

Supplementary Materials for

**The binding mode of orphan glycyl-tRNA synthetase with tRNA supports the synthetase classification and reveals large domain movements**

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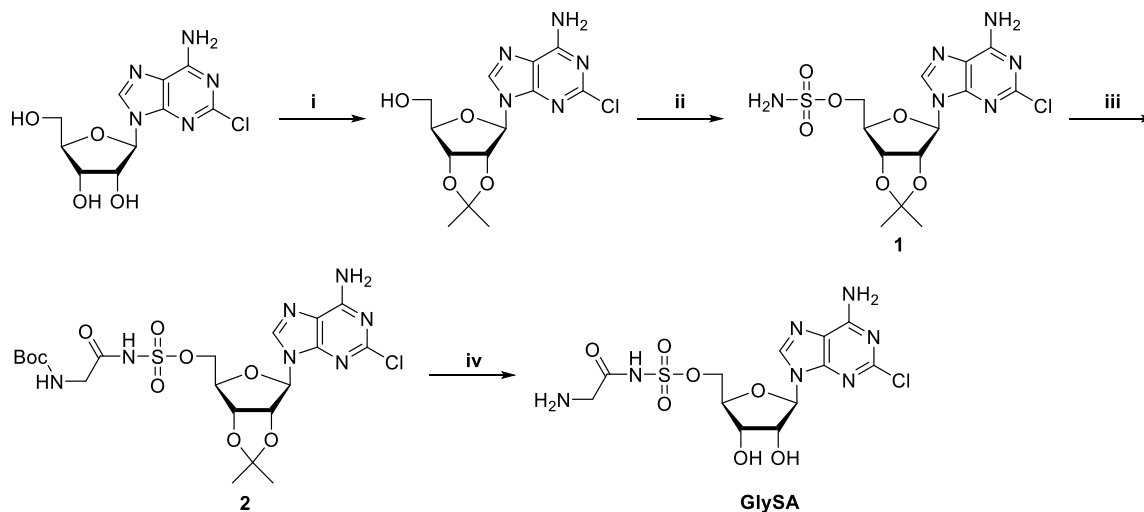
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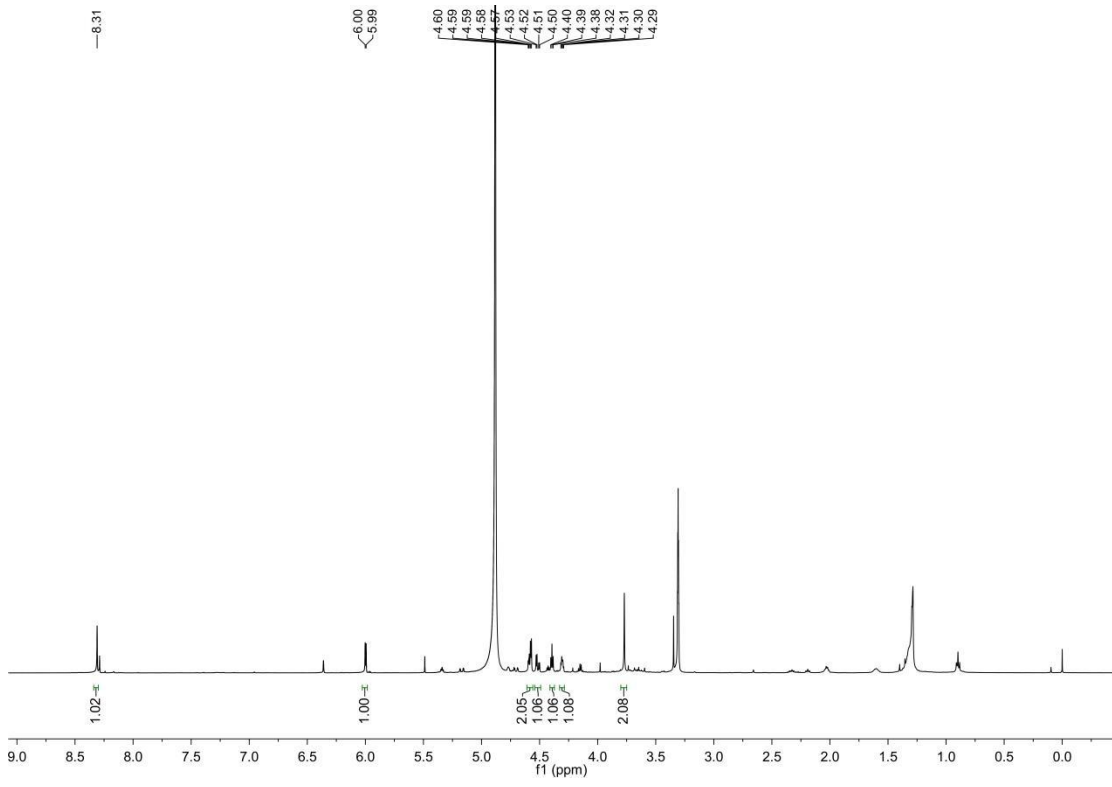
## Supplementary Method

