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Supplemental information

BRPF1 bridges H3K4me3 and H3K23ac

in human embryonic stem cells

and is essential to pluripotency

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Figure S1. BRPF1 is an indispensable gene for the pluripotency and self-renewal of hESCs, related to Figure 1.

H1-BRPF1*2#

H1-BRPF1-1#

41.2

HA.

(A) Overview of the BRPF1 targeting strategy. sgRNA was designed, and a homologous targeting vector containing a puromycin resistance cassette was constructed according to the BRPF1 gene. (B) PCR identification of WT and two BRPF1+ H1 hESC clones. WT hESCs served as a negative control. (C) Karyotype in BRPF1+ H1 hESCs. (D) Immunostaining of the pluripotency and lineage markers OCT4 (pluripotency), GFAP (ectoderm), CALPONIN (mesoderm), and FOXA2 (endoderm) in WT and two BRPF1+ H1 hESC clones. Scale bar, 50 µm. (E) EdU incorporation assay in WT and two BRPF1+ H1 hESC clones. The significance level was determined by unpaired two-tailed Student's t tests. **, P < 0.01. The data represent the mean ± SD (standard deviation) from three independent repeats (n=3). (F) Heatmap and GO analysis of the top 100 genes up- or down-regulated in two BRPF1+ H1 hESC clones compared with WT H1 hESCs. All error bars throughout the figure represent the SD (standard deviation) from three independent replicates (n = 3).



Figure S2. Inducible expression of BRPF1 complements the BRPF1^{-/-} hESCs phenotype, related to Figure 2.

(A) Diagram of the lentiviral-based inducible system for BRPF1 expression (H1-BRPF1-OE). BRPF1 expression was controlled by DOX treatment. (B) Schematic of BRPF1 knockout in H1 hESCs with BRPF1 over expression (KO-OE-BRPF1). (C) The morphology of BRPF1 $^{+}$ H1 hESCs with BRPF1 over-expression (+DOX) or no (-DOX) was shown. Scale bar, 200 µm. The expression of BRPF1 was examined in the indicated cells by qRT-PCR. The significance level was determined by unpaired two-tailed Student's t tests. **, P < 0.01. The data represent the mean ± SD (standard deviation) from three independent repeats (n=3). (D) Examination of the expression of the selected pluripotent and lineage genes in the indicated cells by qRT-PCR. The significance level was determined by unpaired two-tailed Student's t tests. **, P < 0.01. The data represent the mean ± SD (standard deviation) from three independent repeats (n=3). (D) Examination of the expression of the selected pluripotent and lineage genes in the indicated cells by qRT-PCR. The significance level was determined by unpaired two-tailed Student's t tests. **, P < 0.01. The data represent the mean ± SD (standard deviation) from three independent repeats (n=3). (E) Western blot of NANOG, BRPF1 (FLAG), H3K14ac or H3K23ac in the indicated cells. All error bars throughout the figure represent the SD (standard deviation) from three independent replicates (n = 3).



Figure S3. Knock in triple-FLAG into BRPF1 locus in hESCs, Related to Figure 4.

(A) Overview of the BRPF1 locus knock-in (KI) triple-FLAG strategy. (B) Morphology of BRPF1-KI H1 hESCs. Scale bar, 200 µm. (C) PCR identification in WT and two BRPF1-KI H1 hESC clones. WT hESCs served as a negative control. (D) GAPDH protein and FLAG tag were examined in WT and two BRPF1-KI H1 hESC clones by western blot. (E-F) Examination of the expression of the selected (E) pluripotent and (F) lineage genes in WT and two BRPF1-KI H1 hESC clones.



Figure S4. Similar localization of H3K4me3 between WT and BRPF1^{+/-} hESCs, Related to Figure 4.

(A) Heatmap and signal densities of H3K4me3 associated genes in KI and BRPF1^{-/-} hESCs from ChIP-seq. (B) Venn diagram for H3K4me3 associated genes in the indicated cells. (C) Enrichment of H3K4me3 associated with different regions. "C-O" represents regions of close to open; "O-C" represents regions of open to close; "O-C of stemness genes" represents regions of open to close in stemness genes. (D) Genomic views of H3K4me3 enrichment and ATAC-seq analysis in the indicated WT and BRPF1^{-/-} hESCs.