

## Supporting Information

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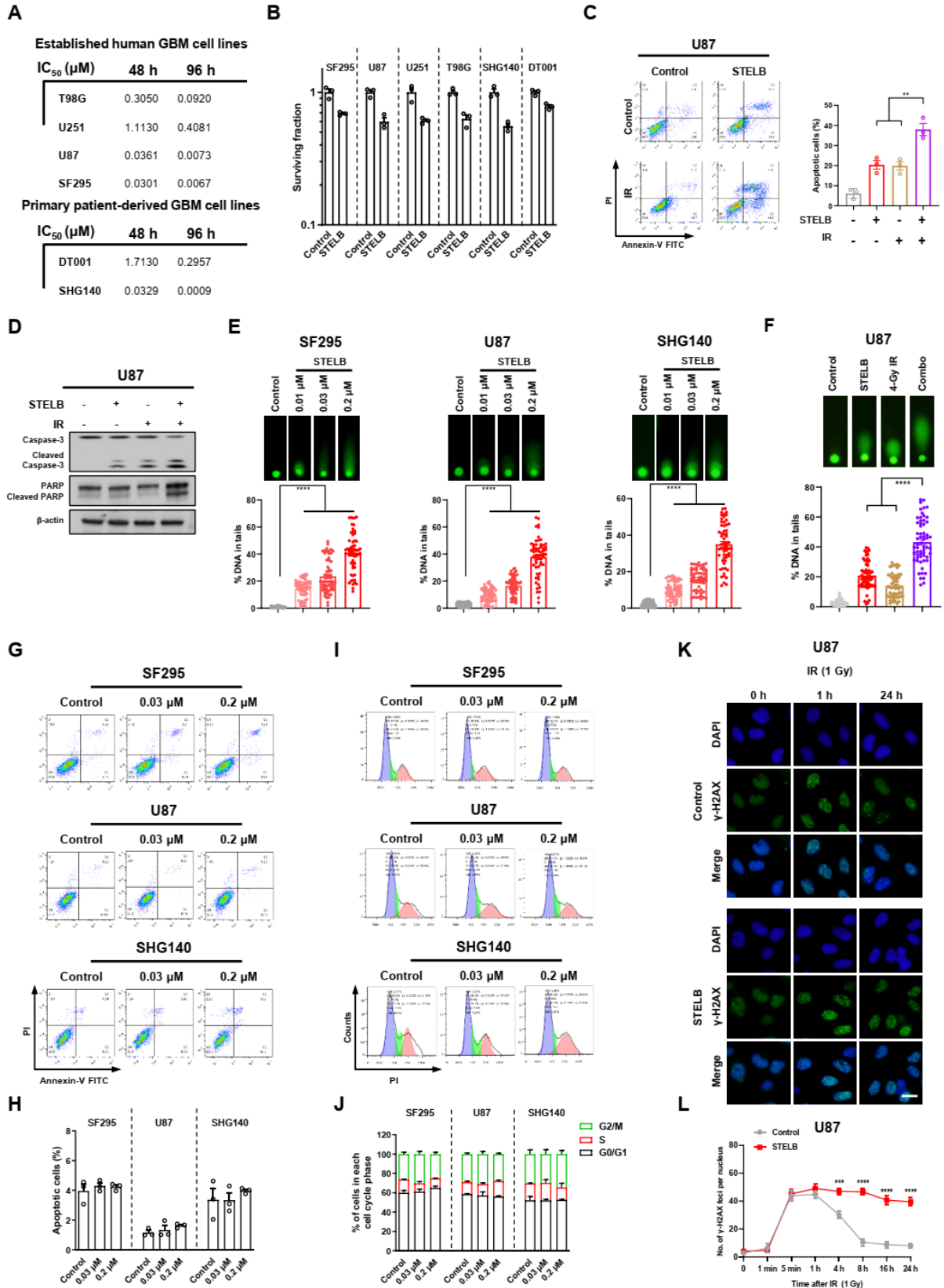
Stelletin B Sensitizes Glioblastoma to DNA-Damaging Treatments by Suppressing PI3K-Mediated Homologous Recombination Repair

