

1136 Supplementary materials for  
1137 **Mini-dCas13X-mediated RNA editing restores dystrophin expression in a**  
1138 **humanized mouse model of Duchenne muscular dystrophy**

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1150 Figure S1. In vitro studies of c.4174C>T mutation correction using the mxABE  
1151 system.

1152 Figure S2. Construction of 10 mxABE expression cassettes.

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1154 Figure S4. Evaluation of SNV counts by transcriptome-wide off-target analysis  
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1159 gene.

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1161 weeks.

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1165 weeks.

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1167 dystrophin expression.

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1169 analysis *in vivo*.

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1171 analysis *in vivo*.

1172 Figure S13. Gene editing 3 weeks after systemic delivery of mxABEs in  
1173 *DMD*<sup>E30mut</sup> mice.

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1179 AAV9-mxABE.

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1187 systemic administration of AAV-mxABE.

1188 Figure S21. Collagen staining shows no obvious fibrosis in hearts of treated or  
1189 untreated DMD<sup>E30mut</sup> mice 6 months after systemic administration of  
1190 AAV-mxABE.

1191 Figure S22. Host immune response to AAV-mxABE treatment indicates no  
1192 overt toxicity after 6 months.

1193 Figure S23. Systemic delivery of AAV-mxABE rescues dystrophin deficiency in  
1194 adult DMD<sup>E30mut</sup> mice.

1195 Figure S24. Immunostaining of dystrophin in adult DMD<sup>E30mut</sup> mice 6 weeks  
1196 after systemic injection of AAV-mxABE.

1197 Tables S1. Target sgRNA and primer sequences.

1198 Supplemental Sequences

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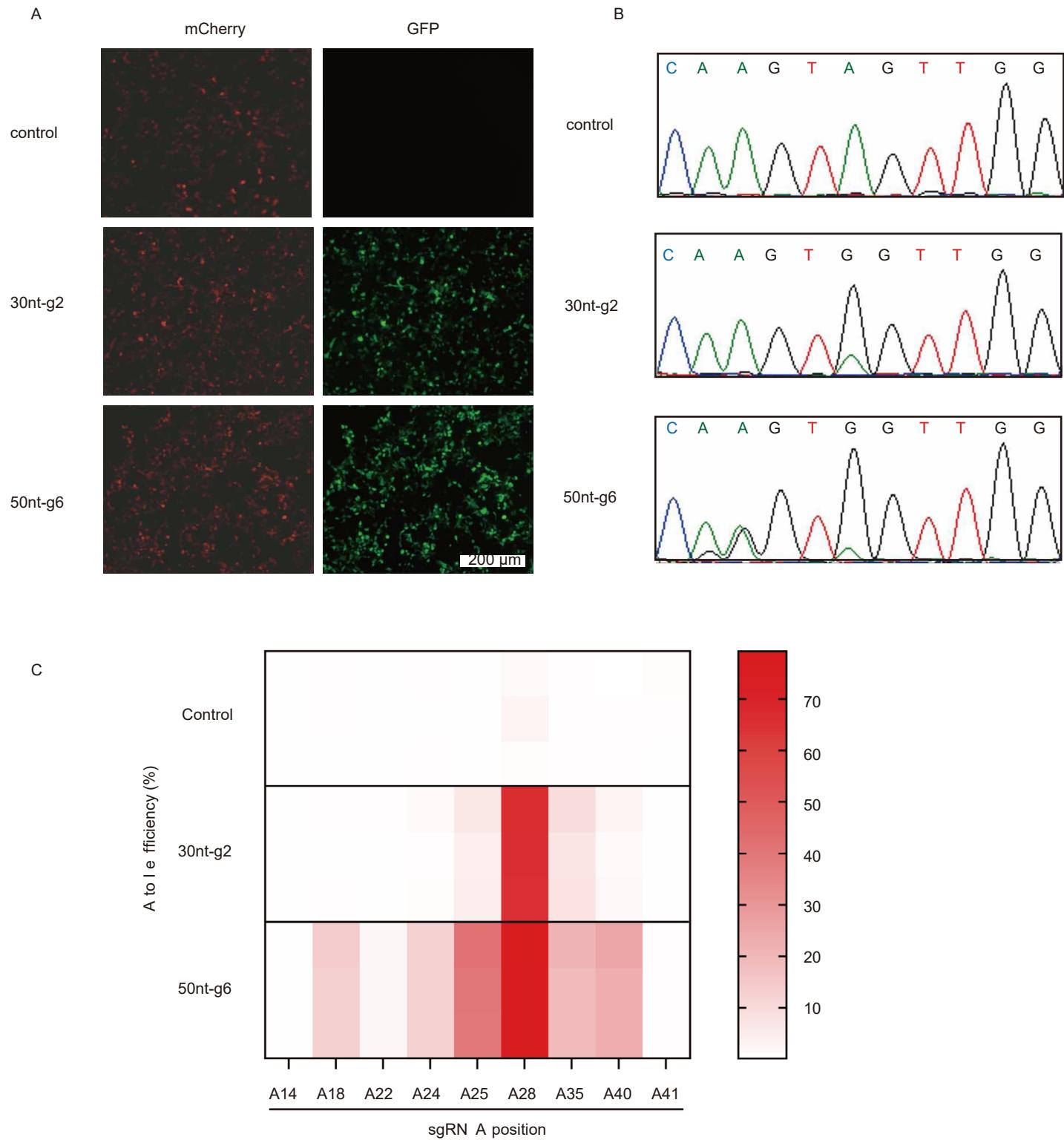
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Figure S1



1224 **SUPPLEMENTAL FIGURES**

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1226 **Figure S1. *In vitro* studies of c.4174C>T mutation correction using**  
1227 **mxABE constructs.**

1228 A. Fluorescence microscopy images of HEK293T cells transfected with  
1229 reporter alone, or reporter and mxABE constructs. Scale bar, 200 µm. B.  
1230 Representative Sanger sequencing trace of reporter transcripts. C.  
1231 Measurements of the representative gRNA (30 nt-g2 and 50 nt-g6) effect on  
1232 bystander A to I editing efficiency (n=3).

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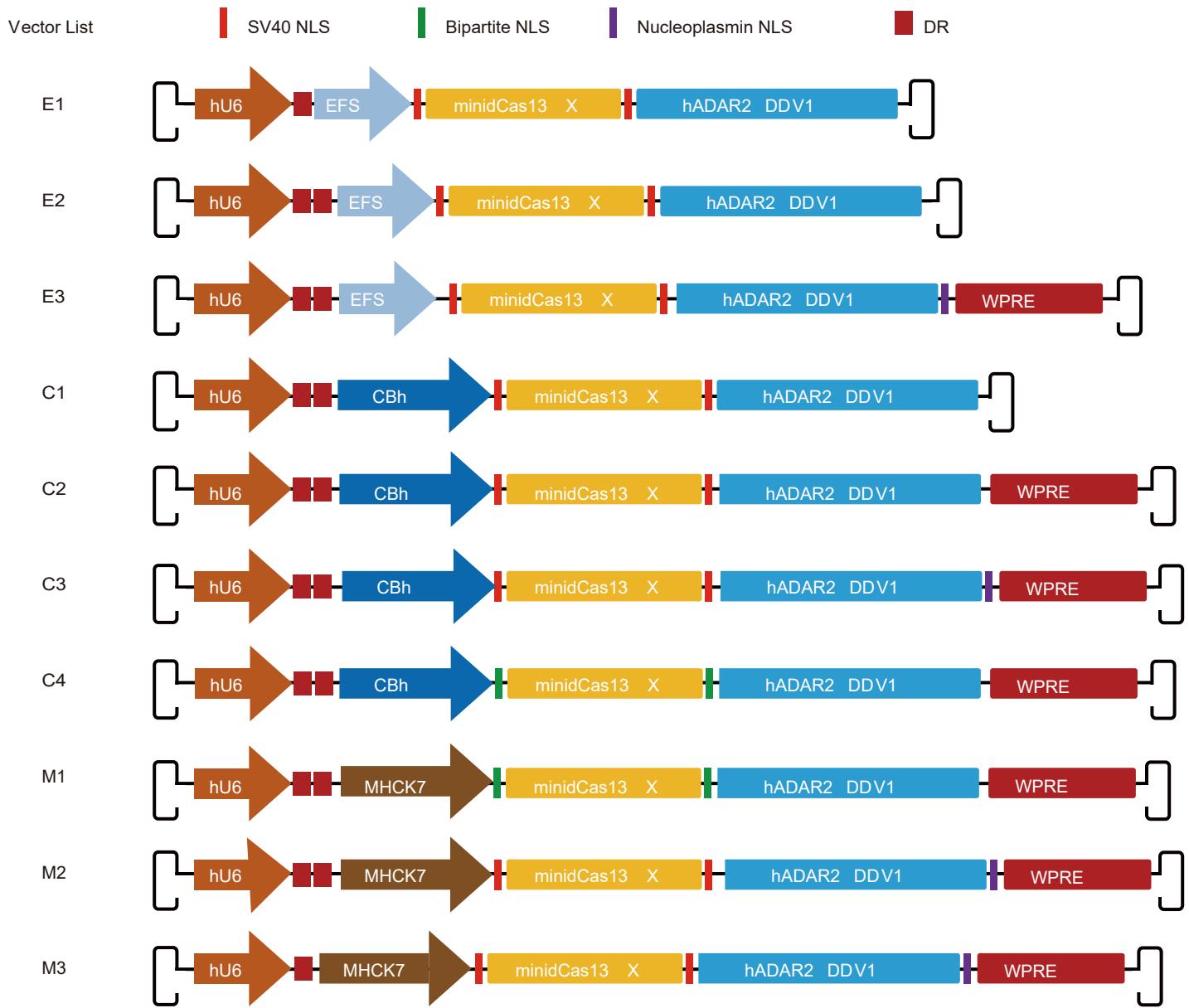
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Figure S2



1268 **Figure S2. Construction of 10 mxABE expression cassettes.**

1269 The mxABE expression cassettes contained different promoters (EFS, CBh,  
1270 MHCK7), direct repeat (DR) numbers (single, dual DR), nuclear localization  
1271 signals (NLS), and were constructed with or without a translational regulatory  
1272 element (WPRE).

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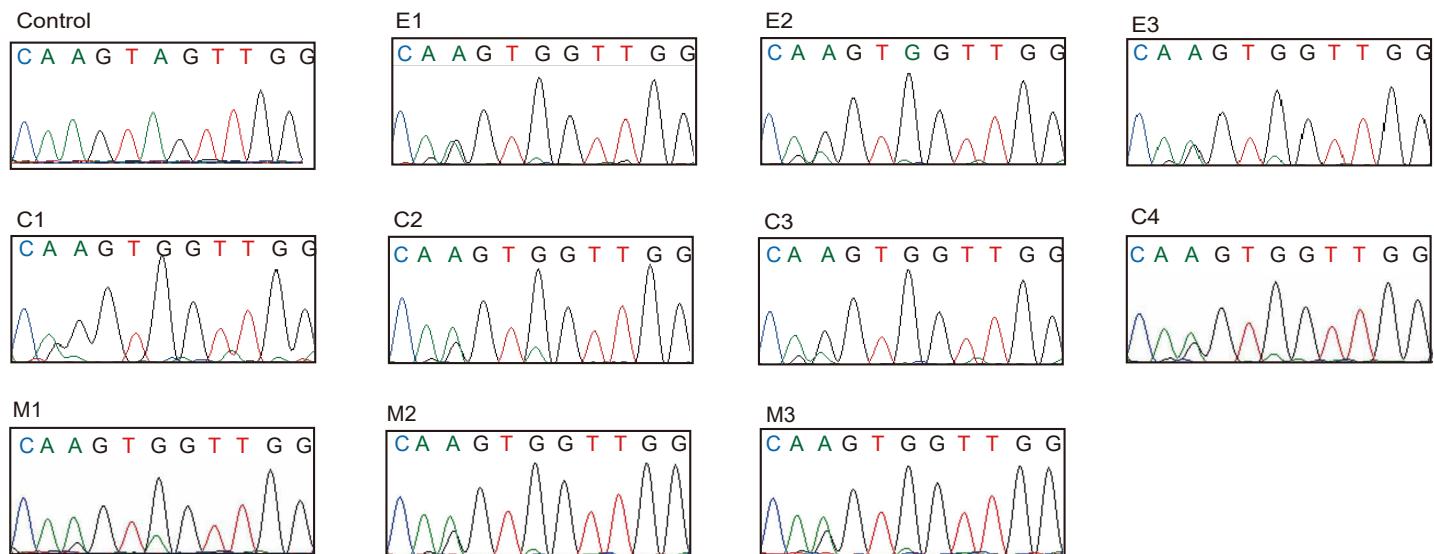
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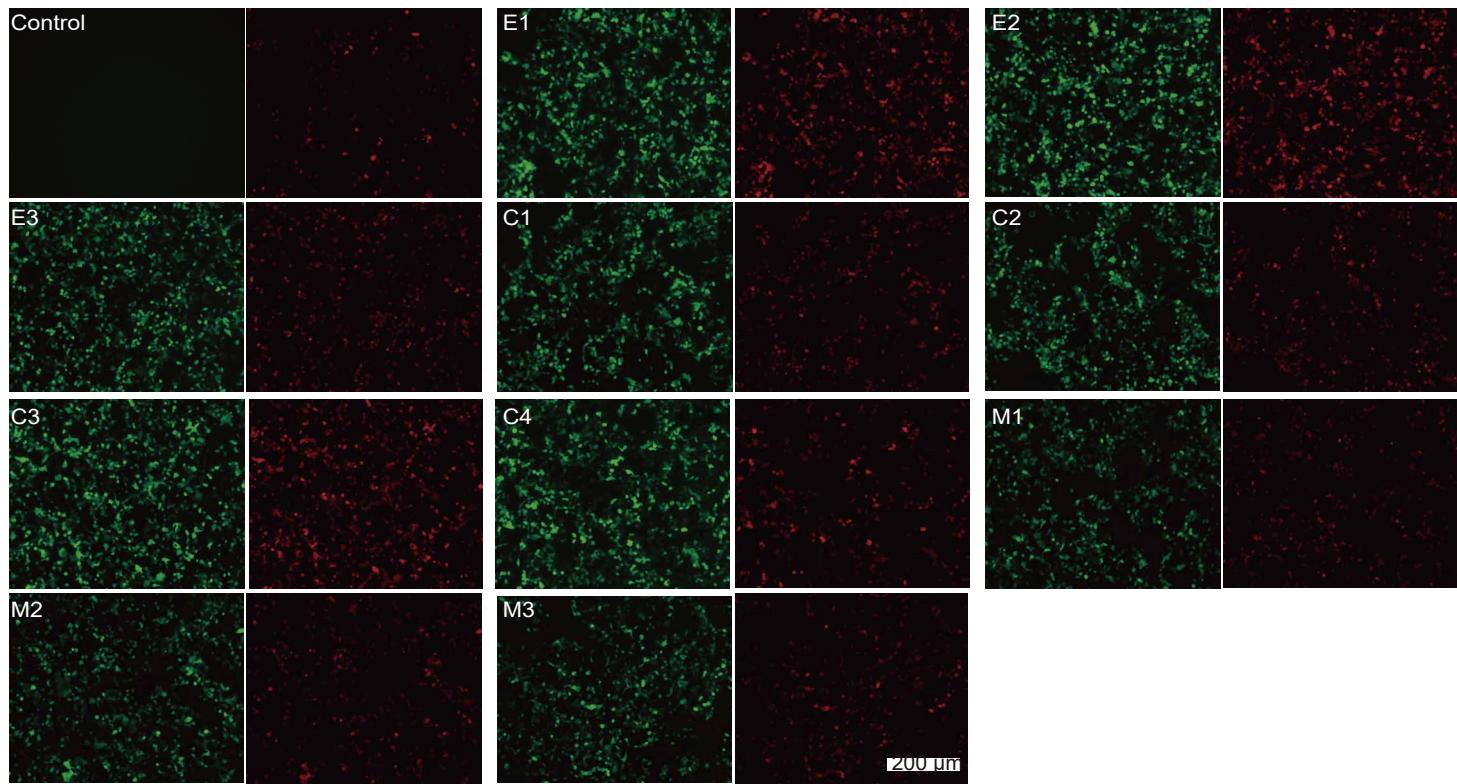
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Figure S3

