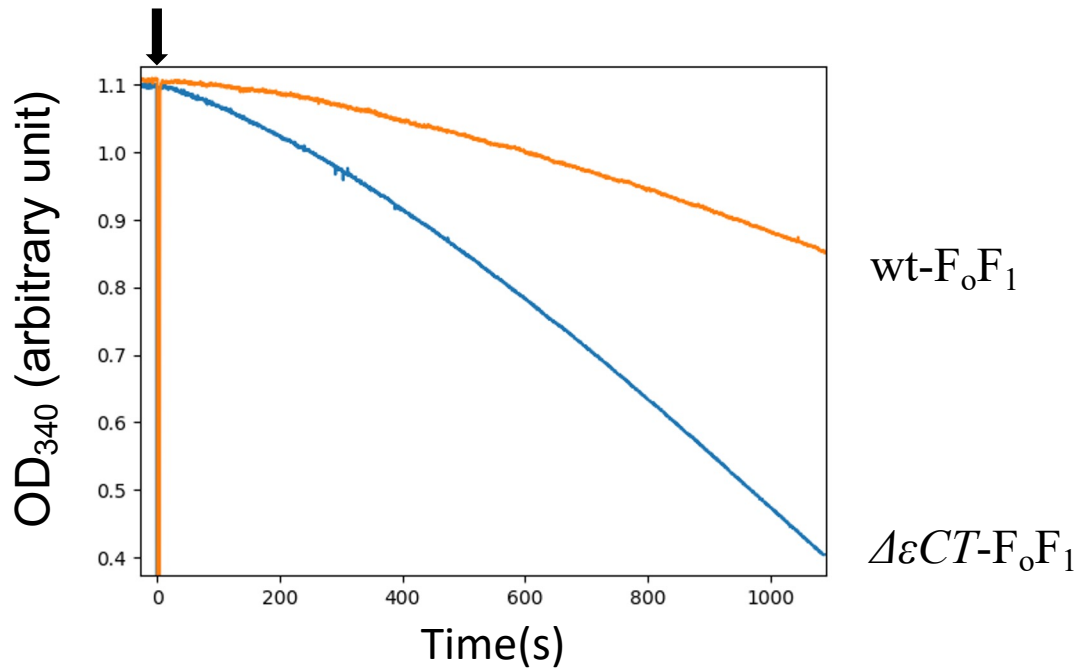
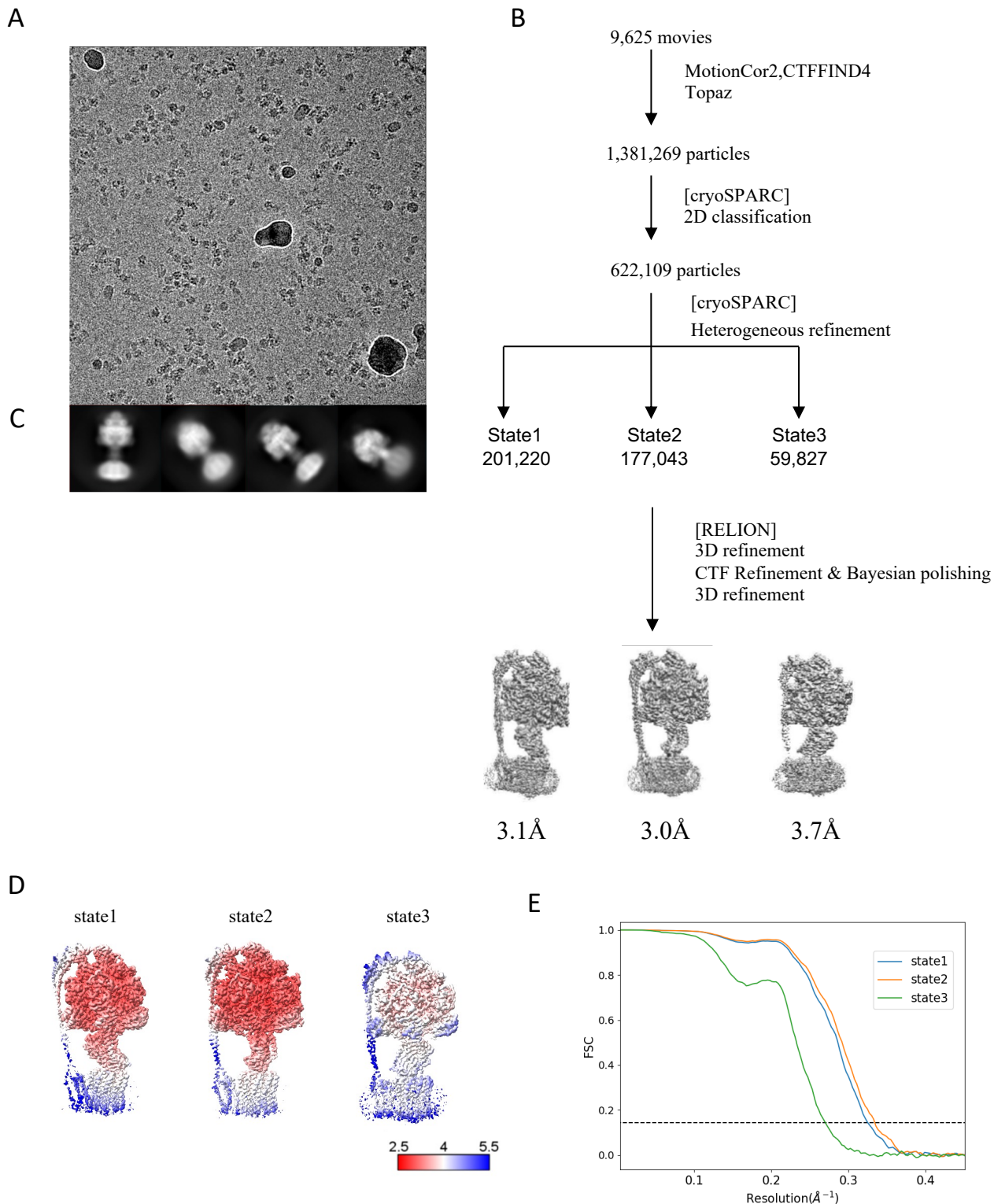


Fig. S1



**Figure S1. ATPase activity of wild-type  $F_0F_1$  and  $\Delta\varepsilon CT$ - $F_0F_1$**   
The activities of wild-type  $F_0F_1$  and  $\Delta\varepsilon CT$ - $F_0F_1$  were measured using the enzyme coupling assay described in the Materials and Methods section.