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### List of Items in Supporting Information

<b>Figure S1</b>	<b>STELB inhibits GBM cell proliferation and induces DNA damage</b>
<b>Figure S2</b>	<b>STELB sensitizes MGMT-negative GBM cells to TMZ</b>
<b>Figure S3</b>	<b>Effects of STELB on HR and NHEJ repair in GBM cells</b>
<b>Figure S4</b>	<b>Effects of STELB on PI3K activity and PI3K protein expression in GBM cells</b>
<b>Figure S5</b>	<b>Effects of genetic and pharmacological inhibition of PI3K on the expression of HR repair key factors BRCA1/2 and RAD51</b>
<b>Figure S6</b>	<b>Effects of STELB on mRNA expression of PI3K isoforms and the stability of exogenous PI3K<math>\alpha</math></b>
<b>Figure S7</b>	<b>Effects of genetic and pharmacological inhibition of PI3K on HR repair efficiency</b>
<b>Figure S8</b>	<b>Effects of STELB on NHEJ-related proteins in vivo</b>
<b>Table S1</b>	<b>The primer sequences used in this study</b>

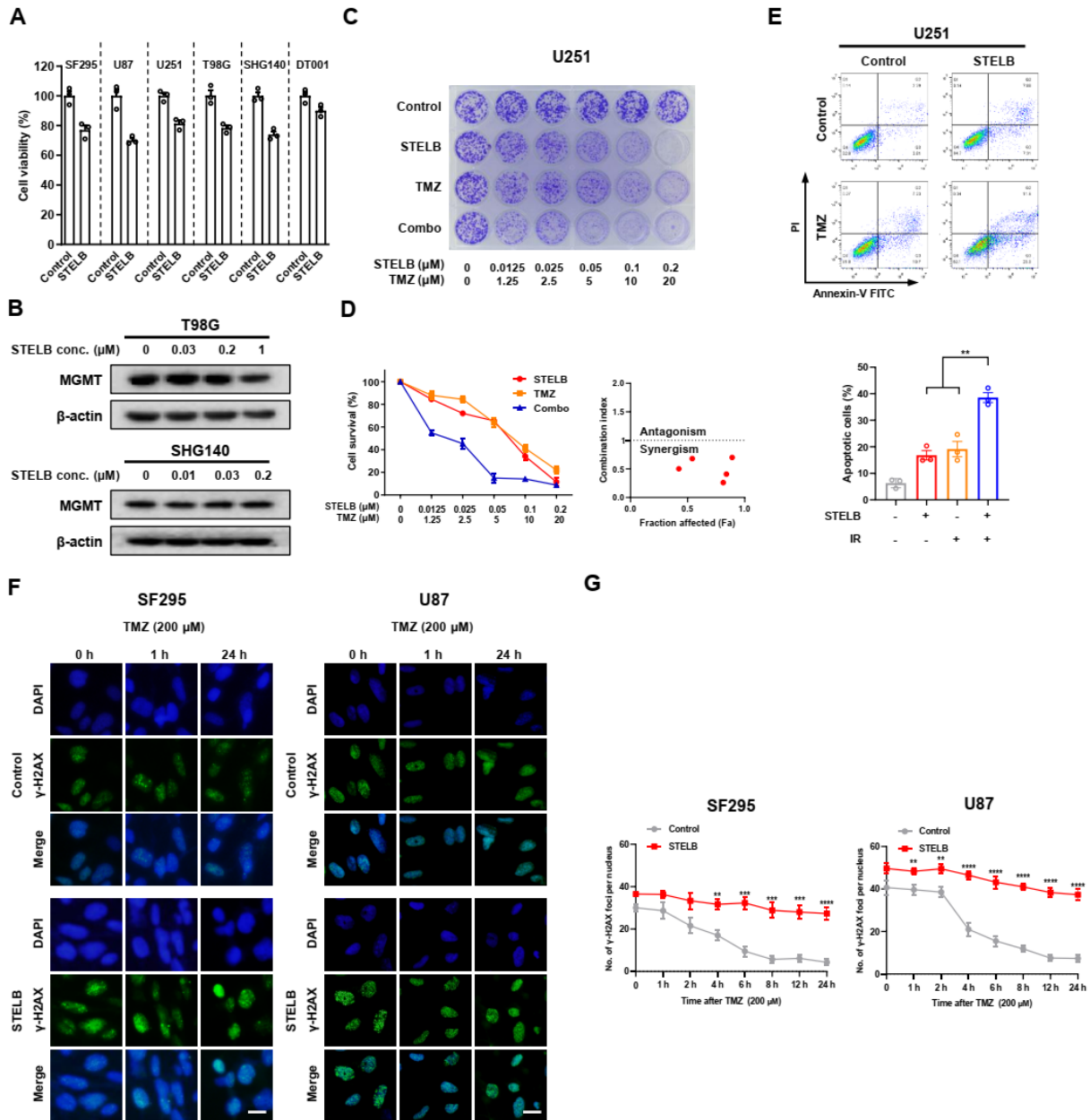
Supplementary Figure 1