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1312 **Figure S3. Representative Sanger sequencing trace of reporter  
1313 transcripts.**

1314 A. Sanger sequencing results showing representative A to I conversion on  
1315 reporter transcripts by different mxABE editors. B. Fluorescence microscopy  
1316 images of HEK293T cells transfected with reporter alone or with reporter and  
1317 mxABE construct. Scale bar, 200  $\mu$ m. Figure S3B and S1A showed similar  
1318 results but from different experiments investigating the effect of gRNA length  
1319 and construct expression elements on mutation correction efficiency  
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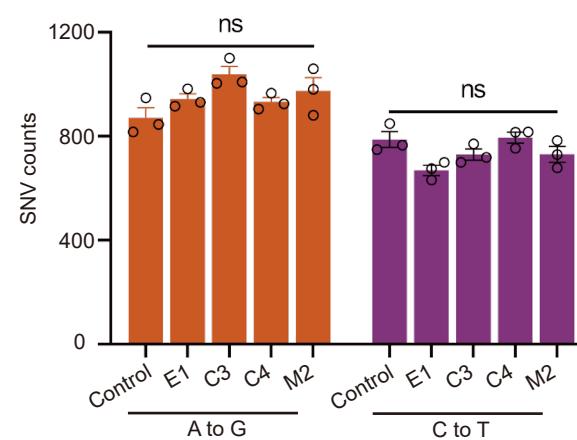
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Figure S4



1356 **Figure S4. Evaluation of SNV counts *in vitro* using transcriptome-wide**  
1357 **off-target analysis.**

1358 The transcriptome-wide off-target effect of different mxABE constructs  
1359 targeting the DMD c.4174C>T mutation were performed *in vitro*. The E1  
1360 mxABE construct was driven by an EFS promoter; C3 and C4 were driven by  
1361 an CBh promoter; and M2 was driven by an MHCK7 promoter. Non-targeting  
1362 mxABE construct was analyzed as a control. Values are shown as mean ±  
1363 SEM (n=3). NS, not statistically significant in multiple comparison test using  
1364 ANOVA (P< 0.01).

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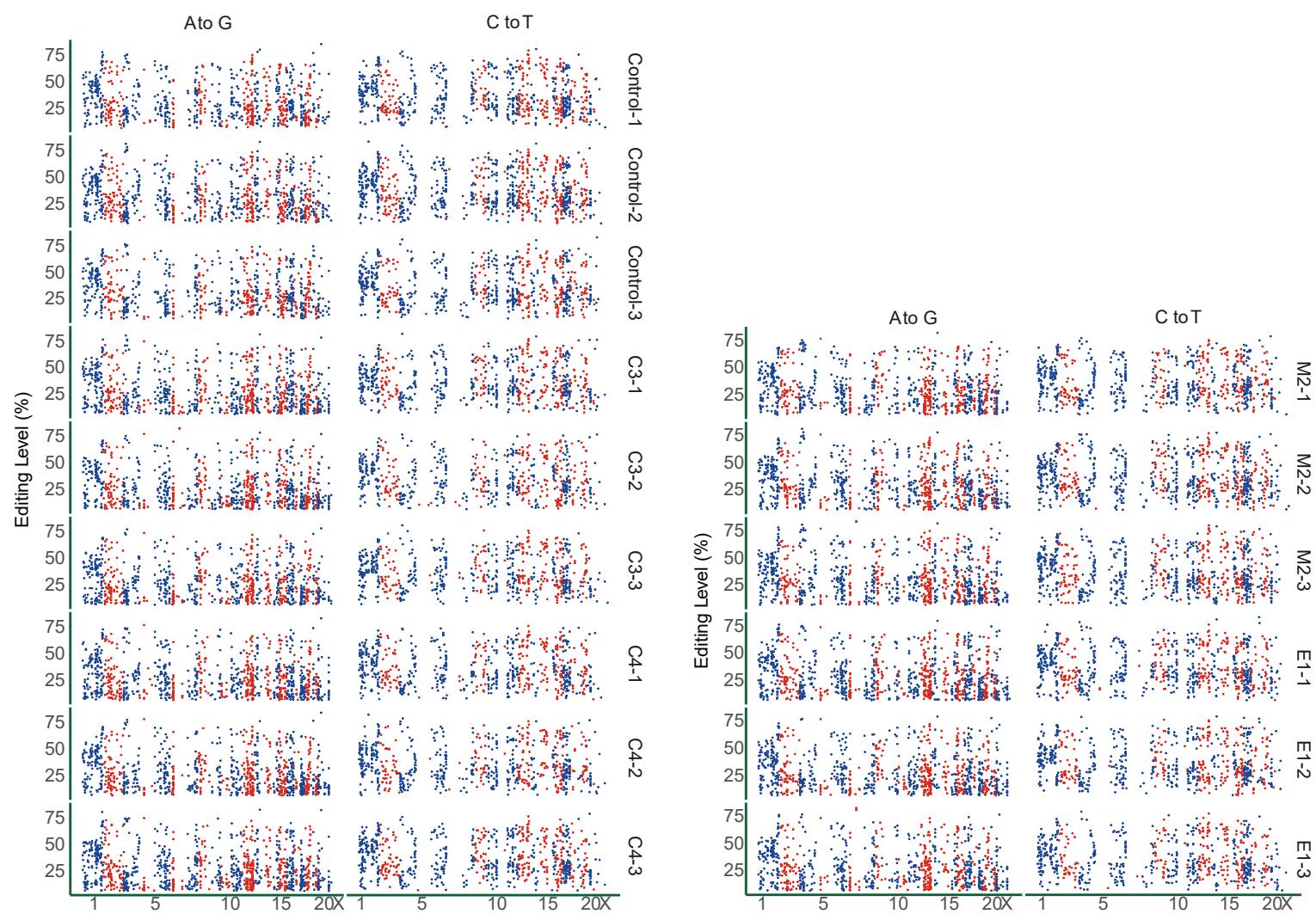
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Figure S5



1400 **Figure S5. Manhattan plots of transcriptome-wide off-target RNA editing  
1401 analysis for different mxABE constructs.**

1402 Non-targeting mxABE construct (control) and E1, C3, C4, and M2 constructs  
1403 were transfected into HEK293T cells. mCherry-positive cells were sorted and  
1404 endogenous transcripts were analyzed by deep sequencing. The x and y axes  
1405 are proportionally enlarged with each Manhattan plot to make the axis legend  
1406 clear (n=3).

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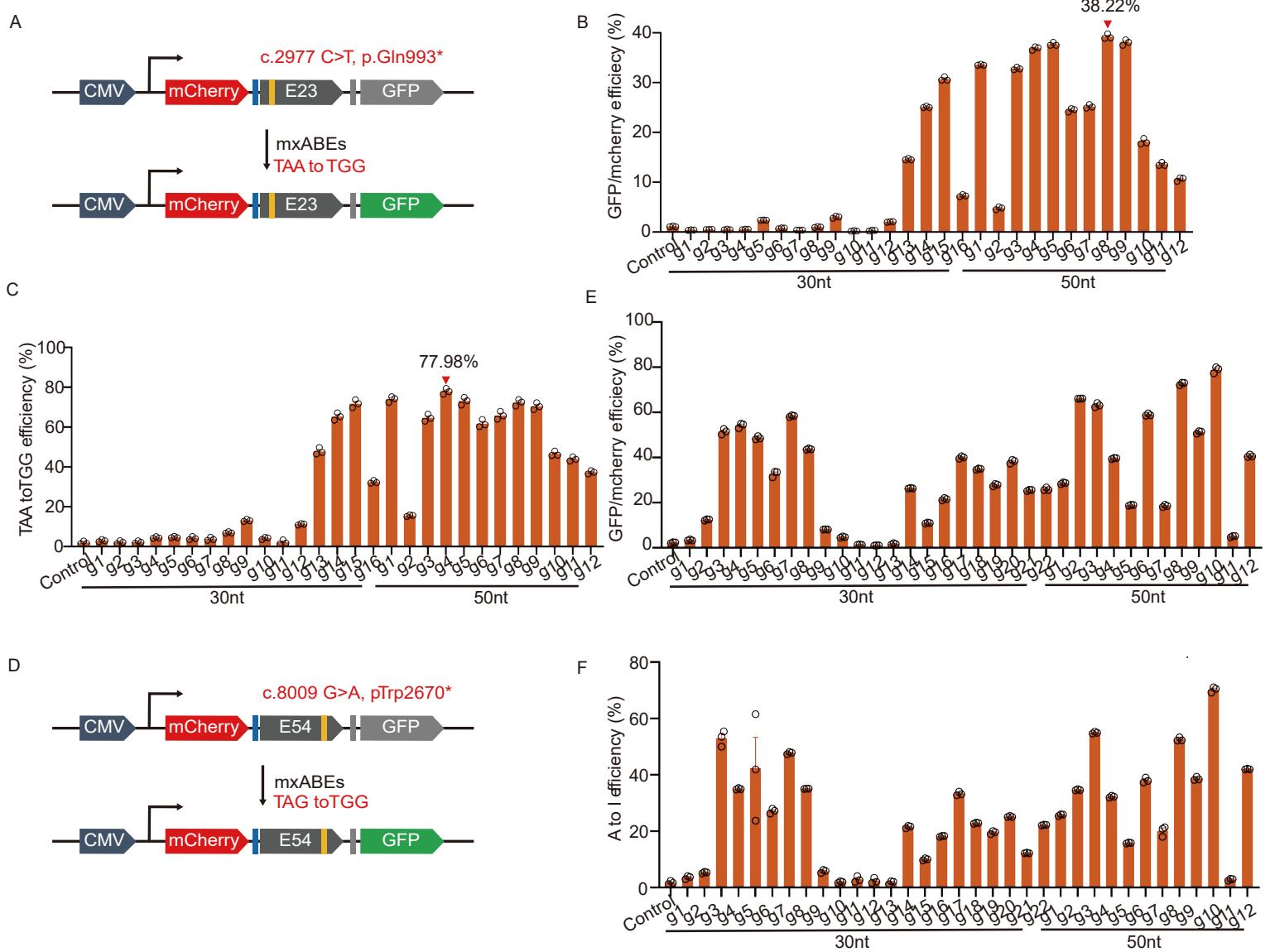
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Figure S6



1444 **Figure S6. mxABE-mediated A to I correction of other mutations in the**  
1445 **DMD gene.**

1446 Schematic diagram of reporter cassettes containing an mCherry sequence  
1447 fused with 2A peptide, mutant human *DMD* exon 23 (A) or exon 54 (D) and  
1448 ATG-removed GFP. Correction of the stop codon within the target sequence  
1449 would allow EGFP expression. Flow cytometry analysis of GFP expression in  
1450 HEK293T cells transfected with 28 gRNAs targeting exon 23 (B) and exon 54  
1451 (E). Deep sequencing of the reporter RNA after GFP rescue experiment in  
1452 exon 23 (C) and exon 54 (F). Data are represented as mean  $\pm$  SEM (n=3).