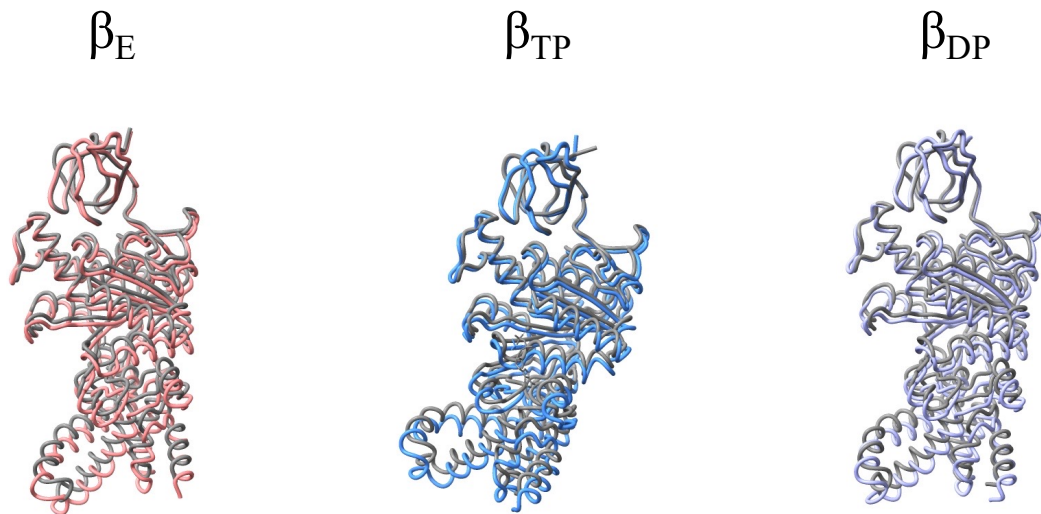
Fig. S2



**Figure S2. Image acquisition and reconstitution of the 3D structure of nucleotide-depleted (*ND*) wild-type (*wt*)- $F_0F_1$**

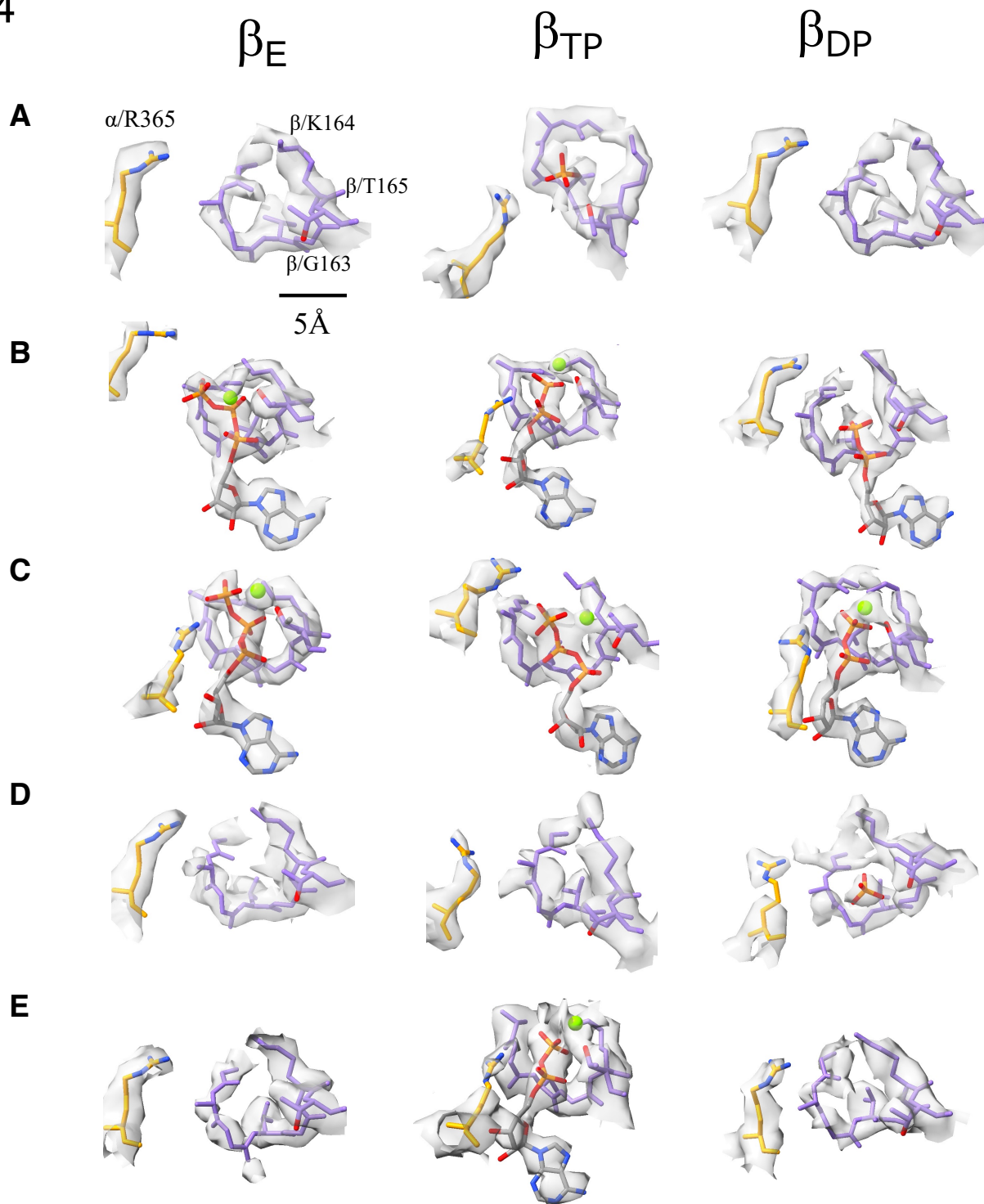
**(A)** Representative motion-corrected and dose-weighted micrographs. **(B)** Workflow for cryo-electron microscopy (cryo-EM) single particle analysis of *ND-wt-F<sub>0</sub>F<sub>1</sub>*. **(C)** 2D class-averaged images. **(D)** Cryo-EM density map. The colors used to represent local resolution are given in the key. **(E)** Fourier shell correlation (FSC) curves for the three rotational states of *wt-ND-F<sub>0</sub>F<sub>1</sub>* using FSC = 0.143 resolution criterion.