**Figure S1. STELB inhibits GBM cell proliferation and induces DNA damage**

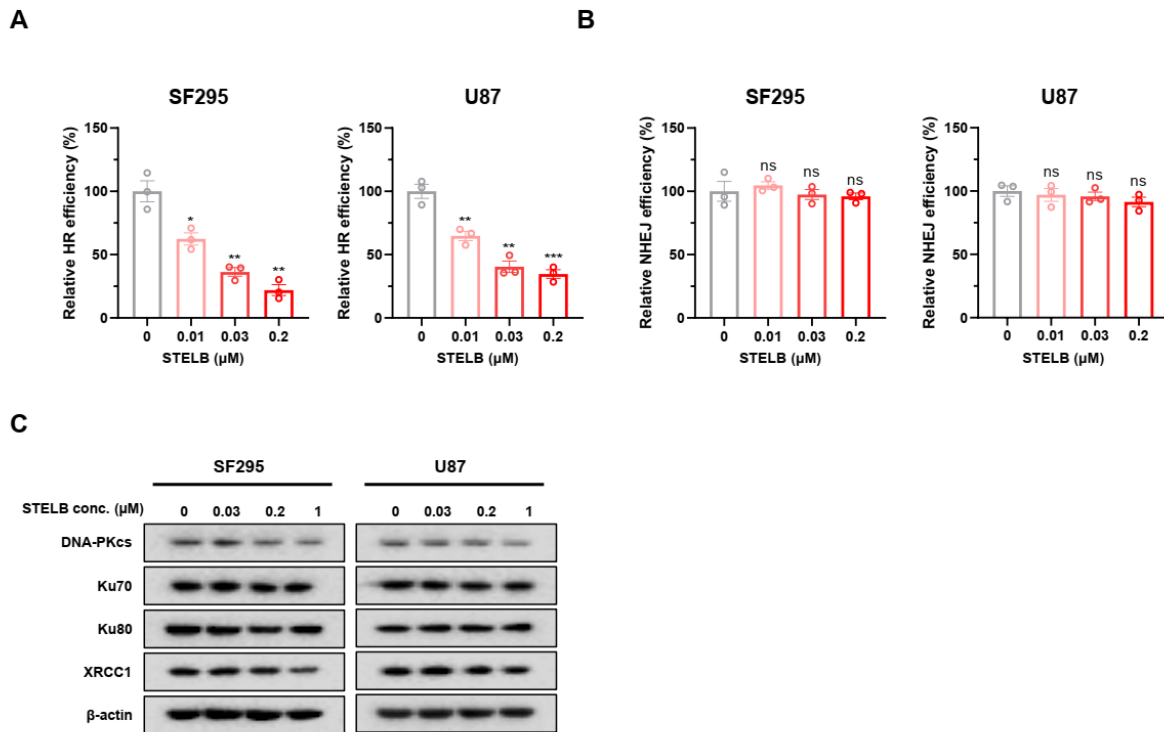
(A) IC<sub>50</sub> for established GBM cell lines and primary patient-derived GBM cell lines treated with increasing concentrations of STELB for 48 h or 96 h. (B) Clonogenic survival assay for GBM cells with or without pretreatment of STELB. (C) Apoptosis assays of U87 cells pretreated with STELB for 24 h and then irradiated at 4 Gy. Apoptosis was detected by Annexin V/PI staining after 72 h. (D) Western blot analysis of Caspase-3 and PARP in U87 cells after treatment with STELB and/or IR for 48 h. (E) Alkaline comet assays of U87 cells treated with different concentrations of STELB for 48 h to measure both single and double stranded DNA breaks. The selected doses are the IC<sub>50</sub> value of STELB in the three cell lines for 48 h, a dose below the IC<sub>50</sub> value, and a higher dose above the IC<sub>50</sub> value. (F) Alkaline comet assays of U87 cells treated with STELB and/or IR for 48 h to measure both single and double stranded DNA breaks. The % DNA in tails was quantified to indicate the degree of DNA damage. (G) SF295, U87 and SHG140 cells treated with STELB for 24 h and then apoptosis was detected by Annexin V/PI staining. (H) FACS quantification of the total apoptotic cell population, including Annexin V<sup>+</sup>/PI<sup>-</sup> early apoptotic cells and Annexin V<sup>+</sup>/PI<sup>+</sup> late apoptotic cells. (I) Cell cycle analysis of SF295, U87 and SHG140 cells treated with different concentrations of STELB for 48 h. (J) The percentage of cell population at G0/G1, S and G2/M phases. (K)  $\gamma$ -H2AX foci assays of U87 cells treated with STELB and/or a 1 Gy dose of IR. The formation and resolution of  $\gamma$ -H2AX foci were assessed using immunofluorescence. Scale bar, 20  $\mu$ m. (L) Quantification of the number of  $\gamma$ -H2AX foci per nucleus at each time point. Representative images are shown. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; \*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001; \*\*\*\*p < 0.0001.