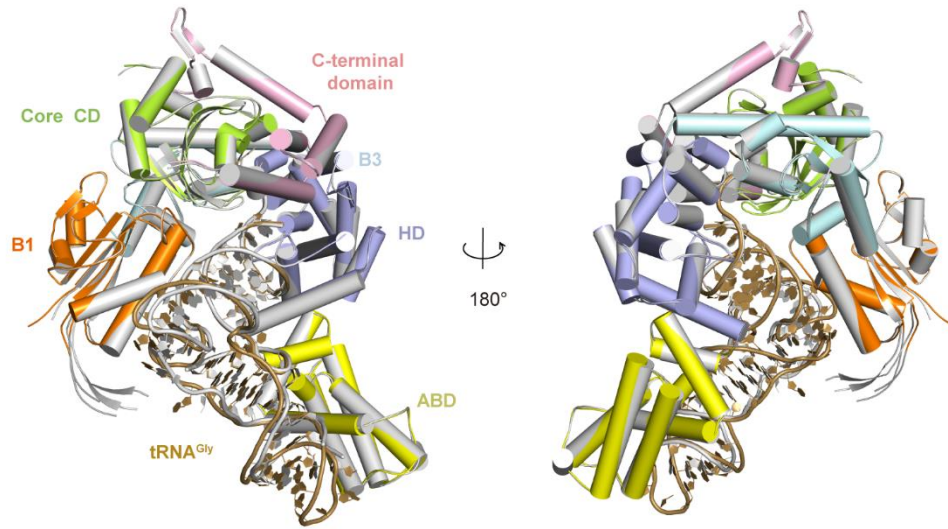
### Synthesis of GlySA



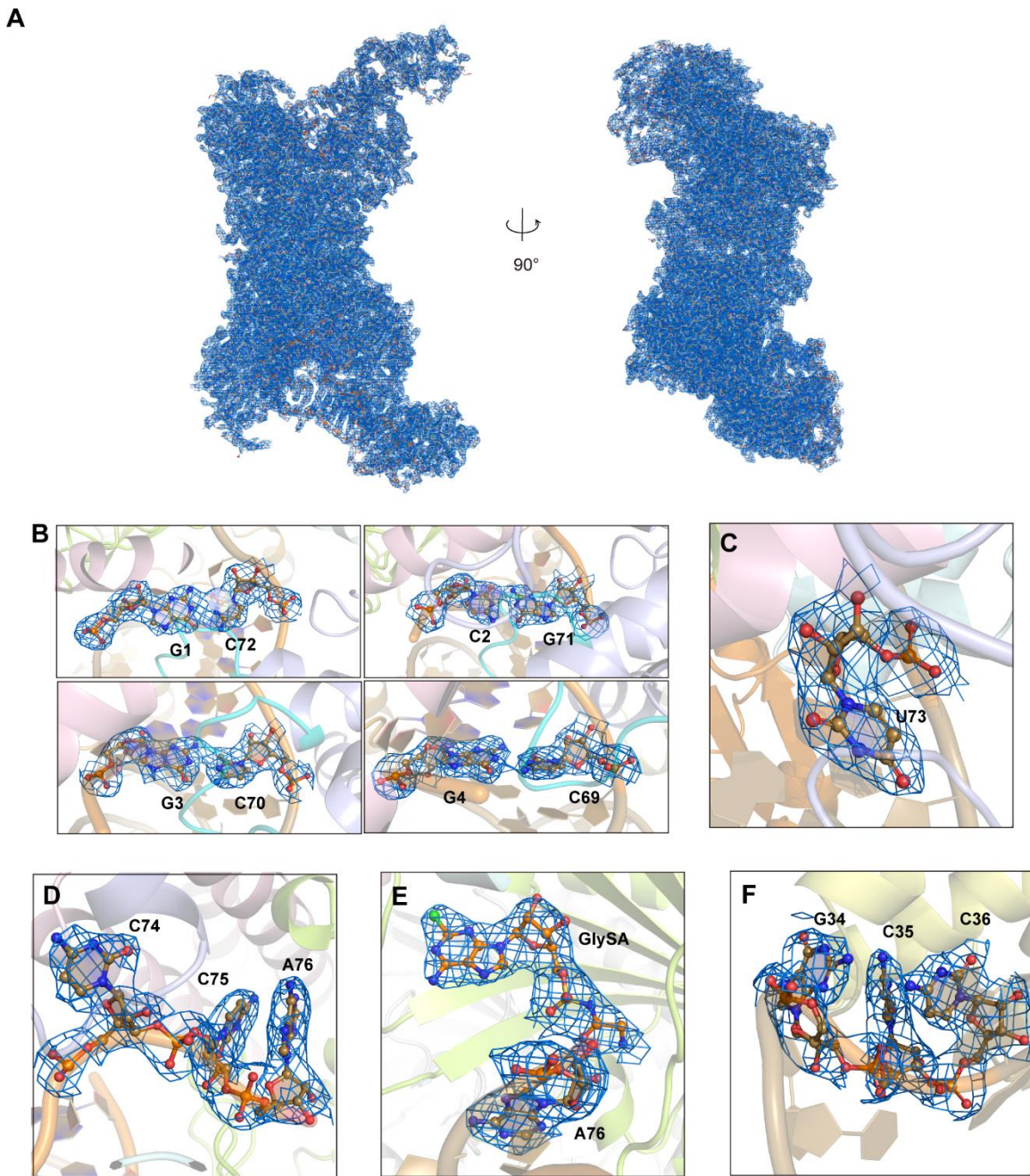
**Scheme S1. The synthesis of intermediate analog.** (i) *p*-toluenesulfonic acid, DMOP, DMF, N<sub>2</sub>, rt, 20 h; (ii) NH<sub>2</sub>SO<sub>2</sub>Cl, DMA, N<sub>2</sub>, rt, 4h; (iii) Boc-Gly-OSu, DBU, DMF, rt, 8h; (iv) TFA: H<sub>2</sub>O=5:1, rt, 2h.

*2-chloro-5'-O-[N-(glycyl)sulfamoyl] adenosine (GlySA)*. GlySA (**26**) was prepared as the procedure shown in Scheme S1. The compound [(3aR,4R,6R,6aR)-6-(6-Amino-2-chloro-9Hpurin-9-yl)-2,2-dimethyl-tetrahydro-2H-furo[3,4-d][1,3]-3-dioxol-4-yl]methyl sulfamate (**1**) was prepared via the similar method described previously (**74**) with high yield. The compound **1** (210 mg, 1.0 equiv) was dissolved in 10 mL DMF, and then DBU (84  $\mu$ L, 1.1 equiv) and Boc-Gly-OSu (150 mg, 1.1 equiv) were added to the reaction mixture. After stirring for 8 h at room temperature, the reaction mixture was diluted with 25 mL brine and extracted with dichloromethane. The organic layers were dried, concentrated and purified by flash chromatography to obtain the compound ((3aR,4R,6R,6aR)-6-(6-amino-2-chloro-9H-purin-9-yl)-2,2-dimethyltetrahydrofuro[3,4-d][1,3]dioxol-4-yl)methyl ((tert-butoxycarbonyl)glycyl)sulfamate (**2**) as a colorless glassy solid. 144 mg compound **2** was dissolved in TFA/H<sub>2</sub>O (5:2, 3 mL) and stirred at room temperature for 0.5 h. The reaction mixture was concentrated in vacuo, and then purified by preparative HPLC to afford the final product GlySA as colorless solid. <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD):  $\delta$  8.31 (s, 1H), 6.00 (d, *J* = 4.3 Hz, 1H), 4.61 – 4.56 (m, 2H), 4.51 (dd, *J* = 11.2, 4.0 Hz, 1H), 4.39 (t, *J* = 5.1 Hz, 1H), 4.30 (dd, *J* = 8.2, 3.9 Hz, 1H), 3.77 (s, 2H). MS (ESI) *m/z*: calcd for C<sub>12</sub>H<sub>17</sub>N<sub>7</sub>O<sub>7</sub>SCl [M + H]<sup>+</sup>, 438.05, found, 438.05.