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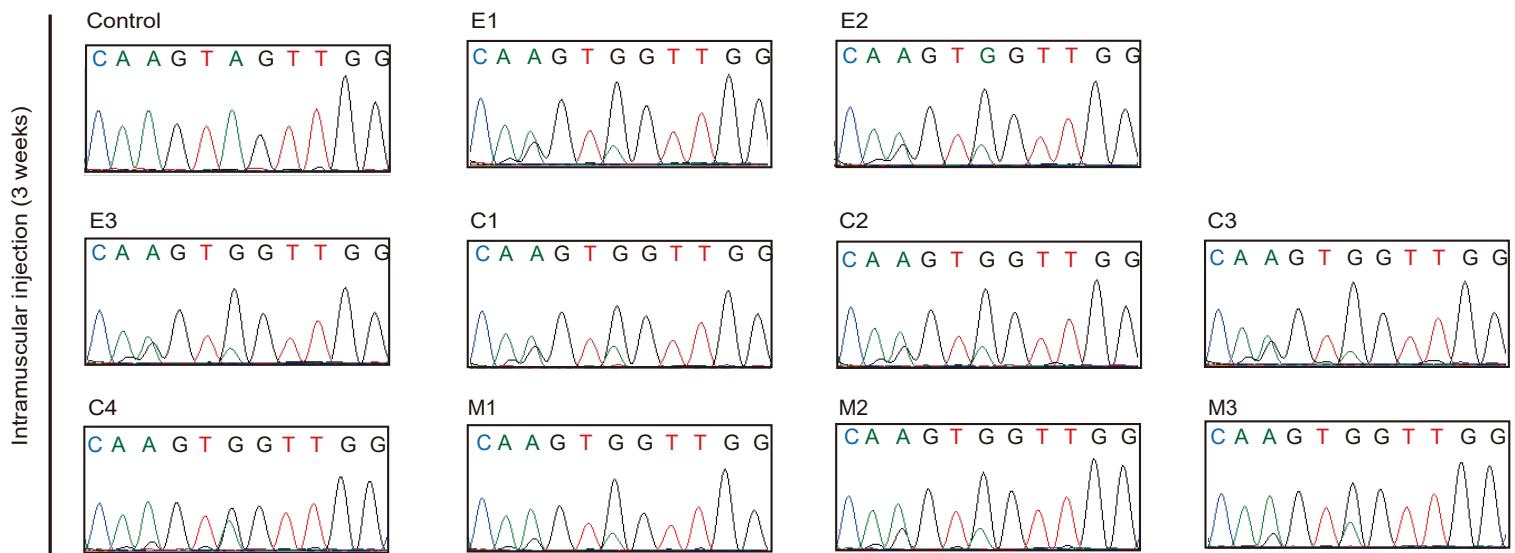
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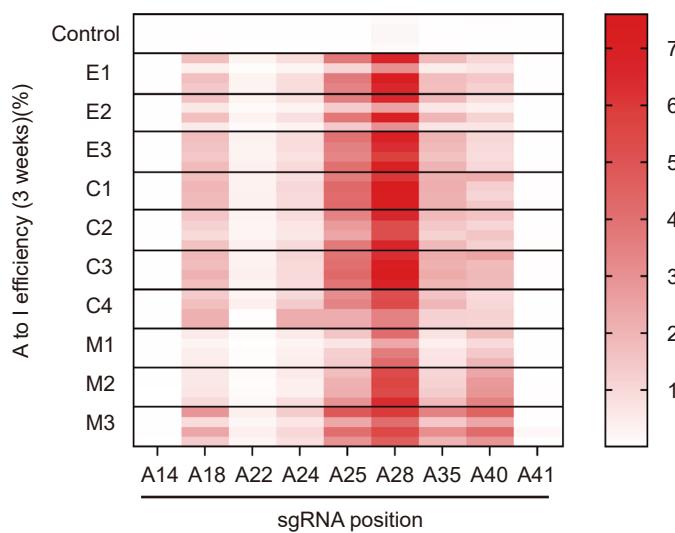
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Figure S7

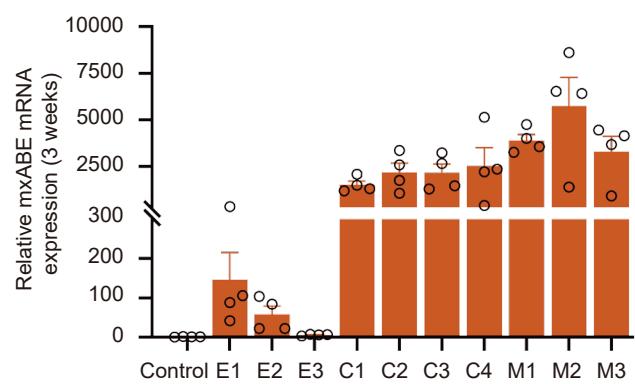
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C



1488 **Figure S7. *In vivo* DMD gene editing by IM injection with mxABEs at 3  
1489 weeks.**

1490 A. Representative Sanger sequencing trace of *DMD* transcripts 3 weeks after  
1491 IM injection with ten AAV9-mxAPE particles. B. Heat map indicates the A>G  
1492 edits in the vicinity of the target. C. qPCR analysis of mxAPE expression in TA  
1493 muscles 3 weeks after intramuscular injection. Values are shown as mean ±  
1494 SEM (n=4).

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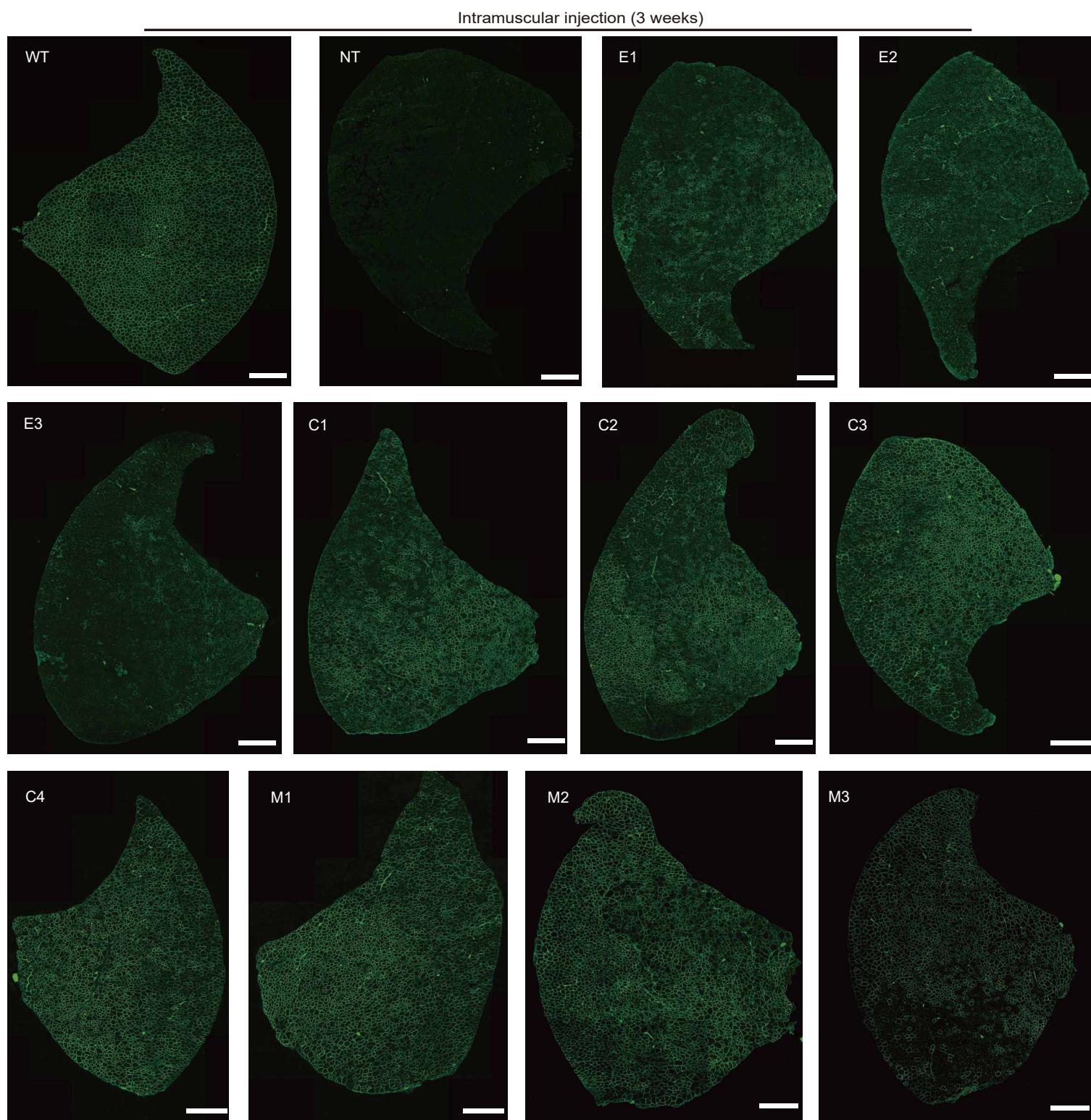
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Figure S8



1532 **Figure S8. Rescue of dystrophin expression following IM injection of**  
1533 **mxABEs in DMD<sup>E30mut</sup> mice.**

1534 Dystrophin immunohistochemistry of entire TA muscle. Control mice were  
1535 injected with saline. Dystrophin (Abcam, ab15277) is shown in green. Scale  
1536 bar, 500 µm.

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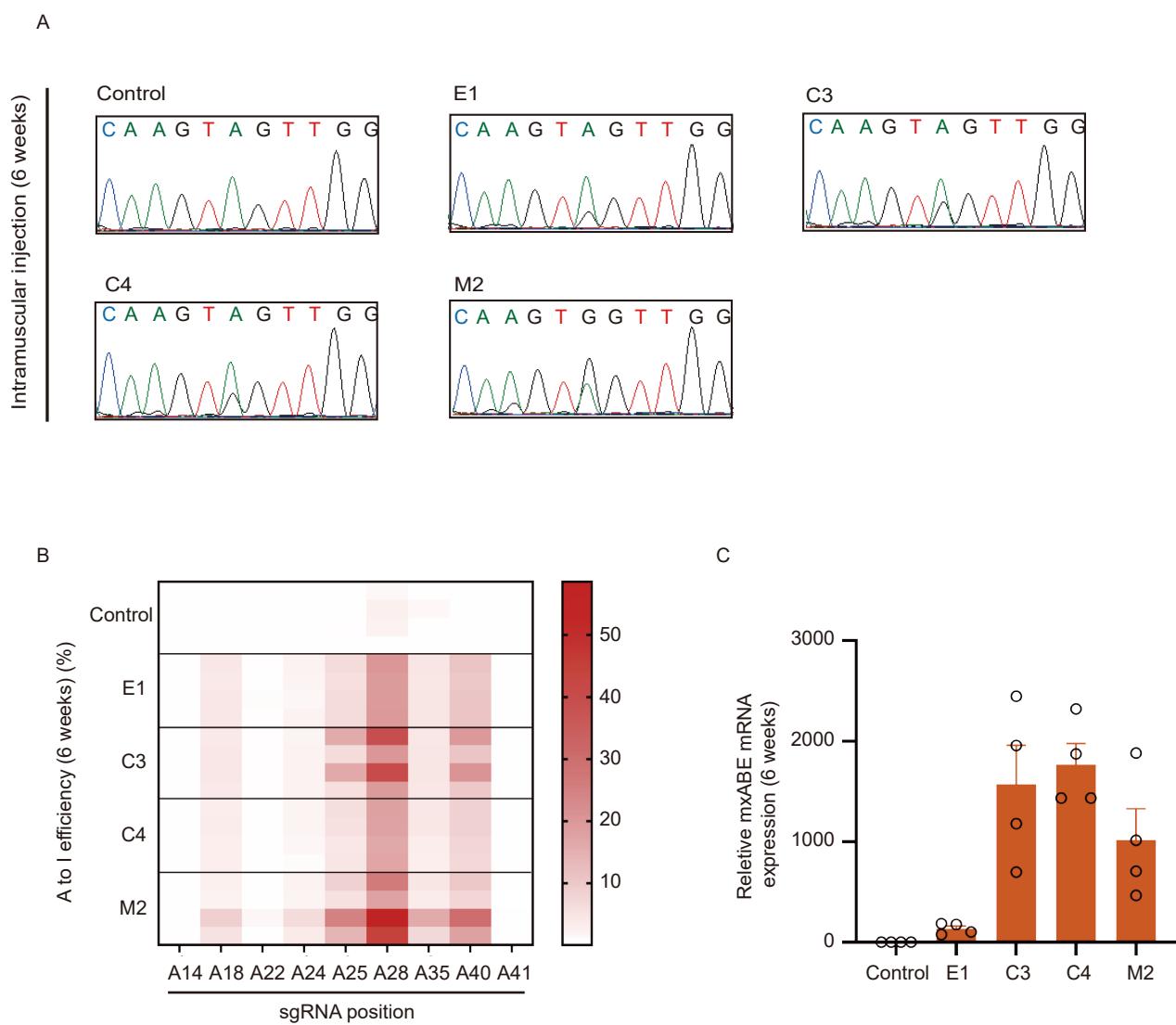
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Figure S9



1576 **Figure S9. In vivo DMD gene editing by IM injection with mxABEs after 6**  
1577 **weeks.**

1578 A. Representative Sanger sequencing trace of *DMD* transcripts 6 weeks after  
1579 IM injection with four AAV9-mxABE particles. B. Heat map indicates the A>G  
1580 edits in the vicinity of the target. C. qPCR analysis of mxABE expression in TA  
1581 muscles 6 weeks after intramuscular injection. Values are shown as mean ±  
1582 SEM (n=4).