Fig. S3



**Figure S3. Structure comparison of *wt-F<sub>0</sub>F<sub>1</sub>* (gray) and *ND-wt-F<sub>0</sub>F<sub>1</sub>* (colored).** The  $\beta_{TP}$ ,  $\beta_{DP}$ , and  $\beta_E$  of *wt-F<sub>0</sub>F<sub>1</sub>* (6N2Y) and *US-ΔεCT-F<sub>0</sub>F<sub>1</sub>* were superimposed in the  $\beta$ -barrel domain (1-80 a .a.), respectively.

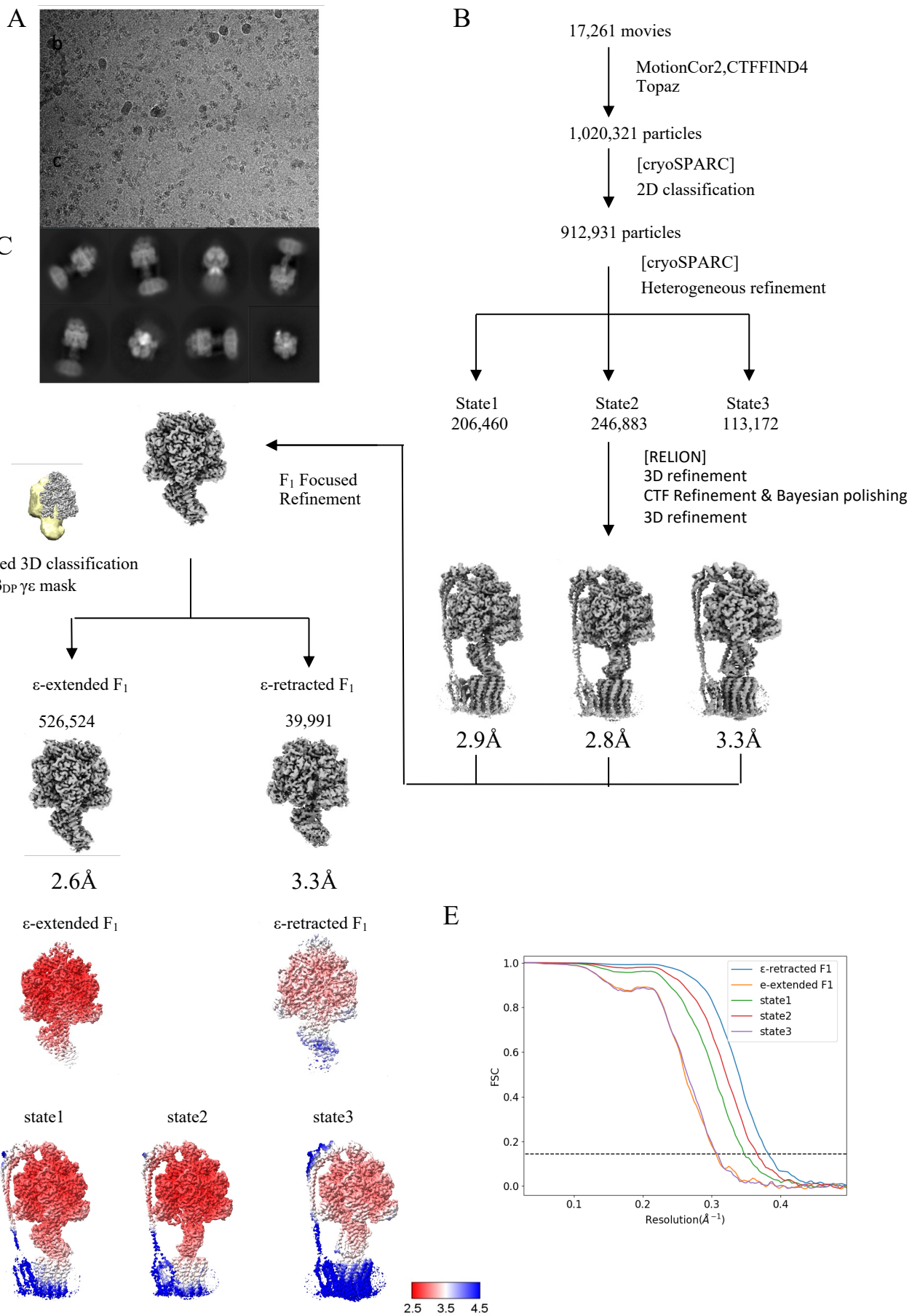
Fig. S4



**Figure S4. Structure of catalytic sites of wild-type  $F_0F_1$**

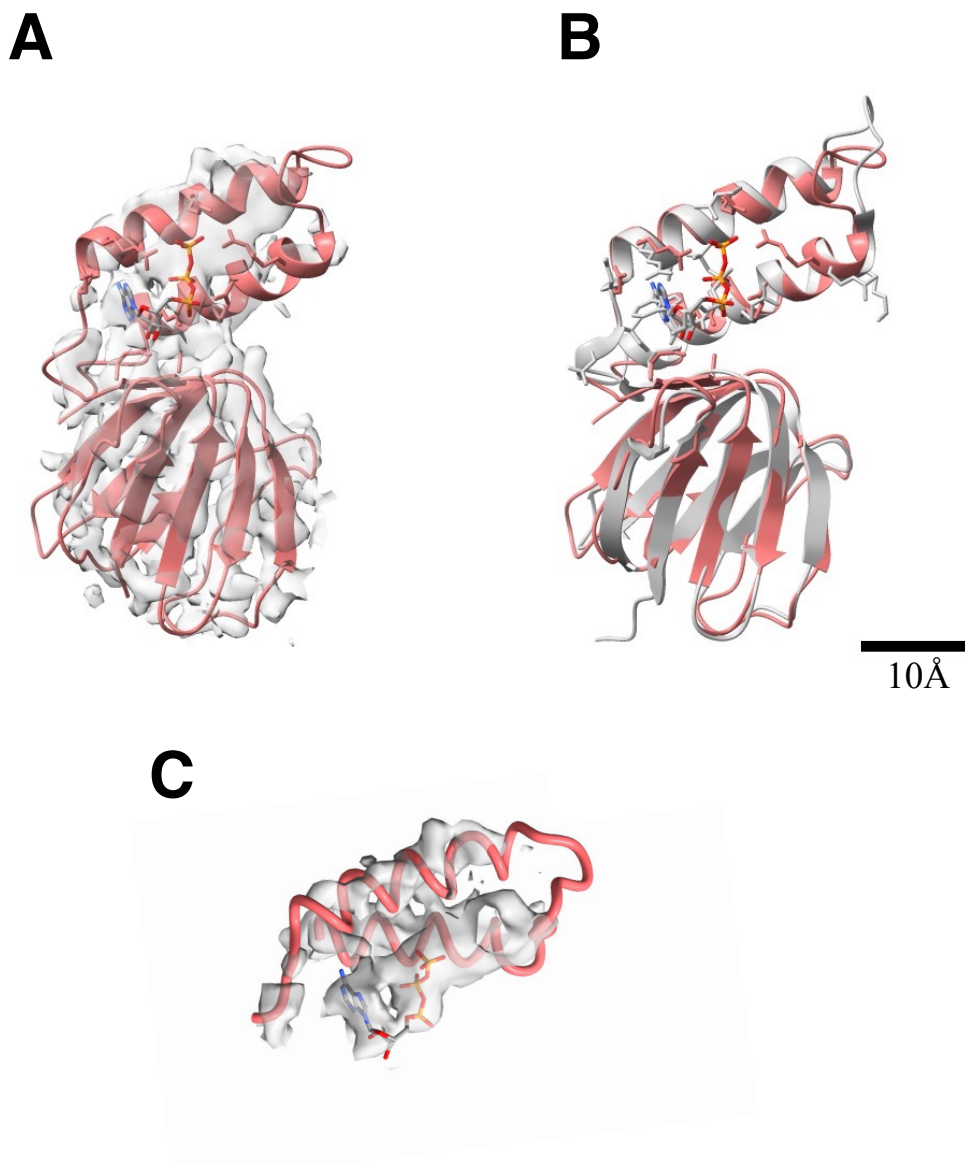
Magnified views of the three catalytic sites ( $\beta_E$ ,  $\beta_{TP}$ , and  $\beta_{DP}$ ) in each structure are shown as follows: **(A)** *ND-wt-F<sub>0</sub>F<sub>1</sub>*, **(B)** *wt-F<sub>0</sub>F<sub>1</sub>*  $\epsilon$  up form with ATP $\gamma$ S, **(C)** *wt-F<sub>0</sub>F<sub>1</sub>*  $\epsilon$  down form with ATP $\gamma$ S, **(D)** *ND- $\Delta$ ACT-F<sub>0</sub>F<sub>1</sub>*, and **(E)** *US- $\Delta$ ACT-F<sub>0</sub>F<sub>1</sub>*. Cryo-electron microscopy maps are represented as semi-transparent structures. Scale bar is 5 Å.

Fig. S5



**Figure S5. Image acquisition and reconstitution of the 3D structure of *wt*-F<sub>0</sub>F<sub>1</sub> under ATP $\gamma$ S saturation condition**

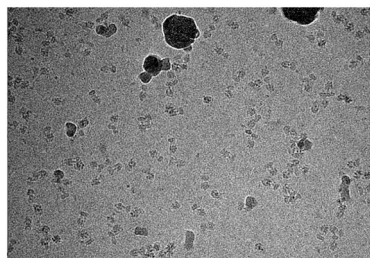