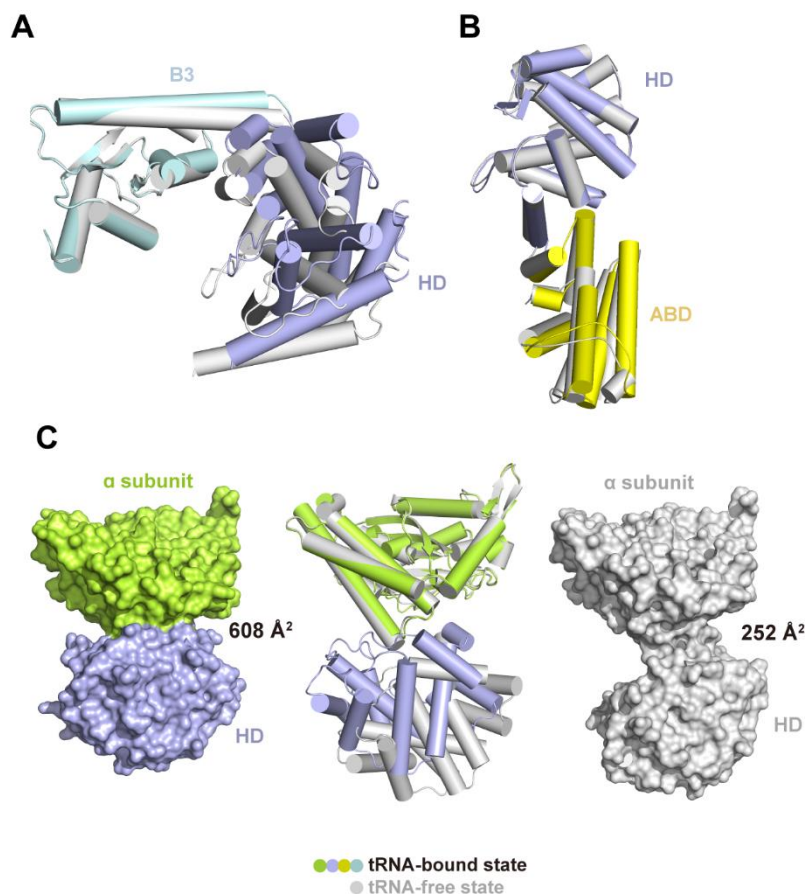




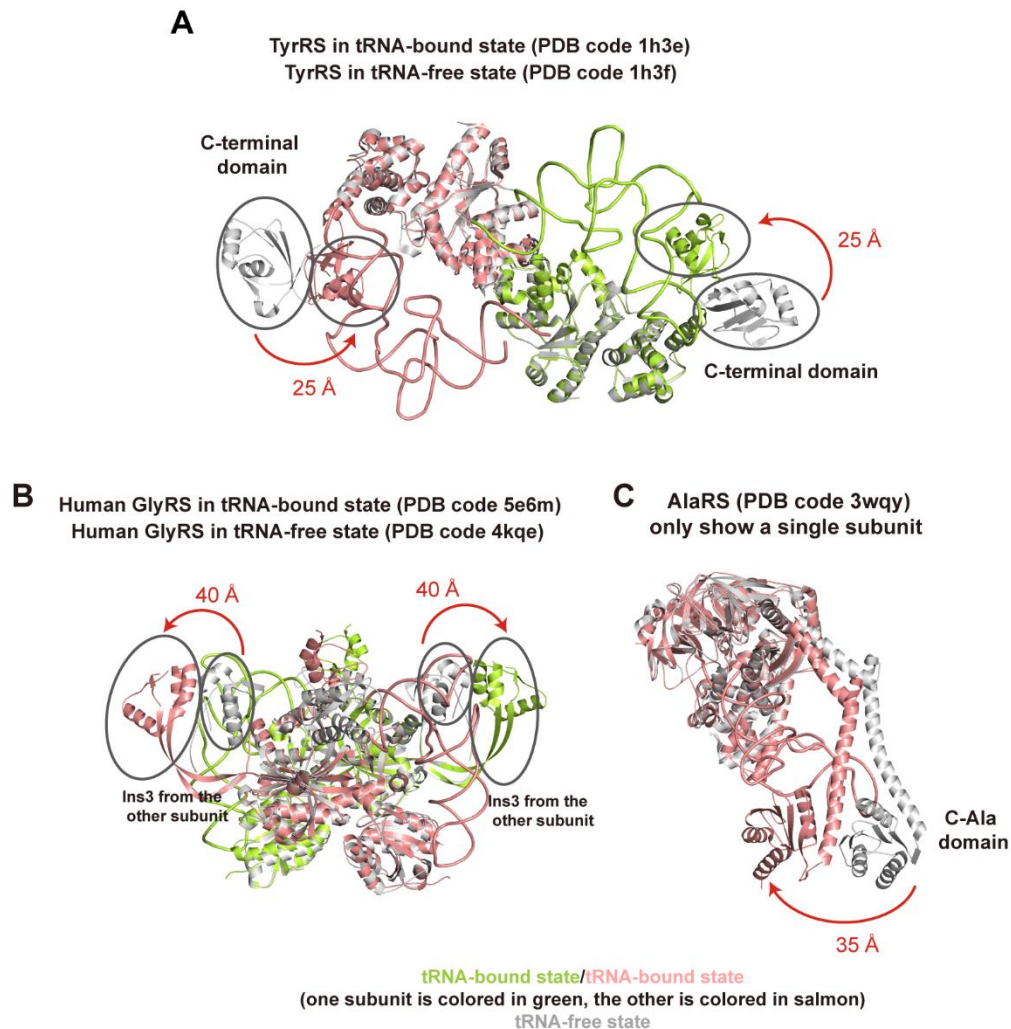
**Fig. S1. Structure superimposition of the two protomers of orphan *EcGlyRS* revealed that both their own structures and their tRNA binding modes are almost identical.** One protomer and its substrate tRNA<sup>Gly</sup> are colored the same as Fig. 1B, while the other protomer and its corresponding tRNA<sup>Gly</sup> are colored in gray.