Supplementary Figure 2



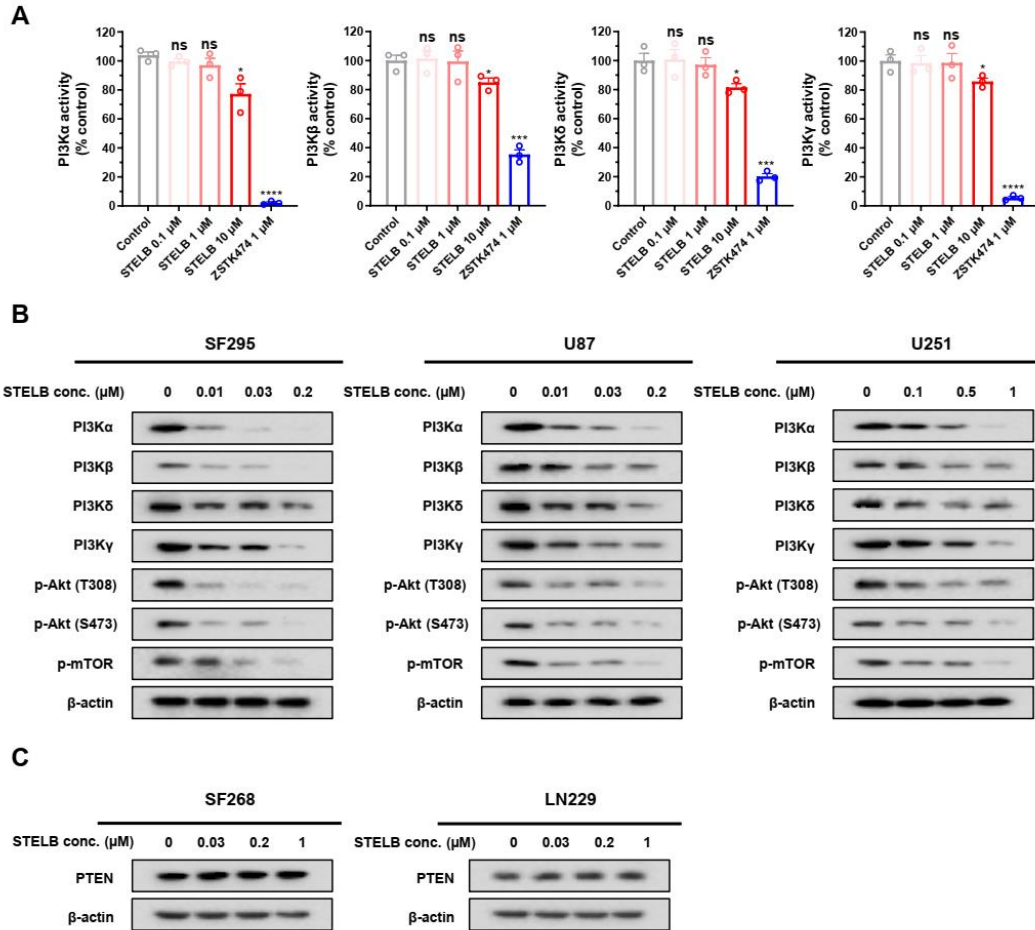
**Figure S2. STELB sensitizes MGMT-negative GBM cells to TMZ**

(A) Cell viability assay for GBM cells with or without pretreatment of STELB. (B) Western blot analysis of MGMT in T98G and SHG140 cells after treatment with STELB for 48 h. (C) Long-term clonogenic assays for U251 cells treated with STELB and/or TMZ for 72 h, allowed to recover for 10-15 days, and then subjected to crystal violet staining. Colony formation was observed under a microscope. (D) Quantification of (C). The absorbance at 570 nm was measured after incubation with 1% SDS for 3 h. Cell survival (%) is expressed as % of the control; CIs were calculated using CalcuSyn.  $CI < 0.9$  represents synergism,  $0.9 < CI < 1.1$  represents additivity, and  $CI > 1.1$  represents antagonism. (E) Apoptosis assays for U251 cells treated with STELB and/or 100  $\mu$ M TMZ for 72 h. Apoptosis was detected by Annexin V/PI staining. FACS quantification of the total apoptotic cell population, including Annexin V<sup>+</sup>/PI<sup>-</sup> early apoptotic cells and Annexin V<sup>+</sup>/PI<sup>+</sup> late apoptotic cells. (F)  $\gamma$ -H2AX foci assays of SF295 and U87 cells treated with STELB and/or 200  $\mu$ M TMZ for 24h, then the STELB/TMZ contained culture media were replaced with common culture media. The formation and resolution of  $\gamma$ -H2AX foci were assessed using immunofluorescence. Scale bar, 20  $\mu$ m. (G) Quantification of the number of  $\gamma$ -H2AX foci per nucleus at each time point. Representative images are shown. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; \*\*\*p < 0.001; \*\*\*\*p < 0.0001.



