Manipulation of sterol homeostasis for the production of 24-epi-

ergosterol in industrial yeast

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Supplementary Fig. 1. A simplified BL biosynthetic pathway from acetyl-CoA. In plants, 24-methylenecholesterol and isofucosterol are catalyzed by DWF1 to synthesize campesterol and β -stiosterol, respectively. BL is proposed to be biologically synthesized from campesterol with 8 enzymatic reactions¹. Unfortunately, several of the BL biosynthetic pathway enzymes have not been identified yet.

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Supplementary Fig. 2. Alignment of the amino acid sequences of DWF1 orthologues. *DWF1* genes from *Arabidopsis thaliana* (*AtDWF1*)(Q39085.2)², *Ajuga reptans* (*ArDWF1*)(BAS68578.1)³, *Brassica rapa* (*BrDWF1*)(VDC72099.1), and *Cannabis sativa* (*CsDWF1*)(XP_030508560.1) were codon-optimized for yeast expression and chemically synthesized. The sequences of BrDWF1 and CsDWF1 were obtained by PSI-BLAST based on AtDWF1 and ArDWF1.



Supplementary Fig. 3. *De novo* biosynthesis of 24-epi-ergosterol in engineered yeast strains. **a** The expression cassettes of *ArDWF1* (YQE224), *AtDWF1* (YQE232), *BrDWF1* (YQE233), and *CsDWF1* (YQE234) were integrated into the genome of YQE102 to evaluate their performance for 24-epi-ergosterol production. **b** HPLC analysis for ergosterol standard, *erg4* Δ -*ArDWF1* strain with ergosterol standard and *erg4* Δ -*ArDWF1* strain respectively. **a** Data are presented as mean values ± SD from three independent biological replicates (n=3), the circles represent individual data points. Source data are provided as a Source Data file.



Supplementary Fig. 4. LCMS analysis of 24-epi-ergosterol and its biosynthetic precursors. a Late sterol biosynthetic pathway including ERG4, ERG5, and DWF1. b HPLC profiles for ergosterol standard, $erg4\Delta$ strain (producing ergosta-5,7,22,24(28)-tetraen-3\beta-ol), $erg5\Delta$ strain (producing ergosta-5,7-dien-3\beta-ol), $erg4\Delta$ -erg5\Delta strain (producing ergosta-5,7,24(28)-trien-3\beta-ol), $erg4\Delta$ -Ar207 strain (producing 24-epi-ergosterol), and $erg4\Delta$ -erg5 Δ -Ar207 strain (producing 24-epi-ergosta-5,7-dien-3\beta-ol). c MS verification of the HPLC peaks as ergosta-5,7,22,24(28)-tetraen-3\beta-ol, ergosta-5,7,24(28)-trien-3\beta-ol, 24-epi-ergosterol, and 24-epi-ergosta-5,7-dien-3\beta-ol, respectively. Source data are provided as a Source Data file.



Supplementary Fig. 5. NMR profiles of ergosterol standard and 24-epi-ergosterol. a ¹H NMR spectroscopy. 24-Epi-ergosterol (500 MHz, CDCl₃) δ 5.57 (dd, 1H), 5.38 (dt, 1H), 5.24-5.11 (m, 2H), 3.63 (tq, 1H), 2.47 (ddd, 1H), 2.28 (ddd, 1H), 2.13-1.92 (m, 2H), 1.88 (dddd, 3H), 1.85-1.73 (m, 1H), 1.76-1.62 (m, 1H), 1.61 (dd, 1H), 1.61-1.49 (m, 1H), 1.52-1.42 (m, 1H), 1.44-1.17 (m, 4H), 1.15-0.97 (m, 3H), 0.96-0.89 (m, 6H), 0.88-0.80 (m, 6H), 0.79 (td, 2H), 0.63 (d, 3H), 0.54 (d, 1H). **b** ¹³C NMR spectroscopy. 24-Epi-ergosterol (126 MHz, CDCl₃) δ 141.47, 141.37, 139.80, 139.77, 135.82, 132.07, 131.98, 119.60, 117.48, 116.30, 116.27, 71.08, 70.47, 55.86, 55.68, 55.06, 54.59, 54.52, 49.46, 46.25, 43.09, 42.94, 42.83, 40.81, 40.62, 40.56, 40.27, 39.20, 39.10, 38.85, 38.39, 38.00, 37.15, 37.04, 36.26, 34.22, 33.66, 33.23, 32.43, 32.01, 31.49, 30.35, 29.66, 28.61, 28.43, 28.11, 27.97, 23.06, 23.04, 21.56, 21.13, 20.22, 20.18, 19.66, 18.84, 18.28, 18.05, 16.30, 15.40, 13.06, 12.06, 12.02, 11.84. Source data are provided as a Source Data file.