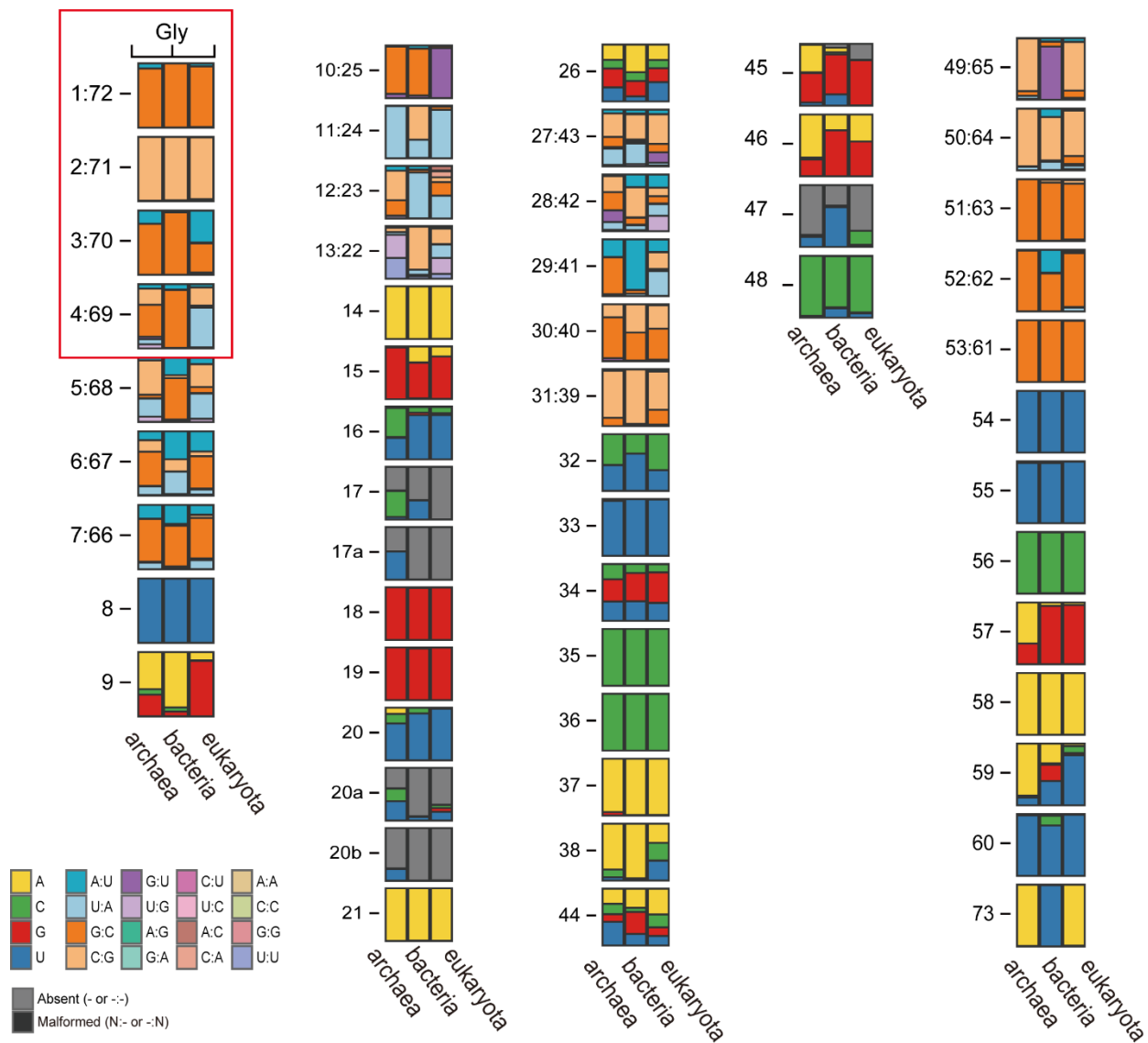
**Fig. S2. Electron density map of orphan *EcGlyRS*·GlySA·tRNA<sup>Gly</sup> complex structure. (A)** 2Fo-Fc omit electron density maps of the protein and nucleic acid chain are drawn as blue meshes contoured at 1.0  $\sigma$ . **(B-F)** 2Fo-Fc omit electron density maps of the first four base pairs of tRNA<sup>Gly</sup> acceptor stem **(B)**, U73 **(C)**, the 3' CCA-end **(D)**, GlySA **(E)** and anticodon triplets **(F)** are shown as blue meshes and contoured at 1.0  $\sigma$ .



**Fig. S3. The HD domain and ABD rotate as a whole upon tRNA<sup>Gly</sup> binding, which also enlarges the interface between the HD domain and the  $\alpha$  subunit.** (A) The HD domain in the  $\beta$  subunit of orphan *EcGlyRS* bound with tRNA rotates related to that of *EcGlyRS* without tRNA when their B3 domains are aligned. (B) The ABD of *EcGlyRS* bound with tRNA aligns well with that of *EcGlyRS* without tRNA when their HD domains are superimposed. Thus, rotation of the  $\beta$  subunit C-terminal part mainly happens at the linker between the B3 and HD domains, and HD domain and ABD rotate as a whole. (C) When  $\alpha$  subunit in tRNA-bound state is superimposed to that in tRNA-free state, the HD domain moves towards  $\alpha$  subunit in tRNA-bound state, resulting in a larger interface between HD domain and  $\alpha$  subunit. In (A-C), orphan *EcGlyRS* in tRNA-bound state is colored the same as Fig. 1B, while orphan *EcGlyRS* in tRNA-free state is colored in gray. The interface areas were calculated using program PISA.