**Figure S3. Effects of STELB on HR and NHEJ repair in GBM cells**

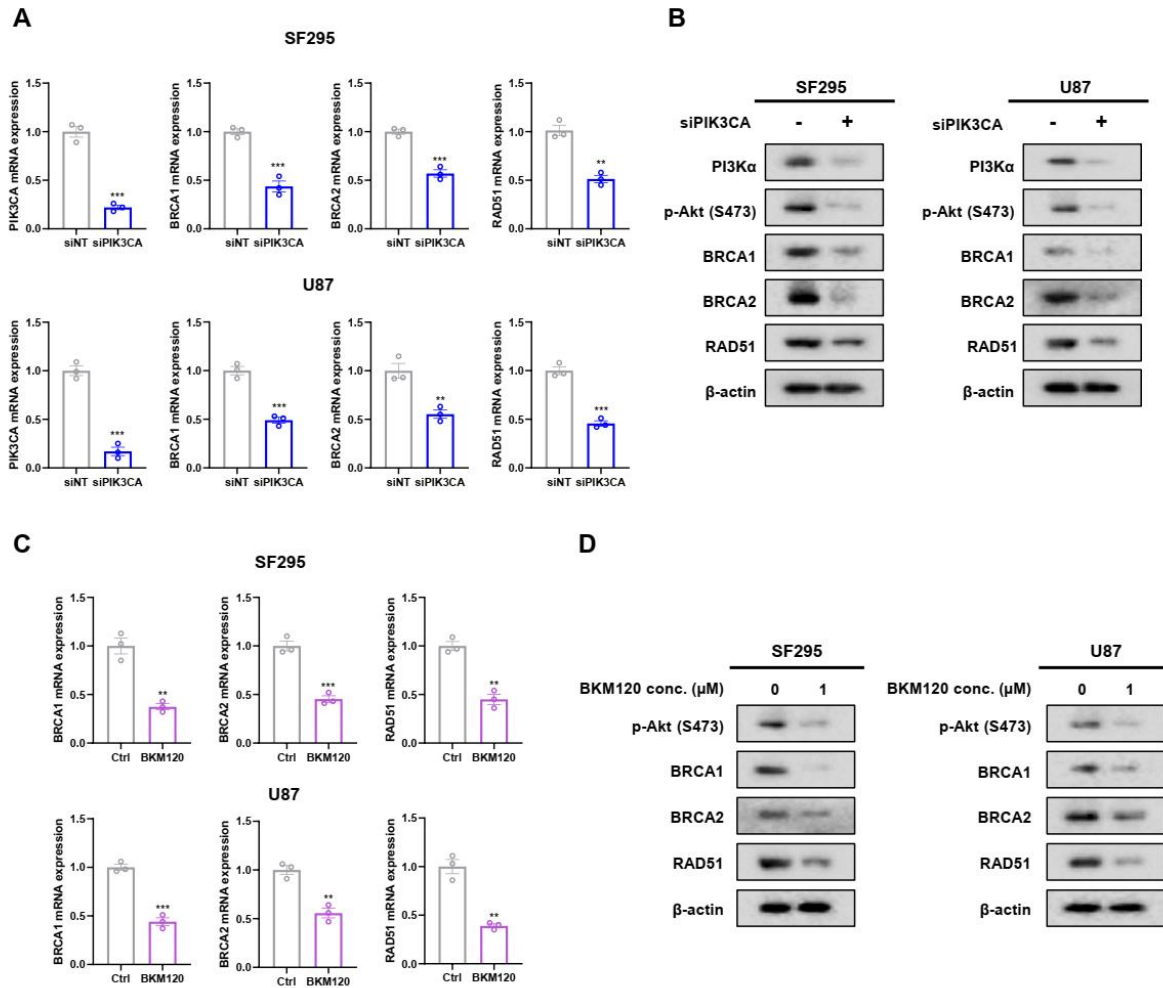
(A) HR reporter assays to evaluate the effect of STELB on HR repair efficiency in SF295 and U87 cells. (B) NHEJ reporter assays to evaluate the effect of STELB on NHEJ repair efficiency in SF295 and U87 cells. (C) Western blot analysis of key proteins involved in the NHEJ repair pathway in SF295 and U87 cells treated with STELB for 48 h. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .



