Supplementary information

Structural insights into the elevator-type transport mechanism of a bacterial ZIP metal transporter

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Figure S1. **Analysis of the N terminal sequence of BbZIP. a** Sequence alignment of the N terminal sequences of selected ZIPs. BbZIP, RMF21814.1 [*Cyanobacteria bacterium J083*], NLY76259.1 [*Firmicutes bacterium*], WP_133483791.1 [*Halomonas ventosae*], WP_062679913.1 [*Achromobacter denitrificans*], WP_014755437.1 [*Pseudomonas*], WP_193090314.1 [*Advenella sp. FME57*], WP_042731040.1 [*Pseudomonas*], and human ZIP11 (Q8N1S5.3) are within the gufA subfamily. ZupT from *Escherichia coli* is a representative member of the ZupT subfamily. The amphipathic helix (α 0a) and α 0 of BbZIP are highlighted in grey and cyan, respectively. The predicted α 0 of the other ZIPs, if there is any, are also highlighted. Note that some ZIPs in the gufA subfamily do not have α 0 or α 0a. **b** Prediction of transmembrane helices of BbZIP by TMHMM

(<u>https://services.healthtech.dtu.dk/service.php?TMHMM-2.0</u>). **c** Prediction of signal peptide of BbZIP by SignalP (<u>https://services.healthtech.dtu.dk/service.php?SignalP-5.0</u>).



Figure S2. **Structure of the loop connecting \alpha7 and \alpha8.** In the apo state structure (*left*), the residues missed in the previous structure (*right*) form a broken helix and a short loop. The conserved proline residue (P279) causes a kink in α 7.



Figure S3. **BbZIP dimer predicted by AlphaFold in top view (***left***) and side view (***right***). The structure was predicted by AlphaFold Colab.**



Figure S4. **Additional evidence supporting the proposed OFC model. a** Cysteine accessibility assay. Selected single cysteine variants were purified in DDM and treated with mPEG5K. EDTA was added to the sample immediately before the treatment to prevent cysteine blockage by Cd²⁺. The reaction was terminated by 100 mM water soluble thiol reacting reagent methyl methanethiosulfonate before analysis in SDS-PAGE. The upper band indicated by the arrow is the PEGylated protein. **b** Hg-mediated chemical crosslinking of the A95C/L217C variant in the native membrane. The membrane fraction of the cells expression the variant was incubated with HgCl₂, terminated by NEM, and applied to Western blot by using a custom monoclonal antibody against BbZIP generated by Creative Biolabs Inc. The arrow indicates the crosslinked product. **c** Size-exclusion chromatography of the Hg-crosslinked of A95C/L217C variant. The eluted sample was applied to SDS-PAGE, and the band shift indicates that the protein is still in the crosslinked state after removal of free Hg²⁺ from the sample. Source data are provided as a Source Data file.



Figure S5. **Mapping of small residues at the interface between the transport domain and the scaffold domain.** The small residues (Gly, Ala, Ser) at the domain interface are colored in red (highly conserved) or yellow (less or non-conserved). The residues in the BMC (transport site) are shown in stick mode.



Figure S6. **Comparison of the BbZIP structures with the structures of human ZIPs predicted by AlphaFold**. All structures were retrieved from the AlphaFold Protein Structure Database (https://alphafold.ebi.ac.uk/). The scaffold domains ($\alpha 2/3/7/8$) of the ZIPs are structurally aligned. For clarity, the extracellular domains and the cytosolic loop between $\alpha 3$ and $\alpha 4$ are not shown. To better distinguish conformational states, the structures are plotted against the distance between a residue at the pore entrance (S106 in BbZIP) and the last residue of the metal chelating motif in $\alpha 5$ (E211 in BbZIP). S106 and E211 are labeled in the BbZIP structure in the apo state (right upper corner) with the scaffold domain colored in green and transport domain in blue. The residue pairs in other ZIPs for distance measurement are highlighted in the sequence alignment and indicated with asterisks. Note that $\alpha 3$ of ZIP8 and ZIP14 is predicted to be adjacent to $\alpha 8$, which is the position for $\alpha 3$ of the other protomer when the transporter forms a homodimer as shown in Figure 5B and Figure S3.



Figure S7. **A putative cytoplasmic gate in the OFC.** *Left*: IFC (PDB: 5TSA). *Right*: the OFC model. The residues involved in gate formation are labeled and shown in stick mode and. Zinc ions in the IFC are depicted as grey spheres. The hydrogen bonds are shown as red dashed lines. The transport domain and the scaffold domain are colored in blue and green, respectively.