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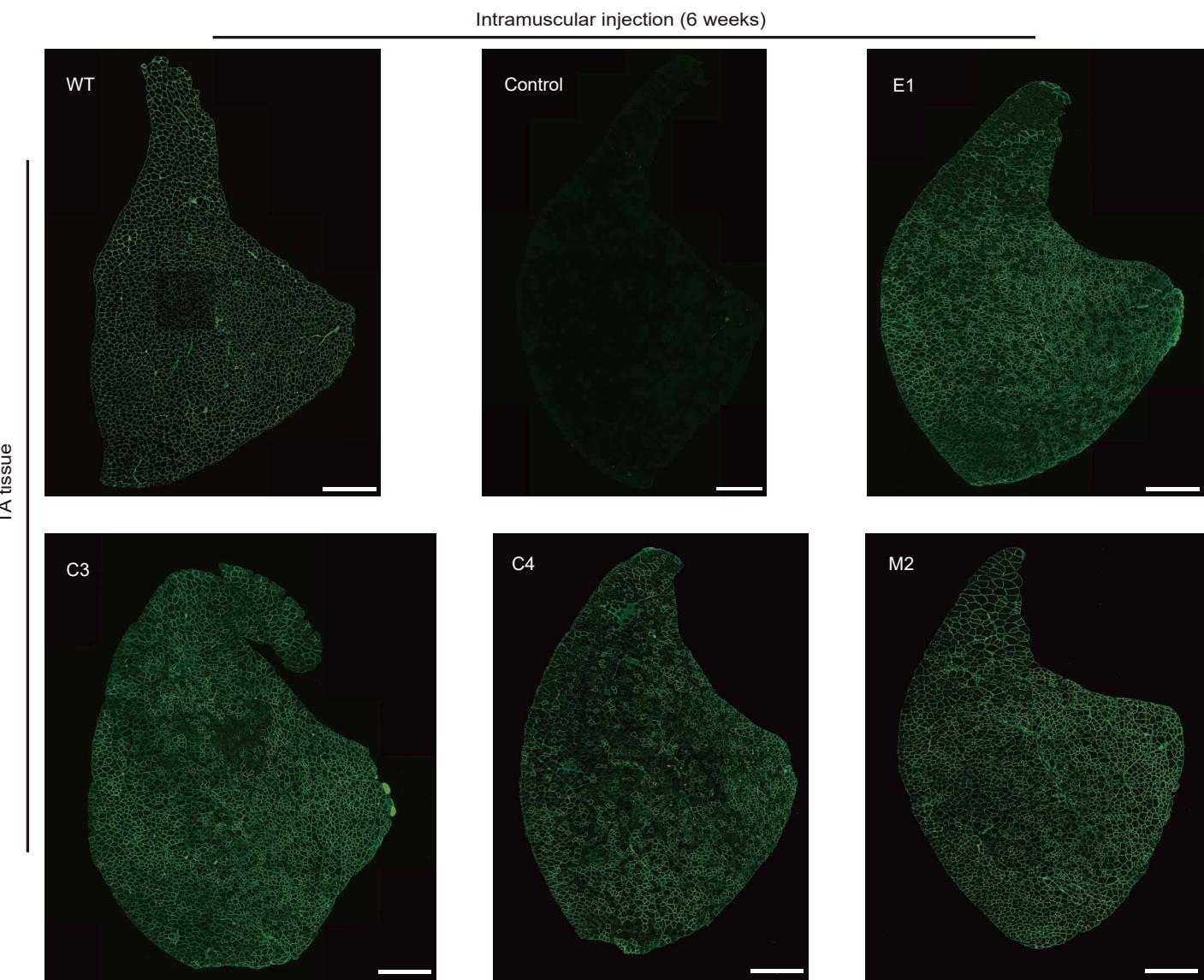
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Figure S10



1620 **Figure S10. Intramuscular AAV9 delivery of gene editing components**  
1621 **rescues dystrophin expression.**

1622 Immunohistochemistry of dystrophin in entire TA muscle 6 weeks after  
1623 intramuscular AAV9 delivery. Control mice were injected with saline.  
1624 Dystrophin (Abcam, ab15277) is showed in green. Scale bar, 500 µm.

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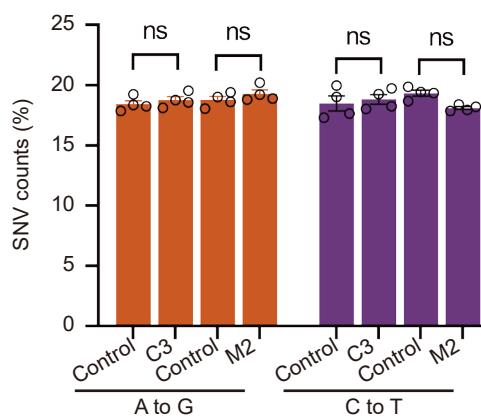
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Figure S11



1664 **Figure S11. Calculation of SNV counts by transcriptome-wide off-target**  
1665 **analysis *in vivo*.**

1666 The transcriptome-wide off-target effect of two AAV9-mxABEs (C3 and M2)  
1667 were analyzed *in vivo*. Control mice were injected with saline. Values are  
1668 shown as mean  $\pm$  SEM (n=4). NS, not statistically significant using unpaired  
1669 two-tailed Student's t tests (P<0.01).

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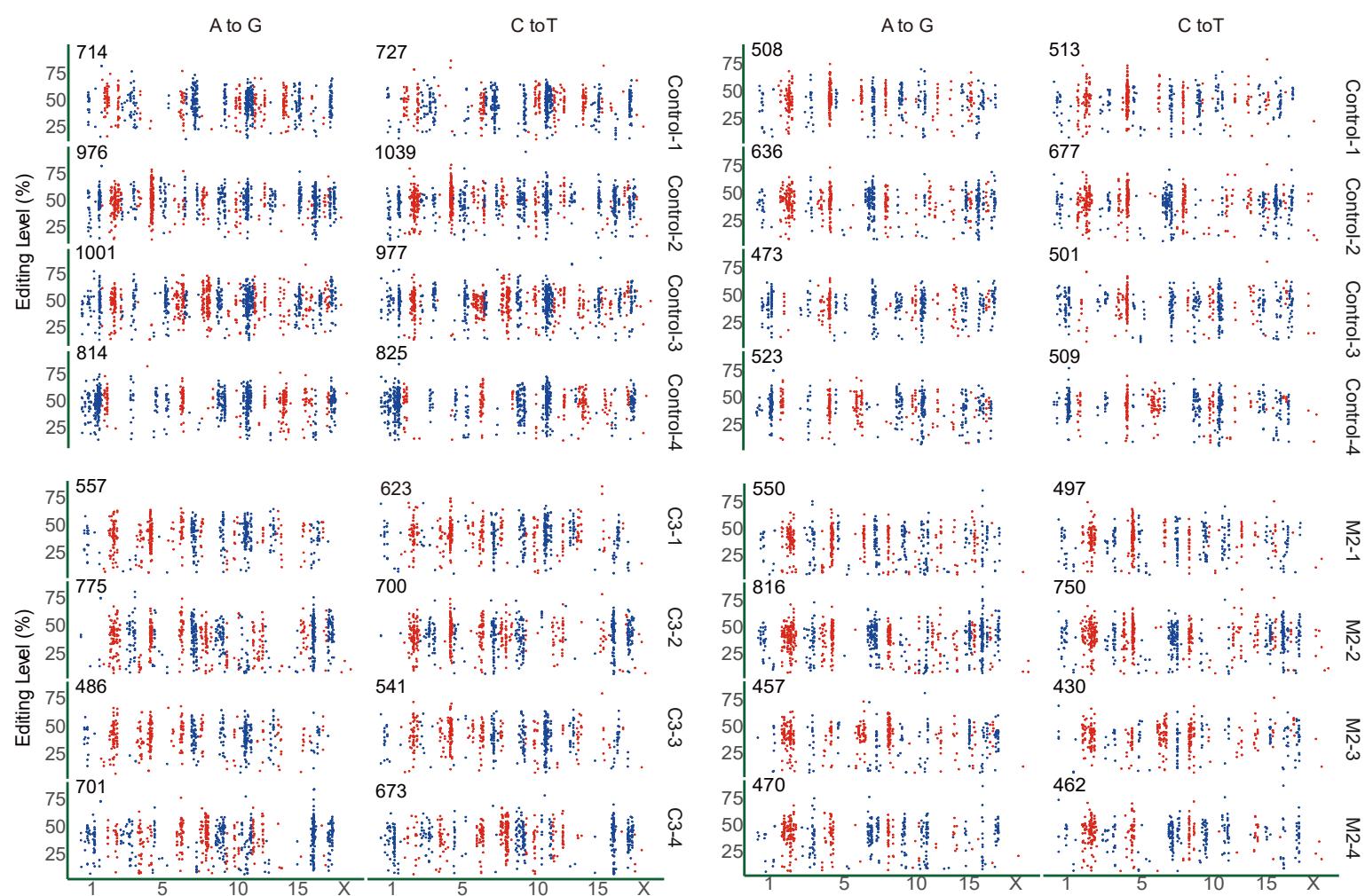
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Figure S12



1708 **Figure S12. Manhattan plots of transcriptome-wide off-target RNA editing**  
1709 **analysis *in vivo*.**

1710 Control mice were injected with saline. The x and y axes are proportionally  
1711 enlarged with each Manhattan plot to make the axis legend clear. n=4.

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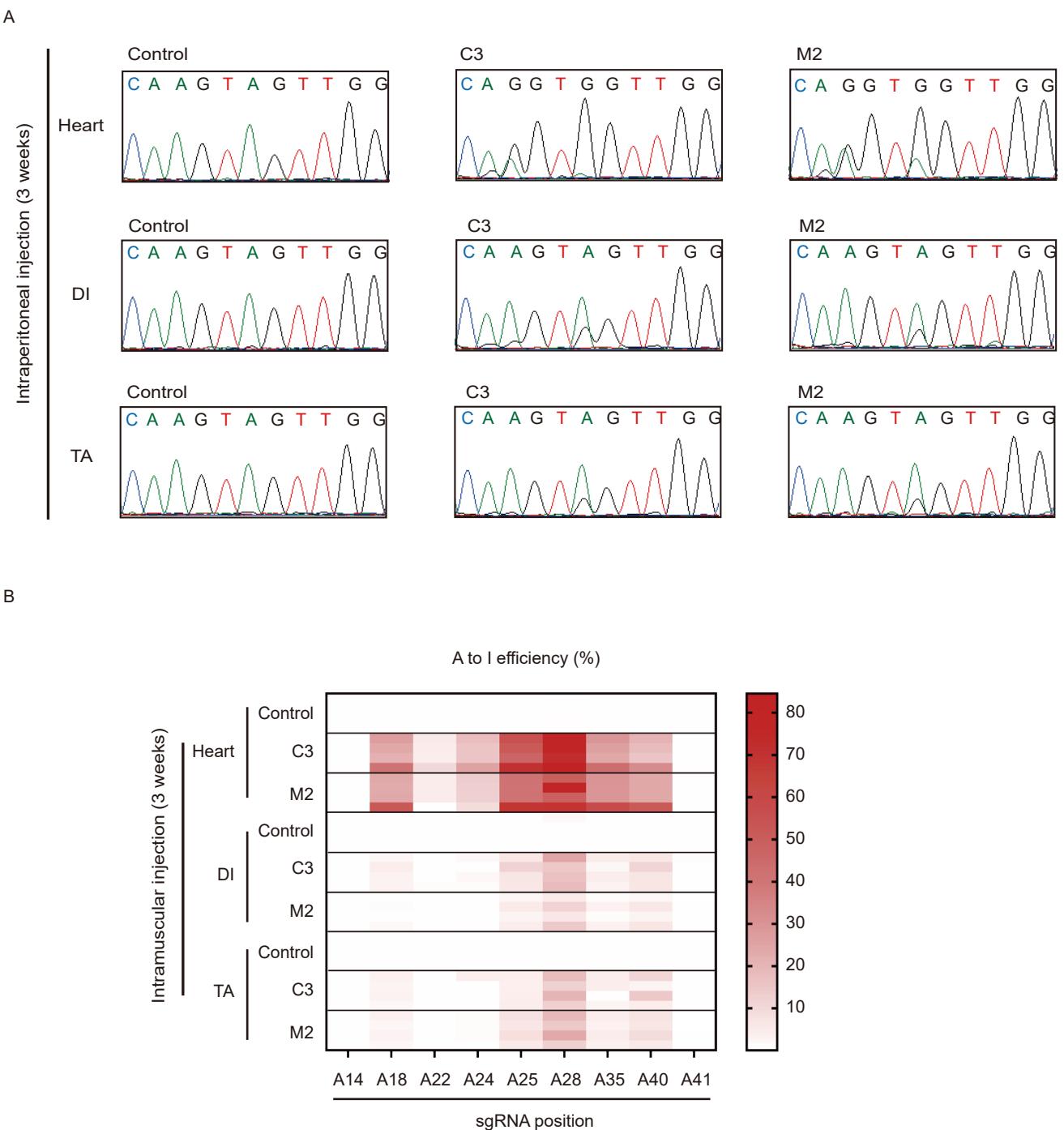
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Figure S13



1752 **Figure S13. Gene editing 3 weeks after systemic delivery of mxABEs in**  
1753 **DMD<sup>E30mut</sup> mice.**

1754 A. Representative Sanger sequencing trace of *DMD* transcripts in TA, DI, and  
1755 cardiac muscle 3 weeks after intraperitoneal (IP) injection with C3 and M2  
1756 particles. B. Heat map indicates the A>G edits in the vicinity of the target in  
1757 indicated tissues. n=4.

