Supporting Information

Subcellular Delivery of Hydrogen Sulfide Using Small Molecule Donors Impacts Organelle Stress

Annie K. Gilbert and Michael D. Pluth*

Department of Chemistry and Biochemistry Materials Science Institute Knight Campus for Accelerating Scientific Impact Institute of Molecular Biology University of Oregon Eugene, OR 97403-1253 United States.

Contact Information:

Michael D. Pluth pluth@uoregon.edu

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1. Fluorescence imaging of **MitoTCM** compared to NaSH



Figure S1. Fluorescence imaging of H₂S produced by **MitoTCM** compared to NaSH in HeLa cells. Cells were incubated with MitoTracker (50 nM) and (a) vehicle (0.5% DMSO), (b) Mito-HS (5 μ M), (c) Mito-HS (5 μ M) and NaSH (200 nM), (d) Mito-HS (5 μ M) and **MitoTCM** (200 nM). Cells were incubated with MitoTracker and vehicle or Mito-HS for 30 minutes, and then either the vehicle, NaSH or **MitoTCM** for 60 min followed by imaging. Bar scale: 25 μ m.

2. Fluorescence imaging of MitoTCM compared to TCM alkyne



Figure S2. Fluorescence imaging of H₂S produced by **MitoTCM** compared to **TCM alkyne** in HeLa cells. Cells were incubated with MitoTracker (50 nM) and (a) vehicle (0.5% DMSO), (b) Mito-HS (5 μ M), (c) Mito-HS (5 μ M) and **MitoTCM** (200 nM), (d) Mito-HS (5 μ M) and **TCM alkyne** (200 nM). Cells were incubated with MitoTracker and vehicle or Mito-HS for 30 minutes, and then either the vehicle, **MitoTCM** or **TCM alkyne** for 60 min followed by imaging. Bar scale: 25 μ m.





Figure S3. Fluorescence imaging of H₂S produced by **LysoTCM** compared to **TCM alkyne** in HeLa cells. Cells were incubated with LysoTracker (50 nM) and (a) vehicle (0.5% DMSO), (b) Lyso-AFP (10 μ M), (c) Lyso-AFP (10 μ M) and NaSH (200 μ M, positive control), (d) Lyso-AFP (10 μ M) and **LysoTCM**, (e) Lyso-AFP (10 μ M) and **TCM alkyne** (200 nM). Cells were incubated with LysoTracker and vehicle or Lyso-AFP for 30 minutes, and then either the vehicle, NaSH, **LysoTCM**, or **TCM alkyne** for 60 min followed by imaging. Bar scale: 25 μ m.

4. Fluorescence imaging of ERTCM compared to TCM alkyne



Figure S4. Fluorescence imaging of H₂S produced by ERTCM compared to TCM alkyne in HeLa cells. Cells were incubated with ERTracker (50 nM) and (a) vehicle (0.5% DMSO), (b) Na-H₂S-ER (10 μ M), (c) Na-H₂S-ER (10 μ M) and NaSH (200 μ M, positive control), (d) Na-H₂S-ER (10 μ M) and ERTCM, (e) Na-H₂S-ER (10 μ M) and TCM alkyne (200 nM). Cells were incubated with ERTracker and vehicle or Na-H₂S-ER for 30 minutes, and then either the vehicle, NaSH, ERTCM, or TCM alkyne for 60 min followed by imaging. Bar scale: 25 μ m.

5. Fluorescence imaging of GolgiTCM compared to TCM alkyne



Figure S5. Fluorescence imaging of H₂S produced by **GolgiTCM** compared to **TCM alkyne** in HeLa cells. Cells were incubated with BODIPYTM TR Ceramide (2 μ g/mL) and (a) vehicle (0.5% DMSO), (b) Golgi-NH (5 μ M), (c) Golgi-NH (5 μ M) and NaSH (200 μ M, positive control), (d) Golgi-NH (5 μ M) and **GolgiTCM**, (e) Golgi-NH (5 μ M) and **TCM alkyne** (200 nM). Cells were incubated with BODIPYTM TR Ceramide and vehicle or Golgi-NH for 30 minutes, and then either the vehicle, NaSH, **GolgiTCM**, or **TCM alkyne** for 60 min followed by imaging. Bar scale: 25 μ m.

