## Supplementary Information

## TRIM27 maintains gut homeostasis by promoting intestinal stem cell self-

renewal

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Supplementary Fig. 1 The constructive strategy and analysis of *Trim27*-knockout mice. a Strategy for the generation of *Trim27*-knockout (*Trim27*-/-) mice. b PCR analysis of the mice. The predicted sizes of PCR products were 450 bp (*Trim27*+/+) and 412 bp (*Trim27*-/-). WT, primers for identification of *Trim27*+/+ mice; KO, primers for identification of *Trim27*-/- mice.

**c** Immunoblots of Trim27 in BMDMs from  $Trim27^{+/+}$ , heterozygous ( $Trim27^{+/-}$ ) and  $Trim27^{-/-}$  mice. **d** Time course of the body weight of  $Trim27^{+/+}$  and  $Trim27^{-/-}$  mice. **e** The morphology of 16-week-old  $Trim27^{+/+}$ ,  $Trim27^{+/-}$  and  $Trim27^{-/-}$  mice and their organs. **f** Hematoxylin and Eosin (H&E) staining of mouse organs from mice as in **e**. Scale bars, 200 µm.



**Supplementary Fig. 2 TRIM27 promotes the development of intestines. a** H&E staining of the small intestine (SI) from 16-week-old  $Trim27^{+/+}$  and  $Trim27^{-/-}$  mice. Bottom, enlarged images of regions outlined in upper panels. Scale bars, 300 µm. White dashed lines mark crypt borders. **b**–**d** Crypt depth (**b**), crypt numbers per mm (**c**) and villus height (**d**) of the SI from mice as in **a**. At least 20 randomly selected crypts (**b**, **d**) or randomly selected fields (**c**) from the SI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed. **e** H&E staining of the large intestine (LI) from 16-week-old  $Trim27^{+/+}$  and  $Trim27^{-/-}$  mice. Bottom, enlarged images of regions outlined in upper panels. Scale bars, 300 µm. White dashed lines mark crypt borders. **f**–**h** Crypt depth (**f**), crypt numbers per mm (**g**) and villus height (**h**) of the LI from mice as in **e**. At least 20 randomly selected crypts (**f**, **h**) or randomly selected fields (**g**) from the LI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed, and the average was calculated. Five mice per group selected crypts (**f**, **h**) or randomly selected fields (**g**) from the LI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed. For **b**–**d and f**–**h**, statistical analyses were performed using unpaired two-tailed Student's *t*-test. Data are mean ± SEM (n=5). \*\**P* < 0.01;

\*\*\*P < 0.001; \*\*\*\*P < 0.0001.



Supplementary Fig. 3 TRIM27 promotes the proliferation of intestinal stem cells (ISCs) in a Notch signaling pathway-independent manner. a Representative image of Lgr5 (green) and EdU signal (red) in the SI and LI from 16-week-old  $Trim27^{+/+}$  and  $Trim27^{-/-}$  mice on day 0.5 after a 4-day EdU administration. Scale bars, 100 µm. **b** The proportion of Lgr5<sup>+</sup>EdU<sup>+</sup> cells in the SI and LI from mice as in **a**. At least 20 randomly

selected crypts from the SI and LI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed. **c**, **d** Quantitative PCR (qPCR) analysis of Notch target gene *Dll1* mRNA of crypts freshly isolated from the SI (**c**) and LI (**d**) from 16-week-old *Trim27*<sup>+/+</sup> and *Trim27*<sup>-/-</sup> mice. **e**, **f** qPCR analysis of Notch target gene *Dll4* mRNA of crypts freshly isolated from the SI (**e**) and LI (**f**) from mice as in **c**. **g**, **h** qPCR analysis of Notch target gene *Hes1* mRNA of crypts freshly isolated from the SI (**g**) and LI (**h**) from mice as in **c**. **i** Gene Set Enrichment Analysis (GSEA) results for Notch signaling pathway in IBS-D patients. FDR, false discovery rate. For **b**, **c**–**h**, statistical analyses were performed using unpaired two-tailed Student's t-test. Data are box-and-whisker plots displaying median, interquartile range (boxes) and minima and maxima (whiskers) (**b**, **c**–**h**, n=5). p > 0.05, not significant (ns); \*p < 0.05; \*\*\*\*P < 0.0001.



Supplementary Fig. 4 TRIM27 interacts with  $\beta$ -catenin and mainly inhibits its K48-linked polyubiquitination. a Immunoprecipitation (IP) of  $\beta$ -catenin by Trim27 in cytoplasm and nucleus of crypts from 16-week-old *Trim27<sup>+/+</sup>* and *Trim27<sup>-/-</sup>* mice. Crypts were lysed and immunoprecipitated with anti-Trim27 antibody. IP products were immunoblotted with the indicated antibodies. **b** *In vivo* ubiquitination assay of Myc- $\beta$ -catenin in *TRIM27<sup>+/+</sup>* and *TRIM27<sup>-/-</sup>* HEK293T cells transfected with the indicated plasmids for 24 h. Cells were lysed and immunoprecipitated with the antibody against Myc. The immunoprecipitated proteins were immunoblotted with the antibody against

HA. **c** qPCR analysis of  $\beta$ -catenin mRNA of crypts freshly isolated from the SI and LI from 16-week-old *Trim27*<sup>+/+</sup> and *Trim27*<sup>-/-</sup> mice. **d** Schematic diagram of Trim27 and  $\beta$ -catenin domains. For **c**, statistical analyses were performed using unpaired two-tailed Student's t-test. Data are box-and-whisker plots displaying median, interquartile range (boxes) and minima and maxima (whiskers) (**c**, n=5). p > 0.05, not significant (ns).