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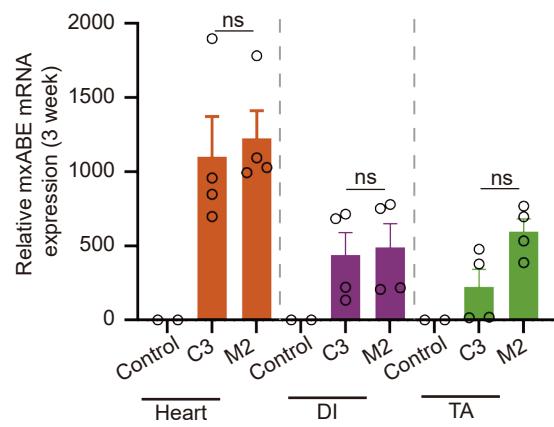
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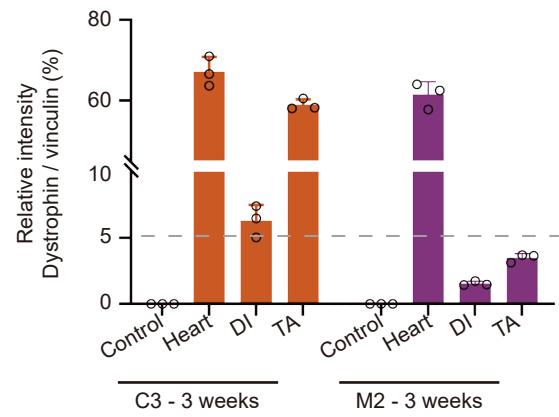
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Figure S14

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1796 **Figure S14. AAV9-mxABE expression and dystrophin protein levels after**  
1797 **systemic delivery in DMD<sup>E30mut</sup> mice.**

1798 A. qPCR analysis of mxABE expression in TA, DI, and heart muscles 3 weeks  
1799 after systemic delivery (n=4). B. Relative dystrophin (Sigma, D8168) intensity  
1800 was calibrated against a vinculin (CST, 13901S) internal control before  
1801 normalizing to the WT control. The results showed restoration of dystrophin in  
1802 TA, DI, and heart muscles of DMD<sup>E30mut</sup> mice 3 weeks after systemic delivery  
1803 of AAV9-packaged C3 and M2 (n=3). Values are shown as mean ± SEM. NS,  
1804 not statistically significant using unpaired two-tailed Student's t tests (P<0.01).

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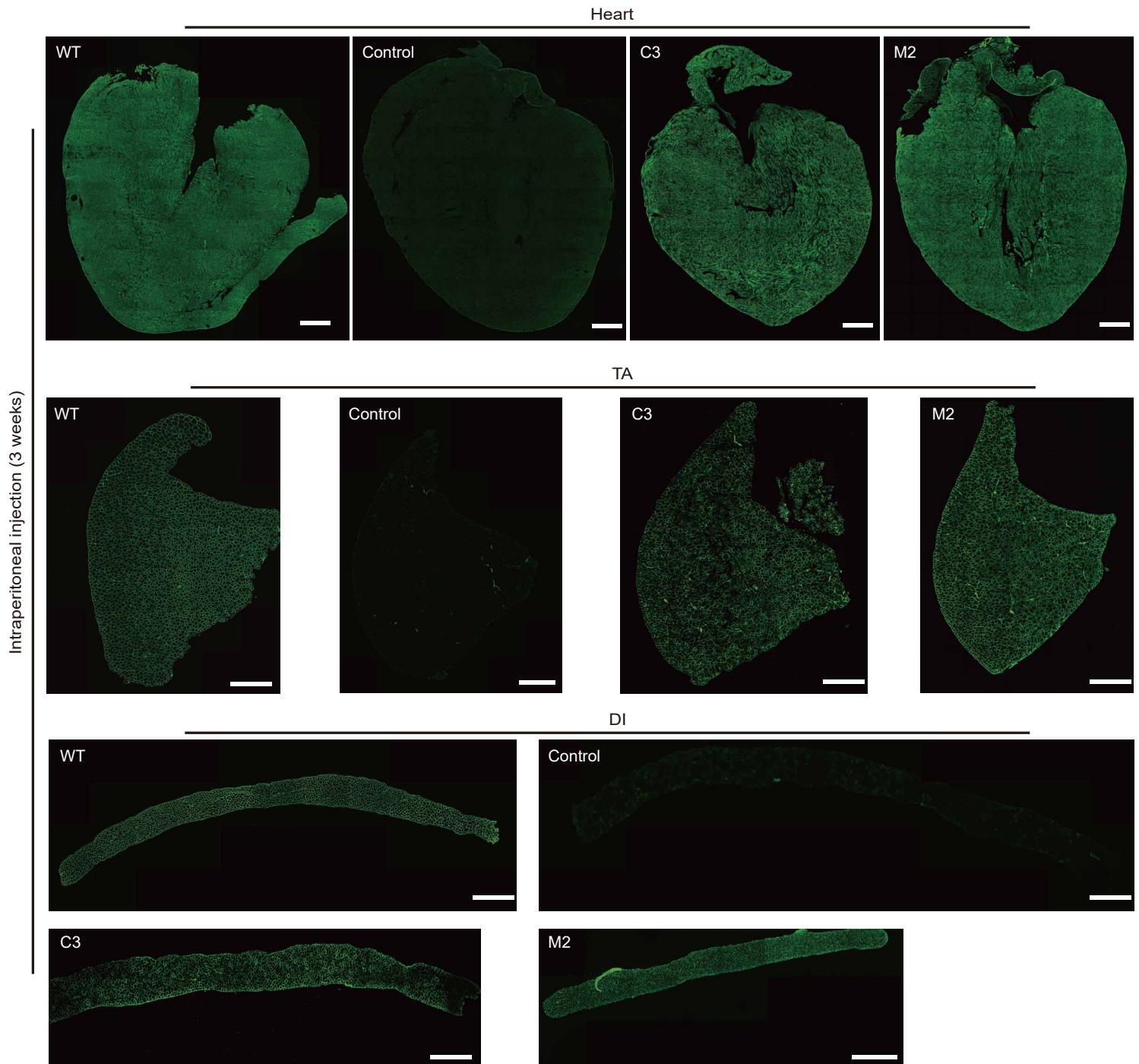
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Figure S15



1840 **Figure S15. Histological analysis of dystrophin expression 3 weeks after**  
1841 **systemic delivery of AAV9-mxABE in DMD<sup>E30mut</sup> mice.**

1842 Whole-muscle scanning of TA, DI, and heart of DMD<sup>E30mut</sup> mice 3 weeks after  
1843 systemic delivery AAV9-C3 and M2 particles. Control mice were injected with  
1844 saline. Dystrophin (Abcam, ab15277) is showed in green. Scale bar, 500 µm.

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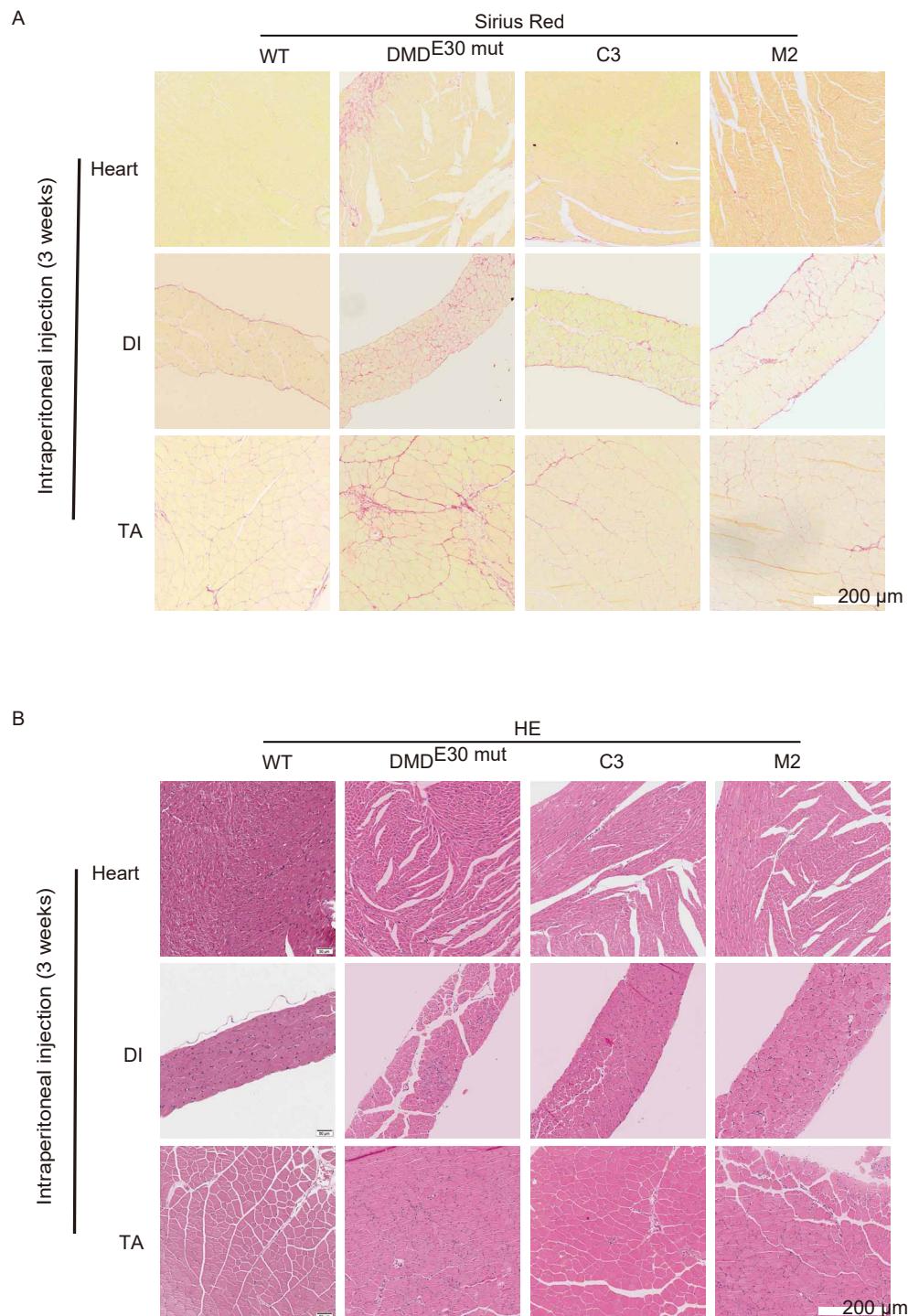
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Figure S16



1884 **Figure S16. Histological analysis 3 weeks after systemic delivery of**  
1885 **AAV9-mxAE.**

1886 Sirius red staining (A) and HE staining (A) of TA, DI, and heart of WT,  
1887 untreated DMD<sup>E30mut</sup> mice and DMD<sup>E30mut</sup> mice treated with AAV9-mxAE 3  
1888 weeks after systemic injection. n=4 for each group. Scale bars, 200 µm.

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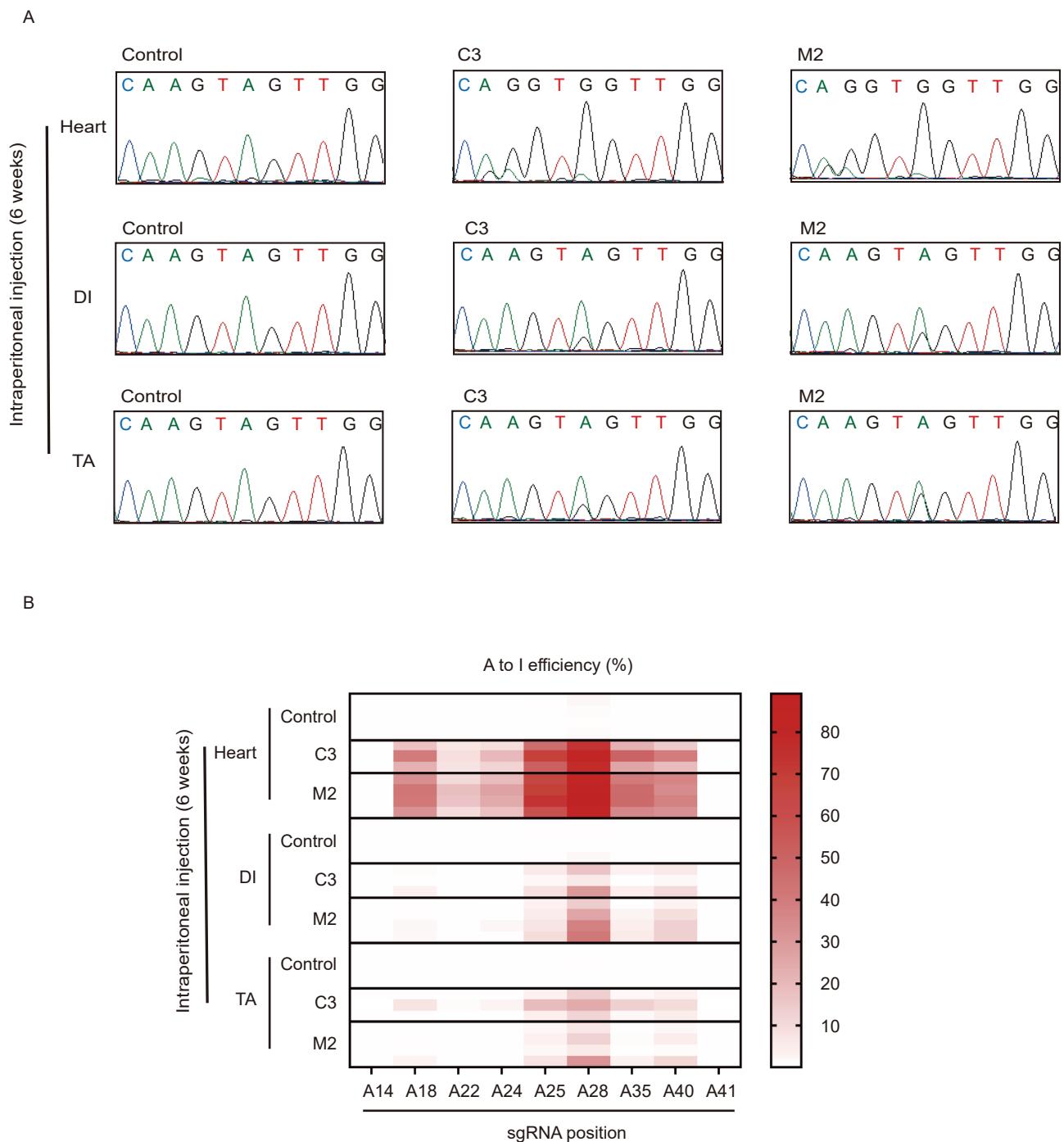
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Figure S17



1928 **Figure S17. Base editing efficiency achieved by systemic delivery of**  
1929 **mxAPE after 6 weeks.**

1930 A. Representative Sanger sequencing trace of *DMD* transcripts in TA, DI, and  
1931 heart tissues 6 weeks after intraperitoneal (IP) injection with C3 and M2  
1932 particles. B. Heat map shows base editing rates in tissues indicated after IP  
1933 injection with C3 (n=3) and M2 (n=4).

