A robust vasculogenic microfluidic model using human immortalized

endothelial cells and Thy1 positive fibroblasts

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Supplemental Figures

Figure S1:



Figure S1: Expression of hTERT in FBs, ImFBs, HUVECs and ImHUVECs.

- (A) Representative confocal images of nucleus (DAPI, in blue) and hTERT (red) staining in FBs, ImFBs, HUVECs and ImHUVECs. Scale bar is 100 μm.
- (**B**) Statistical quantification for the mean fluorescent intensity (MFI) of hTERT in FBs, ImFBs, HUVECs and ImHUVECs. Bars represent mean \pm SD. Two-tailed *t* tests were performed for the statistical comparisons.
- (C) hTERT detection in FBs, ImFBs, HUVECs and ImHUVECs by western blot.

Figure S2:



Figure S2: µVNs made of ImHUVECs from P20 with ImFBs from P5 remain perfusable over 3 weeks.

(A) Representative images of μVNs made of ImHUVECs (green) from passage 20 (P20) and ImFBs from passage 5 (P5) at the same location on day 7, 14 (did not perform dextran perfusion experiment), and 21. μVNs are perfused with 70 kDa MW Texas Red dextran (red).





Figure S3: Evaluation of anti-Fibroblast and Thy1 expression in FBs from different experimental conditions.

(A) FACS measurement of anti-Fibroblast and Thy1 in FBs detached by TrypLE (left) or Trypsin (right).

(B-E) Representative histograms showing the anti-Fibroblast (top) and Thy1 (bottom) expression in parental FBs of ImFBs **(B)**, and FBs from different lot numbers. **(C-E)**.

(F) Representative histograms of anti-Fibroblast and Thy1 expression in ImFBs before and right after Thy1 based cell sorting.

Figure S4:



Figure S4: Thy1+ FBs promote better µVN formation than Thy1- FBs.

(A) Representative images of μ VNs made of human dermal microvascular endothelial cells (HDMEC) with Thy1+ ImFBs or Thy1- ImFBs. Green,

HDMEC. Red, dextran. Scale bar is 150 µm.

- (B) Representative images of dermal μVNs formed with Thy1+ or Thy1- ImFBs at the gap regions between microposts. Arrows point out the microvessel openings and closed structure. Green, HDMEC. Red, dextran. Scale bar is 150 μm.
- (C) Statistical analysis of normalized vessel area (left), junction density (middle), and average vessel length (right) of dermal μ VNs formed with Thy1+ ImFBs or Thy1- ImFBs.

(D-E) Quantification of the number of perfusable microvessels in each image (D) and the percentage of the openings at the gap regions between microposts in each device (E).

- (F) Representative histograms showing Thy1 expression in Thy1+ primary FBs and Thy1- primary FBs after sorting.
- (G) Statistical analysis of normalized vessel area (left), junction density (middle), and average vessel length (right) of μ VNs made of ImHUVECs with Thy1+ or Thy1- primary FBs.
- (H) FACS measurements of Thy1 in ImFBs made from another batch of FBs (ImFB-2) before sorting (left) and after expanding the sorted Thy1+ (top right) and Thy1- (bottom right) cells.
- (I) Representative images of μVNs made of ImHUVECs with Thy1+ ImFB-2 or Thy1- ImFB-2. Green, HDMEC. Red, dextran. Scale bar is 150 μm.
- (J) Statistical analysis of normalized vessel area (left), junction density (middle), and average vessel length (right) of μVNs formed with Thy1+ ImFB-2 or Thy1-ImFB-2.

Bars represent mean \pm SD. Two-tailed *t* tests were performed for the statistical comparisons.

Figure S5:



Figure S5: Reducing seeding density of Thy1- ImFBs cannot rescue the defective μ VNs.

(A-B) Statistical analysis of normalized vessel area (left), junction density (middle), and average vessel length (right) of μ VNs made of ImHUVEC (A) or HDMEC (B) with Thy1- ImFBs at different seeding density.

Bars represent mean \pm SD. Two-tailed *t* tests were performed for the statistical comparisons.

Figure S6:



Figure S6: Expression of angiogenic and lumen formation factors in FBs.

- (A) Heatmap of RT-PCR results of IGFBP2 expression in Thy1+ and Thy1- ImFBs or primary FBs from different samples cultured in 2D. Fold change was relative to Thy1- ImFBs or primary FBs of each sample.
- (**B**) Heatmap of RT-PCR results of genes involved in lumen formation and angiogenesis in Thy1+ ImFBs (top) and Thy1- ImFBs (bottom) cultured in 2D or 3D. Fold change was relative to 2D data of each sample.





Figure S7: IGFBP7 and SPARC are required for µVN formation.

- (A) Representative histograms showing IGFBP7 (left) or SPARC (right) expression in KO control ImFBs and IGFBP7 KO ImFBs, or SPARC KO ImFBs, respectively.
- (B) Statistical analysis of normalized vessel area (left), junction density (middle), and average vessel length (right) of μ VNs made of ImHUVECs with Thy1+ or Thy1- KO control ImFBs, IGFBP7 KO ImFBs, or SPARC KO ImFBs.
- (C) Representative histograms showing IGFBP7 (left) or SPARC (right) expression in double KO control ImFBs and IGFBP7 and SPARC double KO ImFBs.
- (D) Statistical analysis of normalized vessel area (left), junction density (middle),

and average vessel length (right) of μ VNs made of ImHUVECs with Thy1+ or Thy1- double KO control ImFBs, IGFBP7 and SPARC double KO ImFBs. Bars represent mean \pm SD. Two-tailed *t* tests were performed for the statistical comparisons.

Figure S8:



Figure S8: Thy1 expression in FBs decreased when expanding in both 2D and 3D.

- (A) Representative histograms showing Thy1 expression in FBs from P3 (top) and P4 (bottom).
- (B) Schematic representation of the flow cytometry procedure to run from μ VNs. Gel channels containing cells were cut out from the device, and further treated with Liberase to dissociate single cells from matrix.
- (C) Representative histograms showing Thy1 expression in Thy1+ ImFBs and Thy1- ImFBs on day 0 and day 7.
- (**D**) Heatmap of RT-PCR results of Thy1 expression in Thy1+ and Thy1- ImFBs cultured in 3D fibrin gel for 3 days and 7 days. Fold change was relative to day 3 data of each sample.

Figure S9:



Figure S9: Thy1 expression in FBs treated with IGFBP2 or PMA.

- (A) Representative histograms showing Thy1 expression in FBs (left), Thy1+ ImFBs (middle), and Thy1- ImFBs (right) treated with IGFBP2 at 0, 100, or 500 ng/ml for 3 days (top) or 6 days (bottom).
- (B) Representative histograms showing Thy1 expression in ImHUVECs (far left), Thy1+ ImFBs (left), Thy1- ImFBs (right), and FBs (far right) treated with DMSO or PMA for 48h (top) or 96h (bottom).