**(A)** Micrographs. **(B)** Workflow for cryo-electron microscopy (cryo-EM) single particle analysis under ATP $\gamma$ S saturation condition. EM density maps of three states were combined and the F<sub>1</sub> domain was classified into  $\epsilon$ -extended and  $\epsilon$ -retracted forms. **(C)** 2D class-averaged images. **(D)** Cryo-EM density map of  $\epsilon$ -extended and  $\epsilon$ -retracted F<sub>1</sub> domains. The colors used to represent local resolution are given in the key. **(E)** Fourier shell correlation (FSC) curves for  $\epsilon$ -extended and  $\epsilon$ -retracted F<sub>1</sub> domains using FSC = 0.143 resolution criterion.



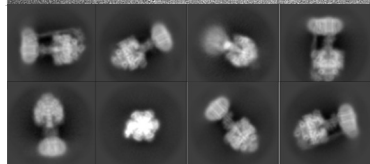
**Figure S6. Structure of down form of  $\epsilon$  subunit with ATP in  $wt-F_0F_1$**   
(A) Density map of  $\epsilon$  subunit in  $F_1$  domain obtained under saturated ATP $\gamma$ S condition. (B) Superposition of the crystal structure of retracted- $\epsilon$  subunit (gray, PDB:2E5Y) and the structure of retracted- $\epsilon$  in  $F_1$  domain obtained under saturated ATP $\gamma$ S condition. (C) Magnified view of ATP binding site of retracted- $\epsilon$  in  $F_1$  domain. ATP was represented as sticks.