Figure S8. Functional analysis of the ZIP4 variants with cysteine substitutions. A portion of cysteine substitutions in cysteine accessibility assay and crosslinking experiments was conducted on conserved residues of BbZIP, including A95, A184, A203, and A214. The corresponding residues in human ZIP4 were replaced with cysteine and the result variants were subjected to zinc transport assay. The relative transport activity of each variant is expressed as the percentage of the activity of the wild type ZIP4. The activity has been calibrated using the expression level estimated by Western blot, which is expressed as the normalized ratio of band intensity of HA-tagged ZIP4 to β -actin. The shown data are from one representative experiment and 2-3 independent experiments with similar results were conducted for each variant. Three biological replicates were included in one experiment. The horizontal bar of the scatter dot plot represents the mean and the vertical bar indicates the standard deviation. The asterisks indicate the significant differences between the variants and the wild type ZIP4 (two-sided Student's ttests: ** P≤0.01). The exact P values are 0.0016, 0.0086, 0.0019, and 0.006 for the variants of A368C, A514C, A532C, and G543C, respectively. The other residues subjected to cysteine substitution are either highly variable or have been functionally characterized in the previous reports. For instance, the functional study of H536 in human ZIP4, which is topologically equivalent to Q207 in BbZIP, has been reported in Ref 46. Source data are provided as a Source Data file.





PDB: 7Z6N





Figure S9. Comparison of the structure solved in this work (PDB entry ID: 8CZJ) and the reported BbZIP structure in metal free state (PDB entry ID: 7Z6N). a Structural comparison after alignment of the scaffold domains. The scaffold domain, the transport domain, and the N-

terminal domain of 8CZJ are colored in green, blue and pink, respectively. 7Z6N (chain B) is shown in grey. Note that the orientation of the transport domains relative to the scaffold domain in the two structures are different with the cytoplasmic side of α 4 being the most variable region as indicated by the arrow. **b** 2FoFc electron density maps (σ =1) of 8CZJ (2.75 Å) and 7Z6N (2.6 Å). The corresponding domains are structurally aligned with labeled structural elements. The density map of the N-terminal domain of 8CZJ, including the transmembrane helix α 0 and amphipathic helix α 0a, is also shown. The image of 7Z6N is adapted from Fig. S1B of Ref 82. **c** Comparison of crystal packing of 8CZJ with 7Z6N. The crystallographic dimer in one asymmetry unit of 8CZJ is colored and the symmetry mates are in grey. The image of 7Z6N in frame is adapted from Fig. S4 of Ref 82. **d** Association of α 0a with α 3 and α 4 through hydrogen bonds (red dashed lines) and hydrophobic interactions, respectively. Hypothetically, α 0a may limit the elevator-like movement of the transport domain (blue) relative to the scaffold domain (green) and therefore function as a negative regulator.



Figure S10. The cryo-EM data processing workflow for BbZIP in Amphipol.



Figure S11. **Sequence alignment for repeat-swap homology modeling. a** Scheme of sequence alignment of the template (BbZIP) and the model. **b** Sequence alignment of the template and the model.



Figure S12. Sensitivity and specificity of the custom monoclonal antibody against BbZIP. The whole cell lysates of the cells transformed with the empty vector or the plasmid containing His₆-tagged BbZIP were applied to Western blot. The His₆-tagged BbZIP was detected with either an anti-His antibody (0.1 μ g/ml) or the anti-BbZIP antibody (0.06 μ g/ml). The result shows a better sensitivity and specificity of the anti-BbZIP antibody than the anti-His antibody. The shown data are from a single experiment. Source data are provided as a Source Data file.

Crystal	Apo BbZIP
Data collection	
Beamline	GM/CA-CAT (23-ID-D)
Wavelength (Å)	1.0331
Space group	P 2 ₁
Unit cell	
a, b, c (Å)	63.0, 117.7, 64.3
α, β, γ (°)	90, 104.2, 90
^a Resolution (Å)	37.1 - 2.75 (2.85 - 2.75)
aRedundancy	7.4 (2.6)
^a Completeness (%)	94.5 (74.8)
^a I/σI	7.8 (0.6)
$^{\mathrm{a,b}}R_{merge}$	0.187 (0.968)
^{a,c} R _{pim}	0.068 (0.57)
$^{d}CC_{1/2}$ of the highest resolution shell	0.491
Refinement	
Unique reflections	22227
Number of Atoms	4289
Protein	4019
Ligands	263
H_2O	7
^e R _{work} /R _{free}	0.233/0.261
<i>Wilson B-factor</i> (Å ²)	63.1
<i>B</i> -factors (Å ²)	
Protein	54.8
MPG	55.5
SO_4^{2-}	78.2
H_2O	52.3
R.m.s. deviations	
Bond lengths (Å)	0.010
Bond angles (°)	1.23
Ramachandran plot (%)	
Favored	97.3
Allowed	2.7
Outliers	0.0

 Table S1. Crystallographic statistics

^aHighest resolution shell is shown in parentheses.