Supplementary Fig. 6. Directed evolution of ArDWF1 for improved production of 24-epi-ergosterol in engineered yeast strains. a The procedure of library construction using homologous recombination in yeast. pRS42H was linearized by *Hin*dIII and *Bam*HI. The mutant library, with 35 bp homologous arms to pRS42H was generated by error-prone PCR. Co-transformation of mutant fragment and linearized plasmid into YQE101 results in recombination and library construction. **b** Positive mutants obtained from directed evolution. Six mutants with a total of 10 mutations were obtained in the first round of directed evolution. The DWF1 activity was defined as the ratio of HPLC peak areas of 24-epi-ergosterol to ergosta-5,7,22,24(28)-tetraen- 3β -ol. **b** Data are presented as mean values \pm SD from three independent biological replicates (n=3), the circles represent individual data points. Significance (*p*-value) was evaluated by two-sided *t*-test, no significance (n.s.) presents *p* > 0.05. Source data are provided as a Source Data file.



Supplementary Fig. 7. The activity of DWF1 combinatorial mutants. The second round of directed evolution via DNA shuffling resulted in the construction of a series of combinatorial mutants, whose activity was evaluated by episomal plasmid transformation into YQE101 (for relative DWF1 activity). Data are presented as mean values \pm SD from three independent biological replicates (n=3), the circles represent individual data points. Significance (*p*-value) was evaluated by two-sided *t*-test, no significance (n.s.) presents *p* > 0.05. Source data are provided as a Source Data file.



Supplementary Fig. 8. Fermentation profiles of YQE224, YQE231, YQE717, and YQE722 in shake flasks. Fermentation of YQE224, YQE231, YQE717, and YQE722 was performed in 250 mL shake flasks with 50 mL YPD medium containing 20 g/L glucose for 96 h, to obtain the time courses of cell growth **a**, titer of 24-epi-ergosterol **b**, titer of late sterols **c**, and the ratio of 24-epi-ergosterol to late sterols **d**. Data are presented as mean values \pm SD from three independent biological replicates (n=3). Source data are provided as a Source Data file.



Supplementary Fig. 9. Fed-batch fermentation for YQE231, YQE717, and YQE722. a YQE231. b YQE717. c YQE722. The green and blue arrows indicated the start of glucose and ethanol feeding, respectively. Data are presented as mean values \pm SD from three independent biological replicates (n=3). Source data are provided as a Source Data file.



Supplementary Fig. 10. Dissolved oxygen curves of fed-batch fermentation for YQE231, YQE717, and YQE722. a YQE231. b YQE717. c YQE722. The green and blue arrows indicated the start of glucose and ethanol feeding, respectively. Source data are provided as a Source Data file.



Supplementary Fig. 11. The percentage of un-acylated sterols in engineered yeast strains. The un-acylated form of 24-epi-ergosterol and ergosta-5,7,22,24(28)-tetraen-3 β -ol were quantified by HPLC. Data are presented as mean values ± SD from three independent biological replicates (n=3), the circles represent individual data points. The statistical evaluation (*p*-value) of the percentage of un-acylated sterols in 36 h (black) and 96 h (red star) were performed by two-sided *t*-test separately, no significance (n.s.) presents *p* > 0.05. Source data are provided as a Source Data file.



Supplementary Fig. 12. Relative transcription level of ARE2, YEH1, YEH2 in YQE717 and YQE231. a ARE2. b YEH1. c YEH2. Samples collected at 12 h, 24 h, 36 h, 48 h, and 72 h during fermentation of the two strains in shake flasks were subject to qPCR analysis. The transcription level of YQE231 for the three genes at 12 h was set to 1. Data are presented as mean values \pm SD from three independent biological replicates (n=3). Source data are provided as a Source Data file.



Supplementary Fig. 13. Quantification of late sterols with different promoter combinations for the expression of *ARE2*, *YEH1*, and *YEH2* in YQE231. Details on the properties of the selected promoters were provided in Supplementary Table 3. Data are presented as mean values \pm SD from three independent biological replicates (n=3), the circles represent individual data points. Significance (*p*-value) of the titer of total late sterols (red) and 24-epi-ergosterol (black) were evaluated by two-sided *t*-test, no significance (n.s.) presents *p* > 0.05. Source data are provided as a Source Data file.