Fig. S7

A



C



B

7,329 movies  
MotionCor2,CTFFIND4  
Topaz

499,788 particles

[cryoSPARC]  
2D classification

418,497 particles

[cryoSPARC]  
Heterogeneous refinement

state1<sup>ND</sup>

state1<sup>US</sup>

state1<sup>US'</sup>

state2<sup>ND</sup>

state2<sup>ND'</sup>

state2<sup>US</sup>

state3<sup>US</sup>

34,672

73,935

34,829

52,165

12,644

84,255

28,776

[RELION]  
3D refinement  
CTF Refinement & Bayesian Polishing  
3D refinement



3.6Å



3.2Å



3.4Å



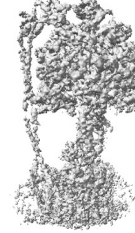
3.4Å



4.5Å



3.2Å



4Å

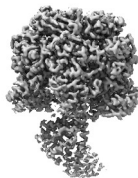
F<sub>1</sub> focused refinement



3.4Å



3.0Å



3.2Å



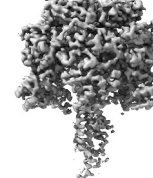
3.3Å



4.5Å



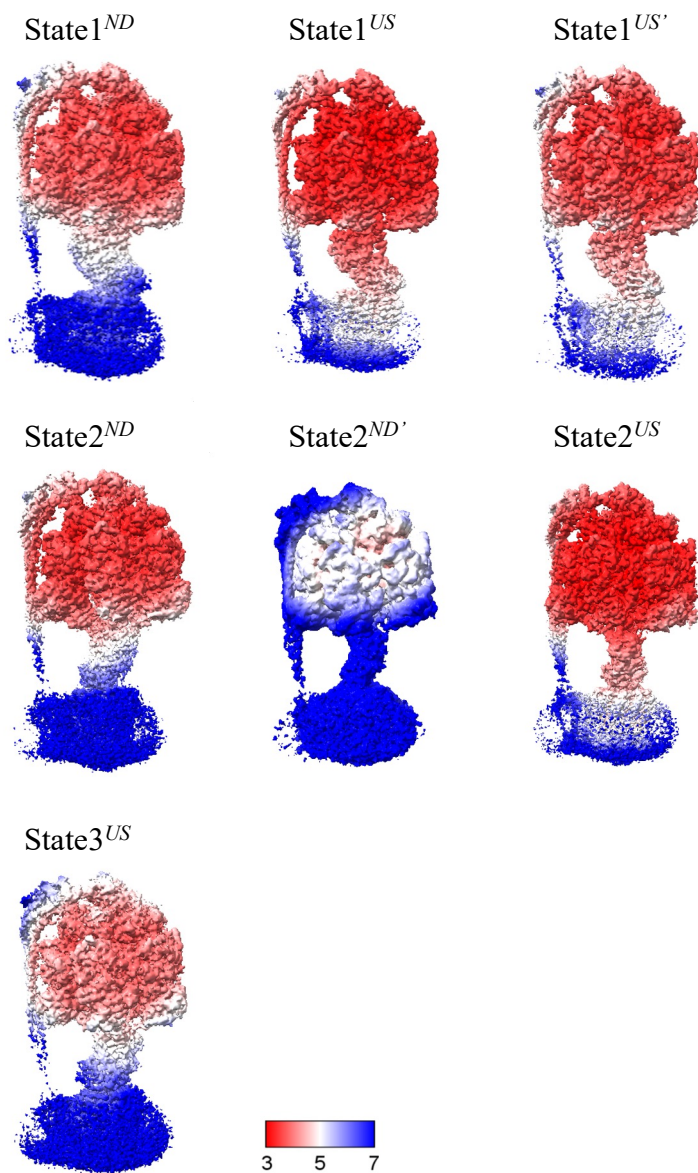
3.1Å



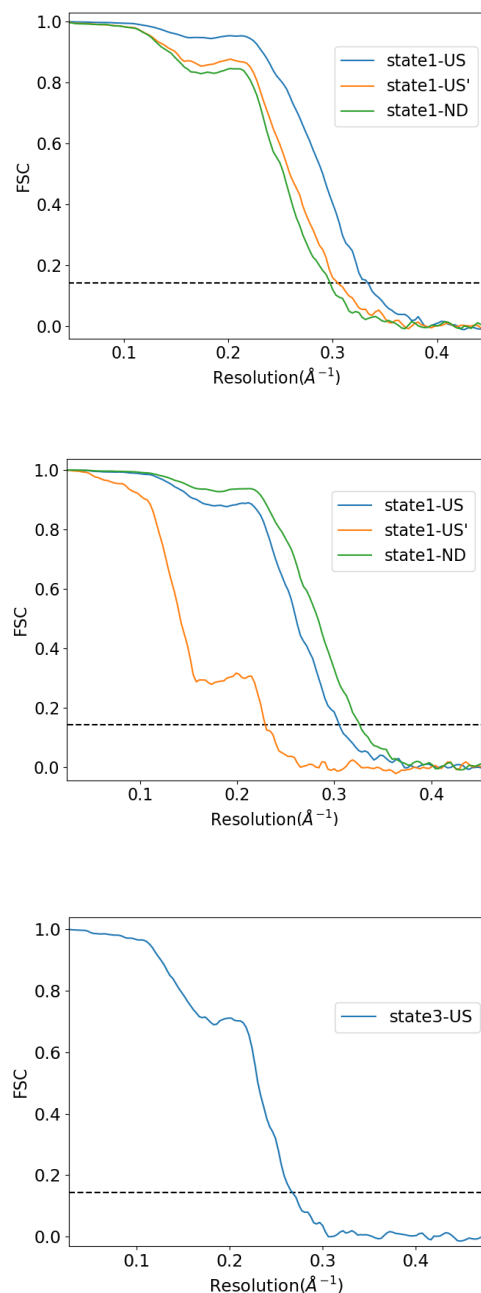
3.8Å



D