6. Selectivity experiment of localized delivery



Figure S6. Fluorescence imaging of H₂S produced by **MitoTCM**, AP39, and **ERTCM** in the presence of Mito-HS in HeLa cells. Cells were incubated with MitoTracker (50 nM) and (a) vehicle (0.5% DMSO), (b) Mito-HS (10 μ M), (c) Mito-HS (10 μ M) and NaSH (200 μ M, positive control), (d) Mito-HS (10 μ M) and **MitoTCM** (200 nM), (e) Mito-HS (10 μ M) and AP39 (200 nM), (f) Mito-HS (10 μ M) and **ERTCM** (200 nM). Cells were incubated with MitoTracker and vehicle or Mito-HS for 30 minutes, and then either the vehicle, **MitoTCM**, AP39, or **ERTCM** for 60 min followed by imaging. Bar scale: 25 μ m.

7. Colocalization negative control experiment



Figure S7. Fluorescence imaging of H₂S produced by **MitoTCM** using Mito-HS in the presence of Lysotracker in HeLa cells. Cells were incubated with LysoTracker (50 nM) and Mito-HS (5 μ M) for 30 minutes, then incubated with **MitoTCM** (200 nM) for 60 min followed by imaging. Pearson's coefficient = 0.30 suggesting minimal colocalization of H₂S produced by MitoTCM and imaged with Mito-HS with LysoTracker. Bar scale: 25 μ m.

8. Monensin Cell Viability Experiment



Figure S8. Cell viability of H9C2 cells in the presence of Monensin. H9C2 cells were treated with either DMSO (0.5%, vehicle) or Monensin (0.5, 1.0, 2.0, or 3 μ M) for 19 h. Cell viability was assessed using a CCK-8 kit. Results are expressed as mean \pm SD (n = 12).

9. NMR Spectra

 1H NMR (600 MHz, CDCl₃) and $^{13}C\{^1H\}$ NMR (151 MHz, CDCl₃) spectra of 1-ethynyl-4-isothiocyanatobenzene.



 1H NMR (500 MHz, DMSO-*d*₆, 60 °C) and $^{13}C\{^1H\}$ NMR (126 MHz, DMSO-*d*₆, 60 °C) spectra of TCM alkyne



¹H NMR (500 MHz, DMSO-*d*₆) and ¹³C{¹H} NMR (126 MHz, DMSO-*d*₆) spectra of (6-bromohexyl)triphenylphosphonium bromide.



 $^{31}P{^{1}H}$ NMR (202 MHz, DMSO-*d*₆) spectrum of (6-bromohexyl)triphenylphosphonium bromide.



¹H NMR (500 MHz, DMSO- d_6) and ¹³C{¹H} NMR (151 MHz, CDCl₃) spectra of (6-azidohexyl)triphenylphosphonium bromide.



³¹P{¹H} NMR (202 MHz, CDCl₃) spectrum of (6-azidohexyl)triphenylphosphonium bromide.

— 19.66

50 130 110 90 70 50 30 10 -10 -30 -50 -70 -90 -110 -130 -150 -170 -190 -210 -230 -2! Chemical Shift (ppm) 1H NMR (500 MHz, CDCl₃) and $^{13}C\{^1H\}$ NMR (126 MHz, CDCl₃) spectra of 4-(2-azidoethyl)morpholine.



¹H NMR (500 MHz, CDCl₃) and ¹³C{¹H} NMR (126 MHz, CDCl₃) spectra of *N*-(2-azidoethyl)-4-methylbenzenesulfonamide.



 1H NMR (500 MHz, CDCl₃) and $^{13}C\{^1H\}$ NMR (126 MHz, CDCl₃) spectra of 4-(2-azidoethyl)-benzenesulfonamide.



¹H NMR (600 MHz, DMSO- d_6 , 60 °C) and ¹³C{¹H} NMR (126 MHz, DMSO- d_6 , 60 °C) spectra of **MitoTCM**.



³¹P{¹H} NMR (202 MHz, DMSO- d_6 , 60 °C) spectrum of **MitoTCM**.

.50 130 110 90 70 50 30 10 -10 -30 -50 -70 -90 -110 -130 -150 -170 -190 -210 -230 -2! Chemical Shift (ppm) ¹H NMR (600 MHz, DMSO- d_6 , 60 °C) and ¹³C{¹H} NMR (151 MHz, DMSO- d_6 , 60 °C) spectra of LysoTCM.



¹H NMR (600 MHz, DMSO- d_6 , 60 °C) and ¹³C{¹H} NMR (151 MHz, DMSO- d_6 , 60 °C) spectra of **ERTCM**.



¹H NMR (600 MHz, DMSO- d_6 , 60 °C) and ¹³C{¹H} NMR (151 MHz, DMSO- d_6 , 60 °C) spectra of **GolgiTCM**.





 1H NMR (500 MHz, CD₃OD) and $^{31}P\{^1H\}$ NMR (202 MHz, CD₃OD) spectra of Mito-HS.





¹H NMR (500 MHz, DMSO-*d*₆) spectrum of Na-H₂S-ER.