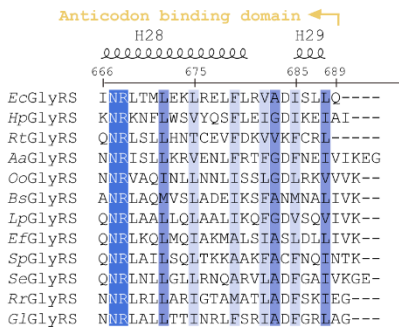
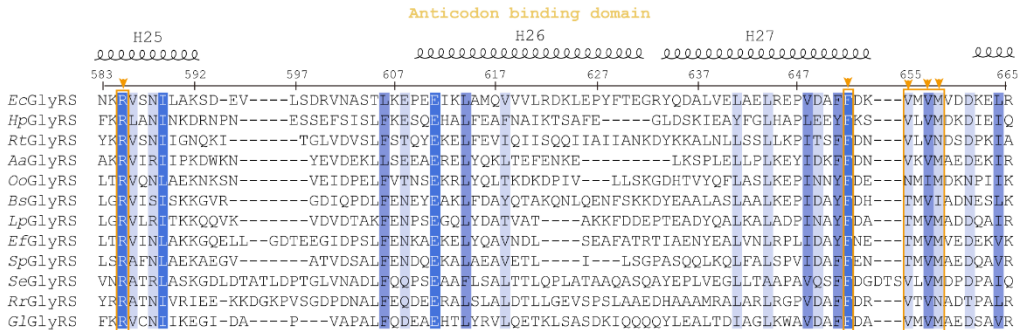
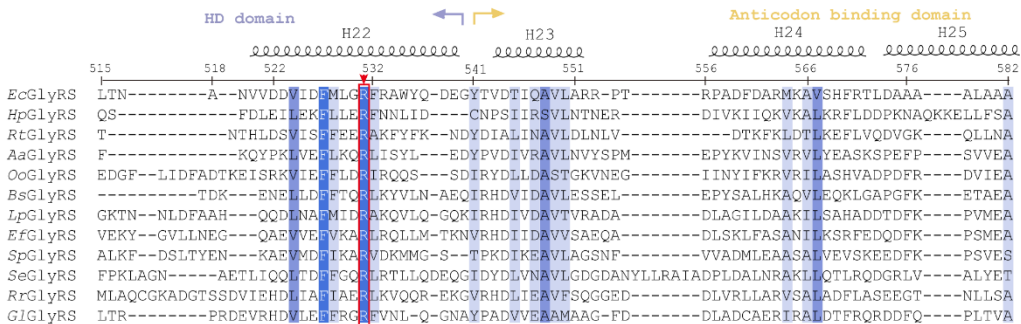
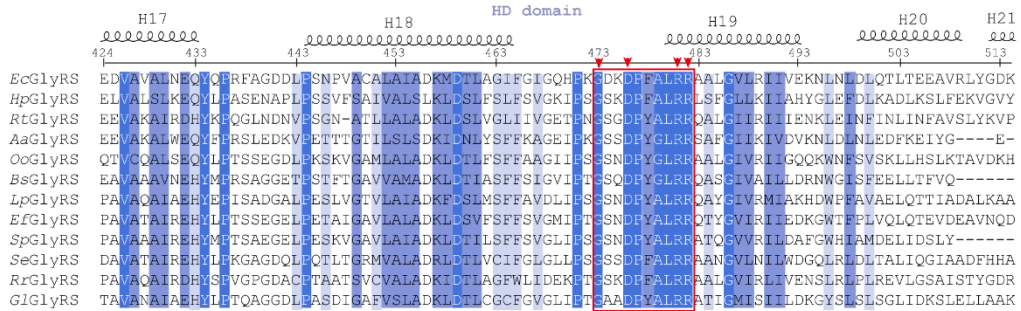
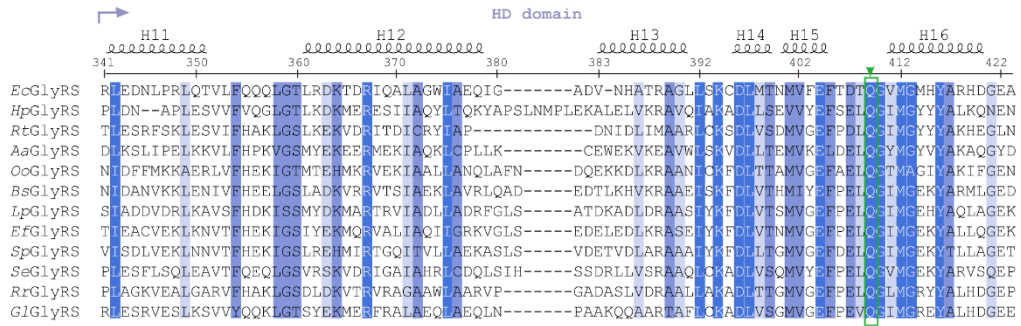


**Fig. S4. Large conformational changes of aaRSs upon tRNA binding.** (A) The largest structural movement of class I aaRSs induced by tRNA binding is the C-terminal domain of TyrRS. It moves about 25 Å to contact and recognize the long variable loop of tRNA<sup>Tyr</sup>. (B) In human GlyRS, the Ins3 domain from the other subunit opens up to contact the elbow region of tRNA<sup>Gly</sup>. (C) The C-Ala domain of AlaRS moves about 35 Å to contact the elbow region of tRNA<sup>Ala</sup>. In (A-C), aaRSs in tRNA-bound state are colored in green for one subunit and salmon for the other, while that in tRNA-free state are colored in gray.



