nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

| Statistics | |
|---------------------------|--|
| For all statistical an | alyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section. |
| n/a Confirmed | |
| The exact | sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| ☐ X A stateme | nt on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| | ical test(s) used AND whether they are one- or two-sided on tests should be described solely by name; describe more complex techniques in the Methods section. |
| X A descript | ion of all covariates tested |
| X A descript | ion of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| X | ription of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) tion (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| | pothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted as as exact values whenever suitable. |
| × For Bayesi | an analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| × For hierard | chical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| | of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |
| | Our web collection on <u>statistics for biologists</u> contains articles on many of the points above. |
| Software and | d code |
| Policy information a | about <u>availability of computer code</u> |
| Data collection | FDTD Lumerical Solution |
| Data analysis | Matlab, Origin |
| For manuscripts utilizing | custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and |

Data

Policy information about <u>availability of data</u>

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

All data supporting the findings of this study are available in the article and its Supplementary Information. Source data for the following figures are provided with this paper. Fig. 2b-2e; Fig. 3d-3e; Supplementary Fig. 1; Supplementary Fig. 4; Supplementary Fig. 7; Supplementary Fig. 8. Source data are provided with this paper.

Human research participants

Policy information about studies involving human research participants and Sex and Gender in Research.

Reporting on sex and gender

Use the terms sex (biological attribute) and gender (shaped by social and cultural circumstances) carefully in order to avoid confusing both terms. Indicate if findings apply to only one sex or gender; describe whether sex and gender were considered in study design whether sex and/or gender was determined based on self-reporting or assigned and methods used. Provide in the source data disaggregated sex and gender data where this information has been collected, and consent has been obtained for sharing of individual-level data; provide overall numbers in this Reporting Summary. Please state if this information has not been collected. Report sex- and gender-based analyses where performed, justify reasons for lack of sex- and gender-based analysis.

Population characteristics

Describe the covariate-relevant population characteristics of the human research participants (e.g. age, genotypic information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design questions and have nothing to add here, write "See above."

Recruitment

Describe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how these are likely to impact results.

Ethics oversight

Identify the organization(s) that approved the study protocol.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

| Ple | ease select the one | below th | at is the best fit for | your research. I | t you | are not sure, | read the app | ropriate sections | before mal | king your s | election. |
|-----|---------------------|----------|------------------------|------------------|-------|----------------|--------------|-------------------|------------|-------------|-----------|
| | Life sciences | | Behavioural & so | cial sciences | | Ecological, ev | olutionary & | environmental sc | iences | | |

For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

Describe how sample size was determined, detailing any statistical methods used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.

Data exclusions

Describe any data exclusions. If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.

Replication

Describe the measures taken to verify the reproducibility of the experimental findings. If all attempts at replication were successful, confirm this OR if there are any findings that were not replicated or cannot be reproduced, note this and describe why.

Randomization

Describe how samples/organisms/participants were allocated into experimental groups. If allocation was not random, describe how covariates were controlled OR if this is not relevant to your study, explain why.

Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.

Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description

Briefly describe the study type including whether data are quantitative, qualitative, or mixed-methods (e.g. qualitative cross-sectional, quantitative experimental, mixed-methods case study).

Research sample

State the research sample (e.g. Harvard university undergraduates, villagers in rural India) and provide relevant demographic information (e.g. age, sex) and indicate whether the sample is representative. Provide a rationale for the study sample chosen. For studies involving existing datasets, please describe the dataset and source.

Sampling strategy

Describe the sampling procedure (e.g. random, snowball, stratified, convenience). Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient. For qualitative data, please indicate whether data saturation was considered, and what criteria were used to decide that no further sampling was needed.

Data collection

Provide details about the data collection procedure, including the instruments or devices used to record the data (e.g. pen and paper, computer, eye tracker, video or audio equipment) whether anyone was present besides the participant(s) and the researcher, and whether the researcher was blind to experimental condition and/or the study hypothesis during data collection.

Timing

Indicate the start and stop dates of data collection. If there is a gap between collection periods, state the dates for each sample cohort.

Data exclusions

If no data were excluded from the analyses, state so OR if data were excluded, provide the exact number of exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.

Non-participation

State how many participants dropped out/declined participation and the reason(s) given OR provide response rate OR state that no participants dropped out/declined participation.