Supplementary Fig. 14. Relative transcription level of *Ar207*, *ACC1*, and *ERG5* in YQE717, YQE729, and YQE734. a *Ar207*. b *ACC1*. c *ERG5*. Samples collected at 12 h, 24 h, 36 h, 48 h, and 72 h during the fermentation of these strains in shake flasks were used for qPCR analysis to determine the transcription level. The transcription level of YQE717 for *Ar207* and *ACC1* as well as YQE729 for *ERG5* at 12 h were set to 1. Data are presented as mean values \pm SD from three independent biological replicates (n=3). Source data are provided as a Source Data file.



Supplementary Fig. 15. HPLC profiles for ergosterol standard and YQE734. Source data are provided as a Source Data file.



Supplementary Fig. 16. Fed-batch fermentation for YQE729 and YQE734. a YQE729. b YQE734. The green and blue arrows indicated the start of glucose and ethanol feeding, respectively. Data are presented as mean values \pm SD from three independent biological replicates (n=3). Source data are provided as a Source Data file.



Supplementary Fig. 17. Dissolved oxygen curves of fed-batch fermentation for YQE729 and YQE734. a YQE729. b YQE734. The green and blue arrows indicated the start of glucose and ethanol feeding, respectively. Source data are provided as a Source Data file.



Supplementary Fig. 18. Molecular mechanisms for improved activity of the evolved ArDWF1 mutants. a Predicted structure of ArDWF1 by AlphaFold2 (https://colab.research.google.com/github/sokrypton/ColabFold/blob/main/AlphaFold 2.ipynb#scrollTo=kOblAo-xetgx). FAD-binding domain and positive sites (K122, Q137, V143, V152, M235, V270, S306, and Y338) were highlighted in green and yellow, respectively. The residues at 143, 338, and 306 in ArDWF1 **b** and Ar207 **c** were compared.

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Species	Bioassay	BL	EBL	HBL	Reference	
D	1 st internode test	100	36	33	4	
Bean	2 nd internode test	100	10	0.1	·	
Radish	Elemention to st	100	10	10	5	
Tomato	Elongation test	100	10	1	5	
Rice (Arborio)		100	10	100	6	
Rice (Bahia)	Rice-lamina	100	5	5	7	
Rice (Tan-		100	10	25	8	
Tomato	Cell-wall-bound Invertase activity	100 (0.2 μM)	79.6 (2 μM)	67.6 (2 μM)	9	
	Sucrose uptake	100 (0.2 μM)	73.5 (2 μM)	71.4 (2 μM)	,	
Barberry (Maria)	Total soluble	100	96.9			
	Free amino acids	100	77.6		10	
Barberry (Red Rocket)	Total soluble	100	65.4		10	
	Free amino acids	100	96.8			

Supplementary Table 1. Activity comparison among BL, EBL, and HBL.

Mutants	Mutation sites					
Ar0105	V143G					
Ar0424	V152A	Q519R				
Ar0440	E61K	K122T	V270A	Y338H		
Ar0527	S306P					
Ar0528	M235T					
Ar1322	Q137R					

Supplementary Table 2. Mutants obtained from the first round of directed evolution.

Strain -	Carbon source							
	Glucose			Ethanol				
YQE717	Ptefi	<i>РтDH3</i>	PFBAI	PTEF1	<i>РтDH3</i>	PFBA1		
	2.159565	3.234124	1.911226	0.430327	0.510003	0.212029		
YQE717 SES	P_{TEF1}	P_{ADH2}	P _{FBA1}	P_{TEF1}	P_{ADH2}	P_{FBA1}		
	2.159565	0.028053	1.911226	0.430327	0.539263	0.212029		
YQE717 SWS	Ptefi	PGASI	PFBA1	P _{TEF1}	PGASI	PFBAI		
	2.159565	1.069321	1.911226	0.430327	0.205553	0.212029		
YQE717 EEE	PJENI	PICLI	P _{ADH2}	PJENI	PICLI	P _{ADH2}		
	0.012039	0.0115	0.028053	0.817225	0.706364	0.539263		
YQE717 ESE	PJENI	Ptefi	P _{ADH2}	PJENI	Ptefi	P _{ADH2}		
	0.012039	2.159565	0.028053	0.817225	0.430327	0.539263		
YQE717 WWW	P_{RPL25}	P _{RPS3}	PGASI	P_{RPL25}	P _{RPS3}	PGASI		
	1.077	1.067463	1.069321	0.178034	0.175343	0.205553		
YQE717 WSW	P_{RPL25}	P_{TEF1}	P_{GASI}	P_{RPL25}	P_{TEF1}	P _{GAS1}		
	1.077	2.159565	1.069321	0.178034	0.430327	0.205553		

Supplementary Table 3. Yeast promoter activity values under different growth conditions. The data were adapted from Keren *et al.*¹¹.

Supplementary references

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