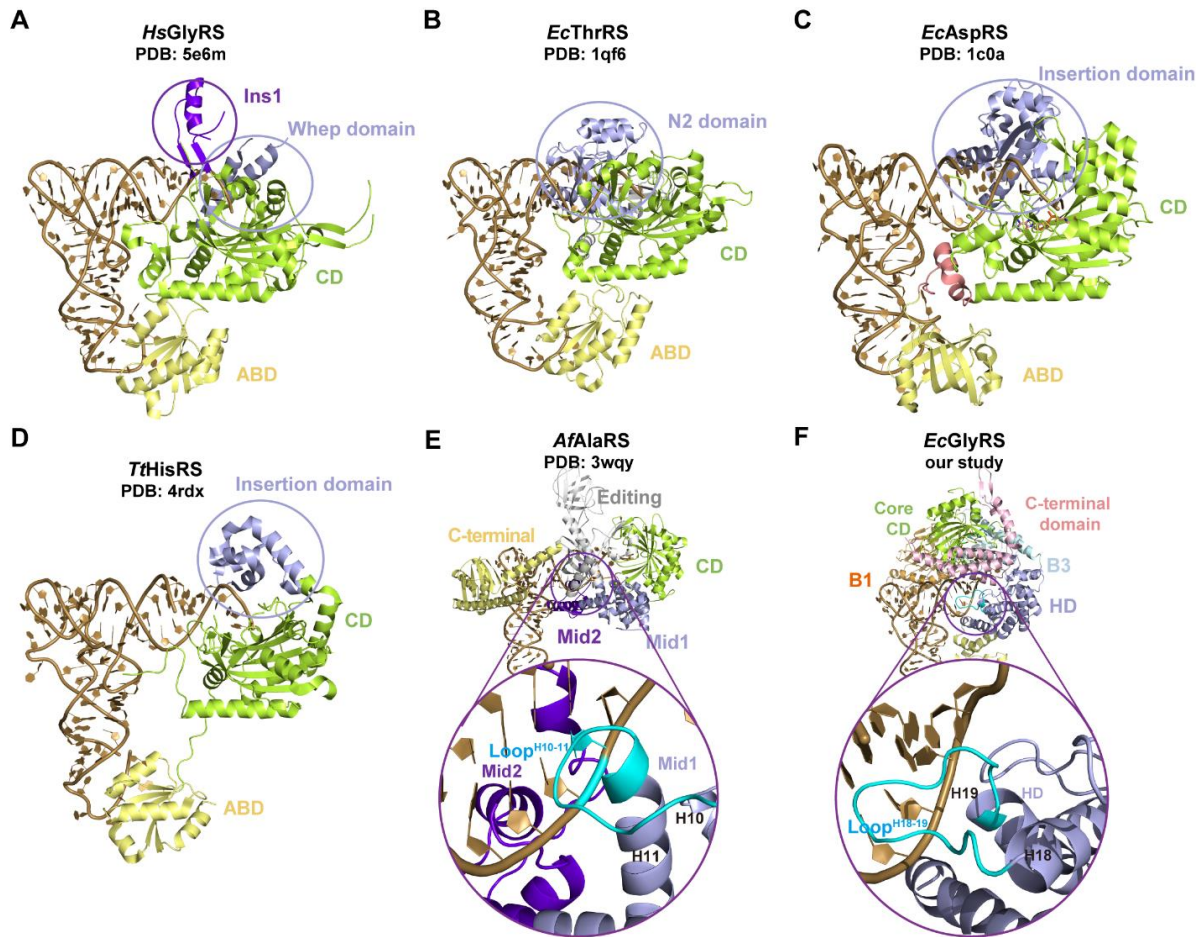
**Fig. S5. Sequence analysis of tRNA<sup>Gly</sup> among archaea, bacteria and eukaryotes using tRNA<sup>viz</sup>.** Sequence feature distribution for all positions of tRNA<sup>Gly</sup> molecules among three domains of life were analyzed using program tRNA<sup>viz</sup> (37). The first four base pairs of the acceptor stem are squared in red.



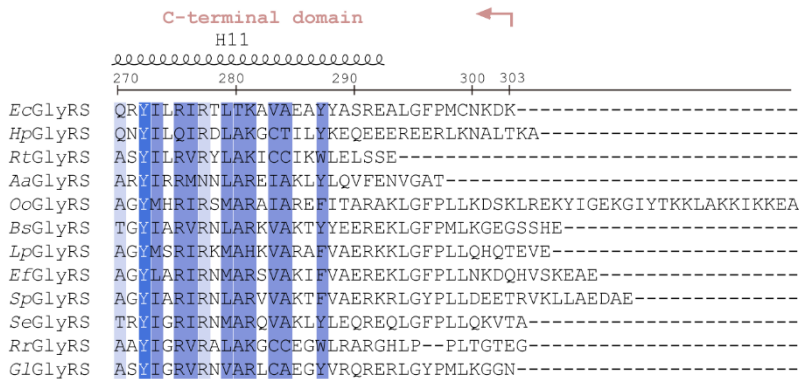
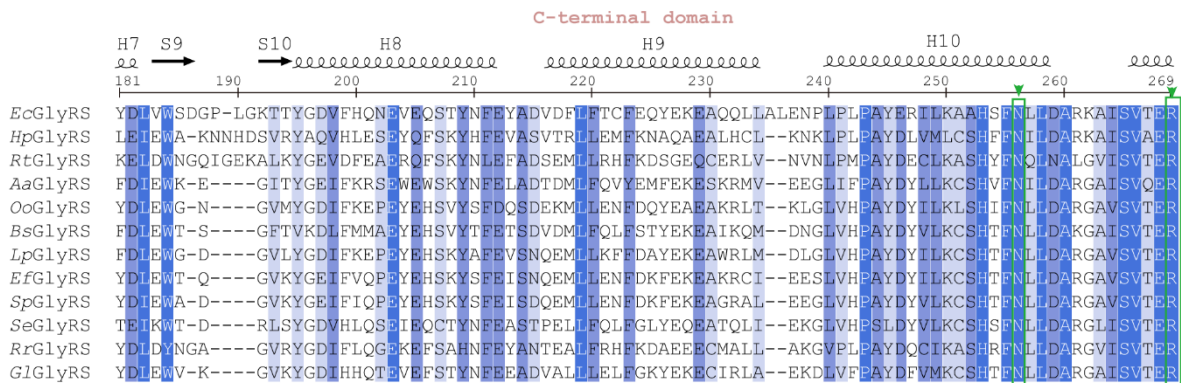
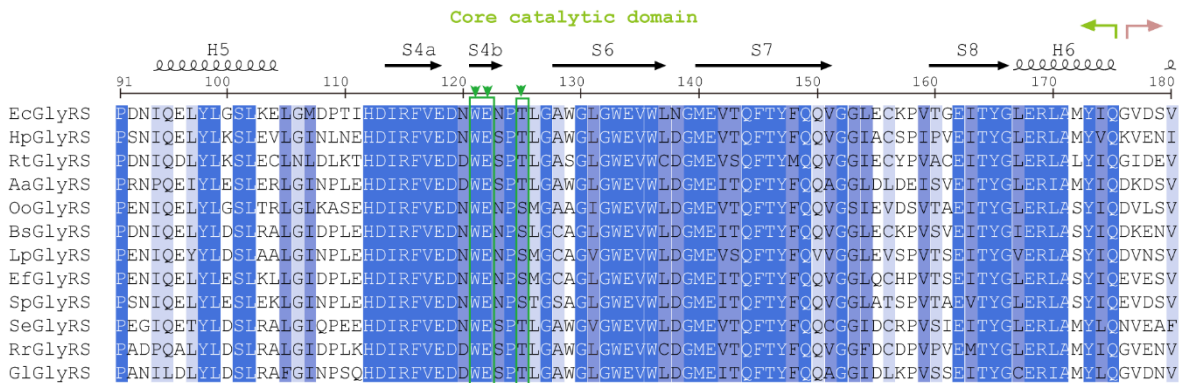
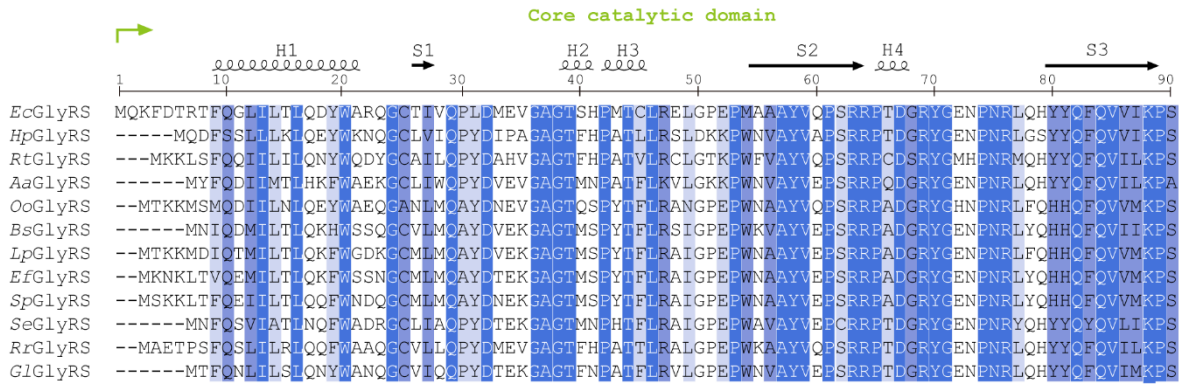


