

#### Figure S1. Phenotypic and functional characteristics of CD69<sup>-</sup>CD103<sup>-</sup>, CD69<sup>+</sup>CD103<sup>-</sup>, CD69<sup>-</sup> CD103<sup>+</sup>, and CD69<sup>+</sup>CD103<sup>+</sup> intestinal CD8<sup>+</sup> T<sub>RM</sub> cells, Related to Figures 1 and 2.

(A) Absolute numbers of gated i.v.-negative CD45.1<sup>+</sup>CD8 $\beta$ <sup>+</sup> P14 T cells in each intestinal tissue compartment.

(**B**–**G**) Bar graphs representing frequencies of CD69<sup>-</sup>CD103<sup>-</sup> (light blue circles), CD69<sup>+</sup>CD103<sup>-</sup> (yellow inverted triangles), CD69<sup>-</sup>CD103<sup>+</sup> (pink squares), or CD69<sup>+</sup>CD103<sup>+</sup> (red circles) cells expressing GzmA (**B**), GzmB (**C**), TNF (**D**), IFN $\gamma$  (**E**), IL-2 (**F**), and CD127 (**G**) in gated i.v.-negative CD45.1<sup>+</sup>CD8 $\beta^+$  P14 T cells from the siIEL, siLPL, cIEL, and cLPL intestinal tissue compartments. For analyses of TNF, IFN $\gamma$ , and IL-2, cells were restimulated *in vitro* for 3h with gp<sub>33-41</sub> peptide.

Data are represented as mean  $\pm$  SEM. Repeated measures one-way ANOVA. \*p<0.05, \*\*p<0.01, \*\*\*\*p<0.001 \*\*\*\*p<0.0001. Data are representative of at least 3 independent experiments with n=5-6 mice per experiment.





D



AMUL-72809 Formation of a pool of free 40S subunits
CO.022513: ribonucleoprotein complex biogenesis
F.MMU-7280275: Celtular reportsoness to stress
R-MMU-7280275: Celtular reportsoness to stress
R-MMU-7280275: Celtular reportsoness to stress
R-MMU-7280275: Celtular reportsones to stress
R-MMU-7280275: Celtular reportsoness
R-MMU-7302989; Transcriptional Regulation by TP43
CO.0006325: Celtular protein-containing complex assembly
GO.0016947: engulation of cell cycle process
R-MMU-16307: DNA damage stimulus

-log10(P)

Figure S2. Transcriptional heterogeneity exhibited by  $CD8^+$  T<sub>RM</sub> cells from small intestine and colon, Related to Figure 3.  $CD8^+$  P14 T cells harvested from siIEL, siLPL, cIEL, and cLPL tissue compartments were FACS-purified more than 21 days after LCMV infection and processed for CITE-seq with the 10x Genomics platform.

(A) UMAP clustering of all CD8 $\beta^+$  P14 T<sub>RM</sub> cells from each intestinal tissue compartment, colored by tissue compartment identity (left) or cluster identity (right).

(B) Bar graph representing the composition of each cell cluster based on intestinal tissue compartment.

(C) Top ten genes differentially expressed by each UMAP cluster, represented as a hierarchically clustered summary heatmap; rows represent individual genes and columns represent each UMAP cluster. Selected genes are highlighted on the left.

**(D)** Comparative gene ontology analyses performed on all UMAP clusters, represented as a summary heatmap.

















# Figure S3. Heterogeneity at the mRNA and protein levels exhibited by $CD8^+ T_{RM}$ cells from small intestine and colon, Related to Figure 3.

(A) Differentially expressed genes shared among  $CD8\beta^+$  P14 T<sub>RM</sub> cells from the siIEL, siLPL, cIEL, and cLPL intestinal tissue compartments, represented as a Circos plot. Inner circle represents gene lists, and purple curves link identical (shared) genes; outer circle represents tissue compartment identity.

(B) Top twenty genes differentially expressed between small intestine vs. colon  $CD8^+$  T<sub>RM</sub> cells, represented as a hierarchically clustered summary heatmap; rows represent individual genes and columns represent each of the 4 intestinal tissue compartments.