Supplementary Fig. 5 Establishment and analysis of  $Trim 27^{fl/fl}$  and trim 27[Lgr5] mice. a Strategy for the generation of trim 27[Lgr5] mice. b PCR analysis of the mice. The predicted sizes of PCR products were 585 bp ( $Trim 27^{fl/fl}$ ), 513 bp ( $Trim 27^{+/+}$ ) and 350 bp (creERT2). c Immunohistochemical analysis of Trim27 in the SI from 16-week-old  $Trim 27^{fl/fl}$  and trim 27[Lgr5] mice. Scale bars, 50 µm. d AB-PAS staining of the SI

from 16-week-old Trim27<sup>fl/fl</sup> and trim27[Lgr5] mice after 2% DSS treatment. Scale bars, 200 µm. e The number of AB-PAS-positive goblet cells in intestinal tissue sections from mice as in d. At least 20 randomly selected villi from the SI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed. f Immunohistochemical analysis of lysozyme in the SI from 16-week-old Trim27<sup>/l/fl</sup> and trim27[Lgr5] mice after 2% DSS treatment. Scale bars, 200 µm. g The number of lysozyme-positive Paneth cells per crypt in intestinal tissue sections from mice as in f. At least 20 randomly selected crypts from the SI of each mouse were analyzed, and the average was calculated. Five mice per group were analyzed. h Ki-67 staining of the SI from 16-week-old Trim27<sup>fl/fl</sup> and trim27[Lgr5] mice after 2% DSS treatment. Scale bars, 100 µm. i Ki-67 positive area in intestinal tissue sections from mice as in h. Five mice per group were analyzed. j Immunoblots of mast cell tryptase, Trim27 and GAPDH in intestinal tissues from 16-week-old Trim27/Lgr5/ mice. k IP of TRIM27 and Axin1 by β-catenin in HEK293T cells. Cells were lysed and immunoprecipitated with anti- $\beta$ -catenin antibody in the presence or absence of SKL2001 (20  $\mu$ M). IP products were immunoblotted with the indicated antibodies. For e, g and i, statistical analyses were performed using two-way ANOVA and Sidak's multiple comparisons test. Data are mean  $\pm$  SEM (e and i, n=5; g, n=8). p > 0.05, not significant (ns); \*p <0.05; \*\*p < 0.01; \*\*\*\*p < 0.0001.



Supplementary Fig. 6 The Wnt/ $\beta$ -catenin signaling activator SKL2001 promotes ISC selfrenewal in *trim27[Lgr5]* mice. **a** Ki-67 staining of the SI from 12–16-week-old *Trim27<sup>/l/fl</sup>* and *trim27[Lgr5]* mice fed with corn oil or SKL2001. Scale bars, 300 µm. **b** Ki-67 positive area in the SI from mice as in **a**. Five mice per group were analyzed. statistical analyses were performed using two-way ANOVA and Sidak's multiple comparisons test. Data are mean ± SEM (b, n=5). p > 0.05, not significant (ns); \*\*\*\*p < 0.0001.

Name	Oligonucleotides (5'-3')
Gapdh-F	GGAGCGAGATCCCTCCAAAAT
Gapdh-R	GGCTGTTGTCATACTTCTCATGG
<i>Tnf</i> -F	CCTCTCTCTAATCAGCCCTCTG
<i>Tnf</i> -R	GAGGACCTGGGAGTAGATGAG
<i>Il1b-</i> F	GCAACTGTTCCTGAACTCAACT
Il1b-R	ATCTTTTGGGGTCCGTCAACT
Gper-F	ATGGATGCGACTACTCCAGC
Gper-R	AAGAGGGCAATCACGTACTGC
Axin2-F	GTCCACCAAACCTATGCCCG
Axin2-R	GAGTGTAAAGACTTGGTCCA
Myc-F	CAACGTCTTGGAACGTCAGA
Myc-R	TCGTCTGCTTGAATGGACAG
<i>Ctnnb1</i> -F	GATTCTGGAATCCATTCTGG
Ctnnb1-R	GGAAAAGCCTTGCTCCCATT
Wnt3a-F	CCTCGGAGATGGTGGTAGA
Wnt3a-R	GTTAGGTTCGCAGAAGTTGG
Dll1-F	CGGGGGCTCTGGCCCGCCTT
Dll1-R	AGCACTGGCGTGACAGCACT
Dll4-F	ACGCCGGTATTGGGCACCAA
Dll4-R	TTGGATGTTGAGTGAGAAGG
Hes1-F	AATGCCGGGAGCTATCTTTCT
Hes1-R	CCAGCCAGTGTCAACACGA
$\beta$ -catenin-F	CCCAGTCCTTCACGCAAGAG
$\beta$ -catenin-R	CATCTAGCGTCTCAGGGAACA
<i>Trim</i> 27 <sup>+/+</sup> -F	CTCCGGAGCCGAGAAGCGGAGCGAG
<i>Trim27</i> <sup>+/+</sup> -R	TCCTGCTCGCAGTACAGCTTCAGAG
<i>Trim27</i> -/F	GAGCAGCTGTGCGGGGGACGGGTCTG
<i>Trim27-/</i> R	TGGAAAGTGGGTCTCCAAATCCAAATC
<i>Trim27<sup>fl/fl</sup></i> -F	GCACGTGCTCTCTCACTAAGCTGT
<i>Trim27<sup>f1/fl</sup></i> -R	GTGGCGCACTCCATTAATCCTCTTGT
CreERT2-F	CGATGCAACGAGTGATGAGG
CreERT2-R	CGCATAACCAGTGAAACAGC

Supplementary Table 1. Oligonucleotides used for qPCR analysis or mouse genotyping PCR.