**Figure S4. Effects of STELB on PI3K activity and PI3K protein expression in GBM cells**

(A) Kinase activities of the four PI3K isoforms after co-incubation with the indicated concentrations of STELB. Pan-PI3K inhibitor ZSTK474 serves as a positive control. (B) Western blot analysis of the effects of STELB treatment on proteins involved in the PI3K pathway in GBM cells. (C) Western blot analysis of PTEN in the PTEN-WT GBM SF268 and LN229 cell lines treated with STELB for 48 h. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; ns indicates not significant; \* $p < 0.05$ ; \*\*\*\* $p < 0.0001$ .

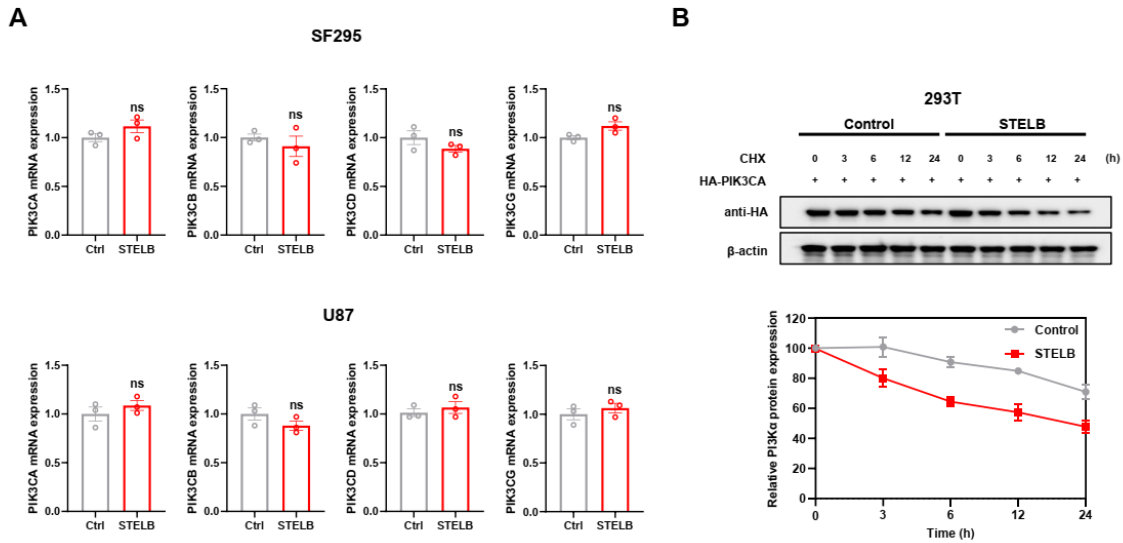




**Figure S5. Effects of genetic and pharmacological inhibition of PI3K on the expression of HR repair key factors BRCA1/2 and RAD51**

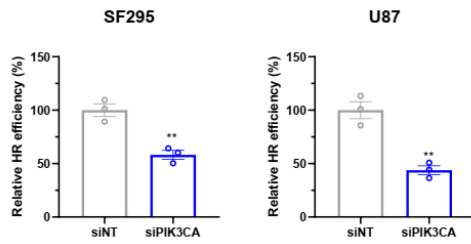
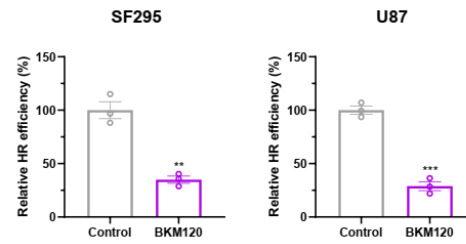
qRT-PCR analysis (A) and Western blot analysis (B) of SF295 and U87 cells transfected with siPIK3CA or siNT to quantify the HR repair key factors BRCA1/2 and RAD51. qRT-PCR analysis (C) and Western blot analysis (D) of SF295 and U87 cells treated with PI3Ki BKM120 to quantify BRCA1/2 and RAD51. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