 ${}^{b}R_{merge} = \sum_{hkl} \sum_{j} |I_{j}(hkl) - \langle I(hkl) \rangle | / \sum_{hkl} \sum_{j} I_{j}(hkl)$, where *I* is the intensity of reflection.

 ${}^{c}R_{pim} = \sum_{hkl} [1/(N-1)]^{1/2} \sum_{j} |I_{j}(hkl) - \langle I(hkl) \rangle| / \sum_{hkl} \sum_{j} I_{j}(hkl)$, where N is the redundancy of the dataset. ${}^{d}CC_{1/2}$ is the correlation coefficient of the half datasets.

 ${}^{e}R_{work} = \sum_{hkl} ||F_{obs}| - |F_{calc}|| / \sum_{hkl} |F_{obs}|$, where F_{obs} and F_{calc} is the observed and the calculated structure factor, respectively. R_{free} is the cross-validation R factor for the test set of reflections (5% of the total) omitted in model refinement.

i	A_i	j	A_j	Probability ^a	ТМ	ТМ	# of interhelical interactions
58	Y	189	F	0.848799	1	4	3
62	G	189	F	0.998361	1	4	
66	G	185	Ι	0.815644	1	4	
58	Y	194	L	0.825278	1	5	14
62	G	201	Т	0.988765	1	5	
65	А	205	А	0.850994	1	5	
65	А	201	Т	0.801456	1	5	
69	А	205	А	0.99679	1	5	
69	А	209	V	0.96778	1	5	
72	L	209	V	0.941264	1	5	
73	G	212	G	0.77212	1	5	
74	А	216	А	0.910318	1	5	
76	М	209	V	0.951789	1	5	
76	М	213	L	0.769472	1	5	
77	A	216	Α	0.822501	1	5	
78	L	216	A	0.870321	1	5	
78	L	220	R	0.784763	1	5	· · ·
59	A	244	A	0.931931	1	6	14
59	A	248	V	0.883283	1	6	
60	V	245	L	0.850106	1	6	
63	G	244	A	0.999786	1	6	
64	A	241	Р	0.988794	1	6	
70	T	236	S	0.772531	1	6	
70	Т	237	G	0.761363	1	6	
71	A	234	V	0.999287	1	6	
71	A	237	G	0.885252	1	6	
74	A	233	A	0.93/339	1	6	
75	L	230	V	0.952012	1	6	
78	L	227	G	0.925817	1	6	
78	L	226	l	0.845856	1	6	
/8	L	230	V	0.841137	1	6	10
104	A	134	G	0.855523	2	3	10
108	I T	133	L	0.998451	2	3	
108	I C	130	A	0.778985	2	3	
111	U I	129	V A	0.989027	2	3	
112	L	130	A	0.998130	2	3	
112		129	V	0.981881	$\frac{2}{2}$	3	
115	A A	129	V	0.94178	2	3	
115	Δ	132	Ğ	0.937740	2	3	
115	G	127	4	0.832135	2	3	
117	Т	127	A A	0.002100	2	3	
95	Δ	218	A	0.783777	2	5	6
99	M	210	P	0.799275	2	5	Ũ
102	A	207	0	0.930451	2	5	
102	A	200	Ľ	0.877758	2	5	
102	A	203	Ā	0.82897	2	5	
106	S	207	0	0.821258	2	5	
99	M	269	Ň	0.981702	2	7	28
99	М	273	V	0.932705	2	7	
100	L	273	V	0.998426	2	7	
100	L	274	S	0.983822	2	7	
100	L	270	Ι	0.867449	2	7	
103	S	270	Ι	0.996602	2	7	
103	S	269	М	0.993979	2	7	
103	S	273	V	0.965434	2	7	
103	S	266	А	0.942894	2	7	
104	А	270	Ι	0.987026	2	7	
107	L	262	М	0.985902	2	7	
107	L	266	А	0.968643	2	7	
108	Ι	266	А	0.970083	2	7	
111	G	263	G	0.999787	2	7	
111	G	259	Р	0.997588	2	7	
111	G	266	А	0.989192	2	7	
114	А	259	Р	0.979581	2	7	

 Table S2. Predicted contacting residues from different transmembrane helices by EVcouplings.