Conservation score: 0~5 6~7 8~9 10

**Fig. S6. Sequence alignments of the HD domain and the ABD of  $\beta$  subunits from orphan GlyRSs.** Protein sequences of *Escherichia coli* GlyRS (*EcGlyRS*, UniProtKB ID: P00961), *Helicobacter pylori* GlyRS (*HpGlyRS*, UniProtKB ID: B5Z7X4), *Rickettsia typhi* GlyRS (*RtGlyRS*, UniProtKB ID: Q68VR4), *Aquifex aeolicus* GlyRS (*AaGlyRS*, UniProtKB ID: O67898), *Oenococcus oeni* GlyRS (*OoGlyRS*, UniProtKB ID: Q04F69), *Bacillus subtilis* GlyRS (*BsGlyRS*, UniProtKB ID: P54381), *Lactocaseibacillus paracasei* GlyRS (*LpGlyRS*, UniProtKB ID: Q038U3), *Enterococcus faecalis* GlyRS (*EfGlyRS*, UniProtKB ID: Q831U3), *Streptococcus pneumoniae* GlyRS (*SpGlyRS*, UniProtKB ID: B8ZL20), *Synechococcus elongatus* (*SeGlyRS*, UniProtKB ID: Q31SB9), *Rhodospirillum rubrum* (*RrGlyRS*, UniProtKB ID: Q2RQ43), *Geobacter lovleyi* GlyRS (*GlGlyRS*, UniProtKB ID: B3E621) were aligned using Clustal Omega program (72). The secondary structures corresponding to *EcGlyRS* are shown above the sequences. The conservation scores were calculated by the program Jalview (73) and presented in various shades of blue. Key residues which recognize the acceptor stem, 3' CCA-end and anticodon of tRNA<sup>Gly</sup> are shown in red, green and orange boxes, respectively.