Supplementary Figure 6



**Figure S6. Effects of STELB on mRNA expression of PI3K isoforms and the stability of exogenous PI3K $\alpha$**

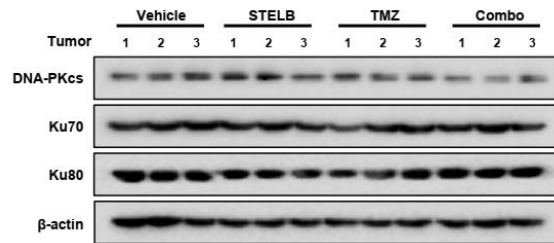
(A) qRT-PCR analysis of SF295 and U87 cells treated with STELB to quantify mRNA levels of four PI3K isoforms, PIK3CA, PIK3CB, PIK3CG, and PIK3CD. (B) Stability analysis of exogenous PI3K $\alpha$  protein in 293T cells transfected with HA-PIK3CA. Cells were treated with 20  $\mu$ M CHX for indicated durations and analyzed by Western blot. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; ns indicates not significant.

**A****B**

**Figure S7. Effects of genetic and pharmacological inhibition of PI3K on HR repair efficiency**

HR reporter assays to detect the effect of siRNA knockdown of PIK3CA (**A**) or BKM120 treatment (**B**) on HR repair efficiency in SF295 and U87 cells. Graphs are shown as the mean  $\pm$  SEM from three independent experiments; P-values, two-tailed unpaired Student's t-test; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

Supplementary Figure 8



**Figure S8. STELB did not influence NHEJ related proteins in vivo.**

Western blot analysis of DNA-PKcs, Ku70, and Ku80 in tumor lysates. Three representative samples were selected from each group.

**Table S1. The primer sequences used in this study.**

Genes	Sequence (5'-3')
PIK3CA	Forward: CGGTGACTGTGTGGGACTTATTGAG Reverse: TGTAGTGTGTGGCTGTTGAACTGC
PIK3CB	Forward: GAGATTGCAAGCAGTGATAGTG Reverse: TAATTTTGGCAGTGATTGTGGG
PIK3CD	Forward: GACCCAGAAGTGAACGACTTTC Reverse: CCTCAAACCTAACGTTGACCAG
PIK3CG	Forward: CACCCAAAAGCATATCCTAAGC Reverse: GTAATGCAGAACATCATCGTCC
BRCA1	Forward: GAACGGGCTTGGAAAGAAAAT Reverse: GTTCACTCTCACACCCAGA
BRCA2	Forward: CAGGTAGACAGCAGCAAGCA Reverse: AAGCCCCTAAACCCCACTTC
RAD51	Forward: CAGATGCAGCTTGAAGCAAA Reverse: TTCTTCACATCGTTGGCATT
MRE11	Forward: TCAGATCTCAGTCAGAGGAGTC Reverse: AGCCATCTGTTCTGCTAAATCT
RAD54B	Forward: TTATCCACGAACTTCGACCTAC Reverse: CCACCAGCTTTTGAACCTAACA
BRIP1	Forward: GTGTGCCAGACTGTGAGCCAAG Reverse: GCCATAAACCCAGTAGAGAGCCAACG
FANCD2	Forward: GCATCTTCCTCCACATTCCTACAC Reverse: CACCGTCTTCTCTAGTTCAGCCATC
FANCM	Forward: AAACGTCAAGGCAGGATAGTTA Reverse: GATAGATGACTTTCGCTGCAAG
18S rRNA	Forward: CAGCCACCCGAGATTGAGCA Reverse: TAGTAGCGACGGGCGGTGTG