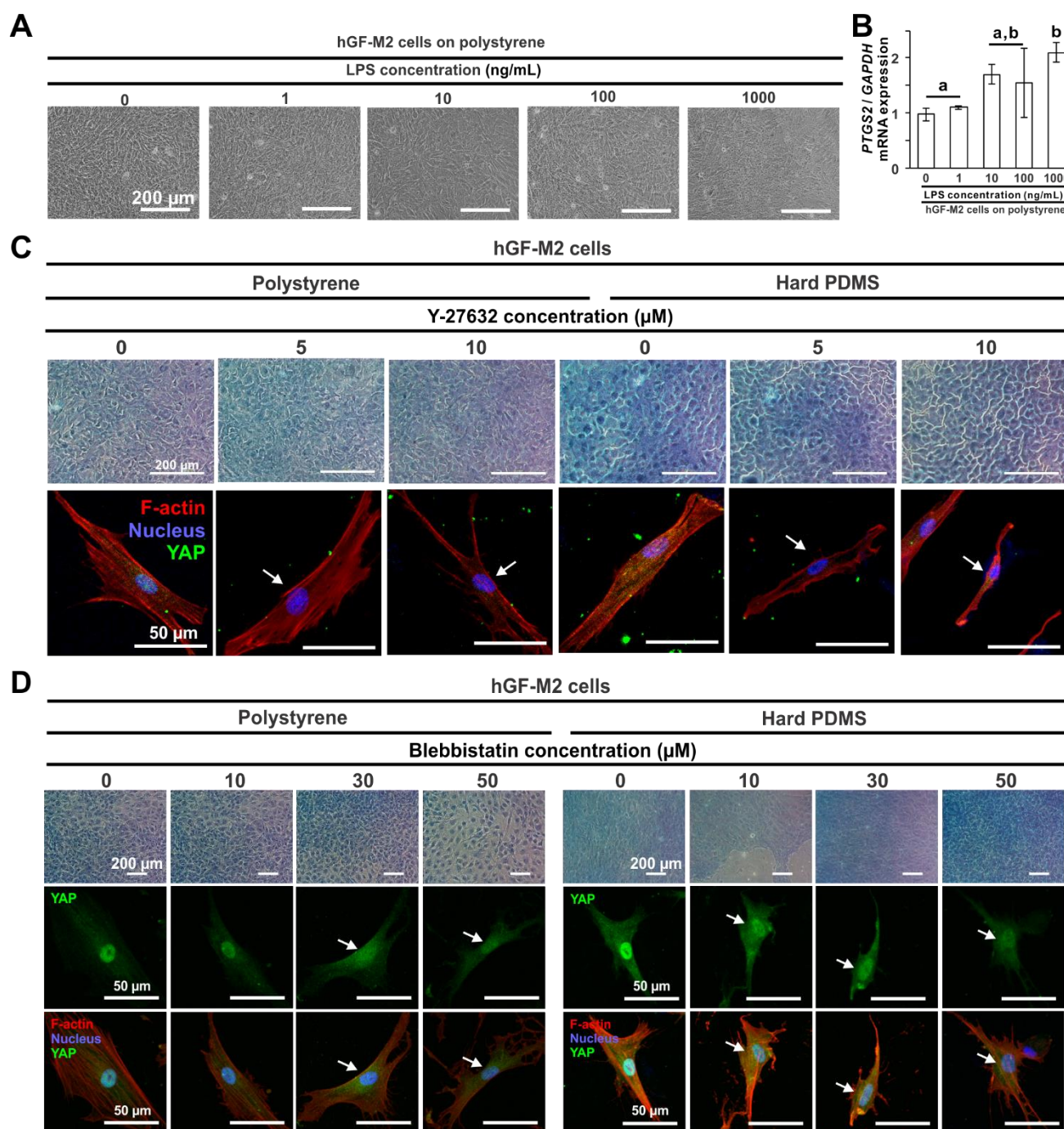
**Supplementary Figure 1. Effects of collagen-coating on hGFs attachment on PDMS substrates**

(A) Representative phase microscopic images in hGF-M2 cells cultured for 24 h on hard PDMS precoated with or without 0.1wt% collagen. (B) Fluorescence confocal laser microscopic images of *F*-actin (red), and nucleus (blue) in hGF-M2 cells cultured for 14 days on 0.1wt% collagen-coated soft or hard PDMS before and after unidirectionally stretching the PDMS substrates. Note: (A) cells cannot adhere to the PDMS substrate without the collagen coating (left panel) and (B) the hGFs are stretched along the direction axis of the PDMS substrates at day 14, regardless of the stiffness. Double-headed arrows in (B) indicate the stretching direction of PDMS substrates. hGFs, Human gingival fibroblasts; PDMS, Polydimethylsiloxane.



**Supplementary Figure 2. Effects of substrate stiffness on proinflammatory responses in the other cell populations of hGFs under non-inflammatory or inflammatory condition**

(A and C) Reverse transcription-polymerase chain reaction (RT-PCR)-based gene expression analysis of *PTGS2* and *IL1B* relative to *GAPDH* in hGF-F1 cells cultured on the 0.1wt% collagen-coated polystyrene culture plate and soft, mid, and hard PDMS for 12 h with or without co-incubation with 1,000 ng/mL of LPS. (B and D) PGE<sub>2</sub> levels per unit DNA in adherent hGF-F1 cells under the corresponding culture conditions described above by enzyme-linked immunosorbent assay (ELISA) analysis of culture supernatants. Data are represented as the mean  $\pm$  standard deviation (SD;  $N = 3$ ). Different letters indicate the statistically significant differences between them ( $P < 0.05$ ; Tukey's honest significant difference [HSD] test). hGFs, Human gingival fibroblasts; LPS, Lipopolysaccharide; PDMS, Polydimethylsiloxane; PGE<sub>2</sub>, Prostaglandin E<sub>2</sub>; *PTGS2*, Prostaglandin G/H synthase 2; *IL1B*, Interleukin-1 $\beta$ ; *IL6*, Interleukin-6; *GAPDH*, Glyceraldehyde-3-phosphate dehydrogenase.