115	А	259	Р	0.988483	2	7	
85	Δ	284	N	0 98924	2	7	
00	0	201	E	0.762208	2	7	
00	Q	280		0.702208	2	/	
89	D	284	N	0.921114	2	7	
89	D	281	Т	0.917349	2	7	
89	D	283	R	0.785205	2	7	
02	I	200	V	0.844068	2	7	
92	L	2//	v	0.044000	2	7	
92	L	281	1	0.822336	2	1	
93	G	281	Т	0.828245	2	7	
96	А	277	V	0.980601	2	7	
96	Δ	278	Ť	0.070021	2	7	
90	A D	270	T	0.979921	2	7	20
80	K	289	1	0.985952	2	8	20
89	D	290	Α	0.836692	2	8	
90	А	293	G	0.999999	2	8	
90	А	289	Т	0 998718	2	8	
03	G	203	Ġ	0.030178	2	õ	
93	U C	293	U .	0.939178	2	0	
93	G	290	A	0.926867	2	8	
93	G	294	L	0.90149	2	8	
94	F	300	L	0.999995	2	8	
94	F	296	Δ	0 954435	2	8	
04	F	207	C	0.706229	2	0	
94	г	297	G	0.790258	2	0	
94	F	293	G	0.7/1431	2	8	
96	А	294	L	0.995867	2	8	
98	М	301	М	0.931046	2	8	
100	T	294	T	0.965934	2	8	
100		200	L L	0.001004	2	0	
101	A	298	F	0.981904	2	8	
101	A	301	М	0.96157	2	8	
101	Α	302	М	0.812075	2	8	
102	А	301	М	0.84909	2	8	
102	E	202	M	0.071876	2	Ŷ	
105	r C	302	IVI	0.9/10/0	2	0	
106	8	301	M	0.93802	2	8	
128	V	260	I	0.940619	3	7	23
128	V	256	L	0.876523	3	7	
129	V	263	G	0.993559	3	7	
120	v	200	D	0.806656	2	7	
129	v	239	r T	0.800030	3	7	
131	L	260	1	0.76513	3	/	
132	G	260	I	0.999996	3	7	
132	G	263	G	0.997772	3	7	
132	G	264	Τ.	0 925151	3	7	
132	T	270	Ĩ	0.008646	3	7	
133	L	270	1	0.998040	5	7	
133	L	266	A	0.980342	3	1	
133	L	263	G	0.934101	3	7	
135	L	260	Ι	0.912802	3	7	
135	T	264	T	0 818489	3	7	
126	C	264	T	0.010407	2	7	
150	U U	204	L	0.994043	5	/	
137	V	274	S	0.999788	3	7	
137	V	275	Н	0.913608	3	7	
137	V	271	F	0.802545	3	7	
137	V	267	G	0 798816	3	7	
137	v T	207	U I	0.790010	2	7	
139	L	264	L	0.999984	3	/	
140	M	268	A	0.991156	3	7	
140	М	271	F	0.870281	3	7	
141	L	271	F	0.946312	3	7	
144	D D	271	F	0.073/17	3	7	
144	D	2/1	Г	0.9/341/	3	/	7
130	A	299	A	0.9/4493	3	8	/
134	G	299	Α	0.976559	3	8	
134	G	298	F	0.907086	3	8	
137	V	295	М	0 995828	3	8	
129	, T	204	T	0.012608	2	Ŷ	
156	L	294	L	0.912008	5	0	
138	L	298	F	0.801334	3	8	
142	G	291	Т	0.958239	3	8	
167	V	222	V	0.969496	4	5	20
170	F	215	V	0.940085	4	5	
171	v	215	V	0 877607	1	5	
171	V	215	v	0.077097	4	5	
1/1	V	219	L	0.802852	4	5	
174	Ι	212	G	0.932936	4	5	
174	Ι	211	Е	0.840814	4	5	
174	I	208	D	0 77545	4	5	
177	LI LI	200	0	0.001014	4	5	
1//	H	207	Q	0.991914	4	2	
1//	H	208	D	0.750139	4	5	