(C–H) Representative flow cytometry plots (bottom) showing expression of Ly108 (C), CD38 (D), CD160 (E), CD314 (F), CD44 (G), and IL-18R (H) by gated i.v.-negative CD45.1<sup>+</sup>CD8 $\beta^+$  P14 T cells from the four intestinal tissue compartments; numbers represent the frequencies of cells within each gate. Bar graphs (top right) indicate the frequencies of P14 T cells expressing each molecule. Histograms (top left) indicate the distribution of expression of each marker by P14 T cells from each of the four intestinal tissue compartments and the spleen; expression by naïve (CD62L<sup>hi</sup>CD44<sup>lo</sup>) CD8 $\beta^+$  T cells from a separate uninfected mouse is shown for comparison.

Data are represented as mean  $\pm$  SEM. Repeated measures one-way ANOVA. \*p<0.05, \*\*p<0.01, \*\*\*\*p<0.001 \*\*\*\*p<0.0001. Data are representative of at least 3 independent experiments with n=5-6 mice per experiment.





F

UMAP 2

UMAP 1







UMAP 2

L

Ċ.

UMAP 1







cIEL



3 Motif J

2

UMAP 1

silet silet det det

JMAP:





aller aller der der





# Figure S4. Epigenetic heterogeneity exhibited by $CD8^+$ T<sub>RM</sub> cells from small intestine and colon, Related to Figure 3.

(A) UMAP clustering analyses of scATAC-seq data from  $CD8\beta^+$  P14 T<sub>RM</sub> cells from the siIEL, siLPL, cIEL, and cLPL intestinal tissue compartments.

(B) Top differentially accessible chromatin regions, represented as a hierarchically clustered summary heatmap; rows represent regions and columns represent each of the 4 intestinal tissue compartments.

**(C)** Venn diagram analysis of preferentially enriched and shared transcription factor motifs identified in accessible chromatin regions from each of the 4 intestinal tissue compartments.

(**D**–**J**) Examples of transcription factor motifs (top) preferentially enriched (**D**–**G**) or shared (**H**– **J**) in accessible chromatin regions from each of the 4 intestinal tissue compartments. Motif activity for selected genes is represented as violin plots (middle) or superimposed onto UMAP embeddings (bottom).



# Figure S5. Deficiency of Eomes has modest effects on effector CD8<sup>+</sup> T cell differentiation in the spleen, Related to Figure 5.

(A) Experimental design.  $CD8^+$  P14 T cells from control ( $CD45.1^+$ ) and *Eomes*<sup>fl/fl</sup>*Cd4-Cre*<sup>+</sup>( $CD45.1.2^+$ , Eomes cKO) mice were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> recipients prior to infection with LCMV. P14 T cells were isolated from spleen at 7 days (**B–I**) or after 21 days (**J–M**) following infection.

(**B** and **E**–**I**) Representative flow cytometry plots (left) and bar graphs (right) displaying frequencies of control (grey squares) vs. Eomes cKO (orange circles) P14 T cells expressing Eomes (**B**), GzmA (**E**), GzmB (**F**), TNF and IFN $\gamma$  (**G**), Ki67 (**H**), or T-bet (**I**) at 7 days following infection; numbers represent the frequencies of cells within each gate. For analyses of TNF and IFN $\gamma$ , cells were restimulated *in vitro* for 3h with gp<sub>33-41</sub> peptide.

(**C** and **J**) Quantification of the proportions (left) or absolute numbers (right) of control vs. Eomes cKO P14 T cells in the spleen at 7 days (**C**) or after 21 days (**J**) following infection.

**(D)** Representative flow cytometry plots (left) displaying KLRG1 and CD127 expression by control vs. Eomes cKO P14 T cells in the spleen at 7 days following infection. Bar graphs representing proportions (top right) or absolute numbers (bottom right) of control vs. Eomes cKO P14 T cells in the spleen exhibiting a terminal effector ('TE', KLRG1<sup>hi</sup>CD127<sup>lo</sup>) or memory precursor ('MP', KLRG1<sup>lo</sup>CD127<sup>hi</sup>) phenotype.