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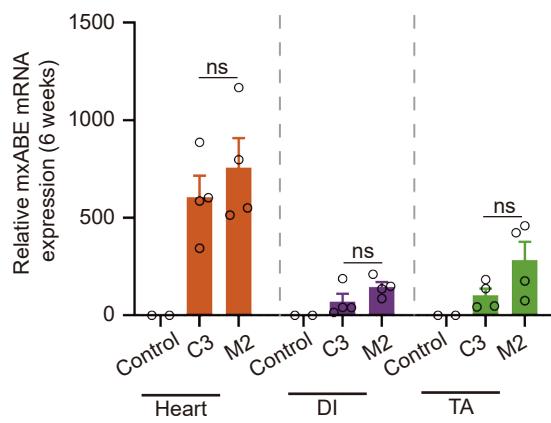
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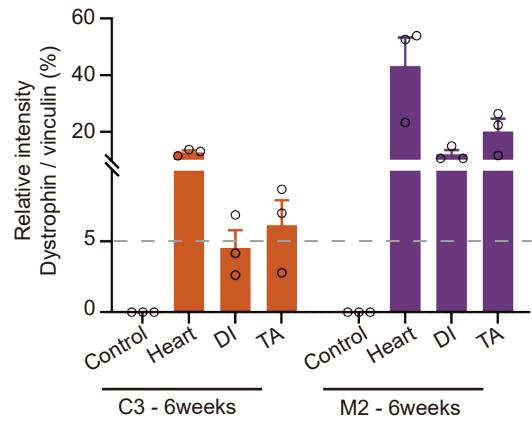
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Figure S18

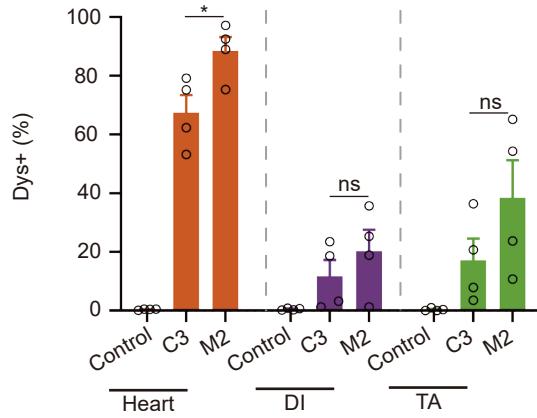
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1972 **Figure S18. mxABE expression and dystrophin restoration level after**  
1973 **6-week administration of AAV-mxABE systemically.**

1974 A. qPCR analysis of mxABE expression in TA, DI, and heart muscles 6 weeks  
1975 after systemic delivery (n=4). B. Relative dystrophin (Sigma, D8168) intensity  
1976 was calibrated against a vinculin (CST, 13901S) internal control before  
1977 normalizing to the WT control. The results showed restoration of dystrophin in  
1978 TA, DI, and cardiac muscles of DMD<sup>E30mut</sup> mice 6 weeks after systemic delivery  
1979 of C3 and M2 AAV-mxABE (n=3). C. Percentage of Dys+ tissue area in TA, DI,  
1980 and heart from treated and untreated DMD<sup>E30mut</sup> mice (n=4). Values are shown  
1981 as mean ± SEM. NS, not statistically significant using unpaired two-tailed  
1982 Student's t test (P<0.01).

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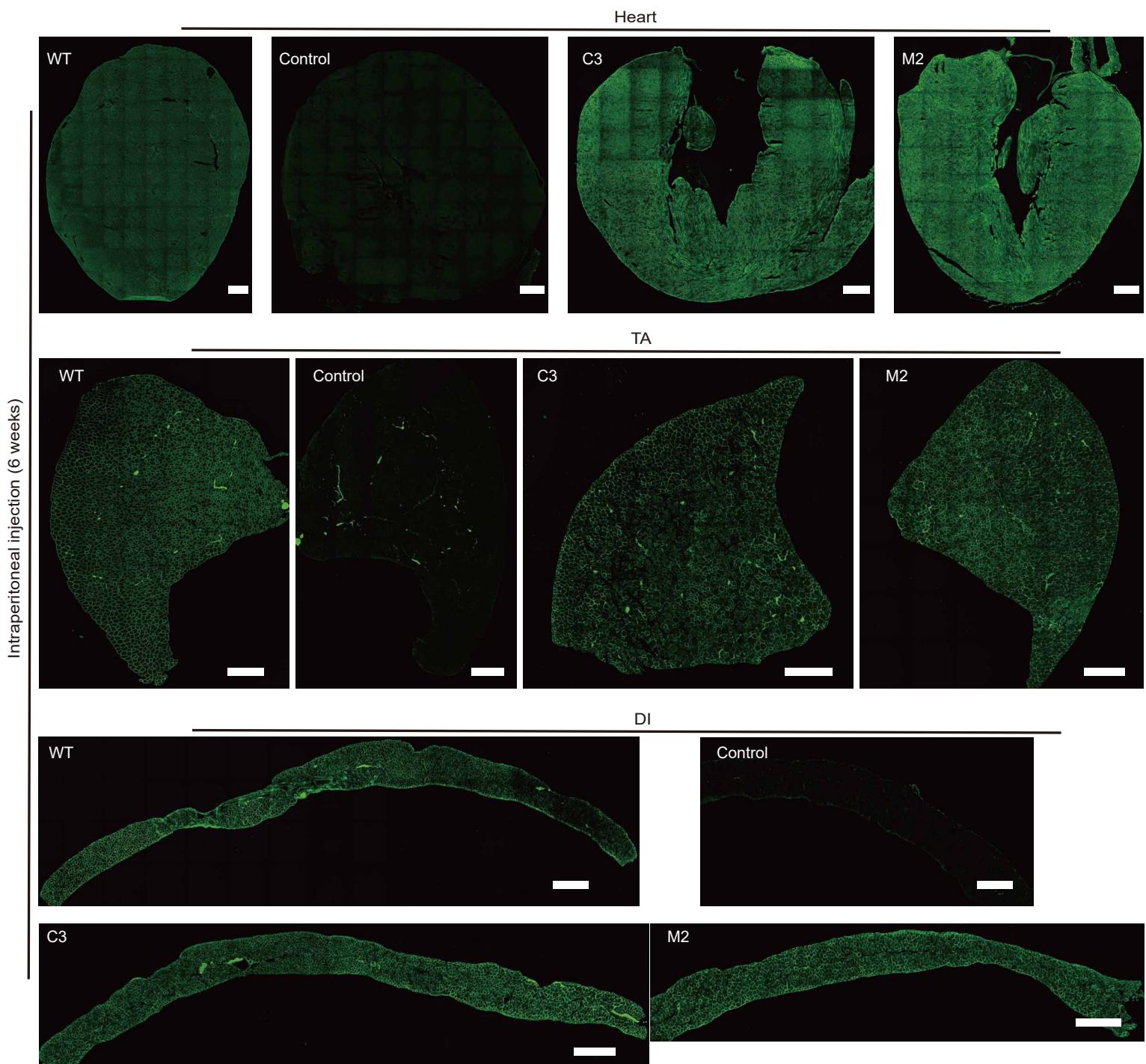
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Figure S19



2016 **Figure S19. Immunostaining of dystrophin in heart, TA, and DI tissues 6**  
2017 **weeks after systemic administration of AAV-mxABE.**

2018 Whole-muscle scanning of TA, DI, and heart muscle of DMD<sup>E30mut</sup> mice 6  
2019 weeks after systemic delivery of AAV9-C3 and M2 particles. Control mice were  
2020 injected with saline. Dystrophin (Abcam, ab15277) is shown in green. Scale  
2021 bar, 500 µm. Images shown in both Figure 6C and Figure S19 were obtained  
2022 from the same tissue at 20x magnification. Figure 6C showed the local region  
2023 staining image rather than the reconstituted whole-tissue scanning image in  
2024 Figure S19.

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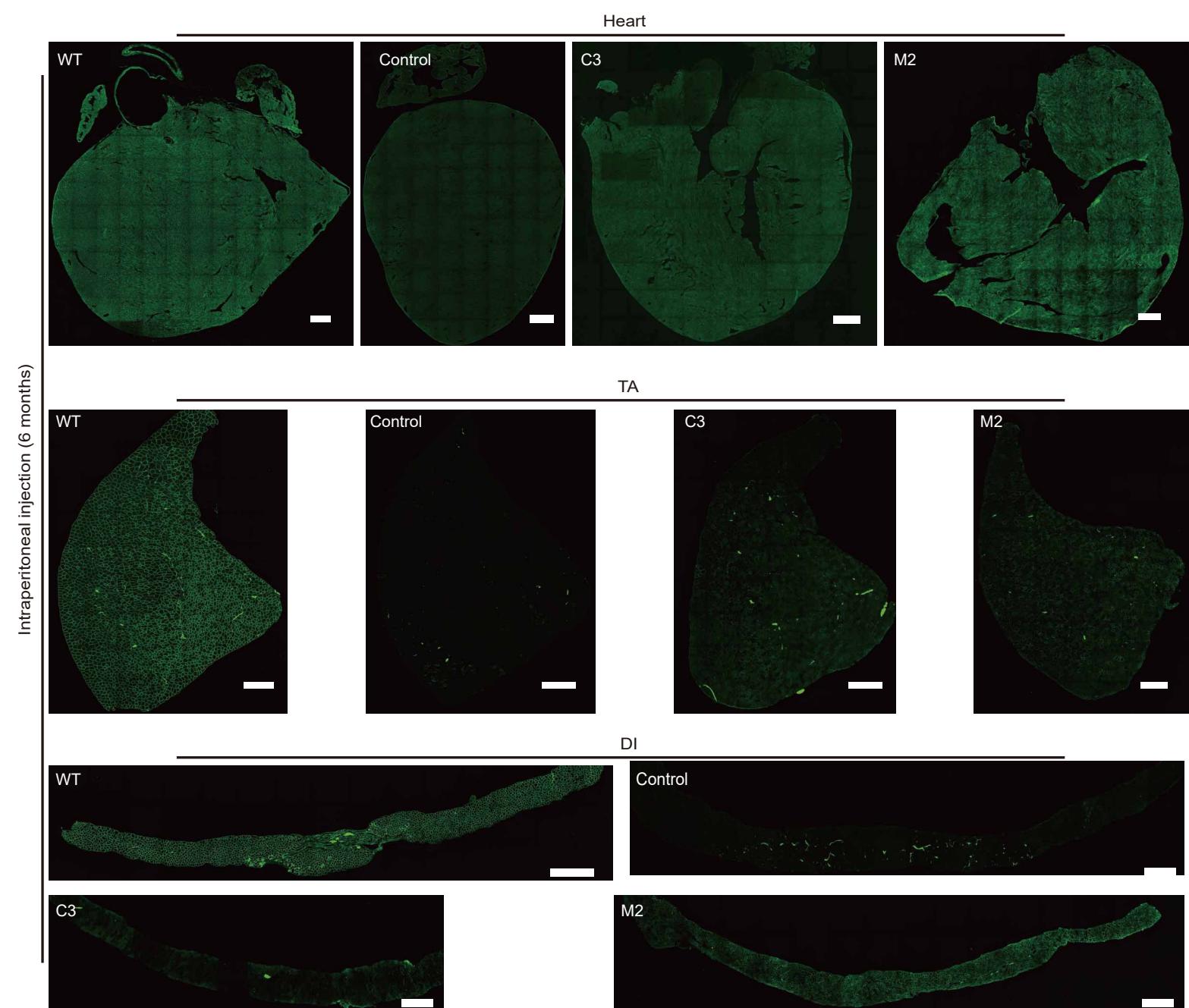
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Figure S20



2060   **Figure S20. Immunostaining of dystrophin in heart, TA, and DI 6 months**  
2061   **after systemic administration of AAV-mxAbe.**

2062   Whole-muscle scanning of TA, DI, and heart muscle of DMD<sup>E30mut</sup> mice 6  
2063   months after systemic delivery of AAV9-C3 and M2 particles. Control mice  
2064   were injected with saline. Dystrophin (Abcam, ab15277) is shown in green.  
2065   Scale bar, 500 μm.