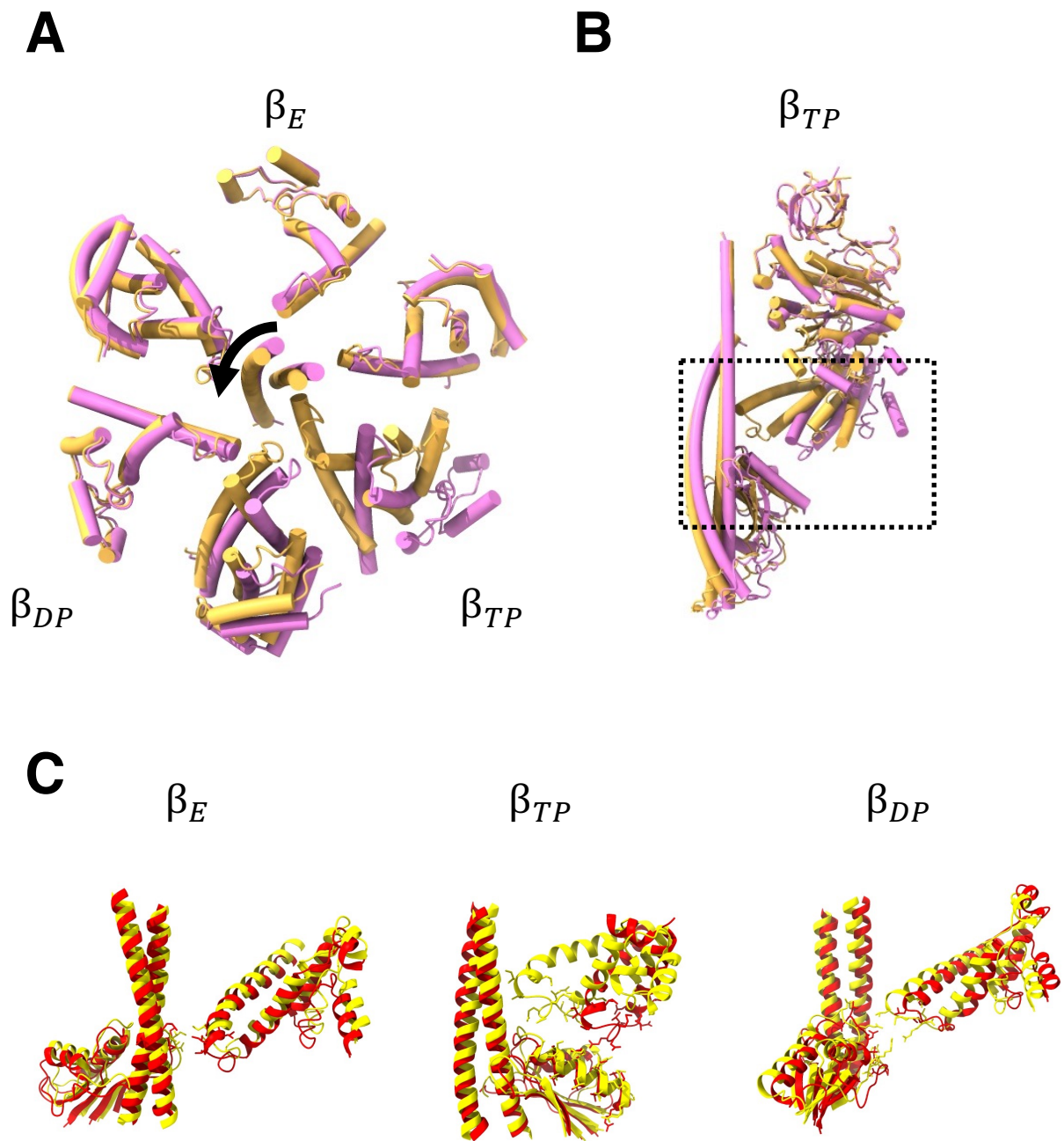
F



**Figure S7. Image acquisition and reconstitution of the 3D structures of nucleotide-depleted (ND)- $\Delta\epsilon CT$ -F<sub>0</sub>F<sub>1</sub> and Uni-Site (US)- $\Delta\epsilon CT$ -F<sub>0</sub>F<sub>1</sub>.**

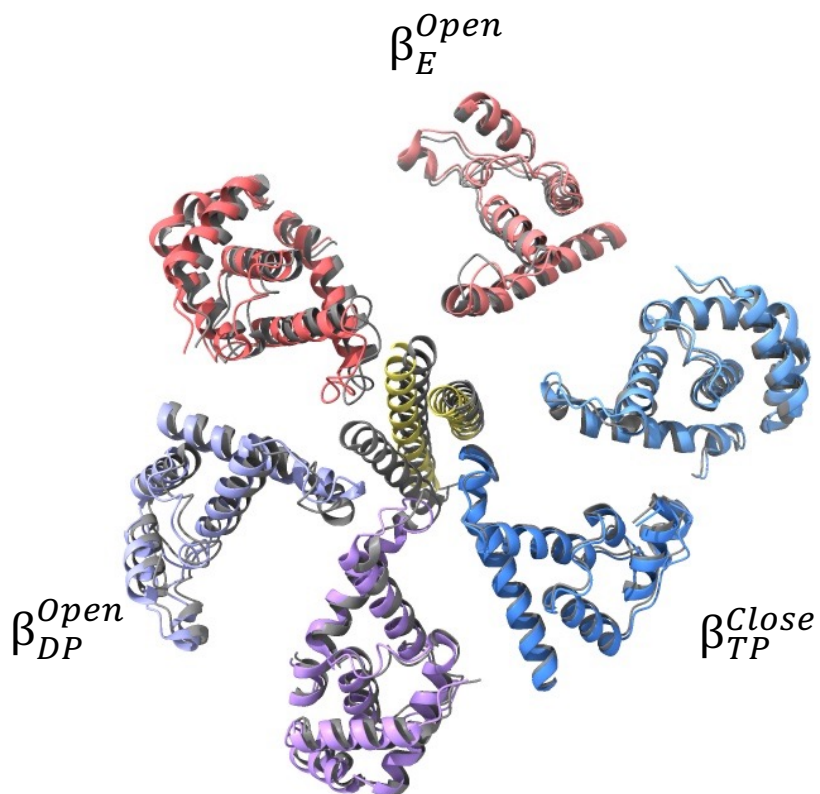
(A) Micrographs. (B) Workflow for cryo-electron microscopy (cryo-EM) single particle analysis under unisite conditions. (C) 2D class-averaged images. (D) Cryo-EM density map of ND- $\Delta\epsilon CT$ -F<sub>0</sub>F<sub>1</sub> and US- $\Delta\epsilon CT$ -F<sub>0</sub>F<sub>1</sub>. The colors used to represent local resolution are given in the key. (E) Fourier shell correlation (FSC) curves for each of the three rotational states of F<sub>0</sub>F<sub>1</sub> obtained from the unisite condition using FSC = 0.143 for resolution criterion.