Randomization

If participants were not allocated into experimental groups, state so OR describe how participants were allocated to groups, and if allocation was not random, describe how covariates were controlled.

Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description

Briefly describe the study. For quantitative data include treatment factors and interactions, design structure (e.g. factorial, nested, hierarchical), nature and number of experimental units and replicates.

Research sample

Describe the research sample (e.g. a group of tagged Passer domesticus, all Stenocereus thurberi within Organ Pipe Cactus National Monument), and provide a rationale for the sample choice. When relevant, describe the organism taxa, source, sex, age range and any manipulations. State what population the sample is meant to represent when applicable. For studies involving existing datasets, describe the data and its source.

Sampling strategy

Note the sampling procedure. Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.

Data collection

Describe the data collection procedure, including who recorded the data and how.

Timing and spatial scale

Indicate the start and stop dates of data collection, noting the frequency and periodicity of sampling and providing a rationale for these choices. If there is a gap between collection periods, state the dates for each sample cohort. Specify the spatial scale from which the data are taken

Data exclusions

If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.

Reproducibility

Describe the measures taken to verify the reproducibility of experimental findings. For each experiment, note whether any attempts to repeat the experiment failed OR state that all attempts to repeat the experiment were successful.

Randomization

Describe how samples/organisms/participants were allocated into groups. If allocation was not random, describe how covariates were controlled. If this is not relevant to your study, explain why.

Blinding

Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.

Did the study involve field work?

Yes 🔀

Field work, collection and transport

Field conditions

Describe the study conditions for field work, providing relevant parameters (e.g. temperature, rainfall).

Location

State the location of the sampling or experiment, providing relevant parameters (e.g. latitude and longitude, elevation, water depth).

Access & import/export

Describe the efforts you have made to access habitats and to collect and import/export your samples in a responsible manner and in compliance with local, national and international laws, noting any permits that were obtained (give the name of the issuing authority, the date of issue, and any identifying information).

Disturbance

Describe any disturbance caused by the study and how it was minimized.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

| Materials & experime n/a Involved in the study Antibodies Eukaryotic cell lines Palaeontology and a Animals and other of Clinical data Dual use research of | n/a Involved in the study ChIP-seq Flow cytometry mrchaeology MRI-based neuroimaging rganisms |
|--|---|
| Antibodies used | Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number. |
| Validation | Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript. |
| Eukaryotic cell lin Policy information about ce Cell line source(s) Authentication Mycoplasma contaminati | Il lines and Sex and Gender in Research State the source of each cell line used and the sex of all primary cell lines and cells derived from human participants or vertebrate models. Describe the authentication procedures for each cell line used OR declare that none of the cell lines used were authenticated. Confirm that all cell lines tested negative for mycoplasma contamination OR describe the results of the testing for |
| Commonly misidentified (See ICLAC register) | ines Mame any commonly misidentified cell lines used in the study and provide a rationale for their use. |
| Palaeontology and | d Archaeology Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the |
| | issuing authority, the date of issue, and any identifying information). Permits should encompass collection and, where applicable, export. |
| Specimen deposition | Indicate where the specimens have been deposited to permit free access by other researchers. |
| Dating methods | If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided. |

|X| Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight

Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals

For laboratory animals, report species, strain and age OR state that the study did not involve laboratory animals.

| Wild animals | Provide details on animals observed in or captured in the field; report species and age where possible. Describe how animals were caught and transported and what happened to captive animals after the study (if killed, explain why and describe method; if released, say where and when) OR state that the study did not involve wild animals. |
|-------------------------|---|
| Reporting on sex | Indicate if findings apply to only one sex; describe whether sex was considered in study design, methods used for assigning sex. Provide data disaggregated for sex where this information has been collected in the source data as appropriate; provide overall numbers in this Reporting Summary. Please state if this information has not been collected. Report sex-based analyses where performed, justify reasons for lack of sex-based analysis. |
| Field-collected samples | For laboratory work with field-collected samples, describe all relevant parameters such as housing, maintenance, temperature, photoperiod and end-of-experiment protocol OR state that the study did not involve samples collected from the field. |
| Ethics oversight | Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not. |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about <u>clinical studies</u>

All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

Clinical trial registration

Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.

Study protocol

Note where the full trial protocol can be accessed OR if not available, explain why.

Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.

Outcomes

Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.