17	'8 N	208	D	0.910642	4	5		
17	'8 N	212	G	0.888044	4	5		
18	1 E	208	D	0.983828	4	5		
18	1 E	204	Ι	0.90104	4	5		
18	1 E	211	Е	0.834965	4	5		
18	5 I	204	Ι	0.980623	4	5		
18	5 I	201	Т	0.959707	4	5		
18	8 S	200	L	0.996096	4	5		
18	8 S	197	G	0.918298	4	5		
18	8 S	196	Ι	0.910892	4	5		
18	9 F	197	G	0.976066	4	5		
17	'1 V	232	V	0.990855	4	6	13	
17	'5 I	239	Μ	0.999986	4	6		
17	'5 I	236	S	0.843348	4	6		
17	'8 N	240	E	0.885684	4	6		
17	'8 N	236	S	0.885544	4	6		
17	'8 N	239	Μ	0.824308	4	6		
17	9 L	243	G	0.999867	4	6		
17	9 L	246	V	0.987151	4	6		
18	1 E	240	E	0.762046	4	6		
18	3 M	247	G	0.999611	4	6		
18	3 M	243	G	0.759774	4	6		
18	6 G	247	G	0.944232	4	6		
18	6 G	248	V	0.759108	4	6		
17	73 T	272	V	0.775614	4	7	5	
18	3 M	261	S	0.763568	4	7		
18	4 A	265	А	0.783626	4	7		
18	6 G	265	А	0.842456	4	7		
18	8 S	262	Μ	0.759294	4	7		
20	8 D	240	E	0.922795	5	6	10	
20	08 D	236	S	0.88534	5	6		
21	2 G	233	А	0.940804	5	6		
21	2 G	236	S	0.784396	5	6		
21	5 V	236	S	0.950061	5	6		
21	6 A	233	А	0.936749	5	6		
21	6 A	229	А	0.854505	5	6		
21	9 L	229	А	0.895863	5	6		
22	0 R	226	Ι	0.999554	5	6		
22	0 R	229	А	0.989852	5	6		
19	7 G	262	М	0.759117	5	7	3	
20	8 D	269	М	0.83822	5	7		
21	1 E	269	М	0.924349	5	7		
27	4 S	294	L	0.976348	7	8	2	
27	5 Н	288	Т	0.817171	7	8		

^a Only the contacting residue pairs with probability greater than 75% are listed.

 Table S3. Primers for mutagenesis in this work.

Uncropped gels/Western blots

(the portions in the dashed frames are shown in Figures)

Fig. 5c



Primary: anti-Histag antibody at 1:5000 from Invitrogen, Product # 37-2900

Fig. 5d



Primary: Customized anti-BbZIP monoclonal antibody at 0.06 μ g/ml (generated by Creative Biolabs Inc.)

Fig. 7a



Primary: anti-Histag antibody at 1:5000 from Invitrogen, Product # 37-2900





Primary: anti-Histag antibody at 1:5000 from Invitrogen, Product # 37-2900









Lane1: marker and vector Lane2: ZIP4 Lane3: 368V Lane4: 368F Lane5: 514V Lane6: 514F Lane7: 532V Lane8: 532F Lane9: 543V Lane10: 543F

ZIP4-HA (upper)

Primary: anti-HA antibody at 1:5000 from Invitrogen, Product # 26183

Secondary: HRP-conjugated Horse anti-mouse IgG at 1:5000 from Cell Signaling Technology, Product # 7076S

β-actin (lower)

Primary: β-actin (13E5) Rabbit mAb at 1:5000 from Cell Signaling Technology, Product #4970S

Supplementary Fig. 4



Primary: anti-Histag antibody at 1:5000 from Invitrogen, Product # 37-2900

Supplementary Fig. 8



Lane1: marker Lane2: vector Lane3: ZIP4 Lane4: 368C Lane5: 514C Lane6: 532C Lane7: 543C Lane8: 546C

ZIP4-HA (upper)

Primary: anti-HA antibody at 1:5000 from Invitrogen, Product # 26183

Secondary: HRP-conjugated Horse anti-mouse IgG at 1:5000 from Cell Signaling Technology, Product # 7076S

β-actin (lower)

Primary: β-Actin (13E5) Rabbit mAb at 1:5000 from Cell Signaling Technology, Product #4970S

Supplementary Fig. 12



Primary: anti-Histag antibody at 0.1 µg/ml from Invitrogen, Product # 37-2900

Secondary: HRP-conjugated Horse anti-mouse IgG at 1:5000 from Cell Signaling Technology, Product # 7076S



Primary: Customized anti-BbZIP monoclonal antibody at 0.06 μ g/ml (generated by Creative Biolabs Inc.)