(K) Representative flow cytometry plots (left) and bar graphs indicating the frequencies (middle) and absolute numbers (right) of control vs. Eomes cKO  $T_{CM}$  (CD62L<sup>hi</sup>CD127<sup>hi</sup>),  $T_{EM}$  (CD62L<sup>lo</sup>CD127<sup>hi</sup>), and t-T<sub>EM</sub> (CD62L<sup>lo</sup>CD127<sup>lo</sup>) cells after 21 days following infection.

(L and M) Representative flow cytometry plots (top) and bar graphs (bottom) displaying CD122 (L) and T-bet (M) expression by control vs. Eomes cKO  $T_{CM}$  (CD62L<sup>hi</sup>CD127<sup>hi</sup>),  $T_{EM}$  (CD62L<sup>lo</sup>CD127<sup>hi</sup>), and t-T<sub>EM</sub> (CD62L<sup>lo</sup>CD127<sup>lo</sup>) cells after 21 days following infection.

Data are represented as mean  $\pm$  SEM. Paired *t*-test. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 \*\*\*\*p<0.0001. Data are representative of at least 3 independent experiments with n=5-6 mice per experiment.



#### Figure S6. Deficiency of Eomes has modest effects on intestinal CD8<sup>+</sup> T cell differentiation, Related to Figure 5.

(A and C-H) CD8<sup>+</sup> T cells from control (CD45.1<sup>+</sup>) and *Eomes*<sup>fl/fl</sup>Cd4-Cre<sup>+</sup>(CD45.1.2<sup>+</sup>, Eomes cKO) P14 mice were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> recipients prior to infection with LCMV; gated i.v.-negative siIEL, siLPL, cIEL, and cLPL CD8<sup>+</sup> T cells were analyzed at 7 days post-infection. Representative flow cytometry plots (top) and bar graphs displaying frequencies of control (grey squares) vs. Eomes cKO (orange circles) P14 T cells expressing Eomes (A), GzmA (C), GzmB (D), TNF and IFNy (E), IL-2 (F), Ki67 (G), or Tbet (H); numbers represent the frequencies of cells within each gate. For analyses of TNF, IFNy, and IL-2, cells were restimulated in vitro for 3h with gp<sub>33-41</sub> peptide. (B) CD8<sup>+</sup> T cells from wildtype CD45.1<sup>+</sup> P14 or wild-type CD45.1.2<sup>+</sup> P14 mice were activated and transduced with empty vector (EV) or Eomes overexpression (Eomes OE) constructs, respectively. Cells were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> hosts that were subsequently infected with LCMV; gated i.v.-negative siIEL, siLPL, cIEL, and cLPL CD8<sup>+</sup> T cells were analyzed at 7 days post-infection. Representative flow cytometry plots (top) displaying CD69 and CD103 expression by EV vs. Eomes OE P14 T cells. Bar graph (bottom) representing frequencies of CD69<sup>+</sup>CD103<sup>+</sup> EV vs. Eomes OE P14 T cells. (I and J) Representative flow cytometry plots (top) and bar graphs (bottom) displaying GzmA (I) and TNF and IFN<sub>Y</sub> (J) expression by control vs. Eomes cKO P14 T cells after 21 days following infection. For analyses of TNF and IFNy, cells were restimulated *in vitro* for 3h with gp<sub>33-41</sub> peptide. (K) Experimental design. Control (CD45.1<sup>+</sup>) and *Eomes*<sup>fl/fl</sup>*Ert2-Cre*<sup>+</sup> (CD45.1.2<sup>+</sup>, Eomes iKO) P14 donor mice were treated with tamoxifen i.p. for 2 days prior to sacrifice. CD8<sup>+</sup> T cells from control and Eomes iKO P14 donor mice were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> recipients 30 minutes

prior to infection with LCMV. Intraperitoneal tamoxifen injections were continued on the day of infection and on days 1 and 2 post-infection; siIEL, siLPL, cIEL, and cLPL compartments were harvested and analyzed after day 21 post-infection.

(L) Quantification of the proportions of control (gray squares) vs. Eomes iKO (red circles) CD8<sup>+</sup>P14 cells in each intestinal tissue compartment.