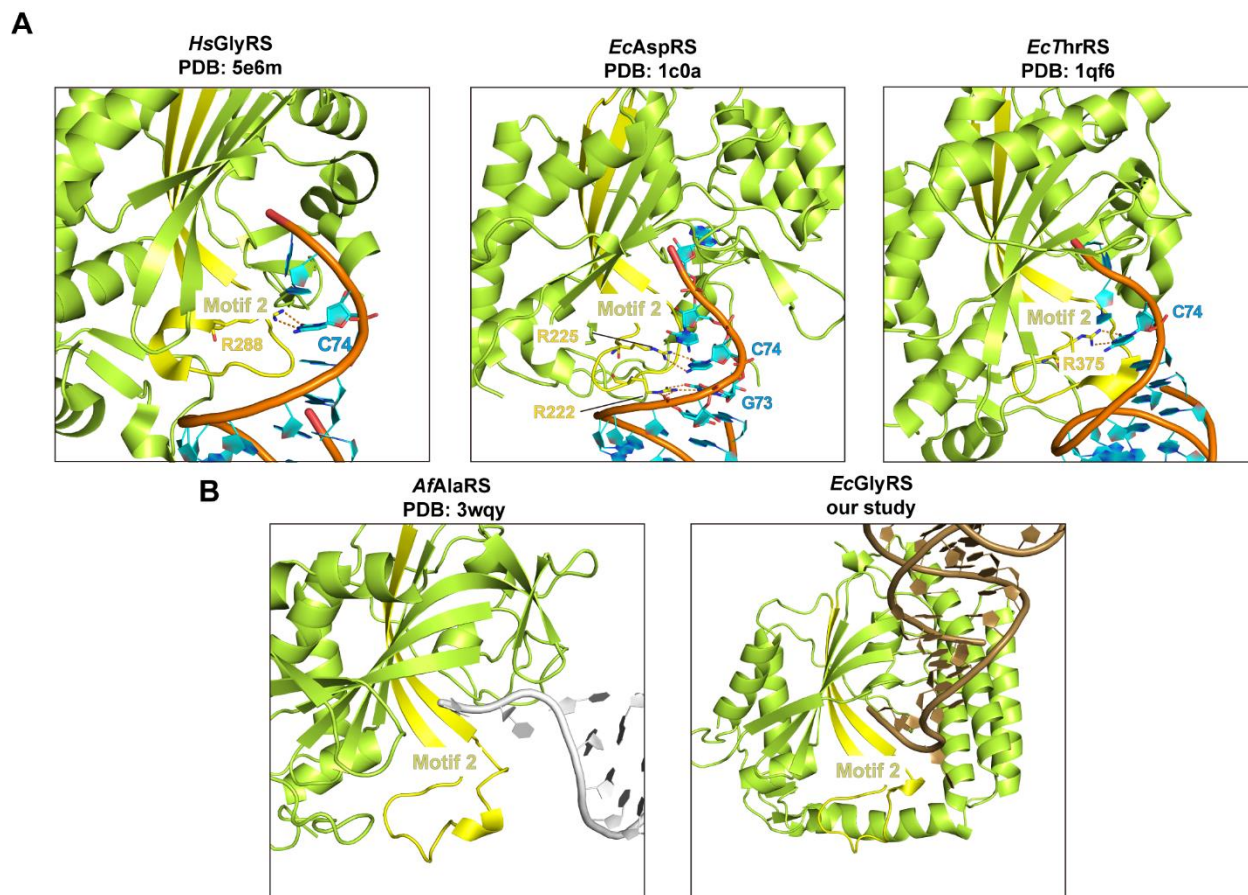


**Fig. S7. Protein sequences beyond aminoacylation domain facilitate to recognize the acceptor stems and 3' termini of tRNAs in many class II synthetases. (A)** In *HsGlyRS* (PDB code 5e6m), the Ins1 and WHEP domains contact the minor groove of the acceptor stem and 3' terminus of tRNA<sup>Gly</sup>, respectively. **(B)** In *ThrRS* (PDB code 1qf6), the N2 domain contacts the minor groove of the acceptor stem of tRNA<sup>Thr</sup>. **(C)** In *AspRS* (PDB code 1c0a), the insertion domain contacts the 3' terminus of tRNA<sup>Asp</sup>. **(D)** In *HisRS* (PDB code 4rdx), the insertion domain contacts the 3' terminus of tRNA<sup>His</sup>. **(E)** In *AlaRS* (PDB code 3wqy), the Mid2 subdomain (purple) and the loop (cyan) before helix 11(H11) of Mid1 subdomain (light blue) clamp the minor and major grooves of the acceptor stem of tRNA<sup>Ala</sup>. **(F)** In orphan *EcGlyRS*, the loop (cyan) similar to that of *AlaRS* from HD domain (light blue) clamps the major groove of the acceptor stem of tRNA<sup>Gly</sup>. In **(A-F)**, the catalytic domains and ABDs are colored in green and yellow, respectively, and accessory sequences assisting the recognition of tRNA are colored in purple or light blue and circled.



Conservation score: 0~5 6~7 8~9 10

**Fig. S8. Sequence alignments of the  $\alpha$  subunits from *EcGlyRS* and homologs.** Protein sequences of *EcGlyRS* (UniProtKB ID: P00960), *HpGlyRS* (UniProtKB ID: B5Z7W3), *RtGlyRS* (UniProtKB ID: Q68VR3), *AaGlyRS* (UniProtKB ID: O67081), *OoGlyRS* (UniProtKB ID: Q04F71), *BsGlyRS* (UniProtKB ID: P54380), *LpGlyRS* (UniProtKB ID: Q038U2), *EfGlyRS* (UniProtKB ID: Q831U2), *SpGlyRS* (UniProtKB ID: B8ZL21), *SeGlyRS* (UniProtKB ID: Q31KD2), *RrGlyRS* (UniProtKB ID: Q2RQ44), *GlGlyRS* (UniProtKB ID: B3E622) were aligned using Clustal Omega program (72). The secondary structures corresponding to *EcGlyRS* are shown above the sequences. The conservation scores were calculated by the program Jalview (73) and presented in various shades of blue. Residues participating in the recognition of the 3' CCA-end of tRNA<sup>Gly</sup> are shown green boxes, respectively.



**Fig. S9. The roles of class II signature motif 2 in the recognition of CCA-end of tRNAs for class II aaRSs. (A)** The motif 2 contributes to stabilizing the 3' CCA-end of tRNA in typical class II aaRSs. The pyrimidine ring of C74 interacts with the conserved Arg in the motif 2 of *HsGlyRS* (PDB code 5e6m), *EcAspRS* (PDB code 1c0a) and *EcThrRS* (PDB code 1qf6). **(B)** In *AfAlaRS* (PDB code 3wqy) and orphan *EcGlyRS*, their substrate tRNAs bind through an angle different to tRNAs of other class II aaRSs. As a result, the motif 2 of *AfAlaRS* and *EcGlyRS* does not contact with 3' CCA-end of their tRNAs. In **(A-B)**, the motif 2 sequences are colored in yellow, and polar interactions between the conserved Arg on motif 2 and the 3' CCA-end of tRNA are labeled in orange dashed lines.

**Table S1. Statistics of X-ray diffraction data collection and structure refinement.**

		<i>EcGlyRS·GlySA·tRNA<sup>Gly</sup></i>
<b>PDB accession code</b>		7YSE
<b>Data collection</b>		
Resolution(Å)		50.00-2.91(3.06-2.91) <sup>a</sup>
Wavelength (Å)		0.979
Space group		P222 <sub>1</sub>
Cell dimensions		
a, b, c (Å)		72.71, 162.123, 324.09
α, β, γ (°)		90.0, 90.0, 90.0
Unique reflections		85473 (12313)
Redundancy		6.6 (7.1)
R <sub>merge</sub> <sup>b</sup>		0.138 (0.749)
CC <sub>1/2</sub>		0.990 (0.774)
Average I/σ(I)		9.4 (2.6)
Completeness (%)		100 (100)
<b>Refinement</b>		
Resolution (Å)		50.00-2.91
Reflections for refinement/test		80999/4393
R <sub>work</sub> <sup>c</sup> /R <sub>free</sub> <sup>d</sup>		0.230/0.260
RMSD bond (Å)		0.004
RMSD angle (°)		1.231
No. atoms		
Protein		14340
RNA		2952
Water		14
Ion		6
Ligand		56
Average B factors (Å <sup>2</sup> )		86.0
Ramachandran plot (%)		
Favored		93.74
Allowed		5.83
Outliers		0.43

<sup>a</sup>Values in parentheses are for the highest resolution shell.

<sup>b</sup> $R_{\text{merge}} = \frac{\sum_h \sum_l |I(h)_l - \langle I(h) \rangle|}{\sum_h \sum_l I(h)_l}$ , where  $I(h)_l$  is the  $l$ th observation of the reflection  $h$  and  $\langle I(h) \rangle$  is the weighted average intensity for all observations  $l$  of reflection  $h$ .

<sup>c</sup> $R_{\text{work}} = \frac{\sum_h ||F_{\text{obs}}(h)| - |F_{\text{cal}}(h)||}{\sum_h |F_{\text{obs}}(h)|}$ , where  $F_{\text{obs}}(h)$  and  $F_{\text{cal}}(h)$  are the observed and calculated structure factors for reflection  $h$  respectively.

<sup>d</sup> $R_{\text{free}}$  was calculated as  $R_{\text{work}}$  using 5.0% of the reflections which were selected randomly and omitted from refinement.

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