**Supplementary Figure 3. Optimization for the concentration of LPS and mechanotransduction inhibitors in hGFs**

(A) Representative phase microscopic images and (B) reverse transcription-polymerase chain reaction (RT-PCR)-based gene expression analysis of *PTGS2* relative to *GAPDH* in hGF-M2 cells cultured on a polystyrene culture plate after co-incubation with 0, 10, 100, and 1,000 ng/mL of LPS for 12 h. (C and D) Representative light microscopic images after methylene blue staining (upper images) and

immunofluorescence confocal laser microscopic images of YAP (green), *F*-actin (red), and/or nucleus (blue) (mid or bottom images) in hGF-M2 cells cultured on 0.1wt% collagen-coated polystyrene or hard PDMS for a total of 24 h, including 2 h of co-incubation with 0, 5, and 10  $\mu$ M Y-27632 (**C**) or 0, 10, 30, and 50  $\mu$ M blebbistatin (**D**). Note that 1000 ng/mL LPS induced the highest *PTGS2* expression in the hGF cell culture (**B**) without loss of cell attachment (**A**) and immunofluorescence confocal laser microscopic images show less or reduced YAP signals in the nucleus of hGF-M2 cells on both substrates after co-incubation with 5  $\mu$ M or more Y-27632 (**C**, arrows) or 30  $\mu$ M or more blebbistatin (**D**, arrows), respectively. Data are represented as the mean  $\pm$  standard deviation (SD;  $N = 3$ ). Different letters indicate the statistically significant differences between them ( $P < 0.05$ ; Tukey's honest significant difference [HSD] test). hGFs, Human gingival fibroblasts; PDMS, Polydimethylsiloxane; LPS, Lipopolysaccharide; *PTGS2*, Prostaglandin G/H synthase 2; GAPDH, Glyceraldehyde-3-phosphate dehydrogenase; ROCK, Rho-associated coiled-coil containing protein kinase; YAP, Yes-associated protein.

Encoded protein name (gene name)	Primers (Fw, forward; Rv, reverse)	Product size (bp)	Accession number
<b>Human</b>			
Prostaglandin G/H synthase 2 ( <i>PTGS2</i> )	Fw: 5'-TCCAGTACCAAAATCGTATTGCT-3' Rv: 5'-AGTGCTTCCAACCTGCAGACAT-3'	370	NM_000963.4
Interleukin-1 $\beta$ ( <i>IL1B</i> )	Fw: 5'-TGGAGCAACAAGTGGTGT-3' Rv: 5'-TTGGGATCTACACTCTCCAGC-3'	157	NM_000576.3
Interleukin-6 ( <i>IL6</i> )	Fw: 5'-TCAATGAGGAGACTTGCCTG-3' Rv: 5'-GATGAGTTGTCATGTCCTGC-3'	261	NM_001371096.1
Nitric oxide synthase 2 ( <i>NOS2</i> )	Fw: 5'-CAGCGGGATGACTTTCCAA-3' Rv: 5'-AGGCAAGATTGGACCTGCA-3'	75	NM_000625.4
Tumor Necrosis Factor- $\alpha$ ( <i>TNFA</i> )	Fw: 5'-GAGGCCAAGCCCTGGTATG-3' Rv: 5'-CGGGCCGATTGATCTCAGC-3'	91	NM_000594.4
Collagen Type 1 Alpha 1 ( <i>COL1A1</i> )	Fw: 5'-AGGCGAACAGGGCGACAGAG-3' Rv: 5'-GGCCAGGGAGACCGTTGAGT-3'	185	NM_000088.4
Elastin ( <i>ELN</i> )	Fw: 5'- AACCAGCCTTGCCCGC-3' Rv: 5'- CCCC AAGCTGCC TGGTG-3'	101	NM_001278939.2
Fibrillin 1 ( <i>FBN1</i> )	Fw: 5'-GAATGCAAGAACCTCATTGGCAC-3' Rv: 5'-TGGCGGTAAACCCATCATTACAC-3'	184	NM_000138.5
Lysyl Oxidase ( <i>LOX</i> )	Fw: 5'-AGCATA CAGGGCAGATGTCAGAG-3' Rv: 5'-CTTGGTCGGCTGGGTAAGAAAT-3'	105	NM_002317.7
Paxillin ( <i>PXN</i> )	Fw: 5'-CCCTGACGAAAGAGAAGCCTAAG-3' Rv: 5'-AGATGCGTGTCTGCTGTTGG-3'	186	NM_001385988.1
Integrin $\beta$ -1 ( <i>ITGB1</i> )	Fw: 5'-TTTGTTTAATGTCTGGTCTTTCTG-3' Rv: 5'-CCCCAAAATTGCAAACAAATACA-3'	69	NM_133376.3
Integrin $\alpha$ -11 ( <i>ITGA11</i> )	Fw: 5'-CAGCTCGCTGGAGAGATACG-3' Rv: 5'-TTACAGGACGTGTTTCGCCTC-3'	186	NM_001004439.2
Glyceraldehyde 3 Phosphate Dehydrogenase ( <i>GAPDH</i> )	Fw: 5'-AATCCCATCACCATCTTCCA-3' Rv: 5'-TGGACTCCACGACGTAACA-3'	82	NM_001357943.2

**Supplementary Table 1.** List of primers for human genes used in SYBR-green-based polymerase chain reaction.