Data are represented as mean  $\pm$  SEM. Paired *t*-test. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 \*\*\*\*p<0.0001. Data are representative of at least 3 independent experiments with n=5-6 mice per experiment.



# Figure S7. Transcriptional analyses identify putative regulators of small intestine CD8<sup>+</sup> $T_{RM}$ cell maintenance, Related to Figures 6 and 7.

(A and B) Representative flow cytometry plots (top) displaying GzmA (A) and GzmB (B) expression by gated i.v.-negative control vs. Eomes iKO P14 T cells in each intestinal tissue compartment at day 31 post-infection; numbers represent the frequencies of cells within each gate. Bar graphs (bottom) indicate the frequencies of control vs. Eomes iKO P14 T cells expressing each molecule. (C and D) Representative flow cytometry plots (top) displaying CD127 (C) and P2RX7 (**D**) expression by gated i.v.-negative control vs. Eomes iKO P14 T cells in each intestinal tissue compartment. Bar graphs indicate the frequencies of control vs. Eomes iKO P14 T cells expressing each molecule (bottom left) and the expression of each molecule on a per-cell basis (gMFI) (bottom right) at day 31 post-infection. (E) Experimental design. CD8<sup>+</sup> T cells from wild-type CD45.1<sup>+</sup> or wild-type CD45.1.2<sup>+</sup> P14 T cells were activated and transduced with empty vector (EV) or Eomes overexpression (Eomes OE) constructs, respectively. Cells were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> hosts that were subsequently infected with LCMV. Small intestine IEL, siLPL, cIEL, and cLPL CD8<sup>+</sup> T cells were isolated at 7 days post-infection. (F) Representative flow cytometry plots (top) displaying P2RX7 expression by gated i.v.-negative EV vs. Eomes OE P14 T cells. Bar graph represents frequencies of cells expressing P2RX7. (G) Experimental design. CD8<sup>+</sup> T cells from control (CD45.1<sup>+</sup>) or Eomes<sup>fl/fl</sup>Ert2-Cre<sup>+</sup>(CD45.1.2<sup>+</sup>, Eomes iKO) P14 mice were adoptively co-transferred into congenic CD45.2<sup>+</sup> recipients prior to infection with LCMV. Mice received 5 doses of tamoxifen along with control mAb or IL-7-aIL-7R mAb complexes i.p. once daily starting at day 21 postinfection; siIEL and siLPL CD8<sup>+</sup> T cells were harvested 10 days later after day 31 post-infection.

(H) Bar graphs representing frequencies of control mAb-treated wild-type vs. control Ab-treated Eomes iKO P14 T cells (left, light purple bars); and frequencies of IL-7- $\alpha$ IL-7R mAb complex-treated wild-type vs. IL-7- $\alpha$ IL-7R mAb complex-treated Eomes iKO P14 T cells (right, dark purple bars) in the siIEL and siLPL tissue compartments. (I) Experimental design. CD8<sup>+</sup> T cells from control (CD45.1<sup>+</sup>) or Eomes iKO (CD45.1.2<sup>+</sup>) P14 mice were activated and transduced with non-targeting shRNA (NT) or shRNA targeting *P2rx7* (P2RX7 KD). Cells were mixed at a 1:1 ratio and adoptively co-transferred into congenic CD45.2<sup>+</sup> hosts that were subsequently infected with LCMV. Mice received 5 doses of tamoxifen i.p. once daily starting on day 12 post-infection; siIEL and siLPL CD8<sup>+</sup> T cells were harvested on day 17 post-infection. (J) Bar graphs representing frequencies of NT-transduced control vs. NT-transduced Eomes iKO P14 T cells (left, light blue bars); and frequencies of P2RX7 KD-transduced control vs. P2RX7 KD-transduced Eomes iKO P14 T cells (right, dark blue bars) in the siIEL and siLPL tissue compartments. Data are represented as mean  $\pm$  SEM. Paired *t*-test. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001 \*\*\*\*p<0.0001. Data are representative of at least 3 independent experiments with n=5-6 mice per experiment.