Fig. S8

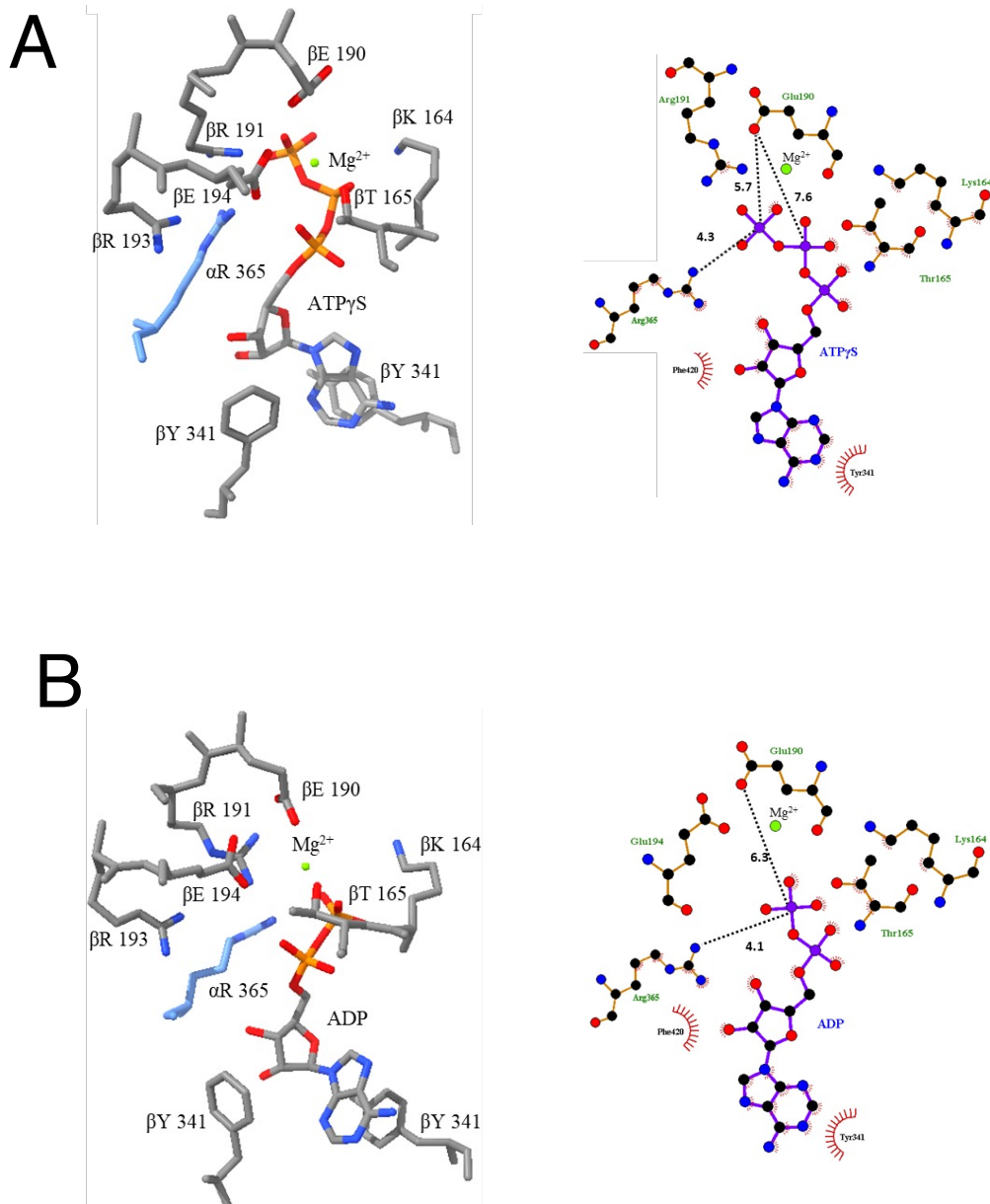


**Figure S8.** Structure comparison of *US-ΔεCT-F<sub>0</sub>F<sub>1</sub>* (orange) and *ND-wt-F<sub>0</sub>F<sub>1</sub>* (pink). (A) slice view of C terminal F<sub>1</sub> domain. (B) Side view of  $\beta_{TP}$  and  $\gamma$  subunit. All subunits were represented as cylinders.

Fig. S9



**Figure S9.** Structure comparison of *US- $\Delta\epsilon$ CT-F<sub>0</sub>F<sub>1</sub>* (colored) and *ND-wt-F<sub>0</sub>F<sub>1</sub>* (gray) in C termini domain of both  $\alpha$  and  $\beta$  subunits, and  $\gamma$  subunit.



**Figure S10. Coordination of nucleotides in the binding sites of wt-F<sub>0</sub>F<sub>1</sub> ε-retracted form with ATPγS.** *Left panels;* binding sites of β<sub>TP</sub> (**A**) and β<sub>DP</sub> (**B**) in wt-F<sub>0</sub>F<sub>1</sub> ε-retracted form with ATPγS (drawn with colored atoms and bonds and blue main chain) *Right panels;* Schematic representations of the coordination of ATP group bound to the side chains, respectively. The distances between the atoms are shown in dotted lines. All distances are in Å. Two-dimensional diagram generated by Ligplot.