Dual use research of concern

Policy information about dual use research of concern

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

| No | Yes |
|-------------|----------------------------|
| X | Public health |
| \boxtimes | National security |
| \times | Crops and/or livestock |
| \times | Ecosystems |
| \boxtimes | Any other significant area |

Experiments of concern

Does the work involve any of these experiments of concern:

| | · · · · · · · · · · · · · · · · · · · |
|----------|---|
| No | Yes |
| X | Demonstrate how to render a vaccine ineffective |
| \times | Confer resistance to therapeutically useful antibiotics or antiviral agents |
| X | Enhance the virulence of a pathogen or render a nonpathogen virulent |
| X | Increase transmissibility of a pathogen |
| X | Alter the host range of a pathogen |
| X | Enable evasion of diagnostic/detection modalities |
| \times | Enable the weaponization of a biological agent or toxin |
| X | Any other potentially harmful combination of experiments and agents |

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| | Confirm that both raw and final | processed data have been deposited in a public database such as GEO | |
|--|---------------------------------|---|---|
| | Committe that both raw and mian | i processed data riave been deposited in a public database such as ofto | ٠ |

Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

Data access links

May remain private before publication.

For "Initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.

Files in database submission

source data. xlsx

Genome browser session (e.g. UCSC)

Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.

Methodology

Antibodies

Replicates Describe the experimental replicates, specifying number, type and replicate agreement.

Sequencing depth Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.

Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot

Peak calling parameters

Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files

Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.

Software

Data quality

Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details

Flow Cytometry

Plots

Confirm that:

The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

All plots are contour plots with outliers or pseudocolor plots.

A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.

Instrument Identify the instrument used for data collection, specifying make and model number.

Software Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a

community repository, provide accession details.

Cell population abundance Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the

samples and how it was determined.

Describe the gating strategy used for all relevant experiments, specifying the preliminary FSC/SSC gates of the starting cell Gating strategy population, indicating where boundaries between "positive" and "negative" staining cell populations are defined.

Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

Design type

Indicate task or resting state; event-related or block design.

| Design specifications | | e number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial ftrials are blocked) and interval between trials. | | | | |
|--|-----------------|---|--|--|--|--|
| Behavioral performance measure | | ber and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used h that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across | | | | |
| Acquisition | | | | | | |
| Imaging type(s) | Specify: fu | nctional, structural, diffusion, perfusion. | | | | |
| Field strength | Specify in | Tesla | | | | |
| Sequence & imaging parameters | | e pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, ness, orientation and TE/TR/flip angle. | | | | |
| Area of acquisition | State whe | ther a whole brain scan was used OR define the area of acquisition, describing how the region was determined. | | | | |
| Diffusion MRI Used | ∑ Not ∪ | sed | | | | |
| Preprocessing | | | | | | |
| Preprocessing software | | n software version and revision number and on specific parameters (model/functions, brain extraction, smoothing kernel size, etc.). | | | | |
| Normalization | | rmalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for OR indicate that data were not normalized and explain rationale for lack of normalization. | | | | |
| Normalization template | | mplate used for normalization/transformation, specifying subject space or group standardized space (e.g. ch, MNI305, ICBM152) OR indicate that the data were not normalized. | | | | |
| Noise and artifact removal | | rocedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and gnals (heart rate, respiration). | | | | |
| Volume censoring | Define your sof | tware and/or method and criteria for volume censoring, and state the extent of such censoring. | | | | |
| Statistical modeling & infere | nce | | | | | |
| Model type and settings | | (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and to (e.g. fixed, random or mixed effects; drift or auto-correlation). | | | | |
| Effect(s) tested | | effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether prial designs were used. | | | | |
| Specify type of analysis: WI | nole brain [| ROI-based Both | | | | |
| Statistic type for inference (See Eklund et al. 2016) | Specify voxel-w | ise or cluster-wise and report all relevant parameters for cluster-wise methods. | | | | |
| Correction | Describe the ty | pe of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo). | | | | |
| Models & analysis | | | | | | |
| n/a Involved in the study Functional and/or effective Graph analysis Multivariate modeling or p | , | s | | | | |
| Functional and/or effective conn | ectivity | Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information). | | | | |
| Graph analysis | | Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.). | | | | |
| Multivariate modeling and predic | ctive analysis | Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics. | | | | |