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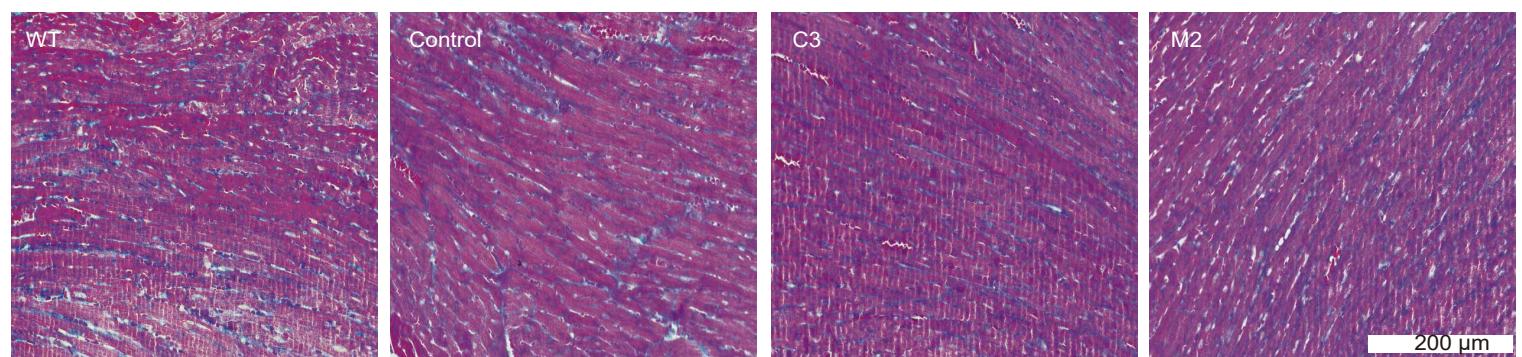
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Figure S21



2104      **Figure S21. Collagen staining shows no obvious fibrosis in treated and**  
2105      **untreated hearts 6 months after systemic administration of AAV-mxABE.**  
2106      Scale bar, 200  $\mu$ m.

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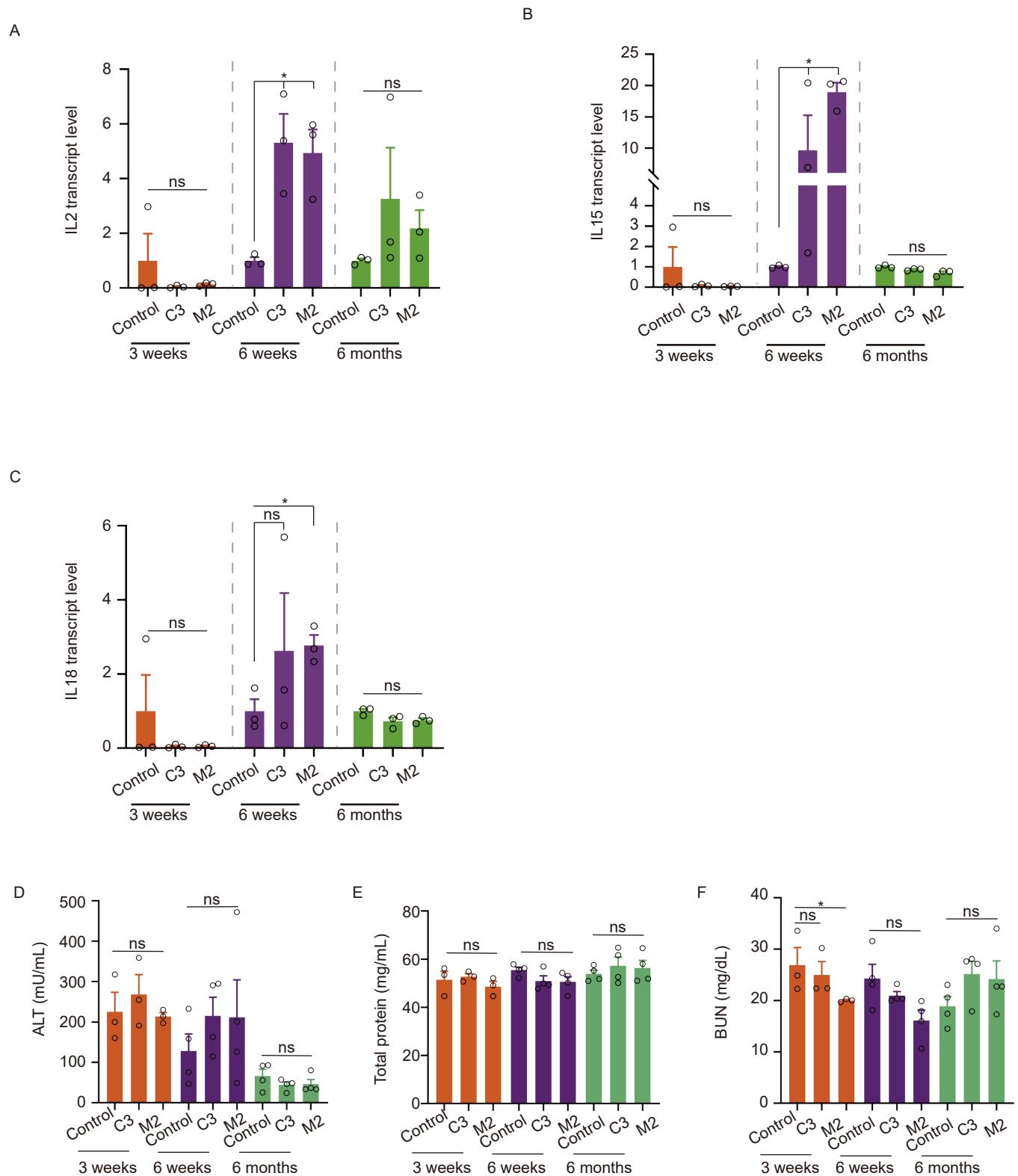
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Figure S22



2148 **Figure S22. Host immune response to AAV-mxAbe treatment indicates**  
2149 **no overt toxicity after 6 months.**

2150 (A-C) IL2, IL15, and IL18 transcript levels at baseline for untreated mice, 3  
2151 weeks, 6 weeks, and 6 months for treated mice (n=3). (D-F) No obvious  
2152 change in ALT, total protein and BUN level after AAV-mxAbe treatment (n=3).  
2153 Significance is indicated by asterisk ( $P < 0.05$ ). NS represents not statistically  
2154 significant using unpaired two-tailed Student's t test.

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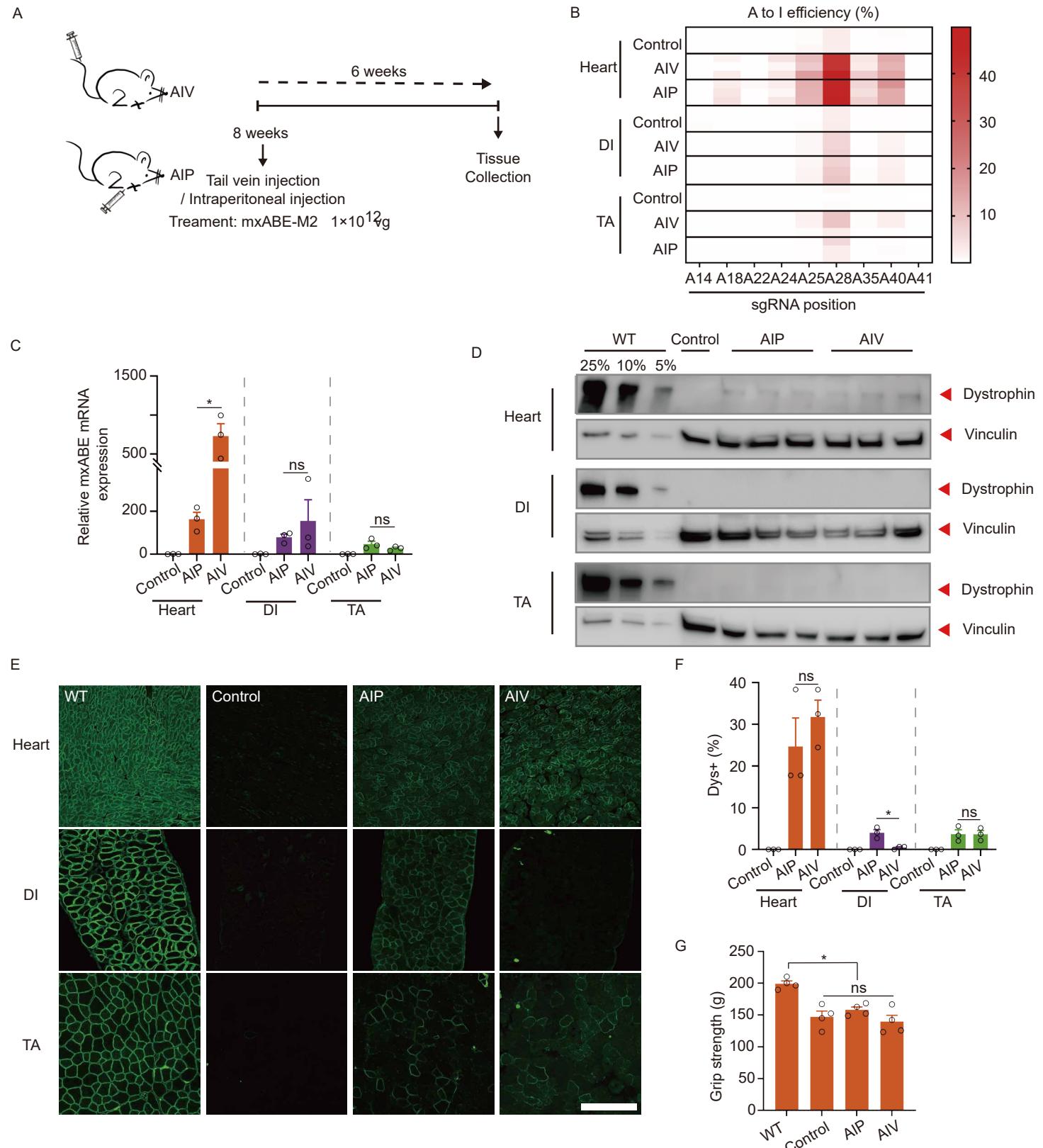
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Figure S23



2192 **Figure S23. Systemic delivery of AAV-mxABE rescued dystrophin  
2193 deficiency in adult DMD<sup>E30mut</sup> mice.**

2194 A. Experimental design for mxABE administration and therapeutic analysis in  
2195 adult DMD<sup>E30mut</sup> mice. Both intravenous (AIV group) and intraperitoneal (AIP  
2196 group) injection of AAV-mxABE were evaluated. B. Heatmap shows base  
2197 editing rate in heart, DI, and TA muscles (n=3). C. mxABE expression level in  
2198 heart, DI, and TA muscles. D. Dystrophin restoration level analyzed by western  
2199 blot in heart, DI, and TA muscles of treated and untreated mice. E. Tissue  
2200 section immunostaining for dystrophin in heart, DI, and TA muscles of treated  
2201 and untreated mice. Scale bar, 200  $\mu$ m. F. Quantification of Dys+  
2202 immunostaining in (E) (n=3). G. Forelimb grip strength was measured in WT,  
2203 DMD<sup>E30mut</sup> mice, and DMD<sup>E30mut</sup> mice treated with AAV9-M2 (n=4).  
2204 Significance is indicated by asterisk ( $P < 0.05$ ). NS represents not statistically  
2205 significant using unpaired two-tailed Student's t test.

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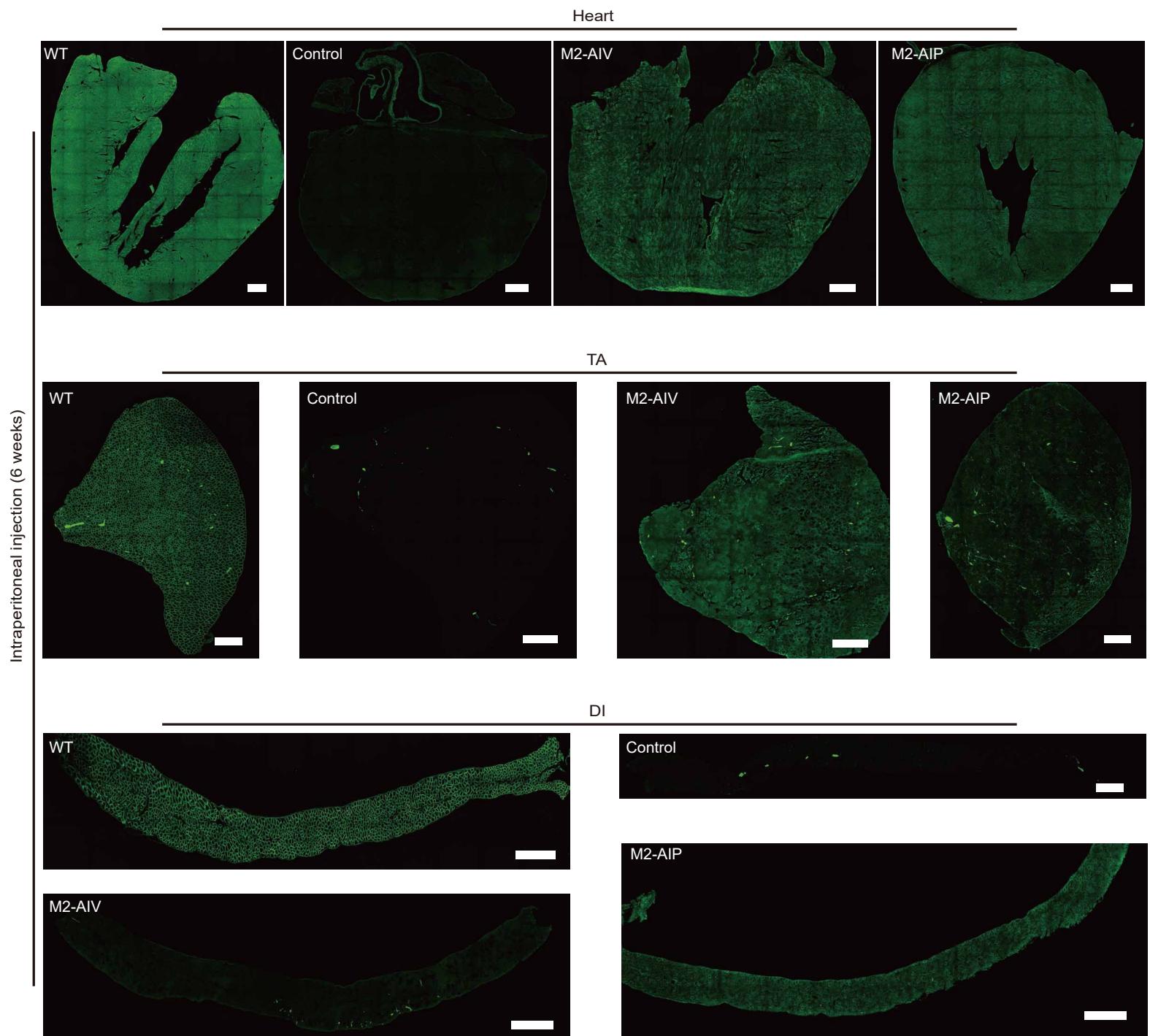
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Figure S24