**Supplementary Table1. ATPase activity of *wt*-F<sub>0</sub>F<sub>1</sub> and  $\Delta\varepsilon CT$  - F<sub>0</sub>F<sub>1</sub> at the indicated time after initiation of reaction.** The activities of wild-type F<sub>0</sub>F<sub>1</sub> and  $\Delta\varepsilon CT$ -F<sub>0</sub>F<sub>1</sub> were measured using the enzyme coupling assay described in the Materials and Methods section.

time	30 s	600 s	1000 s
<i>wt</i> -F <sub>0</sub> F <sub>1</sub>	11 s <sup>-1</sup>	41 s <sup>-1</sup>	51 s <sup>-1</sup>
$\Delta\varepsilon CT$ -F <sub>0</sub> F <sub>1</sub>	49 s <sup>-1</sup>	107 s <sup>-1</sup>	130 s <sup>-1</sup>

**Supplementary Table 2. Cryo-EM data collection, refinement and validation statistics for *wt-ND* F<sub>0</sub>F<sub>1</sub>.**

	state1	state2	state3
	F <sub>0</sub> F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>
EMDB ID	33251	33252	33253
PDB ID	7XKH		
<b>Data collection and processing</b>			
Magnification	50,000	50,000	50,000
Microscope	CRYOARM300	CRYOARM300	CRYOARM300
Voltage(kV)	300	300	300
Total dose (e <sup>-</sup> /Å <sup>2</sup> )	50	50	50
Pixel size(Å/pix)	1.01	1.01	1.01
Defocus range(μm)	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0
symmetry imposed	C1	C1	C1
Initial particle	1,381,269	1,381,269	1,381,269
Final Particle	201,220	177,043	59,827
Map resolution(Å)	3.1	3	3.7
FSC threshold	0.143	0.143	0.143
<b>Refinement</b>			
Initial model used	This study	This study	This study
Model resolution	3.1		
FSC threshold	0.5	0.5	0.5
<b>Model composition</b>			
Nonhydrogen atoms	25,563		
Protein residues	3,317		
Ligands	Pi		
<b>R.m.s deviations</b>			
Bond length (Å)	0.002		
Bond Angles (°)	0.443		
<b>Validation</b>			
MolProbity score	1.04		
EMRinger score	2.56		
Clashscore	2.56		
Rotamer outlier (%)	0		
CaBALM outlier (%)	1.67		
<b>Ramachandran plot</b>			
Favored (%)	98.36		
Allowed (%)	1.64		
Disallowed (%)	0		

**Supplementary Table 3. Cryo-EM data collection, refinement and validation statistics for *wt*-F<sub>0</sub>F<sub>1</sub> obtained under 4 mM ATP $\gamma$ S condition.**

	$\epsilon$ -extended	$\epsilon$ -retracted	state1	state2	state3
	F <sub>1</sub> domain	F <sub>1</sub> domain	F <sub>0</sub> F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>
EMDB ID	33264	33265	33266	33267	33268
PDB ID	7XKQ	7XKR			
<b>Data collection and processing</b>					
Magnification	50,000	81,000	81,000	81,000	81,000
Microscope	Titan Krios	Titan Krios	Titan Krios	Titan Krios	Titan Krios
Voltage(kV)	300	300	300	300	300
Total dose (e <sup>-</sup> /Å <sup>2</sup> )	50	50	50	50	50
Pixel size(Å/pix)	0.88	0.88	0.88	0.88	0.88
Defocus range( $\mu$ m)	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0
symmetry imposed	C1	C1	C1	C1	C1
Initial particle	1,020,321	1,020,321	1,020,321	1,020,321	1,020,321
Final Particle	526,524	39,991	206,460	246,883	113,172
Map resolution(Å)	2.6	3.3	2.9	2.8	3.3
FSC threshold	0.143	0.143	0.143	0.143	0.143
<b>Refinement</b>					
Initial model used	This study	This study	This study	This study	This study
Model resolution	2.7	3.5			
FSC threshold	0.5	0.5	0.5	0.5	0.5
<b>Model composition</b>					
Nonhydrogen atoms	25,254	25,306			
Protein residues	3,257	3,259			
Ligands	5MG, 4ATP, 2ADP	6MG, 6ATP, ADP			
<b>R.m.s deviations</b>					
Bond length (Å)	0.003	0.003			
Bond Angles (°)	0.545	0.507			
<b>Validation</b>					
MolProbity score	1.1	1.27			
EMRinger score	4.29	3.31			
Clashscore	3.13	4.92			
Rotamer outlier (%)	0	0			
CaBALM outlier (%)	1.3	1.85			
<b>Ramachandran plot</b>					
Favored (%)	98.36	97.96			
Allowed (%)	1.64	2.01			
Disallowed (%)	0	0			

**Supplementary Table 4. Cryo-EM data collection, refinement and validation statistics for  $\Delta CT-F_0F_1$  obtained under unisite.**

	state1 ND		state1 US		state1 US'	
	F <sub>0</sub> F <sub>1</sub>	F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>	F <sub>1</sub>	F <sub>0</sub> F <sub>1</sub>	F <sub>1</sub>
EMDB ID	33277	33258	33278	33259	33279	33260
PDB ID		7XKO		7XKP		
<b>Data collection and processing</b>						
Magnification	81,000	81,000	81,000	81,000	81,000	81,000
Microscope	Titan Krios	Titan Krios	Titan Krios	Titan Krios	Titan Krios	Titan Krios
Voltage(kV)	300	300	300	300	300	300
Total dose (e <sup>-</sup> /Å <sup>2</sup> )	60	60	60	60	60	60
Pixel size(Å/pix)	0.88	0.88	0.88	0.88	0.88	0.88
Defocus range(μm)	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0	-0.8 to -2.0
symmetry imposed	C1	C1	C1	C1	C1	C1
Initial particle	499,788	499,788	499,788	499,788	499,788	499,788
Final Particle	34,672	34,672	73,935	73,935	34,829	34,829
Map resolution(Å)	3.6	3.4	3.2	3	3.4	3.2
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143
<b>Refinement</b>						
Initial model used	This study	This study	This study	This study	This study	This study
Model resolution		3.5				
FSC threshold	0.5	0.5	0.5	0.5	0.5	0.5
<b>Model composition</b>						
Nonhydrogen atoms		24,070		24,594		
Protein residues		3,127		3,188		
Ligands		Pi		ATP, MG		
<b>R.m.s deviations</b>						
Bond length (Å)		0.003		0.002		
Bond Angles (°)		0.49		0.48		
<b>Validation</b>						
MolProbity score		1.47		1.25		
EMRinger score		2.61		3.37		
Clashscore		5.68		3.86		
Rotamer outlier (%)		0		0		
CaBALM outlier (%)		1.9		1.87		
<b>Ramachandran plot</b>						
Favored (%)		97.08		97.67		
Allowed (%)		2.92		2.33		
Disallowed (%)		0		0		