2236 **Figure S24. Immunostaining of dystrophin in adult DMD<sup>E30mut</sup> mice 6  
2237 weeks after systemic injection of AAV-mxAbe.**

2238 Scale bar, 500 μm.

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**Supplementary Table S1: Target sgRNA and primer sequence.**

Experiment	Primer name	Primer sequence (5'-3')
sgRNA for generation DMD <sup>E30mut</sup> model	DMD-T7sgRNA1	CTTCTTCTATTTAGGCTGTG
	DMD-T7sgRNA2	AGCTTATATCACTGACAAGG
Genotyping of DMD <sup>E30mut</sup> mice	DMD <sup>E30mut</sup> -2333F	ATTCATATAGGGCTTCAGTCC
	DMD <sup>E30mut</sup> -2333R	CATCTGTTTAATAGTGTGCAT
RT-PCR primer flanking exon 30	RT-DMD-358F	AATCAGATTGCTATTGGCACA
	RT-DMD-358R	CCCTTGGTGGCATCCTT
RNA base editing sgRNA for exon 30	30nt-g1	GCTGCCAACCACTTGTCAATGAATGTGAGG
	30nt-g2	AGCTGCCAACCACTTGTCAATGAATGTGAG
	30nt-g3	TGCAATATAAGCTGCCAACCACTTGTCAAT
	30nt-g4	CTGCAATATAAGCTGCCAACCACTTGTCAA
	30nt-g5	AAGCTGCCAACCACTTGTCAATGAATGTGA
	30nt-g6	TAAGCTGCCAACCACTTGTCAATGAATGTG
	30nt-g7	ATAAGCTGCCAACCACTTGTCAATGAATGT
	30nt-g8	TATAAGCTGCCAACCACTTGTCAATGAATG
	30nt-g9	ATATAAGCTGCCAACCACTTGTCAATGAAT
	30nt-g10	AATATAAGCTGCCAACCACTTGTCAATGAA
	30nt-g11	CAATATAAGCTGCCAACCACTTGTCAATGA
	30nt-g12	GCAATATAAGCTGCCAACCACTTGTCAATG
	50nt-g1	TGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTCCTGGA
	50nt-g2	CTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTCCTGG
	50nt-g3	CCACCTTGTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGG
	50nt-g4	TCCACCTTGTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAG
	50nt-g5	TCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTCCTG
	50nt-g6	GTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTCCT
	50nt-g7	TGTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTCC

	50nt-g8	TTGCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACTC
	50nt-g9	CTTGTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGACT
	50nt-g10	CCTTGTCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGAC
	50nt-g11	ACCTTGCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGGA
	50nt-g12	CACCTTGCTGCAATATAAGCTGCCAACCACTTGTCAATGAATGTGAGGG
RT-PCR primer for	RT-rep765F	CCACAACGAGGACTACACCA
Reporter	RT-rep765R	TCCTTGAAGTCGATGCCCTT
qPCR primer	GAPDH-119F	TCAACGACCCCTTCATTGACC
	GAPDH-119R	CTTCCCGTTGATGACAAGCTTC
	Cas13X-140F	GCCATGCAGAAATAATATCTCGG
	Cas13X-140R	CATTCTCCTTCAGCCTAAACCC
RNA base editing sgRNA for exon 23	50nt-g1	CTGAGATAGTATAGGCCACTccATTGCTCTTGCAAGAGAACTTTGTAAAGC
	50nt-g2	GCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTGTAAAG
	50nt-g3	TGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTGTAAA
	50nt-g4	GTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTGTAA
	50nt-g5	GGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTGTAA
	50nt-g6	TGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTGT
	50nt-g7	GTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTTG
	50nt-g8	CACAGTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAAC
	50nt-g9	TCACAGTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAA
	50nt-g10	AGTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTTT
	50nt-g11	CAGTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACTT
	50nt-g12	ACAGTGGTGCTGAGATAGTATAGGCCACTCCATTGCTCTTGCAAGAGAACT
	30nt-g1	TAGTATAGGCCACTCCATTGCTCTTGCAAG
	30nt-g2	ATAGTATAGGCCACTCCATTGCTCTTGCAAG
	30nt-g3	GATAGTATAGGCCACTCCATTGCTCTTGCAAG
	30nt-g4	AGATAGTATAGGCCACTCCATTGCTCTTGCAAG
	30nt-g5	GAGATAGTATAGGCCACTCCATTGCTCTTGCAAG
	30nt-g6	TGAGATAGTATAGGCCACTCCATTGCTCTTGCAAG

	30nt-g7	CTGAGATAGTATAGGCCACTCCATTGCTCT
	30nt-g8	GCTGAGATAGTATAGGCCACTCCATTGCTC
	30nt-g9	TGCTGAGATAGTATAGGCCACTCCATTGCT
	30nt-g10	GTGCTGAGATAGTATAGGCCACTCCATTGC
	30nt-g11	GGTGCTGAGATAGTATAGGCCACTCCATTG
	30nt-g12	AGTATAGGCCACTCCATTGCTCTGCAGAG
	30nt-g13	TAGGCCACTCCATTGCTCTGCAGAGAACT
	30nt-g14	ATAGGCCACTCCATTGCTCTGCAGAGAAC
	30nt-g15	TATAGGCCACTCCATTGCTCTGCAGAGAA
	30nt-g16	GTATAGGCCACTCCATTGCTCTGCAGAGA
RNA base editing sgRNA for exon 54	50nt-g1	TGTAATTACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTC
	50nt-g2	GTAATTACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCT
	50nt-g3	CACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTATC
	50nt-g4	TCACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTAT
	50nt-g5	TTCAACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTA
	50nt-g6	ATTCAACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTT
	50nt-g7	AATTCAACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGT
	50nt-g8	TAATTCAACACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTG
	50nt-g9	ATACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTATCA
	50nt-g10	TACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTATCAT
	50nt-g11	ACCTTTATGAATGCTTCTCCAAGAGGCATTGATATTCTCTGTTATCATG
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	30nt-g1	CACACCTTTATGAATGCTTCTCCAAGAGGC
	30nt-g2	TCACACCTTTATGAATGCTTCTCCAAGAGGC
	30nt-g3	TTCACACCTTTATGAATGCTTCTCCAAGAGGCAGA
	30nt-g4	ATACCTTTATGAATGCTTCTCCAAGAGGC
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	30nt-g6	ACCTTTATGAATGCTTCTCCAAGAGGCAT
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	30nt-g8	TTCTCCAAGAGGCATTGATATTCTCTGTTA
	30nt-g9	TATGAATGCTTCTCCAAGAGGCATTGATAT
	30nt-g10	TTATGAATGCTTCTCCAAGAGGCATTGATA
	30nt-g11	TTTATGAATGCTTCTCCAAGAGGCATTGAT
	30nt-g12	TTTTATGAATGCTTCTCCAAGAGGCATTGA
	30nt-g13	CTTTTATGAATGCTTCTCCAAGAGGCATTG
	30nt-g14	CCTTTTATGAATGCTTCTCCAAGAGGCATT
	30nt-g15	CTTCTCCAAGAGGCATTGATATTCTCTGTT
	30nt-g16	GCTTCTCCAAGAGGCATTGATATTCTCTGT
	30nt-g17	TGCTTCTCCAAGAGGCATTGATATTCTCTG
	30nt-g18	AATGCTTCTCCAAGAGGCATTGATATTCTC
	30nt-g19	GAATGCTTCTCCAAGAGGCATTGATATTCT
	30nt-g20	TGAATGCTTCTCCAAGAGGCATTGATATTTC
	30nt-g21	ATGAATGCTTCTCCAAGAGGCATTGATATT
	30nt-g22	ATGCTTCTCCAAGAGGCATTGATATTCTCT

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2302     **Supplemental sequences AAV constructs' sequences of C1 to C4, E1 to**  
2303     **E4 and M1 to M3**

2304     C1 to C4 AAV constructs are CBh-driven mxAPE. CBh promoter sequence is  
2305     highlighted in **red**

2306     E1 to E4 AAV constructs are EFS-driven mxAPE. EFS promoter sequence is  
2307     highlighted in **green**

2308     M1 to M3 AAV constructs are MHCK7-driven mxAPE. MHCK7 promoter  
2309     sequence is highlighted in **blue**

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2313 CCTGCAGGCAGCTGCGCGCTCGCTCGCTCACTGAGGCCGCCGGCG  
2314 TCGGGCGACCTTGGTCGCCCGGCCTCAGTGAGCGAGCGAGCGCA  
2315 GAGAGGGAGTGGCCAACTCCATCACTAGGGGTTCTGCAGGCCTCTAGAg  
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2321 ACTCctGCTGGAGCAGCCCCCGATTGTGGGTGATTACAGCTTTTTc**cg**  
2322 ttacataacttacggtaaatggcccgcctggctgaccgcacgcacccgcattgacgtcaatagta  
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2332 agggtaaggatggtggtgggttt  
2333 **agttGGaccggGCCACCATGGCCCCAAGAAGAAGaGGAAGGTGCTGAGC**  
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2335 TGCTGCTGTTAGAGACATCCTGGCCCAGCTGGGAAGAATCCCCGCCGA  
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2343 CCTCGTGCTGCTGAGCCTGCAGGGGAAGGGCGACGATGCCATGCCAA  
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2345 GTGACCGATAAGGATAACCACGTGTTCCCTGCCCGCTCGTGGAGC

2346 AGCACGGCATCGGCAGAAAGGCCTCAAGCAGCGGATCGATGGACGGG  
2347 TGAAGCACGTGCGGGCGTGTGGAGAAGAAGAACGCCACCAATG  
2348 AAATGACCCTGCACGAGAAGGCCAGAGACATCCTGCAGTACGTGAACGA  
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2352 AGACCAGCACCATCAATGAGATGCACCAAGGTGGTGTGCACCAGATCCT  
2353 GAACCGGCTGTGCAGAATCGCGACCAGAAGCTGTACGATTACGTGGGA  
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2355 AGGAGCACATCAGCATCCGGAGAGGATTCTGAGAAAGAAGTTCTGGTA  
2356 CGATAGCAAGAAGGGATTCGCAAAGCTGGTGGAGGAACACCTGGAGTCC  
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2358 GACGCCATCGGCAGATTGAGGGCGCCAACCCCGCCCTGTACGAGACC  
2359 CTGGCCAGAGATCGGCTGTGCCTCATGATGCCAGTACTCCTGGGCA  
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2378 AGTCCAAGCTGGCGCAAGGAGTACCAAGGCCAACGGCGTCTGT  
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2381 cgacgtgcctgattatgcataccatatgtgtcccgactatgccGCGGCCGCTGATTACAAA  
2382 GACGATGACGATAAGtaaATCGAATTCCGCTCGAGATAATCAACCTCTGGA  
2383 TTACAAAATTGTGAAAGATTGACTGGTATTCTTAACATATGTTGCTCCTTT  
2384 ACGCTATGTGGATACGCTGCTTAATGCCTTGTATCATGCTATTGCTTCCC  
2385 GTATGGCTTCATTTCTCCTCCTGTATAAATCCTGGTTAGTTCTGCCAC  
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3625 ATAAATCTGGAGCCGGTGAGCGTGGAGCCGGTATCATTGCAGCACT  
3626 GGGGCCAGATGGTAAGCCCTCCGTATCGTAGTTATCTACACGACGGGG  
3627 AGTCAGGCAACTATGGATGAACGAAATAGACAGATCGCTGAGATAGGTGC  
3628 CTCACTGATTAAGCATTGGTAACTGTCAGACCAAGTTACTCATATATACTT  
3629 TAGATTGATTAAAACCTCATTTTAATTAAAAGGATCTAGGTGAAGATCCT  
3630 TTTTGATAATCTCATGACCAAAATCCCTAACGTGAGTTTCGTTCCACTG  
3631 AGCGTCAGACCCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTT  
3632 TCTGCGCGTAATCTGCTGCTTGCAAACAAAAAAACCACCGCTACCAGCG  
3633 GTGGTTGTTGCCGGATCAAGAGCTACCAACTCTTTTCCGAAGGTAAC  
3634 TGGCTTCAGCAGAGCGCAGATAACCAAATACTGTTCTTAGTGTAGCCGT  
3635 AGTTAGGCCACCACTTCAAGAACTCTGTAGCACCGCCTACATACCTCGCT  
3636 CTGCTAATCCTGTTACCAAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCT  
3637 TACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCCAGCGGGTCG  
3638 GGCTGAACGGGGGGTTCGTGCACACAGCCCAGCTGGAGCGAACGACC  
3639 TACACCGAACTGAGATACTACAGCGTGAGCTATGAGAAAGCGCCACGCT  
3640 TCCCGAAGGGAGAAAGGCCGGACAGGTATCCGGTAAGCGGCAGGGTCGG  
3641 AACAGGGAGAGCGCACGAGGGAGCTTCCAGGGGAAACGCCCTGGTATCT  
3642 TTATAGTCCTGTCGGTTGCCACCTCTGACTTGAGCGTCGATTTTGT  
3643 GATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAACCGCGG  
3644 CCTTTTACGGTTCCCTGGCCTTTGCTGGCCTTGCTCACATGT  
3645  
3646  
3647